

СПИСЪК

на цитиранията на научните трудове на чл.-кор. Христо Найденски в научни публикации, патенти и дисертации (без автоцитати)

1). Georgiev, L., M. Chochkova, I. Totseva, K. Seizova, E. Marinova, G. Ivanova, M. Ninova, H. NAJDENSKI and T. Milkova. Anti-tyrosinase, antioxidant and antimicrobial activities of hydroxycinnamoylamides. Medicinal Chemistry Research. 2013, 22(9), 4173-4182. ISSN:10542523.

1. Arciszewska, Żaneta, et al. "Caffeic Acid/Eu (III) complexes: Solution equilibrium studies, structure characterization and biological activity." *International journal of molecular sciences* 23.2 (2022): 888.
2. Błaszczuk, Natalia, Angelina Rosiak, and Joanna Kałużna-Czaplińska. "The Potential Role of Cinnamon in Human Health." *Forests* 12.5 (2021): 648.
3. Botta, G., Bizzarri, B. M., Garozzo, A., Timpanaro, R., Bisignano, B., Amatore, D., ... & Saladino, R. (2015). Carbon nanotubes supported tyrosinase in the synthesis of lipophilic hydroxytyrosol and dihydrocaffeoyl catechols with antiviral activity against DNA and RNA viruses. *Bioorganic & medicinal chemistry*, 23(17), 5345-5351.
4. Chaaban, Ibrahim, et al. "Synthesis and evaluation of new phenolic derivatives as antimicrobial and antioxidant agents." *Monatshefte für Chemie-Chemical Monthly* 149.1 (2018): 127-139.
5. Crespo, María Inés, et al. "Inhibitory effects of compounds isolated from *Lepechinia meyenii* on tyrosinase." *Food and chemical toxicology* 125 (2019): 383-391.
6. Dagmara, Wróbel-Biedrawa, et al. "Anti-melanoma potential of two benzoquinone homologues embelin and rapanone-a comparative in vitro study." *Toxicology in Vitro* (2020): 104826.
7. Divar, M.; Tadayyon, S.; Khoshneviszadeh, M.; Pirhadi, S.; Attarroshan, M.; Mobaraki, K.; Damghani, T.; Mirfazli S.; Edraki N. Benzyl-Triazole Derivatives of Hydrazinecarbothiamide Derivatives as Potent Tyrosinase Inhibitors: Synthesis, Biological Evaluation, Structure-Activity Relationship and Docking Study. *Chemistry Select*. Vol. 8, Issue 823. 2023, Feb. ISSN: 23656549. DOI: 10.1002/slct.202203382.
8. Dobritsch, Melanie, et al. "MATE transporter-dependent export of hydroxycinnamic acid amides." *The Plant Cell* 28.2 (2016): 583-596.
9. Feng, Jia-Hao, et al. "Synthesis and biological evaluation of clovamide analogues with catechol functionality as potent Parkinson's disease agents in vitro and in vivo." *Bioorganic & medicinal chemistry letters* 29.2 (2019): 302-312.
10. Furlan, V.; Bren, U. *Helichrysum italicum*: From Extraction, Distillation, and Encapsulation Techniques to Beneficial Health Effects. *Foods* 2023, 12, 802. <https://doi.org/10.3390/foods12040802>.
11. Ghafari, Shahrzad, et al. "Novel morpholine containing cinnamoyl amides as potent tyrosinase inhibitors." *International journal of biological macromolecules* 135 (2019): 978-985.
12. Godlewska-Żyłkiewicz, Beata, et al. "Biologically Active Compounds of Plants: Structure-Related Antioxidant, Microbiological and Cytotoxic Activity of Selected Carboxylic Acids." *Materials* 13.19 (2020): 4454.

13. Gunia-Krzyżak, Agnieszka, et al. "Cinnamic acid derivatives in cosmetics: current use and future prospects." *International journal of cosmetic science* 40.4 (2018): 356-366.
14. Hosseinpour, H., Iraj, A., Edraki, N., et al. , A Series of Benzylidenes Linked to Hydrazine-1-carbothioamide as Tyrosinase Inhibitors: Synthesis, Biological Evaluation and Structure–Activity Relationship., 2020, *Chemistry and Biodiversity*, 17(8), e2000285
15. Kwak, Seon-Yeong, et al. "Chemical modulation of bioactive compounds via oligopeptide or amino acid conjugation." *Peptide Science* 100.6 (2013): 584-591.
16. Lee, Sangwon, et al. "Inhibitory effects of N-(acryloyl) benzamide derivatives on tyrosinase and melanogenesis." *Bioorganic & medicinal chemistry* 27.17 (2019): 3929-3937.
17. Lesma, G., et al. "Cannabinoid-free Cannabis sativa L. grown in the Po valley: evaluation of fatty acid profile, antioxidant capacity and metabolic content." *Natural product research* 28.21 (2014): 1801-1807.
18. Liaqat, Atika, et al. "Anti-Bio Gram of the Most Common Dietary Additives (Spices) Against Common Problematic Organisms." *Annals of Punjab Medical College (APMC)* 11.4 (2017): 276-282.
19. Mansouri, Hadjer, and Sidi Mohamed Mekelleche. "Radical scavenging activity of hydroxycinnamic acids in polar and nonpolar solvents: A computational investigation." *Journal of Theoretical and Computational Chemistry* 19.08 (2020): 2050032.
20. Mendes, Eduarda, Maria de Jesus Perry, and Ana Paula Francisco. "Design and discovery of mushroom tyrosinase inhibitors and their therapeutic applications." *Expert opinion on drug discovery* 9.5 (2014): 533-554.
21. Mercogliano M., Iesce M.R., Alfieri M.L., Buommino E., DellaGreca M. Hands-on synthesis of furanamides and evaluation of their antimicrobial activity. (2023) *Natural Product Research*, 37 (20), pp. 3484 – 3491. DOI: 10.1080/14786419.2022.2087220
22. Monteiro, Luís S., et al. "An efficient one-pot synthesis of polyphenolic amino acids and evaluation of their radical-scavenging activity." *Bioorganic chemistry* 89 (2019): 102983.
23. Monteiro, Luís S., et al. "Synthesis and preliminary biological evaluation of new phenolic and catecholic dehydroamino acid derivatives." *Tetrahedron* 73.43 (2017): 6199-6209.
24. Monteiro, L. S., & Paiva-Martins, F. (2022). Amino acids, amino acid derivatives and peptides as antioxidants. In *Lipid Oxidation in Food and Biological Systems: A Physical Chemistry Perspective* (pp. 381-404). Cham: Springer International Publishing.
25. Noel, Amandine, et al. "Poly (ferulic acid-co-tyrosine): Effect of the Regiochemistry on the Photophysical and Physical Properties en Route to Biomedical Applications." *Macromolecules* 47.20 (2014): 7109-7117.
26. Otero, Elver, et al. "Triclosan-caffeic acid hybrids: Synthesis, leishmanicidal, trypanocidal and cytotoxic activities." *European journal of medicinal chemistry* 141 (2017): 73-83.
27. Popoola, Olugbenga Kayode. "The chemical and biological characterization of South African helichrysum species." (2015), PhD thesis, University of the Western Cape, SA.
28. Popoola, O.K.; Marnewick, J.L.; Iwuoha, E.I.; Hussein, A.A. Methoxylated Flavonols and ent-Kaurane Diterpenes from the South African *Helichrysum rutilans* and Their Cosmetic Potential. *Plants* 2023, 12, 2870. <https://doi.org/10.3390/plants12152870>.
29. Rao, Pasupuleti Visweswara, and Siew Hua Gan. "Cinnamon: a multifaceted medicinal plant." *Evidence-Based Complementary and Alternative Medicine* 2014 (2014).
30. Sall, C., et al. "Towards smart biocide-free anti-biofilm strategies: Click-based synthesis

- of cinnamide analogues as anti-biofilm compounds against marine bacteria." *Bioorganic & medicinal chemistry letters* 28.2 (2018): 155-159.
31. Si, Huijie, et al. "Inhibitory effects of 4-chlorocinnamaldehyde on the activity of mushroom tyrosinase." *Medicinal Chemistry Research* 26.7 (2017): 1377-1381.
 32. Sun, J., Song, Y. L., Zhang, J., Huang, Z., Huo, H. X., Zheng, J., ... & Tu, P. F. (2015). Characterization and Quantitative Analysis of Phenylpropanoid Amides in Eggplant (*Solanum melongena* L.) by High Performance Liquid Chromatography Coupled with Diode Array Detection and Hybrid Ion Trap Time-of-Flight Mass Spectrometry. *Journal of agricultural and food chemistry*, 63(13), 3426-3436.
 33. Taofiq, Oludemi, et al. "Hydroxycinnamic acids and their derivatives: Cosmeceutical significance, challenges and future perspectives, a review." *Molecules* 22.2 (2017): 281.
 34. Wang, Ji-Rui, et al. "Variations in the Components and Antioxidant and Tyrosinase Inhibitory Activities of *Styphnolobium japonicum* (L.) Schott Extract during Flower Maturity Stages." *Chemistry & biodiversity* 16.3 (2019): e1800504.
 35. Wang Y., Xiong B., Xing S., Chen Y., Liao Q., Mo J., Chen Y., Li Q., Sun H. Medicinal Prospects of Targeting Tyrosinase: A Feature Review. *Curr Med Chem.* 2023;30(23):2638-2671. doi: 10.2174/0929867329666220915123714. PMID: 36111760.
 36. Wen, Xin, et al. "Physicochemical characteristics and phytochemical profiles of yellow and red *Physalis* (*Physalis alkekengi* L. and *P. pubescens* L.) fruits cultivated in China." *Food Research International* 120 (2019): 389-398.
 37. Wróbel-Biedrawa, Dagmara, et al. "Anti-melanoma potential of two benzoquinone homologues embelin and rapanone-a comparative in vitro study." *Toxicology in Vitro* 65 (2020): 104826.
 38. Xie, L.-W., Li, S.-X., Xie, Y.-X., et al. Bioassay-guided fractionation of constituents targeting mediators of inflammation from *Lycii* Cortex as inhibitors of NF- κ B. *Chinese Mat. Med.*, 2014, 39 (4), 689-694.
 39. Xie, Lian-Wu, et al. "Activity-guided isolation of NF- κ B inhibitors and PPAR γ agonists from the root bark of *Lycium chinense* Miller." *Journal of ethnopharmacology* 152.3 (2014): 470-477.
 40. Yuan, Ye, et al. "Tyrosinase inhibitors as potential antibacterial agents." *European Journal of Medicinal Chemistry* 187 (2020): 111892.
 41. Zucca, Paolo, et al. "Antimicrobial, antioxidant and anti-tyrosinase properties of extracts of the Mediterranean parasitic plant *Cytinus hypocistis*." *BMC research notes* 8.1 (2015): 1-9.
 42. Zhu, J., Tang, Y., Lu, L., Qiu, X., & Pan, L. (2024). Reactive extrusion of caffeic acid functionalized ϵ -polylysine with low-density polyethylene as an antimicrobial and antioxidant film. *Reactive and Functional Polymers*, 105861.
 43. Zhu, P., Li, R., & Lu, A. (2024). Electrode impedance modeling based on XGboost algorithm for analyzing the antioxidant properties of juice. *Journal of Food Measurement and Characterization*, 1-12.
- 2). Nikolova, S., Tzvetkov, Y., NAJDENSKI, H., Vesselinova, A. Isolation of pathogenic yersiniae from wild animals in Bulgaria (2001) J. Vet. Med., Series B, 48 (3), pp. 203-209.**
44. Abd-El Salam, M., and N. Benkerroum. "North African brined cheeses." *Brine Cheese*

- (2006): 139-187.
45. Annamalai, Thirunavukkarasu. Investigating the cold tolerance and osmotolerance mechanisms in *Yersinia enterocolitica*. University of Connecticut, 2006.
 46. Arrausi-Subiza, M., et al. "Prevalence of *Yersinia enterocolitica* and *Yersinia pseudotuberculosis* in wild boars in the Basque Country, northern Spain" *Acta Veterinaria Scandinavica* 58, 1, 20 (2016): Article number 4
 47. Arrausi-Subiza, M., Ibabe, J. C., Atxaerandio, R., Juste, R. A., & Barral, M. Evaluation of different enrichment methods for pathogenic *Yersinia* species detection by real time PCR. *BMC veterinary research*, 10(1), 2014, 192.
 48. Balada-Llasat, J.-M., Panilaitis, B., Kaplan, D., Mecsas, J. Oral inoculation with Type III secretion mutants of *Yersinia pseudotuberculosis* provides protection from oral, intraperitoneal, or intranasal challenge with virulent *Yersinia* (2007) *Vaccine*, 25 (8), pp. 1526-1533.
 49. Bancierz-Kisiel, A., Platt-Samoraj, A., Szczerba-Turek, A., Syczyło, K., Szweda, W. The first pathogenic *Yersinia enterocolitica* bioserotype 4/O: 3 strain isolated from a hunted wild boar (*Sus scrofa*) in Poland *Epidemiology and Infection* 2015, 143, 13, 2758-2765
 50. Bancierz-Kisiel, A., Platt-Samoraj, A., Szczerba-Turek, A., Syczyło, K., & Szweda, W. (2015). The first pathogenic *Yersinia enterocolitica* bioserotype 4/O: 3 strain isolated from a hunted wild boar (*Sus scrofa*) in Poland. *Epidemiology and infection*, 1-8.
 51. Blackburn, Jason Kenna. "Evaluating the spatial ecology of anthrax in North America: Examining epidemiological components across multiple geographic scales using a GIS-based approach." (2006)
 52. Boynukara, Banur, and Timur Gülhan. "Su Samurlarında (*Lutra lutra*) Görülen Hastalıklar." *Etlik Veteriner Mikrobiyoloji Dergisi* 27.1 (2016): 53-62.
 53. Carella, E., Romano, A., Domenis, L., et al. "Characterisation of *Yersinia enterocolitica* strains isolated from wildlife in the northwestern Italian Alps". *Journal of Veterinary Research (Poland)*, 2022, 66(2), pp. 141-149.
 54. Caspari, K. Untersuchungen zum Vorkommen von humanpathogenen *Yersinia enterocolitica* bei Schlachtschweinen aus verschiedenen Haltungsformen. Dissertation, Aus dem Institut für Lebensmittelqualität und -sicherheit des Zentrums für Lebensmittelwissenschaften der Tierärztlichen Hochschule Hannover, 2005.
 55. Chassang, Lucile, et al. "Antemortem diagnosis and surgical management of splenitis due to *Yersinia pseudotuberculosis* infection in a pet rabbit (*Oryctolagus cuniculus*)." *Journal of exotic pet medicine* 29 (2019): 182-187.
 56. Christophersen, Olav Albert, and Anna Haug. "Why is the world so poorly prepared for a pandemic of hypervirulent avian influenza?." *Microbial ecology in health and disease* 18.3-4 (2006): 113-132.
 57. Conover, M. R., & Vail, R. M. (2014). *Human diseases from wildlife*. CRC Press.
 58. Crockett, Kathryn. A historical analysis of *Bacillus anthracis* as a biological weapon and its application to the development of nonproliferation and defense strategies. George Mason University, 2006.
 59. Defabachew, Eyassu Seifu. Application of the lactoperoxidase system to improve the quality and safety of goat milk and goat cheese. Diss. University of Pretoria, 2006.
 60. DeLong, D., Bacterial diseases. *The Laboratory Rabbit, Guinea Pig, Hamster, and Other Rodents*. 2012, 301-363.

61. Dembek, Zygmunt F. "The history and threat of biological weapons and bioterrorism." Hospital Preparation for Bioterror. Academic Press, 2006. 17-35.
62. Donati, D., et al. "Tropical and travel-associated diseases." J Immunol 177 (2006): 3294-3302.
63. Fisher, M.L., Castillo, C., Mecsas, J. Intranasal inoculation of mice with *Yersinia pseudotuberculosis* causes a lethal lung infection that is dependent on *Yersinia* outer proteins and PhoP (2007) Infection and Immunity, 75 (1), pp. 429-442.
64. Fratini, Filippo, et al. "Experimental infection with *Yersinia pseudotuberculosis* in European brown hare (*Lepus europaeus*, Pallas)." Asian Pacific Journal of Tropical Medicine 10.3 (2017): 285-291.
65. Fredriksson-Ahomaa, M. Epidemiology of human *Yersinia pseudotuberculosis* infection [Epidemiologic von *Yersinia pseudotuberculosis* Infektionen beim Menschen] (2009) Archiv fur Lebensmittelhygiene, 60 (2), pp. 82-87.
66. Fredriksson-Ahomaa, M., Wacheck, S., Koenig, M., Stolle, A., Stephan, R. Prevalence of pathogenic *Yersinia enterocolitica* and *Yersinia pseudotuberculosis* in wild boars in Switzerland (2009) International Journal of Food Microbiology, 135 (3), pp. 199-202.
67. Frimodt-Møller, N., Hammerum, A.M. Food safety revisited (2006) Journal of Infectious Diseases, 194 (9), pp. 1191-1193.
68. Fujita, Osamu, et al. "Development of a real-time PCR assay for detection and quantification of *Francisella tularensis*." Japanese journal of infectious diseases 59.1 (2006): 46.
69. Garcês, A., & Pires, I. (2021). Secrets of the astute Red Fox (*Vulpes vulpes*, Linnaeus, 1758): an inside-ecosystem secret agent serving one health. Environments, 8(10), 103.
70. Georgiev, D. Habitats of the Otter (*Lutra lutra* L.) in some Regions of Southern Bulgaria. IUCN Otter Spec. Group Bull, 2005, 22, 1, 6-13.
71. Goodwin, Steven Mark. Chemistry and genetics of plant cuticle function as a permeability barrier. Diss. Purdue University, 2006.
72. Gratz, Norman. Vector-and rodent-borne diseases in Europe and North America: distribution, public health burden, and control. Cambridge University Press, 2006.
73. Hamel D, Silaghi C, Lescai D, Pfister K. Epidemiological aspects on vector-borne infections in stray and pet dogs from Romania and Hungary with focus on *Babesia* spp. Parasitol. Res., 110 (4), 2012, 1537-1545.
74. Hudson, Krischan Jerry. The mechanisms and functions of the *Yersinia pseudotuberculosis* outer membrane adhesins invasin and YadA in phagocytic uptake and systemic infections. University of Virginia, 2006.
75. Imnadze, T., Malania, L., Chakvetadze, N., Burjanadze, I., Abazashvili, N., Zhgenti, E., ... & Kosoy, M. (2021). Evidence of extensive circulation of *Yersinia enterocolitica* in rodents and shrews in natural habitats from retrospective and perspective studies in south Caucasus. Pathogens, 10(8), 939.
76. Jalava, K., S. Hallanvuo, U.-M. Nakari, et al. Multiple outbreaks of *Yersinia pseudotuberculosis* infections in Finland. J. Clin. Microbiol. 2004, 42, 6, 2789-2791.
77. Janda, J. Michael, and Sharon L. Abbott. "The genus *Hafnia*: from soup to nuts." Clinical microbiology reviews 19.1 (2006): 12-28.
78. Kisková J, Hrehová Z, Janiga M, Lukáč M, Haas M. Bacterial prevalence in the Dunnock (*Prunella modularis*) in sub-alpine habitats of the Western Carpathians, Slovak Republic,

- Ornis Fennica, 89 (1), 2012, 2-80.
79. Kiskova, J., Hrehova, Z., Janiga, M., LUKÁČ, M., Haas, M., & JURĚŮVĚOVÁ, M. (2011). *Yersinia* species in the dunnoek (*Prunella modularis*) in sub-alpine habitats of the Western Carpathians. *Pol J Microbiol*, 60(1), 79-83.
 80. Laukkanen, R. Enteropathogenic *Yersinia* in pork production. PhD Thesis, University of Helsinki, 2010.
 81. Laukkanen, R., Martínez, P.O., Siekkinen, K.-M., Ranta, J., Maijala, R., Korkeala, H. Transmission of *Yersinia pseudotuberculosis* in the pork production chain from farm to slaughterhouse (2008) *Applied and Environmental Microbiology*, 74 (17), pp. 5444-5450.
 82. Leibiger, R., Niedung, K., Geginat, G., Heesemann, J., Trülsch, K. *Yersinia enterocolitica* Yop mutants as oral live carrier vaccines (2008) *Vaccine*, 26 (51), pp. 6664-6670.
 83. Mak, Windsor, Raymond TF Cheung, and Shu Leong Ho. "Biological basis of the racial disparities in health and diseases: an evolutionary perspective." *Racial and Ethnic Disparities in Health and Health Care* (2006).
 84. Martin, Yvonne. Pseudodistomin E: Versuche zur Totalsynthese über das Konzept der Tandem Wittig-[3+ 2]-Cycloaddition. Diss. Universität Würzburg, 2006.
 85. Martínez, P.O. Prevalence of enteropathogenic *Yersinia* in pigs from different european countries and contamination in the pork production chain. Department of Food Hygiene and Environmental Health, Faculty of Veterinary Medicine, University of Helsinki, Finland, Academic Dissertation, 2010
 86. Mattix, Marc E., et al. "Clinicopathologic aspects of animal and zoonotic diseases of bioterrorism." *Clinics in laboratory medicine* 26.2 (2006): 445-489.
 87. Niskanen, T. Diagnostics and epidemiology of *Yersinia pseudotuberculosis*. Academic Dissertation. Department of Food and Environmental Hygiene, Faculty of Veterinary Medicine, University of Helsinki, Finland, 2009
 88. Novotný, M., Fečková, M., Janiga, M., Lukáš, M., Novotná, M., Kovalčíková, Z. High incidence of *Yersinia enterocolitica* (Enterobacteriaceae) in Alpine Accentors *Prunella collaris* of the Tatra Mountains (2007) *Acta Ornithologica*, 42 (2), pp. 137-143.
 89. Nowakiewicz, Aneta, et al. "Free-living species of carnivorous mammals in Poland: Red fox, beech marten, and raccoon as a potential reservoir of *Salmonella*, *Yersinia*, *Listeria* spp. and coagulase-positive *Staphylococcus*." *PLoS One* 11.5 (2016): e0155533.
 90. Odyniec, Marta, et al. "Bioserotypes, Virulence Markers, and Antimicrobial Susceptibility of *Yersinia enterocolitica* Strains Isolated from Free-Living Birds." *BioMed Research International* (2020):8936591.
 91. Odyniec, M., Bancerz-Kisiel, A. "Assessment of the Role of Free-Living and Farmed Fallow Deer (*Dama dama*) as A Potential Source of Human Infection with Multiple-Drug-Resistant Strains of *Yersinia enterocolitica* and *Yersinia pseudotuberculosis*". *Pathogens*, 2022, 11(11), 1266.
 92. Olano, Juan P., C. J. Peters, and David H. Walker. "Distinguishing tropical infectious diseases from bioterrorism." *Tropical Infectious Diseases* (2006): 1386.
 93. Oliveira M, Pedroso N, Sales-Luís T, et al. Evidence of Antimicrobial Resistance in Eurasian Otter (*Lutra lutra* Linnaeus, 1758) Fecal Bacteria in Portugal. In: *Wildlife: destruction, conservation, and biodiversity* (J.D. Harris and P.L. Brown, eds.), 2009, pp. 201-221.

94. Oliveira, M., Sales-Luís, T., Duarte, A., Nunes, S.F., Carneiro, C., Tenreiro, T., Tenreiro, R., Santos-Reis, M., Tavares, L., Vilela, C.L. First assessment of microbial diversity in faecal microflora of Eurasian otter (*Lutra lutra* Linnaeus, 1758) in Portugal (2008) *European Journal of Wildlife Research*, 54 (2), pp. 245-252.
95. Oliveira, M., Sales-Luís, T., Semedo-Lemsaddek, T., et al. Antimicrobial resistant *Aeromonas* isolated from Eurasian Otters (*Lutra lutra* Linnaeus, 1758) in Portugal. *Persp. Anim. Ecol. Reprod.*, 2010, 123-144.
96. Powell, Jillian Elizabeth. Bacteriocins and bacteriocin producers present in kefir and kefir grains. Diss. Stellenbosch: University of Stellenbosch, 2006.
97. Ray, Prasanta K. Disaster Preparedness Against Accidents Or Terrorist Attack. New Age International, 2006.
98. Reinhardt, Marie, et al. "Yersinia pseudotuberculosis prevalence and diversity in wild boars in Northeast Germany." *Applied and environmental microbiology* 84.18 (2018).
99. Sagarna, X.G. Los carnívoros silvestres como reservorios de enfermedades de interés en sanidad animal y salud pública. Universidad del Pais Vasco, Tesis Doctorales, Vitoriaa Gasteiz, 2010.
100. Syczyło, Kinga, et al. "The prevalence of *Yersinia enterocolitica* in game animals in Poland." *PloS one* 13.3 (2018).
101. Tschäpe, Helmut, et al. "Population Structure of." *J. Bacteriol* 188.14 (2006): 5319.
102. Vanantwerpen, G., Houf, K., Van Damme, I., et al. Estimation of the within-batch prevalence and quantification of human pathogenic *Yersinia enterocolitica* in pigs at slaughter. *Food Control*, 2013, 1, 34, 9-12.
103. Vanantwerpen, G., Van Damme, I., De Zutter, L., et al. Seroprevalence of enteropathogenic *Yersinia* spp. in pig batches at slaughter. *Prev. Vet. Med.*, 2014, 116, 1-2, 193-196.
104. von Altrock, A. Seinige, D. Kehrenberg, C *Yersinia enterocolitica* isolates from wild boars hunted in Lower Saxony, Germany *Applied and Environmental Microbiology* 2015, 81, 14, 4835-4840
105. Wacheck, S. Mikrobiologische und sensorische Untersuchung tiefgefrorenen Wildbrets im Hinblick auf die Festlegung mikrobiologischer Richtwerte. Dissertation, Institut für Hygiene und Technologie der Lebensmittel tierischen Ursprungs der tierärztlichen Fakultät der Ludwig-Maximilians-Universität München, 2008
106. Wacheck, S., Fredriksson-Ahomaa, M., König, M., Stolle, A., Stephan, R. Wild boars as an important reservoir for foodborne pathogens (2010) *Foodborne Pathogens and Disease*, 7 (3), pp. 307-312.
107. Wagner, Michael M., Louise S. Gresham, and Virginia Dato. "Case detection, outbreak detection, and outbreak characterization." *Handbook of biosurveillance* (2006): 27.
108. Wang, Hua. Investigation of the interactions between sanitizers, surface characteristics, washing conditions, and bacteria for improving microbial safety of fresh produce. University of Illinois at Urbana-Champaign, 2006.
109. Williams, B. H., Huntington, K. B., & Miller, M. (2018). Mustelids. In *Pathology of Wildlife and Zoo Animals* (pp. 287-304). Academic Press.
110. Yunis, Edmond J., et al. "Stem cells in aging: Influence of ontogenic, genetic and environmental factors." *Journal of stem cells* 1.2 (2006): 125.

111. Zimmer, B., et al. "Automated methods for antibiotic susceptibility testing." *Clinical Microbiology and Infection* 12 (2006): 4.
- 3). Taskova, R., Mitova, M., NAJDENSKI, H., Tzvetkova, I., Duddeck, H. Antimicrobial activity and cytotoxicity of *Carthamus lanatus* (2002) *Fitoterapia*, 73 (6), pp. 540-543.**
112. Abdolmaleki, M., S. Bahraminejad and S. Abbasi. Antifungal activity of some plant crude extracts on four phytopathogenic fungi. *J. Med. Plants*. 2011, 10(38), 148-155.
113. Abdou, H.S., Sh, S., & Booles, H.F. Effect of pomegranate pretreatment on genotoxicity and hepatotoxicity induced by carbon tetrachloride (CCl₄) in male rats. *J. Med. Plants Res.*, 2012, 6(17), 3370-3380.
114. Akya, Alisha, et al. "Effect of *Vigna radiata*, *Tamarix ramosissima* and *Carthamus lanatus* extracts on *Leishmania major* and *Leishmania tropica*: An in vitro study." *Chinese Herbal Medicines* 12.2 (2020): 171-177.
115. Al-Saedi, S.R., 2023. Antibacterial Activity of the Aqueous and Methanol Extracts of *Nigella Sativa* Seeds in Mice. *Journal of Kerbala for Agricultural Sciences*, 10(2), pp.51-57.
116. Benlafya, K., Karrouchi, K., Charkaoui, Y., El Karbane, M., & Ramli, Y.. Antimicrobial activity of aqueous, ethanolic, methanolic, cyclohexanic extracts and essential oil of *Nigella sativa* seeds. *Journal of Chemical & Pharmaceutical Research*, 2014, 6(8), 9-11.
117. Bukhari IA. The central analgesic and anti-inflammatory activities of the methanolic extract of *Carthamus oxycantha*. *J. Physiol. Pharmacol.* 3, 64, 2013, 369-375.
118. Campos, Marina Pereira de. "Análise do potencial antimicrobiano de extrato, frações e substâncias puras obtidas de *Piper solmsianum* CDC VAR. *solmsianum* (PIPERACEAE)." (2006).
119. Celik, T.A. Potential genotoxic and cytotoxic effects of plant extracts. *A Compendium of Essays on Alternative Therapy*, (A. Bhattacharya, ed.), 2012, 233-250.
120. De Campos, M.P. *Análise do potencial antimicrobiano de extrato, frações e substâncias puras obtidas de Piper solmsianum CDC VAR. solmsianum (Peperaceae)*, Dissertação, Universidade do Vale do Itaiai, Centro de Ciências da Saúde, 2006
121. Dilshad, Muhammad, et al. "New lipooxygenase and cholinesterase inhibitory sphingolipids from *Carthamus oxyacantha*." *Natural Product Research* 30.16 (2016): 1787-1795.
122. Esmaili, S., et al. "Effects of hydro-alcoholic extract of some medicinal plants on control of *Alternaria solani* fungus causing tomato early blight disease." *Iranian Journal of Medicinal and Aromatic Plants* 35.4 (2019).
123. Feroz, S.,J. Ali. "Preliminary phytochemical analysis, antioxidant, and antimicrobial evaluation of *Carthamus lanatus*." *Innovare Journal of Science* 4 (2016): 1-3.
124. Gasimova, Shahla A. "The study of chemical composition of fatty oil from *Carthamus lanatus* (L.) Boiss. seeds."
125. Gilani, A.H., Bukhari, I.A., Khan, R.A., Khan, A.-U., Ullah, F., Ahmad, V.U. Cholinomimetic and calcium channel blocking activities of *Carthamus oxycantha* (2005) *Phytotherapy Research*, 19 (8), pp. 679-683.
126. Gong, Weifan, et al. "Research Progress on Chemical Constituents and Pharmacological Activities of *Carthamus* L." (2017).
127. Hassan, Shurooq Falah, and Intisar Hadi Al-Yasari. Effect of alcoholic extract of

- medicinal plants *Nigella sativa* and *Foeniculum vulgare* in the growth of *Staphylococcus aureus*. *J. Genet. Env. Resources Conserv.*, 9.1 (2021): 202-206.
128. Hassan, Z., Ahmad, V.U., Hussain, J., Zahoor, A., Siddiqui, I.N., Rasool, N., Zubair, M. Two new carthamosides from *Carthamus oxycantha* (2010) *Natural Product Communications*, 5 (3), pp. 419-422.
 129. Hosseinzadeh, H., Fazly Bazzaz, B.S., Haghi, M.M. Antibacterial activity of total extracts and essential oil of *Nigella sativa* L. seeds in mice (2007) *Pharmacologyonline*, 2, 429-435.
 130. Ivancheva, S., M. Nikolova and R. Tsvetkova. Pharmacological activities and biologically active compounds of Bulgarian medicinal plants. In: *Phytochemistry: Advances in Research*, (F. Imperato, ed.), 2006: 87-103.
 131. Karima, Saffidine, Sahli Farida, and Zerroug Mohamed Mihoub. "Antimicrobial activity of an Algerian medicinal plant: *Carthamus caeruleus* L." *Pharmacognosy Communications* 3.4 (2013).
 132. Kasper, Cornelia. *Neue Ansätze in der Zellkulturtechnik*. Diss. 2006.
 133. Khalil, Hany Ezzat, and Anas AlAhmed. Phytochemical Analysis and Free Radical Scavenging Activity of *Carthamus oxyacantha* Growing in Saudi Arabia: A Comparative Study. *Int. J. Pharm. Sci. Rev. Res.*, 45(1):51-55.
 134. Kim, Jiseon, et al. "Assessment of Metabolic Profiles in Florets of *Carthamus* Species Using Ultra-Performance Liquid Chromatography-Mass Spectrometry. *Metabolites* 10.11 (2020): 440, 1-19.
 135. Li, B.-Q., Wu, C.-C., Wu, Z.-W. The development roadmap analysis on China's Forest Parks (2009) *Shengtai Xuebao/ Acta Ecologica Sinica*, 29 (5), pp. 2749-2756.
 136. Liao, Y.W., Chen, C.R., Hsu, J.L., et al. Norcucurbitane triterpenoids from the fruits of *Momordica charantia* var. *abbreviata*. *Nat. Prod. Commun.*, 2013, 8(1), 79-81.
 137. Martinez, M.J.A., R.M. Lazaro, L.M. Bedoya Del Olmo, et al. Anti-infectious activity in the anthemideae tribe. *Studies in Natural Products Chemistry*, 2008, 35, 445-516.
 138. Moosavi, M.R. Nematicidal effect of some herbal powders and their aqueous extracts against *Meloidogyne javanica*. *Nematropica*, 2012, 42(1), 48-56.
 139. Rafiq, M., A. Javaid, A.Shoaib. Antifungal activity of methanolic leaf extract of *Carthamus oxycantha* against *Rhizoctonia solani*. *Pak. J. Bot* 53 (2021): 3, 1133-1139.
 140. Rafiq, Muhammad, Arshad Javaid, and Amna Shoaib. "Possible antifungal and antibacterial constituents in inflorescence extract of *Carthamus oxycantha*." *Mycopath* 15.2 (2017): 89-95.
 141. Roy, A. and C. Saraf. Ethnomedicinal approach in biological and chemical investigation of phytochemicals as antimicrobials, *Pharmaceutical Reviews*, 2006, 4, 2.
 142. Salem, N., Msaada, K., Dhifi, W., et al. Effect of drought on safflower natural dyes and their biological activities. *EXCLI J.*, 2014, 13, 1-18
 143. Schinor, E.C., Salvador, M.J., Ito, I.Y., Dias, D.A. Evaluation of the antimicrobial activity of crude extracts and isolated constituents from *Chresta scapigera* (2007) *Brazilian Journal of Microbiology*, 38 (1), 145-149.
 144. Singh, A. Phytomedicinal investigation for antimicrobials based on chemical and biological properties of herbal medicines: an overview. *Novel Sci. Int. J. Pharm. Sci.*, 2012, 1(7).
 145. Sultan, S. M., Dikshit, N., & Sivaraj, N. (2015). Diversity, distribution and conservation of

- Saffron Thistle (*Carthamus lanatus* L.) in mid-high altitude temperate zone of Jammu and Kashmir, India: A DIVA-GIS study. *Tropical Ecology*, 56(3), 303-310.
146. Tanis, H., Aygan, A., Digrak, M. Antimicrobial activity of four *Nigella* species grown in southern turkey (2009). *Int. Journal of Agriculture and Biology*, 11 (6), pp. 771-774.
 147. Toker, Z., Keskin, C. Composition of essential oil of *Carthamus glaucus* Bieb. subsp. *Glaucus* (2008) *Asian Journal of Chemistry*, 20 (2), pp. 1651-1653.
 148. Yagci-Tuzun, C., Hacıoglu, B. T., Bulbul, A. S., Arslan, Y., & Subasi, I. (2019). Comparative anatomical studies on some species of *carthamus* l. in Turkey. *Fresenius Environmental Bulletin*, 28(5), 4072-4079.
 149. Zhang, W., Cheng, Z., Liu, Y., Wu, J., He, D. The bactericidal and aromatic substance of volatile gas of the plant. *Ecology and Environment*, 2007, 16, 3, 1455-1459.
 150. Zhou Li, Huang Ji, Xu Han, Zhang Xin. Anti-microbial activities and active ingredients of compositae plants. *Acta Botanica Boreali-Occidentalia Sinica*, 2006, 26, 9.
- 4). Trusheva, B., M. Popova, V. Bankova, I. Tsvetkova, C. Naydensky, A.G. Sabatini. A new type of European propolis, containing bioactive labdabes. Rivista Italiana E.P.P.O.S. 36, 3 – 7 (2003)**
151. Al-Ghamdi, Ahmad A., et al. Chemical compositions and characteristics of organic compounds in propolis from Yemen. *Saudi J. Biol. Sci.*, 24.5 (2017): 1094-1103.
 152. Aliboni, A., D'Andrea, A., & Massanisso, P. Propolis specimens from different locations of Central Italy: chemical profiling and gas chromatography– mass spectrometry (GC– MS) quantitative analysis of the allergenic esters benzyl cinnamate and benzyl salicylate. *J. Agr. Food Chem.*, 2010, 59(1), 282-288.
 153. Almohammadi, Ohood Hasan, et al. "Chemical composition of propolis from the Baha region in Saudi Arabia." *Czech Journal of Food Sciences* 36.2 (2018): 109-118.
 154. Aminimoghadamfarouj, Noushin, and Alireza Nematollahi. "Propolis diterpenes as a remarkable bio-source for drug discovery development: a review." *International journal of molecular sciences* 18.6 (2017): 1290.
 155. Bakdash, Abdulsallam, et al. "Chemical composition of propolis from the Baha region in Saudi Arabia." *Czech J. Food Sci* 36.2 (2018): 00-10.
 156. Barrero, A. F., Herrador, M., Arteaga, P., et al. Communic acids: occurrence, properties and use as chirons for the synthesis of bioactive compounds. *Molecules*, 2012, 17(2), 1448-1467.
 157. Carrillo J.L.R. Fundamentos científicos de la apiterapia la miel de abeja y el propóleo. 19 Congreso Internacional de Actualización Apícola, 72-80 6-8 Junio, 2012, Oaxaca, Mexico
 158. Cupull-Santana, R.D., Cortés-Rodríguez, R., Olazábal-Manso, E.E., & Hernández-Medina, C.A. Actividad antifúngica de propóleos obtenidos en tres provincias de Cuba sobre hongos contaminantes en cultivo de tejidos vegetales. *Acta Universitaria*, 2013, 23(6), 3-9.
 159. da Silva J.C.S. Própolis: teor em fenóis totais e actividades antimicrobiana e inibitória da enzima hialorunidase Instituto Politecnico Escola Superior Agraria De BragançaAq, 2012, PhD Thesis.
 160. da Silva Nascimento, Rosilene. "Desenvolvimento de métodos analíticos para a análise de própolis utilizando técnicas espectrométricas e análise multivariada." (2013).
 161. Değirmencioğlu, H. T. (2018). Evaluation of phenolic profile, botanical origin, antioxidant and antimicrobial activities of Turkish propolis (Master's thesis, Sağlık

Bilimleri Enstitüsü).

162. Dezmirean, D. S., Paşca, C., Moise, A. R., & Bobiş, O. (2020). Plant sources responsible for the chemical composition and main bioactive properties of poplar-type propolis. *Plants*, 10(1), 22.
163. Fernández, M. C. Estudio químico de propóleos rojos cubanos. Editorial Universitaria. PhD Thesis, Universidad de la Habana. 2007.
164. Fokt, H., Pereira, A., Ferreira, A.M., et al. How do bees prevent hive infections? The antimicrobial properties of propolis. *Curr. Res. Technol. Educ. Topics Appl. Microbiol. Microb. Biotechnol.*, 2010, 1, 481-4493.
165. Gil-Gonzales, J., Durango-Restrepo, D.L., Rojano, B.A., Martin-Loaiza, G. Antioxidant activity and Chemical Composition of Colombian Propolis. In: *Natural Antioxidants and Biocides from Wild Medicinal Plants* Edited by C. Cespedes, D. Sampietro, D. Seigler, M. Rai. CABI, 2013, 92-116
166. Martínez, J., Garcia, C., Durango, D., et al. Characterization of propolis from municipality of Caldas obtained through two collection methods. *Revista MVZ Córdoba*, 2012, 17(1), 2861-2869.
167. Miguel, M.G. Chemical and biological properties of propolis from the western countries of the Mediterranean basin and Portugal. *Int. J. Pharm. Pharmac. Sci.*, 2013, 5(3), 403-409.
168. Miguel, M. G., & Antunes, M. D. Is propolis safe as an alternative medicine? *J. Pharm. & Bioal. Sci.*, 2011, 3(4), 479-495.
169. Miguel, Maria Graça, and Maria Dulce Antunes. "Is propolis safe as an alternative medicine?." *Journal of pharmacy & bioallied sciences* 3.4 (2011): 479.
170. Milet-Pinheiro, Paulo, et al. "A Semivolatile Floral Scent Marks the Shift to a Novel Pollination System in Bromeliads." *Current Biology* 31.4 (2021): 860-868.
171. Mokam, C. E. D., Tamfu, A. N., Kuissu, M. M., Kucukaydin, S., Mezui, C., Enow-Orock, E. G., & Tan, P. V. (2024). Phenolic profile and chronic gastric ulcer healing effects of Cameroonian propolis. *Clin. Tradit. Med. Pharmacol.*, 5(2), 200140.
172. Ohkura, N., K. Maruyama, F. Kihara-Negishi. Possible antithrombotic properties of propolis. *J. Apitherapy* 7.1 (2020): 1-9.
173. Paul, S., Emmanuel, T., Matchawe, C., Alembert, T. T., Elisabeth, Z. O. M., Maurice, T. A. G. A. T. S. I. N. G., & Joel, Y. A. G. Pentacyclic triterpenes and crude extracts with antimicrobial activity from Cameroonian brown propolis samples. *J. Appl. Pharm. Sci.*, (2014), 4(07), 001-009.
174. Rabiya, A. H. Treatment of periodontal disease with high functional paste made from nanoemulsion gel "NBF". *Guident*, 2013, 7(1), 102-103.
175. Righi, A.A. Extratos brutos e constituintes de própolis brasileiras: avaliação dos efeitos nos carrapatos *Rhipicephalus sanguineus*, *Rhipicephalus microplus* e *Amblyomma cajennense*, PhD Thesis, Instituto de Biociências, Sao Paulo, 2013.
176. Roldan, A.M., J. Enrique, R. Chahboun. Procedimiento para la obtención de ácido trans-comúnico y derivados a partir de *Cupressus sempervirens* L. (patent), ES 2 284 341 B1, 2008, 10-16.
177. Rushdi, A. I., Adgaba, N., Bayaqoob, N. I., Al-Khazim, A., Simoneit, B. I., El-Mubarak, A. H., & Al-Mutlaq, K. F. Characteristics and chemical compositions of propolis from Ethiopia. *SpringerPlus*, 2014, 3(1), 1-9.

178. Santana, René Dionisio Cupull. Actividad antifúngica de propóleos obtenidos en tres provincias de Cuba sobre hongos contaminantes en cultivo de tejidos vegetales. (2014).
179. Shehu A., Rohin M.A.K., Aziz A.A., Ismail S. Antifungal, characteristic properties and composition of bee glue (propolis). J. Chem. Pharm. Res., 2015, 7(3): 1992-1996.
180. Sulaeman, A., Kalsum, N., & Mahani, M. (2019). Trigona propolis and its potency for health and healing process. In: The role of functional food security in global health (pp. 425-448). Academic Press.
181. Tamfu, A. N., et al. "A new isoflavonol and other constituents from Cameroonian propolis and evaluation of their anti-inflammatory, antifungal and antioxidant potential" Journal of Natural Products and Resources.
182. Tran, Trong D., et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020):4988.
183. Wagh, V.D, Propolis: a wonder bees product and its pharmacological potentials. Adv. Pharm. Sci., 2013, Art. ID 308249, 1-11.
184. Wagh, V.D., & Borkar, R.D. Indian propolis: a potential natural antimicrobial and antifungal agent. Int. J. Pharm. Pharmac. Sci., 2012, 4, 12-17.
185. Zammit, E.J., Theuma, K.B., Darmanin, S., et al. Totarol content and cytotoxicity varies significantly in different types of propolis. Res. J. Pharm., Biol. Chem. Sci., 2013, 4, 3, 1047-1057.
186. Zingue, Stéphane, et al. "Ethanol-extracted Cameroonian propolis exerts estrogenic effects and alleviates hot flushes in ovariectomized Wistar rats. BMC Compl. Altern. Med., 17.1 (2017): 1-17.

5). Orozova, P., Chikova, V., Kolarova, V., Nenova, R., Konovska, M., NAJDENSKI, H. Antibiotic resistance of potentially pathogenic Aeromonas strains. Trakia J. Sci., 2008, 6, 71-77.

187. Abou-Elela, G.M., N.A. El-Sersy, H. Abd-Elnaby, and S.H Wefki. Distribution and bio-diversity of faecal indicators and potentially harmful pathogens in North Delta (Egypt) - Australian Journal of Basic and Applied Sciences, 2009, 3, 4, 3374-3385.
188. Akpudo, M. O., Jimoh, O., Adeshina, G. O., & Olayinka, B. O. (2023). Biofilm Formation and Antimicrobial Susceptibility Pattern of Staphylococcus aureus Clinical Isolates from Two Healthcare Facilities in Zaria. Nigerian J. Pharm. Res., 19(1), 59-70.
189. Al-Ruwaili, M.A. Bacterial assessment and antibiotic susceptibility of pathogenic. Microbes in Applied Research: Current Advances and Challenges, 2012, 33-38.
190. Anowai, C. O., O. O. Agarry, and B. C. Akin-Osanaiye. Antibiotic susceptibility profile of Staphylococcus aureus isolated from food handlers in Abuja, Nigeria Review.
191. Bright Singh, I. S., Philip Rosamma, and K. Sreedharan. "Virulence Potential And Antibiotic Susceptibility Pattern Of Motile Aeromonads Associated With Freshwater Ornamental Fish Culture Systems: A Possible Threat To public Health." (2012).
192. Châu, Đ.T.M.,LH. Mai, T.T. Dung.Antimicrobial susceptibility of bacterial pathogen causing internal white spot disease in snakehead fish (Channa striata) in Tra Vinh province. Tạp chí Khoa học Trường Đại học Cần Thơ 54, 2018, (2):108-115
193. Diêu, HK. Antibacterial activity of some medicinal plants in the Mekong Delta of VietNam against common fish pathogens., Tạp chí Khoa học 2010:15b222-229.
194. Diêu, H.K.. Evaluation the genetic diversity and anti-bacterial activity of

- Melaleuca leucadendra leaf. *Tạp chí Khoa học Cần Thơ* 2011:19a 143-148.
195. dos Santos, F. G. B., Gouveia, G. V., de França, C. A., de Souza, M. G., & da Costa, M. M. Microbiota bacteriana com potencial patogênico em pacamã e perfil de sensibilidade a antimicrobianos. *Revista Caatinga*, 27(2), 2014, 176-183.
 196. Etalong, V., Tytler, B. A., Profit, I., et al. (2021). Antibiotic susceptibility pattern of *Escherichia coli* isolated from food, cooking utensils and palms of food handlers in some restaurants in Zaria, Nigeria. *AROC Pharm. Biotechnol.*, 1(2), 28-34.
 197. Ezech, C. K., Eze, C. N., Dibua, M. E. U., & Emencheta, S. C. (2023). A meta-analysis on the prevalence of resistance of *Staphylococcus aureus* to different antibiotics in Nigeria. *Antimicrobial Resistance & Infection Control*, 12(1), 40.
 198. Garba, S., et al. Vancomycin resistant *Staphylococcus aureus* from clinical isolates in Zaria Metropolis, Kaduna State. *Clin Infect Dis* 2.105 (2018): 2.
 199. Hamid, Nur Hidayahanum. Evaluation of natural immunostimulants for growth promotion and protection against *Aeromonas hydrophila* in juvenile red hybrid Tilapia (*Oreochromis Sp.*). Diss. Universiti Putra Malaysia, 2014.
 200. Igbinosa, I.H., E.O. Igbinosa, A.I. Okoh. Antibigram characterization and putative virulence genes in *Aeromonas* species isolated from pig fecal samples. *Env. Sci. Pollution Res.*, 23.12 (2016): 12199-12205.
 201. Igwe J.C. et al. Impact of outer membrane protein OmpC and OmpF on antibiotics resistance of *E. coli* isolated from UTI and diarrhoeic patients in Zaria, Nigeria, *Clin. Microbiol.*, Open Access. (2016).
 202. Igwe, J.C., et al. Tetracycline resistant genes in *E. coli* isolated from UTI and diarrhea patients in Zaria, Nigeria. *Clin. Microbiol.*, Open Access (2015).
 203. Igwe, J., et al. Virulent characteristics of multidrug resistant *E. coli* from Zaria, Nigeria. *Clin Microbiol Open Access* 5 (2016).
 204. Igwe, J. C., Onaolapo, J. A., Ehimidu, J. O., Bolaji, R. O., Tytler, A. B., Ojiego, B. O., ... & Salihu, M. S. (2016). Antibiotic susceptibility profile of *E. coli* serotype O157: H7 in ABUTH, Zaria, Nigeria. *Int J Trop Dis Health*, 11(1), 1-8.
 205. Islam, M. S., Hossain, M. M. M., Neowajh, M. S., Kholil, M. I., Khatun, A., Rikta, S., & Imteazzaman, A. M. (2015). Antimicrobial resistance of *Aeromonas* spp. isolated from the carp farm following Streptomycin treatment. *Int. J. Fish. Aquatic Studies*, 2, 99-102.
 206. Kalgo, Z. M., Amin, B. M., Muhammed, B., & Saka, H. K. (2022). Antibigram of Bacteria Isolated from Patients with Lower Respiratory Tract Infection at the general outpatient department of some hospitals in Kebbi State, Nigeria. *Microbes, Infection and Chemotherapy*, 2, e1549-e1549.
 207. Lavanya, C., Neeraja, T. and Ramana, T.V., 2023. Antimicrobial resistance study of pathogenic aeromonas spp. Isolated from freshwater fish, catla catla of andhra pradesh, india: a major threat to public health. *Journal of Experimental Zoology India*, 26(2).
 208. Loka, Jayasree, et al. "Multiple antibiotic resistance pattern of *Vibrio harveyi* from luminous vibriosis affected cultured tiger shrimp, *Penaeus monodon* in Andhra Pradesh, India." *Intern. J. Curr. Microbiol. Appl. Sci.*, 4.11 (2015): 523-535.
 209. M Aly, S., F Khalil W., M. Ghaleb S. Antibacterial activity, biochemical effect and tissue residue of fourth generation cephalosporin used in treatment of Nile tilapia fish against bacterial infection." *Egyptian J. Aquatic Biol. Fisheries*, 24.3 (2020): 29-43.
 210. Mahyuni, S., T. Sofihidayati. Kadar saponin dan aktivitas antibakteri ekstrak daun

- Filicium decipiens* (Wight & Arn.) Thwaites Terhadap *Staphylococcus aureus*, *Escherichia coli* DAN *Candida albicans*. *Fitofarmaka: Jurnal Ilmiah Farmasi* 8.2 (2018): 92-109.
211. Morshdy, A. E., Tharwat, A. E., Merwad, A., Abdallah, N. A., & Saber, T. (2023). Prevalence, phenotypic-genotypic resistance and biofilm formation of *Staphylococcus aureus* in chicken meat with reference to its public health hazard. *Slovenian Veterinary Research/Slovenski Veterinarski Zbornik*, 60.
 212. Mudryk, Z.J., Kosiorek, A., & Perliński, P. In vitro antibiotic resistance of *Vibrio*-like organisms isolated from seawater and sand of marine recreation beach in the southern Baltic Sea. *Hydrobiologia*, 2013, 702(1), 141-150.
 213. Mudryk, Zbigniew J., et al. Occurrence of potentially human pathogenic bacteria in the seawater and in the sand of the recreational coastal beach in the southern Baltic Sea. *Oceanological and Hydrobiological Studies* 43.4 (2014): 366-373.
 214. Mudryk, Z.J., P. Perlinski, J. Gackowska. Antibiotic resistance of *Aeromonas* spp. isolated from seawater and sand of marine recreation beach in the southern Baltic Sea. *Baltic Coastal Zone. J. Ecol. Protect. Coastline* 19 (2015).
 215. Nofiani, R., S. Nurbetty, A. Sapar. Antimicrobial activities of methanol extract from unidentified sponge associated bacteria in Lemukutan Island, Kalimantan Barat. *Jurnal Ilmu dan Teknologi Kelautan Tropis* 1.2 (2009).
 216. Obajuluwa, A.F., Udobi, C.E., Onaolapo, J.A., et al. Comparative studies of the antibacterial activities of the extracts of parts of the African Locust Bean (*Parkia Biglobosa*) tree against hyper beta lactamase producing staphylococci (phenotypic MRSA) isolates from orthopaedic patients. *Int. J. Pharma & Bio Sciences*, 2010, 1(4).
 217. Obajuluwa, A. F., Parom, S. K., & Kubau, S. K. (2024). Prevalence and Antibigram of Methicillin Resistant *Staphylococcus aureus* Nasal Carriage among Apparently Healthy University Staff and Students in Kaduna, Nigeria. *J. Appl. Sci. Env. Manag.*, 28(3), 967-974.
 218. Okafor, C. N., et al. "Antibiotic susceptibility profile of *Escherichia coli* serotype O157: H7 from River Kaduna, Nigeria." *African J. Nat.Sci., (AJNS)* 21 (2019).
 219. Olowo-Okere, A., Atata, R. F., Abass, A., Shuaibu, A. S., Yahya, U. H., & Tanko, N. (2017). Incidence and antibiotic susceptibility profile of *Staphylococcus aureus* isolates from wounds of patients at specialist hospital, Sokoto, Nigeria. *Journal of medical Bacteriology*, 6(3-4), 44-50.
 220. Onaolapo, J. A., et al. Antimicrobial susceptibility pattern of *Staphylococcus aureus* isolates from orthopaedic patients in Abuth, Zaria. *J Food Ind. Microbiol*, 2.106 (2016):2.
 221. Raheema, R. H., & Abed, K. A. (2019). Molecular identification of Virulence genes of *Staphylococcus aureus* isolated from different clinical isolates. *Indian Journal of Public Health Research & Development*, 10(2), 212-218.
 222. Rihibiha, D.D., Hatmanti, A., Antariksa, M.M. et al. (2018). Skrining aktivitas antibakteri isolat bakteri simbiosis teripang dari perairan lombok. prosiding pertemuan ilmiah nasional penelitian dan pengabdian masyarakat i (PINLITAMAS 1), 1(1), 603-607.
 223. Skórczewski, P., Mudryk, Z. J., Miranowicz, J., et al. Antibiotic resistance of *Staphylococcus*-like organisms isolated from a recreational sea beach on the southern coast of the Baltic Sea as one of the consequences of anthropogenic pressure. *Oceanological and Hydrobiological Studies*, 2014, 43(1), 41-48.
 224. Sreedharan, K., R. Philip, I.S.B. Singh. Virulence potential and antibiotic susceptibility pattern of motile aeromonads associated with freshwater ornamental fish culture systems: a

- possible threat to public health. *Braz. J. Microbiol.*, 43.2, (2012): 754-765.
225. Stratev, D. O.A. Odeyemi. Antimicrobial resistance of *Aeromonas hydrophila* isolated from different food sources: A mini-review. *J. Inf. Publ.Health* 9.5 (2016): 535-544.
 226. Velichkova, K., I. Sirakov, S. Denev. "In vitro antibacterial effect of *Lemna minuta*, *Chlorella vulgaris* and *Spirulina* sp. extracts against fish pathogen *Aeromonas hydrophila*." *Aquaculture, Aquarium, Conservation & Legislation* 12.3 (2019): 936-940.
 227. Wiyoko, T., Agrita, T. W., Hermana, N. S. P., & Indrawati, A. (2024). Detection of Resistance Gene from *Aeromonas hydrophila* Isolated from Catfish Farming in Jambi. *Jurnal Penelitian Pendidikan IPA*, 10(2), 583-588.
 228. Zdanowicz, M., Z.J. Mudryk, P. Perliński. Abundance and antibiotic resistance of *Aeromonas* isolated from the water of three carp ponds. *Vet. Res. Commun.*, 44.1 (2020): 9-18.
- 6). Popova, M., V. Bankova, S. Spassov, I. Tsvetkova, C. NAYDENSKI, M. V.Silva, M. Tsartsarova. New Bioactive Chalcones in Propolis from El Salvador. *Z. Naturforsch.* 56c, 593 – 596 (2001).**
229. Alday, Efrain, et al. "Advances in pharmacological activities and chemical composition of propolis produced in Americas." *Beekeeping and Bee Conservation—Advances in Research* (2016).
 230. Al-Saheb, Rawan, et al. "Synthesis of new pyrazolone and pyrazole-based adamantyl chalcones and antimicrobial activity." *Bioscience Reports* 40.9 (2020):BSR20201950
 231. Bin, Z., X. Bing. Research progress on propolis pathogenic biological effect. *Chinese J. Pathogen Biol.*, 2011, 6 (7), 547-552.
 232. Beltrán Herrera, E. A., & Figueroa Deras, M. N. (2021). Evaluación de la actividad antimicrobiana in vitro de extractos de propoleos de diferentes zonas de El Salvador (Doctoral dissertation, Universidad de El Salvador).
 233. Brahma, P., Basumatary, S., & Baruah, S. (2024). Micromorphology, phytochemical screening, and GC-MS analysis of bioactive compounds isolated from *Papilionanthe teres* (Roxb.) Schltr.: A potent medicinal plant. *Med. Plants*, 2024, 16,1, 145-155.
 234. Buchta, V., Černý, J., & Opletalová, V. In vitro antifungal activity of propolis samples of Czech and Slovak origin. *Cent. Eur. J. Biol.*, 2011, 6(2), 160-166.
 235. Cai Shuang, Shi Qing, Li Yujing. The inhibitory effect of propolis varnish on the growth and adhesion of *S. mutans* and *S. sobrinus*. *J. Pract. Stomatol.*, 2006, 22(2), 171-174
 236. Correa, R., Fenner, B.P., Buzzi, F.D.C., Cechinel Filho, V., Nunes, R.J. *Zeitschrift fur Naturforschung - Section C Journal of Biosciences* 63(11-12), 830-836 (2008)
 237. Ferreira, J.M., et al. "New propolis type from north-east Brazil: chemical composition, antioxidant activity and botanical origin." *J. Sci. Food Agric.*, 2017, 97, 11, 3552-3558.
 238. Joya, M, M. Gil, G. Bastidas-Pacheco. Actividad fungistática y fungicida de extractos etanólicos de propóleos sobre el crecimiento in vitro de cepas del género *Candida*. *Revista Tecnología en Marcha* 30.3 (2017): 3-11.
 239. Kwesiga, G. (2022). Synthesis of isoflavonoids from African medicinal plants with activity against tropical infectious diseases (Doctoral dissertation, Universität Potsdam).
 240. Li Yiping. (2010). Research on the antibacterial properties of propolis and its application in toothpaste. *Daily Chemical Science*, 33(7), 25-29.
 241. Li Yiping. (2009). Research on the antibacterial properties of propolis extracted by supercritical CO₂ and its application in toothpaste. *Toothpaste Industry*, 19(2), 17-21.

242. Li Pingping, Lin Juhong, & Liu Mingfang. (2010). Effects of water-soluble propolis on cariogenic plaque biofilm morphology and acid production. *Journal of Dental Endodontics and Periodontology*, (5), 277-280.
243. Li, Y. P., Chen, Y. P., Chen, X. L., Tao, Y. S., & Zhang, R. P. (2022). A New Phenanthrene Derivative with α -Glucosidase Inhibitory Activity from *Pleione maculata*. *Chemistry of Natural Compounds*, 58(1), 6-8.
244. Méndez Ramírez, G. R. (2020). Evaluación de las características fisicoquímicas y perfil cromatográfico por TLC de propoleo procedentes de cinco localidades de El Salvador (Doctoral dissertation, Universidad de El Salvador).
245. Moorthi SS, Chinnakali K, Nanjundan S, et al. 1-(4-Hydroxyphenyl)-3-(4-methoxyphenyl)-prop-2-en-1-one. *Acta Crystallographica section e: Structure reports online* 61: O480-O482 Part 2 (2005).
246. Nan Yao, Guo Jia, Zheng Lianxiang, & Zhou Lidong. (2006). Research progress on the chemical components of propolis. *World Science and Technology: Modernization of Traditional Chinese Medicine*, 8(1), 61-71.
247. Ohsaki, A., R. Yokoyama, H. Miyatake, F. Fukuyama. *Chem. Pharm. Bull.* 54(12), 1728 – 1729 (2006)
248. Ping, L. Antibacterial propolis in toothpaste. *Daily Chem. Sci.*, 2010, 33 (7), 25-29.
249. Ping, L.P., L. Fang. Effects on acid production of water-soluble propolis on cariogenic bacteria and plaque biofilm morphology. *Conserv. Dent. J.*, 2010, 5, 277-280
250. Poblócka-Olech L., Krauze-Baranowska M. Pharmacological Activity of Chalcones Borgis - *Postępy Fitoterapii* 2007, 4, 194-201
251. Quintero-Mora ML, Londoño-Orozco A, Hernández-Hernández F, Manzano-Gayosso P, López-Martínez R, Soto-Zárate CI, Carrillo-Miranda L, Penieres-Carrillo G, García-Tovar CG, Cruz-Sánchez TA. *Rev Iberoam Micol.* 25(1):22-6 (2008).
252. Ravishankar, T., Chinnakali, K., Nanjundan, S., Selvamalar, C. J., Ramnathan, A., Usman, A., & Fun, H. K. (2003). 3-(3, 4-Dimethoxyphenyl)-1-(4-hydroxyphenyl) prop-2-en-1-one. *Acta Crystallograph., Section E: Structure Reports Online*, 59(8), o1143-o1145.
253. Ruddock, P. (2005). Phytochemical mixtures and NHPs as antimicrobials against antibiotic resistant *Neisseria gonorrhoeae* and other pathogens (Doctoral dissertation, University of Ottawa (Canada)).
254. RD'Arcy, B. Antioxidants in Australian Floral Honeys –Identification of health-enhancing nutrient components. A report for the Rural Industries Research and Development Corporation, 2005, RIRDC Publication No 05/040, 2005, p.94.
255. Salatino, A., Fernandes-Silva, C. C., Righi, A. A., & Salatino, M. L. F. Propolis research and the chemistry of plant products. *Nat. Prod. Rep.*, 2011, 28(5), 925-936.
256. Sathiya Moorthi, S., Chinnakali, K., Nanjundan, S., Selvam, P., Fun, H. K., & Yu, X. L. (2005). 3-(3-Hydroxyphenyl)-1-(4-methoxyphenyl) prop-2-en-1-one. *Acta Crystallographica Section E: Structure Reports Online*, 61(3), o743-o745.
257. Seidel, V., Peyfoon, E., Watson, D.G. et al. Comparative study of the antibacterial activity of propolis from different geographical and climatic zones. *J. Phytotherapy Res.*, 22 (9), 1256-1263 (2008).
258. Selvi, M.T., Chinnakali, K., Nanjundan, S. et al. 1-(4-Aminophenyl)-3-(3,4-dimethoxyphenyl)-prop-2-en-1-one. *Acta Crystallographica Section E: Structure Reports Online*, 2003, 59(8), o1146-o1148.

259. Senthana, S., Srinivasan, S., Kabilan, S. Synthesis, Molecular Structure, Spectral, Thermal, and DFT Studies of an Organic Crystal: 1-(benzo[d][1,3]dioxol-5-yl)-3-phenylprop-2-en-1-One. *Molecular Crystals and Liquid Crystals* 2015, 609, 249-265
260. Sforcin, Jose M. "Biological properties and therapeutic applications of propolis." *Phytotherapy research* 30.6 (2016): 894-905.
261. Suárez, M.D.L.C.P., Carballo, M.M.R., Cruz, E.G., Banqueris R.F.; I.B.Núñez; L.A. Hechavarria. Propoleum Gel for subprothesis stomatitis with associated candidiasis. *Multimed* 2006; 10(2) <http://www.multimedgrm.sld.cu/articulos/2006/v10-2/1.html>
262. Suárez, M.D.L.C.P., Cruz, E.G., Carballo, M.M.R., et al. Propoleoterapia en la estomatitis subprotesis con candidiasis asociada. *Hospital Universitario "Celia Sánchez Manduley"*, Clínica Estomatológica Manzanillo, Granma, 2010, 1-15.
263. Sudha, S., Sundaraganesan, N., Vanchinathan, K., et al. Spectroscopic (FTIR, FT-Raman, NMR and UV) and molecular structure investigations of 1, 5-diphenylpenta-1, 4-dien-3-one: A combined experimental and theoretical study. *J. Mol. Struct.*, 2012. 1030, 191-203.
264. Sudha, S., Sundaraganesan, N., Vanchinathan, K., et al. Spectroscopic (FTIR, FT-Raman, NMR and UV) and molecular structure investigations of 1,5-diphenylpenta-2,4-dien-1-one: A combined experimental and theoretical approach. 2013, *Mol. Simulatio*, 4, 39, 330-349.
265. Suleman, Tasneem. The antimicrobial and chemical properties of South African propolis. Doctoral Dissertation, 2016.
266. Sumathi, A., et al. "FT-IR, FT-Raman and UV-Visible Analysis of (2E, 6E)-2, 6-Dibenzylidene-4-(4-Hydroxyphenyl) Cyclohexanone-DFT Method." *Journal of Applicable Chemistry* 5.2 (2016): 346-368.
267. Tran, T. D. et al. Lessons from exploring chemical space and chemical diversity of propolis components. *International journal of molecular sciences* 21.14 (2020): 4988.1-35.
268. Umaña, E., Solano, G., Zamora, G. and Tamayo-Castillo, G., 2023. Costa Rican Propolis Chemical Compositions: Nemorosone Found to Be Present in an Exclusive Geographical Zone. *Molecules*, 28(20), p.7081.
269. Vanchinathan, K., Bhagavannarayana, G., Muthu, K., & Meenakshisundaram, S.P. Synthesis, crystal growth and characterization of 1, 5-diphenylpenta-1, 4-dien-3-one: An organic crystal. *Physica B: Condensed Matter*, 2011, 406(22), 4195-4199.
270. Vasilev, R.F., Kancheva, V.D., Fedorova, G.F., et al. Antioxidant activity of chalcones: The chemiluminescence determination of the reactivity and the quantum chemical calculation of the energies and structures of reagents and intermediates. *Kinetics and Catalysis*, 2010, 51(4), 507-515.
271. Wali, A., et al. Bee Propolis (Bee's glue): A Phytochemistry Review. *J. Crit. Rev.*, 4.4 (2017).
272. Yan Xiong. In vitro antibacterial activity of domestic-made water-soluble propolis against oral anaerobic bacteria from the infected root canal. *Journal of Chongqing Medical University*, 2008, 33(12)
273. Yang Shuzhen, Peng Litao, Yao Xiaolin, & Pan Siyi. (2009). Research progress on the antifungal effect of propolis. *Food Industry Science and Technology*, (11), 349-352.
274. Yao, N., G. Jia, Zh. Lian-Xiang, Zh. Li-Dong. *Advances in Studies on Chemical Constituents of Propolis*. World Science and Technology-Modernization of Traditional

- Chinese Medicine and Materia Medica, 2006, 8, 1.
275. Zhang Hongmei, Lin Juhong, & Jiang Lin. (2008). In vitro antibacterial experiment of domestic water-soluble propolis on anaerobic bacteria in infected root canals. *Journal of Chongqing Medical University*, 33(12), 1516-1519.
 276. Zheng Bin, & Xu Binghong. (2011). Research progress on the anti-pathogenic effects of propolis. *Chinese Journal of Pathogen Biology*, 6(7), 547-552.
 - 7). NAJDENSKI, H., Golkocheva, E., Vesselinova, A., Bengoechea, J.A., Skurnik, M. Proper expression of the O-antigen of lipopolysaccharide is essential for the virulence of *Yersinia enterocolitica* O:8 in experimental oral infection of rabbits. (2003) *FEMS Immunology and Medical Microbiology*, 38 (2), pp. 97-106.**
 277. Clarke, B.R., L. Cuthbertson, and C. Whitfield. Nonreducing terminal modifications determine the chain length of polymannose O antigens of *Escherichia coli* and couple chain termination to polymer export via an ATP-binding cassette transporter. *J. Biol. Chem.*, 2004, 279, 34, 35709-35718.
 278. Clarke, B.R., Richards, M.R., Greenfield, L.K., et al. In vitro reconstruction of the chain termination reaction in biosynthesis of the *Escherichia coli* O9a O-polysaccharide the chain-length regulator, WbdD, catalyzes the addition of methyl phosphate to the non-reducing terminus of the growing glycan. *J. Biol. Chem.*, 2011, 286(48), 41391-41401.
 279. De Castro, C. (2021). Characterisation of the lipopolysaccharide and peptidoglycan and their structural determinants when bound to major proteins involved in its transport across the periplasm. PhD dissertation in Chemical Sciences, University of Naples Federico II.
 280. Fang, X., M. Zhang, S. Li, C. Du, C. Sun, W. Han, L. Zhou and L. Lei. Differential gene expression in the pathogenic strains of *Actinobacillus pleuropneumoniae* serotypes 1 and 3. *Journal of Microbiology and Biotechnology*. 2010, 20(4), 789-797.
 281. Feodorova, V.A., Golova, A.B. Antigenic and phenotypic modifications of *Yersinia pestis* under calcium and glucose concentrations simulating the mammalian bloodstream environment (2005). *J. Med. Microbiol.*, 54 (5), 435-441.
 282. Greenfield LK, Whitfield C. Synthesis of lipopolysaccharide O-antigens by ABC transporter-dependent pathways. *Carbohydr. Res.*, 2012, 356, 12-24.
 283. Guo, H., Lokko, K., Zhang, Y., Yi, W., Wu, Z., Wang, P.G. Overexpression and characterization of Wzz of *Escherichia coli* O86:H2 (2006) *Prot. Expr. Pur.*, 48 (1), 49-55
 284. Guo, H., Yi, W., Shao, J., Lu, Y., Zhang, W., Song, J., Wang, P.G. Molecular analysis of the O-antigen gene cluster of *Escherichia coli* O86:B7 and characterization of the chain length determinant gene (wzz) (2005) *Appl. Environm. Microbiol.*, 71 (12), 7995-8001.
 285. Hering, N. A. (2011). Die epitheliale Barriere als Zielstruktur pathogener und probiotischer Bakterien: Wirkmechanismen von *Y. enterocolitica* und *E. coli* Nissle (Doctoral dissertation).
 286. Kaszowska, M., Jachymek, W., Lukasiewicz, J., et al. The unique structure of complete lipopolysaccharide isolated from semi-rough *Plesiomonas shigelloides* O37 (strain CNCTC 39/89) containing (2S)-O-(4-oxopentanoic acid)- α -D-Glcp (α -D-Lenose). *Carbohydr. Res.*, 2013, 378, 98-107.
 287. Kukkonen, M., Structure-function relationships in the omptin family of enterobacterial proteases/adhesins. Acad. Dissert. in Gen. Microbiol., Helsinki 2003.

288. Liu, D. (Ed.). (2011). Molecular detection of human bacterial pathogens. CRC press.
289. Matthias Ullrich. Bacterial polysaccharides. Current innovations and future trends (2009) Horizon Scientific Press
290. Pinta, E. Biosynthesis of *Yersinia enterocolitica* serotype O: 3 lipopolysaccharide outer core., Publications of The University Of Turku, Annales Universitatis Turkuensis, 927, 2010.
291. Reyes, R.E., S.H. Ramírez, G.C. Solís, H.M. Ortiz, R.C. Jimenez. Mecanismos involucrados en la variabilidad del antígeno O de bacterias Gram negativas. Revista Latinoamericana de Microbiología, 2009, 52, 32-43.
292. Rosa E. Reyes, Saad H.R., Galicia C.S., Herrera M.O., Jimenez R.C. Mecanismos involucrados en la variabilidad del antígeno O de bacterias Gram negativeas (2009) Rev Latinoam Microbiol, 51 (1-2), pp. 31-43.
293. Stevenson, R. M. W. (2008). Investigating survival mechanisms of *Yersinia ruckeri* in rainbow trout, *Oncorhynchus mykiss* (Doctoral dissertation, University of Guelph).
294. Tang, K.-H., Guo, H., Yi, W., Tsai, M.-D., Wang, P.G. Investigation of the conformational states of Wzz and the Wzz·O-antigen complex under near-physiological conditions (2007) Biochemistry, 46 (42), pp. 11744-11752.
295. Tran E.N.H., Morona R. Residues located inside the *Escherichia coli* FepE protein oligomer are essential for lipopolysaccharide O-antigen modal chain length regulation Tran. Microbiology (United Kingdom), 4, 159, 2013, 701-714.
296. Tran, E.N.H., Papadopoulos, M., & Morona, R. Relationship between O-antigen chain length and resistance to colicin E2 in *Shigella flexneri*. Microbiology, 2014, 160(Pt 3), 589-601.
297. Triet, Tran H., et al. Development and potential use of an *Edwardsiella ictaluri* wzz mutant as a live attenuated vaccine against enteric septicemia in *Pangasius hypophthalmus* (Tra catfish). Fish & Chellfish Immunology 87 (2019): 87-95.
298. Vanantwerpen, G. (2014). Prevalence and risk factors of enteropathogenic *Yersinia* spp. in pigs at slaughter age (Doctoral dissertation, Ghent University).
299. Wang, X., Duan, R., Liang, J.Gu, W., Jing, H. *Yersinia*, Laboratory Models for Foodborne Infections, 2017, 427 – 437.
300. Whitfield, C., Fridrich, E., & Reid, A. N. (2006). Periplasmic Events in the Assembly of Bacterial Lipopolysaccharides. The Periplasm, 214-234.
301. Xie F, Zhang MJ, Li SQ, et al. Differential gene expression in the pathogenic strains of *Actinobacillus pleuropneumoniae* serotypes 1 and 3. J. Microbiol. Biotechnol., 2010, 20, 4, 789-797
302. Yang, Z., Li, X., Qi, X., et al. Identification and functional analysis of the chain length determinant gene *ste8* involved in the biosynthesis of ebosin by *Streptomyces* sp. J. Microbiol. Biotechnol., 2013, 11, 23, 1500-1508
303. Zhang, Y., Li, X., Qi, X., et al. Identification and functional analysis of the gene *ste9* involving in Ebosin biosynthesis from *Streptomyces* sp. 139. FEMS Microbiol.Let. 2014, 350(2), 257-264
304. Zhao, G., Liu, J., Liu, X., Chen, M., Zhang, H., Wang, P.G. Cloning and characterization of GDP-perosamine synthetase (Per) from *Escherichia coli* O157:H7 and synthesis of GDP-perosamine in vitro (2007) Biochem. Biophys. Res. Comm., 363 (3), 525-530.

305. Дентовская, С. В. (2012). Молекулярно-генетические механизмы образования и функциональная значимость липополисахарида *Yersinia pestis* (Doctoral dissertation, диссертация на соискание ученой степени доктора медицинских наук: 03.02. 03-микробиология 03.01. 04-биохимия. М).
306. Паунова-Кръстева, Ц.С. Фенотипни вариации свързани с полизахаридните антигени при *Escherichia coli* O157. Дисертация за присъждане на образователната и научна степен „доктор”. София 2015
- 8). Kamenarska, Z., Dimitrova-Konaklieva, S., Stefanov, K., NAJDENSKI, H., Tzvetkova, I., Popov, S. Comparative study of the volatile compounds from some Black Sea brown algae (2002) Botanica Marina, 45 (6), pp. 502-509.**
307. Alcaide, María, et al. Pressure adaptation is linked to thermal adaptation in salt-saturated marine habitats. *Environmental microbiology* 17.2 (2015): 332-345.
308. Alves, C., Oliveira, T., Pio, C., Silvestre, A.J.D., Fialho, P., Barata, F., Legrand, M. Characterisation of carbonaceous aerosols from the Azorean Island of Terceira (2007) *Atmospheric Environment*, 41 (7), 1359-1373.
309. Amini, F. (2020). Heavy Metal Concentrations in *Padina gymnospora* and *Padina tetrastrum* (Dictyotaceae, Ochrophyta,) and Sediments of Bushehr Coastline (Bushehr Province, Iran). *Plant, Algae, and Environment*, 4(1), 497-507.
310. Bazzi, F., Aslani, E., & Heidari, F. (2018). Effect of Cyanobacterial Extract on Medicinal Plant *Mentha piperita* L. *Journal of Phycological Research*, 2(1), 154-163.
311. Borik, Rita M. Volatile compounds extraction, fractionation and identification from the red alga *Corallina officinalis*. *World Applied Sciences Journal* 30.6 (2014): 741-746.
312. Brito, Leonor, Mary Isabel Segnini, and Oscar Crescente. "Identificación de Algunos Constituyentes Químicos de *Gelidium Serrulatum* (Gelidiales: Rhodophyta) Mediante Cromatografía de Gases-Espectrometría de Masas." *Boletín del Instituto Oceanográfico de Venezuela* 55.2 (2016).
313. Cajnko, Miša Mojca, Uroš Novak, and Blaž Likozar. Cascade valorization process of brown alga seaweed *Laminaria hyperborea* by isolation of polyphenols and alginate." *J. Appl. Phycol.*, 31.6 (2019): 3915-3924.
314. Davari, M., Modaressi, Z., & Aghashariatmadari, Z. (2018). Taxonomic Study on Cyanobacteria Species in Natural Habitats of *Tanacetum parthenium* Emphasising on *Wolffia* and *Cylindrocapsa* Morphological Characters. *Plant, Algae, and Environment*, 2(2), 237-253.
315. Demirel, Z., F.F. Yilmaz-Koz, N.U. Karabay-Yavasoglu, et al. Antimicrobial and antioxidant activities of solvent extracts and the essential oil composition of *Laurencia obtusa* and *Laurencia obtusa* var. *pyramidata*. *Romanian Biotechnol. Lett.*, 2011, 16, 1.
316. Demirel, Z., Yilmaz-Koz, F.F., Karabay-Yavasoglu, U.N., Ozdemir, G., Sukatar, A. Antimicrobial and antioxidant activity of brown algae from the Aegean Sea [Russian Source] (2009) *Journal of the Serbian Chemical Society*, 74 (6), 619-628.
317. Demirel, Zeliha, et al. Antimicrobial and antioxidant activities of solvent extracts and the essential oil composition of *Laurencia obtusa* and *Laurencia obtusa* var. *pyramidata*. *Romanian Biotechnological Letters* 16.1 (2011): 5927-5936.
318. Deprá, M.C., Dias, R.R., Nascimento, T.C., Silva, P.A., Zepka, L.Q. and Jacob-Lopes, E., 2023. Essential Oils Obtained from Algae: Biodiversity and Ecological Importance.

- Essential Oils: Extraction Methods and Applications, 465-476.
319. Erakin, S., Güven, K.C. The volatile petroleum hydrocarbons in marine algae around Turkish coasts (2008) *Acta Pharmaceutica Scientia*, 50 (3), 167-182.
 320. Farré-Armengol, G., Filella, I., Llusà, J., Peñuelas, J. Pollination mode determines floral scent (Article) *Biochemical Systematics and Ecology* 2015,61, 44-53
 321. Fauziyah, Nahdlotul, et al. Postharvest processing of *Sargassum duplicatum* for tea products. *J. Appl. Phycol.*, 33.2 (2021): 1209-1216.
 322. Felício, Rafael de. Produtos naturais marinhos: identificação de metabólitos fenólicos halogenados na macroalga *Bostrychia tenella* (Rhodomelaceae, Rhodophyta) e potencial biológico de micro-organismos endofíticos associados. Diss. Univers. de São Paulo, 2010.
 323. Ferraces-Casais, P., Lage-Yusty, M.A., Rodríguez-Bernaldo, De Q-A., et al. Rapid identification of volatile compounds in fresh seaweed. *Talanta*, 2013, 115, 798-800.
 324. Firouzi, J., Gohari, A.R., Rustaiyan, A., et al. Composition of the essential oil of *Nizamuddiniana zanardinii*, a brown alga collected from Oman Gulf. *J. Essent. Oil Bear. Plants*, 2013, 16(5), 689-692.
 325. García, F.S. Evolución de los compuestos volátiles y características sensoriales en *Ulva rigida* durante su almacenamiento. Estudio y Evaluación del Potencial Alimentario del Alga Verde *Ulva* spp. *De Los Esteros Gaditanos*, 99.12: 135.
 326. Ghahraman, Z., Riahi, H., Aghashariatmadari, Z., & Heidari, F. (2020). Morphological and ecologic diversity of thermophilic cyanobacteria in Maragheh mineral springs. *Journal of Phycological Research*, 4(2), 572-581.
 327. Guschina, I.A., Harwood, J.L. Lipids and lipid metabolism in eukaryotic algae (2006) *Progress in Lipid Research*, 45 (2), 160-186.
 328. Güven, KASIM C., Burak Coban, and H. U. S. E. Y. I. N. Erdugan. A chemical research on three red algae *Gracilaria bursa-pastoris*, *Phyllophora crispa* and *Laurencia obtusa* var. *pyramidata*. *Asian Journal of Chemistry* 26.18 (2014): 6118-6120.
 329. Li S., Hu M., Tong Y., Xia Z., Tong Y., Sun Y., Cao J., Zhang J., Liu J., Zhao S., He P. A review of volatile compounds in edible macroalgae. *Food Research International*, 2023, 165, art. no. 112559.
 330. López-Pérez, O., A. Picon, and M. Nuñez. Volatile compounds and odour characteristics of seven species of dehydrated edible seaweeds. *Food Res. Intern.*, 99 (2017): 1002-1010.
 331. López-Pérez, Olga, et al. "Volatile compounds and odour characteristics of five edible seaweeds preserved by high pressure processing: Changes during refrigerated storage." *Algal Research* 53 (2021): 102137.
 332. López-Pérez, Olga, et al. "Volatile compounds and odour characteristics during long-term storage of kombu seaweed (*Laminaria ochroleuca*) preserved by high pressure processing, freezing and salting." *LWT* 118 (2020): 108710.
 333. Lu-lu, L. I. U., & Bao-bei, W. A. N. G. (2021). Volatile compounds in algae and their extraction methods. *Food and Machinery*, 37(2), 220-228.
 334. Mohamed S.F. Extraction Optimization, Analysis and Biological Activity of Volatile Compounds from *Padina pavonia* Collected from Farasan Island's Coasts, Jazan, Saudi Arabia. *Tropical Journal of Natural Product Research*, 2023, 7 (3), 2574 – 2579.
 335. Mohebbi, F., Ghoroghi, A., Riahi, H., Nekoeifard, A., & Seidgar, M. (2017). A Biosystematics Study of *Microcystis* (Cyanobacteria), A Bloom-Forming Cyanobacterium

- from Aras Reservoir (North-West Iran). *Plant, Algae, and Environment*, 1(1), 21-28.
336. Park, Na-Bi, et al. "Antimicrobial activity of *Myagropsis yendoii* extract." *Korean Journal of Fisheries and Aquatic Sciences* 43.6 (2010): 642-647.
 337. Salehi Balashahri, M. (2021). Molecular Identification of *Caulerpa selago* as a New Record For The Persian Gulf. *J. Phycol. Res.*, 5(2), 713-724.
 338. Sánchez-García, Fini, et al. "Effect of different cooking methods on sea lettuce." *Journal of the Science of Food and Agriculture* 101.3 (2021): 970-980.
 339. Sánchez-García, Fini, et al. "Effect of different cooking methods on sea lettuce (*Ulva rigida*) volatile compounds and sensory properties. *Journal of the Science of Food and Agriculture*, 2021, 101, 3, 970-980.
 340. Sánchez-García, Fini, et al. Evolution of Volatile Compounds and Sensory Characteristics of Edible Green Seaweed (*Ulva rigida*) during Storage at Different Temperatures. *Journal of the Science of Food and Agriculture* 99.12 (2019) 5475-5482.
 341. Saxena, Manjula K., Singh Neerja, and Dobhal Mp. Review potent pharmaceutical products from aquatic plants—review. *Asian J. Pharm. Clin. Res.* (2021): 48-63.
 342. Shams, M., & Karimian Shamsabadi, S. (2019). Identification of algae as pollution bioindicators in Shakh-Kenar, Gavkhouni Wetland, Isfahan. *J. Phycol. Res.*, 3(2), 386-394.
 343. Sohrabipour, Jelveh. Fatty Acids Composition of Marine Macroalgae. *Journal of Phycological Research* 3.2 (2019): 348-374.
 344. Sukatar, A., Karabay-Yavaşoglu, N.U., Ozdemir, G., Horzum, Z. Antimicrobial activity of volatile component and various extracts of *Enteromorpha linza* (Linnaeus) J. Agardh from the coast of Izmir, Turkey (2006) *Annals of Microbiology*, 56 (3), 275-279.
 345. Thinakaram, T., and K. Sivakumar. Antifungal activity of certain seaweeds from Puthumadam coast. *Int. J. Res. Rev. Pharm. Appl. Sci.*, 2013, 3, 3, 341-350.
 346. Urllass S., Wu Y., Nguyen T.T.L., Winberg P., Turner M.S., Smyth H. Unravelling the aroma and flavour of algae for future food applications. *Trends in Food Science and Technology*, 2023, 138, 370 – 381.
 347. Wiesemeier, T. (2007). Untersuchungen der aktivierten und induzierten chemischen Verteidigung von marinen Makroalgen (Doctoral dissertation).
 348. Xu, N.-J., Y.-I. He, J. Tang, X.-J. Yan. Volatile metabolites in *Gracilaria lemaneiformis* at high temperature. *Oceanologia and Limnologia Sinica*, 2009, 40, 2, 221-227.
 349. Xu Nianjun, He Yanli, Tang Jun, & Yan Xiaojun. (2009). GC-MS analysis of metabolites of high-temperature stress in *Gracilaria lemaneiformis*. *Ocean and Limnology*, (2), 221-227.
 350. Yılmaz, F.F. et al. Antimicrobial and Antioxidant Activities of *Porphyridium cruentum*. *Hacettepe Üniversitesi Eczacılık Fakültesi Dergisi*, 1: (2017) 1-7.
 351. Yu, K. X., Wong, C. L., Ahmad, R., & Jantan, I. (2015). Larvicidal activity, inhibition effect on development, histopathological alteration and morphological aberration induced by seaweed extracts in *Aedes aegypti* (Diptera: Culicidae). *Asian Pacific journal of tropical medicine*, 8(12), 1006-1012.
 352. Zhu, X., Healy, L.E., Sevindik, O. et al. "Impacts of novel blanching treatments combined with commercial drying methods on the physicochemical properties of Irish brown seaweed *Alaria esculenta*". *Food Chemistry*, 2022, 369, 130949.

9). Kamenarska, Z., Stefanov, K., Dimitrova-Konaklieva, S., NAJDENSKI, H., Tsvetkova, I., Popov, S. Chemical composition and biological activity of the brackish-water green alga *Cladophora rivularis* (L.) Hoek (2004) *Botanica Marina*, 47 (3), pp. 215-221.

353. Abdel-Aal, E. I., Haroon, A. M., & Mofeed, J. (2015). Successive solvent extraction and GC–MS analysis for the evaluation of the phytochemical constituents of the filamentous green alga *Spirogyra longata*. *Egyptian J. Aquat. Res.*, 41(3), 233-246.
354. Abdel-Hamid, M. I., E.I. Abdel-Aal, M. Abdel-Mogib. Isolation and characterization of new *Botryococcus braunii* (Trebouxioophyceae) isolates. *Renewable Energy* 141 (2019): 782-790.
355. Ahmed, H.H., M.M. Hegazi, H.I. Abd-Alla, E.F. Eskander and M.S. Ellithey. Antitumour and antioxidant activity of some red sea seaweeds in Ehrlich ascites carcinoma in vivo. *Zeitschr. Naturforsch.- Sect. C J. Biosci.*, 2011, 66 C(7-8), 367-376.
356. Aoun, Z.B., Said, R.B., Farhat, F. Anti-inflammatory, antioxidant and antimicrobial activities of aqueous and organic extracts from *Dictyopteris membranacea* (2010) *Botanica Marina*, 53 (3), pp. 259-264.
357. Boedeker, Christian, et al. Molecular, biochemical and morphological data suggest an affiliation of *S pongiochrysis hawaiiensis* with the *Trentepohliales* (Ulvophyceae, Chlorophyta). *Phycological Research* 61.2 (2013): 133-144.
358. Cornish, M. L., & Garbary, D. J. Antioxidants from macroalgae: Potential applications in human health and nutrition. *Algae*, 2010, 25(4), 155-171.
359. Dang, V. T., Speck, P., Doroudi, M., et al. Variation in the antiviral and antibacterial activity of abalone *Haliotis laevigata*, *H. rubra* and their hybrid in South Australia. *Aquaculture*, 2011, 315(3), 242-249.
360. Dang, V.T., Y. Li, P. Speck and K. Benkendorff. Effects of micro and macroalgal diet supplementations on growth and immunity of greenlip abalone, *Haliotis laevigata*. *Aquaculture*. 2011, 320(1-2), 91-98.
361. De Felício, R., de Albuquerque, S., Young, M.C.M., Yokoya, N.S., Deboni, H.M. Trypanocidal, leishmanicidal and antifungal potential from marine red alga *Bostrychia tenella* J. Agardh (Rhodomelaceae, Ceramiales) (2010) *J. Pharm. Biomed. Analysis*, 52 (5), 763-769.
362. de Felício, Rafael, et al. "Trypanocidal, leishmanicidal and antifungal potential from marine red alga *Bostrychia tenella* J. Agardh (Rhodomelaceae, Ceramiales)." *Journal of pharmaceutical and biomedical analysis* 52.5 (2010): 763-769.
363. Devi, G. K., Manivannan, K., & Anantharaman, P. Evaluation of antibacterial potential of seaweeds occurring along the coast of Mandapam, India against human pathogenic bacteria. *Journal of Coastal Life Medicine*, 2014, 2(3), 196-202.
364. Erickson, Amy A., et al. Palatability of macroalgae that use different types of chemical defenses. *J. Chem. Ecol.*, 32.9 (2006): 1883-1895.
365. Felício, Rafael de. Produtos naturais marinhos: identificação de metabólitos fenólicos halogenados na macroalga *Bostrychia tenella* (Rhodomelaceae, Rhodophyta) e potencial biológico de micro-organismos endofíticos associados. Diss. Univers. São Paulo, 2010.
366. Gerasimenko, N. I., et al. Antimicrobial and hemolytic activity of low-molecular metabolites of brown seaweed *Laminaria cichorioides* (Miyabe). *Appl. Biochem. Microbiol.*, 46.4 (2010): 426-430.

367. Gerasimenko, N. I., Martyyas, E. A., & Busarova, N. G. Composition of lipids and biological activity of lipids and photosynthetic pigments from algae of the families Laminariaceae and Alariaceae. *Chem. Nat. Comp.*, 2012, 48(5), 737-741.
368. Gerasimenko, N. I., Martyyas, E. A., Logvinov, S. V., & Busarova, N. G. Biological activity of lipids and photosynthetic pigments of *Sargassum pallidum* C. Agardh. *Applied Biochemistry and Microbiology*, 2014, 50(1), 73-81.
369. Ghazal, Fekry M., Mohamed A. Deyab, and Manal AH El-Gamal. "Allelopathic activity of algal blooms against some plant pathogenic fungi in Egypt." *Egyptian Journal of Phycology* 7.1 (2006): 79-92.
370. Güven, Kasım Cemal, Burak Coban, and Osman Özdemir. *Pharmacology of Marine Macroalgae. Encyclopedia of Marine Biotechnology* 1 (2020): 585-615.
371. Imbs, T. I., et al. Comparative study of the chemical composition of ethanol extracts from brown algae and their effects on seedling growth and productivity of soya *Glycine max* (L.) MERR. *Russian Journal of Bioorganic Chemistry* 37.7 (2011): 871-876.
372. Imbs, T.I., E.L. Chajkina, L.A. Dega, A.P. Vashchenko, M.M. Anisimov. Comparative studying of a chemical compound of ethanol extracts of brown seaweed and their influence on growth of sprouts and productivity of *Glycine max* (l). *Merr. Chemistry of Plant Raw Material*, 2010, 1, 143-148.
373. Kumari, P. (2017). Seaweed lipidomics in the era of 'omics' biology: A contemporary perspective. *Systems Biology of Marine Ecosystems*, 49-97.
374. Kumari, P., et al. *Algal lipids, fatty acids and sterols. Functional ingredients from algae for foods and nutraceuticals.* Woodhead Publishing, 2013. 87-134.
375. Kumari, P.S. *Lipidomics in the Era of 'Omics' Biology: A Contemporary Perspective.* *Systems Biology of Marine Ecosystems.* Springer, Cham, 2017. 49-97.
376. Liu, J., S. Willför, A. Mihranyan. On importance of impurities, potential leachables and extractables in algal nanocellulose for biomedical use. *Carbohydrate Polymers* 172 (2017): 11-19.
377. Mariya, V., & Ravindran, V. S. Biomedical and Pharmacological significance of marine macro algae-review. *Indian J. Geo-Marine Sciences*, 2013, 42(5), 527-537.
378. Martyyas, E. A., Gerasimenko, N. I., Busarova, N. G., et al. Seasonal changes in biological activity of lipids and photosynthetic pigments of *Saccharina cichorioides* (Miyabe) (Laminariaceae Family). *Russian J. Bioorg. Chem.*, 2013, 39(7), 720-727.
379. Michalak, I., B. Messyas. Concise review of *Cladophora* spp.: macroalgae of commercial interest. *J. Appl., Phycol.*, 33.1 (2021): 133-166.
380. Milchakova, N. A. *Marine plants of the Black Sea. An illustrated field guide.* Digit Print, Sevastopol. 2011. 144.
381. Mohamed S, Hashim SN, Rahman HA. Seaweeds: a sustainable functional food for complementary and alternative therapy. *Trends Food Sci. Technol.*, 2012, 23(2), 83-96.
382. Munir, Mubashrah, et al. Pharmaceutical aptitude of *Cladophora*: A comprehensive review. *Algal Research* 39 (2019): 101476.
383. Nasir, Masoumeh, et al. Sterols from the red algae, *Gracilaria salicornia* and *Hypnea flagelliformis*, from Persian Gulf. *Pharmacognosy Magazine* 7.26 (2011): 97.
384. Orhan, I., et al. Turkish freshwater and marine macrophyte extracts show in vitro antiprotozoal activity and inhibit FabI, a key enzyme of *Plasmodium falciparum* fatty acid biosynthesis." *Phytomedicine* 13.6 (2006): 388-393.

385. Paradas, W. C. Mecanismos de armazenamento, biossíntese e liberação de metabólitos secundários em macroalgas vermelhas (Rhodophyta), PhD Thesis, Universidade Federal Fluminense, 2013.
386. Piotrowicz, Z., Tabisz, Ł., Łęska, B., Messyasz, B., Pankiewicz, R. Comparison of the Antioxidant Properties of Green Macroalgae from Diverse European Water Habitats by Use of Several Semi-Quantitative Assays. *Molecules*, 2022, 27(12), 3812.
387. Prazukin, A.V., E.V. Anufrieva, N.V. Shadrin. Is biomass of filamentous green algae *Cladophora* spp.(Chlorophyta, Ulvophyceae) an unlimited cheap and valuable resource for medicine and pharmacology? A review. *Rev. Aquacult.*, 12.4 (2020): 2493-2510.
388. Rickert, E., et al. Seasonal variations in surface metabolite composition of *Fucus vesiculosus* and *Fucus serratus* from the Baltic Sea. *PLoS One* 11.12 (2016): e0168196.
389. Rosales, Alan Rodrigo López, and Maestro En Ciencias En Energía Renovable. "Potencial de cepas de microalgas aisladas de la costa de Yucatán para la producción de biodiesel." (2017).
390. Saranya, C., Parthiban, C., & Anantharaman, P. Evaluation of antibacterial and antioxidant activities of seaweeds from Pondicherry coast. *Adv. Appl. Sci. Res.*, 2014, 5, 4, 82-90.
391. Snegireva, A. V., Shchetinin, M. P., & Meleshkina, L. E. (2021, October). Chickpea sprouting as a way to increase the garnish nutritional value. In *AIP Conference Proceedings* (Vol. 2419, No. 1). AIP Publishing.
392. Spavieri, J. et al. Antiprotozoal, antimycobacterial and cytotoxic potential of some British green algae. *Phytotherapy Research* 24.7 (2010): 1095-1098.
393. Stabili, L., et al. Biotechnological potential of the seaweed *Cladophora rupestris* (Chlorophyta, Cladophorales) lipidic extract. *New biotechnology* 31.5 (2014): 436-444.
394. Suchý, Václav, et al. Relict Pleistocene calcareous tufa of the Chlupáčova sluj Cave, the Bohemian Karst, Czech Republic: A petrographic and geochemical record of hydrologically-driven cave evolution. *Sedimentary Geology* 385 (2019): 110-125.
395. Suryanarayanan, T.S. Fungal endosymbionts of seaweeds. *Biol. Mar. Fungi*, 2012, 53, 53-69.
396. Tabakaeva, O. V., A. V. Tabakaev. "Amino acids from potentially commercial Far-East brown algae *Costaria costata* and *Undaria pinnatifida*." *Chem. Nat. Comp.*, 52.2 (2016): 376-378.
397. Tan, S. P., O'Sullivan, L., Prieto, M.L., et al. 13 Seaweed antimicrobials: isolation, characterization, and potential use in functional foods. *Bioactive Compounds from Marine Foods: Plant Anim. Sourc.*, 2013, 269.
398. Todorov, D., Hinkov, A., Shishkova, K., & Shishkov, S. Antiviral potential of Bulgarian medicinal plants. *Phytochemistry Reviews*, 2014, 13(2), 525-538.
399. Tolpeznikaite, E., Bartkevics, V., Ruzauskas, M., Pilkaityte, R., Viskelis, P., Urbonaviciene, D., ... & Bartkiene, E. (2021). Characterization of macro-and microalgae extracts bioactive compounds and micro-and macroelements transition from algae to extract. *Foods*, 10(9), 2226.
400. Van Doan, H., Doolgindachbaporn, S., Suksri, A. Effects of low molecular weight agar and *Lactobacillus plantarum* on growth performance, immunity, and disease resistance of basa fish (*Pangasius bocourti*, Sauvage 1880). *Fish & Shellfish Immunology*, 2014, 41, 2, 340–34

401. Анисимов, М. М., et al. "Противомикробная активность экстрактов и компонентов морских водорослей." Растительные ресурсы 48.1 (2012): 139-154.
402. Анисимов, М. М., Мартыяс, Е. А., Чайкина, Е. Л., & Герасименко, Н. И. Противомикробная, гемолитическая и фиторегулирующая активность липидных экстрактов из морских водорослей. Химия растительного сырья, 2010, 4, 125-130.
403. Герасименко, Н. И., Чайкина, Е. Л., Бусарова, Н. Г., & Анисимов, М. М. (2010). Противомикробная и гемолитическая активности низкомолекулярных метаболитов бурой водоросли *Laminaria sichorioides* Miyabe. Прикладная биохимия и микробиология, 46(4), 467-471.
404. Имбс, Т. И., Чайкина, Е. Л., Дега, Л. А., Ващенко, А. П., & Анисимов, М. М. (2010). Сравнительное изучение химического состава этанольных экстрактов бурых водорослей и их влияния на рост проростков и урожайность сои *glycine max* (L.) Merr. Химия растительного сырья, (1), 143-148.
405. Мартыяс, Е. А., Герасименко, Н. И., Бусарова, Н. Г., и сътр. Биологическая активность липидов и фотосинтетических пигментов *Saccharina sichorioides* (Miyabe)(сем. *Laminariaceae*). Сезонные изменения активности. Химия растительного сырья, 2012, 1, 123-131.
406. Солоненко, Анатолій Миколайович. Водорості гіпергалійних водойм північно-західного узбережжя Азовського моря та їх участь в утворенні мулових сульфідних пелоїдів: дис. доктора біологічних наук: 03.00. 05. Diss. Міністерство освіти і науки України, 2015.
407. Табакаева, О. В., & Табакаев, А. В. (2016). Биологически активные вещества потенциально промысловых бурых водорослей Дальневосточного региона. Вопросы питания, 85(3), 126-132.
408. Федянина, Л. Н., В. А. Лях, and Е. С. Смертина. Оценка эффективности профилактического действия хлеба с добавлением экстракта бурых водорослей. Вестник Красноярского государственного аграрного университета 5 (140) (2018).
- 10). Orozova, P., Chikova, V., NAJDENSKI, H. Antibiotic resistance of pathogenic for fish isolates of *Aeromonas* spp. 16, 3, Bulgarian Journal of Agricultural Science, 2010, 376-386. ISI IF:0.26**
409. Adeyemi, J.A., K. O. Ogunlowo, O. O. Oyedara. Bacterial Microflora in the Gut, Gill and Skin of African Catfish, *Clarias gariepinus* (Burchell, 1822) collected from Earthen Ponds in Oke-Baale, Osogbo, Nigeria. NISEB Journal 13.3&4 (2019).
410. Ali, H., et al. Putative virulence and antimicrobial susceptibility of *Aeromonas* spp. isolated from marketed fish intended for human consumption in Bangladesh. Asian J. Microbiol. Biotech. Env 16.485 (2014): e495.
411. Ali, S. et al. Identification, characterization and antibiotic sensitivity of *aeromonas hydrophila*, a causative agent of epizootic ulcerative syndrome in wild and farmed fish from potohar, Pakistan. Pakistan J. Zool 48.3 (2016): 899-901.
412. Boumerdassi, H., Djouadi, L. N., Ouzari, H. I., & Nateche, F. (2021, June). Isolation and Characterization of Ichthyopathogenic Bacteria from a Northwestern Algerian Dam. In Euro-Mediterranean Conference for Environmental Integration (pp. 719-722). Cham: Springer Nature Switzerland.
413. Carvalho, Daiane, et al. "Isolation and antimicrobial susceptibility of bacteria from

- injuries caused by ictio in jundia (*Rhamdia quelen*)." B. Inst. Pesca (2016): 195-202.
414. Chethurajupalli, L., Tambireddy, N., Ramana, T. V., Dasari, M., Boda, S., Narra, M. S., ...& Bheemeswararao, K. (2023). Detection of virulence genes, antimicrobial susceptibility and pathogenicity of *Aeromonas veronii* isolates from *Labeo rohita* and *Catla catla*. *Indian J. Animal Res.*, 57(12), 1707-1716.
 415. Dash, J.P, L. Mani, S.K. Nayak. Antibacterial activity of *Blumea axillaris* synthesized selenium nanoparticles against multidrug resistant pathogens of aquatic origin. *Egyptian Journal of Basic and Applied Sciences* 9.1 (2022): 65-76.
 416. Dinçtürk, E. (2019). Alabalıklarda bakteriyel balık patojenlerinin teşhisinde yeni izolasyon ve identifikasyon yöntemlerinin uygulanması (Doctoral dissertation, İzmir Katip Celebi University (Turkey)).
 417. El-Barbary, M.I. Serum biochemical and histopathological changes associated with *Aeromonas hydrophila* isolated from *Oreochromis niloticus* and *Sparus aurata* with multiple antibiotic resistance index. *J. Biol. Sci* 17 (2017): 222-234.
 418. Guz, L., Nowakiewicz, A., Puk, K., Zięba, P., Gnat, S., & Matuszewski, Ł. (2021). Virulence and Antimicrobial Resistance Pattern of *Aeromonas* spp. Colonizing European Pond Turtles *Emys orbicularis* and Their Natural Environment. First Study from Poland. *Animals*, 11(10), 2772.
 419. Hamouda, A.H., E.M. Moustafa, and M.M. Zayed. Overview on the most prevailing bacterial diseases infecting *oreochromis niloticus* at aswan fish hatchery, egypt. *Adv. Anim. Vet. Sci* 7.11 (2019): 950-961.
 420. Ibryamova, S., Toschkova, S., Pavlova, B., Stanachkova, E., Ivanov, R., Natchev, N., ... & Ignatova-Ivanova, T. (2022). A study of the microbiology of the intestinal tract in different species of Teleost fish from the Black Sea. *BioRisk*, 18, 105-113.
 421. Ictio, O.D.L.C. P., & Jundia, E. (2016). Isolamento e suscetibilidade antimicrobiana de bactérias. *Bol. Inst. Pesca, São Paulo*, 42(1), 195-202.
 422. Igbinosa, I.H., E.O. Igbinosa, A.I. Okoh. Antibigram characterization and putative virulence genes in *Aeromonas* species isolated from pig fecal samples. *Environm. Sci. Pollut. Res.*, 23.12 (2016): 12199-12205.
 423. Igbinosa, I.H. E.O. Igbinosa, A. I. Okoh. Detection of antibiotic resistance, virulence gene determinants and biofilm formation in *Aeromonas* species isolated from cattle. *Environ Sci Pollut Res.*, DOI 10.1007/s11356-015-4934-4.
 424. Islam, M. S., Hossain, M. M. M., Neowajh, M. S., Kholil, M. I., Khatun, A., Rikta, S., & Imteazzaman, A. M. (2015). Antimicrobial resistance of *Aeromonas* spp. isolated from the carp farm following Streptomycin treatment. *Int. J. Fish. Aquatic Studies*, 2, 99-102.
 425. Kanchan, C., et al. Antibiotic resistance of *Aeromonas hydrophila* isolated from diseased catfish. The 6th International conference on sciences and social sciences: mutual community engagement toward global understanding and sustainable well-being. 2016.
 426. Korun, J. Kültür Levrek (*Dicentrarchus labrax*, L.) Balıklarından İzole Edilen *Vibrio harveyi* Suşlarının Antimikrobiyal Duyarlılıkları Üzerine Bir Çalışma. *Nevşehir Bilim ve Teknoloji Dergisi* 5 (2016): 411-421.
 427. Lavanya, C., Neeraja, T. and Ramana, T.V., 2023. Antimicrobial resistance study of pathogenic *Aeromonas* spp. isolated from freshwater fish, *Catla catla* of Andhra Pradesh, India: a major threat to public health. *J. Exp. Zool. India*, 26(2).
 428. Lavanya, C., Neeraja, T., Babu, P. H., Ramana, T. V., Balasubramanian, A., Meshram,

- S., & Sruthi, P. (2021). Antimicrobial resistance of *Aeromonas* species isolated from cultured Indian major carp, *Labeo rohita*: Possible public health concern. *International Journal of Bio-resource and Stress Management*, 12(5), 506-515.
429. Majolo, Cláudia, et al. Antimicrobial activity of some essential oils against *Streptococcus agalactiae*, an important pathogen for fish farming in Brazil. *J. Essent. Oil Res.*, 30.5 (2018): 388-397.
 430. Majolo, Claudia, et al. Chemical composition of *Lippia* spp. essential oil and antimicrobial activity against *Aeromonas hydrophila*. *Aquacult. Res.*, 48.5, 2017, 2380-2387.
 431. Majolo, Cláudia, et al. Essential Oils from Five Brazilian Piper Species as Antimicrobials Against Strains of *Aeromonas hydrophila*. *J. Essent. Oil Bearing Plants* 22.3 (2019): 746-761.
 432. Nurrahmad, N.R. Pengaruh vaksinasi outer membrane protein 52 kda *Aeromonas hydrophila* terhadap perubahan indeks eritrosit ikan nila (*Oreochromis niloticus*) yang di tantang *Aeromonas hydrophila*. Diss. Universitas airangga, 2019.
 433. Nurrahmad, N. R., Yuliani, M. G. A., Ernawati, R., Chusniati, S., Herupradoto, E. B. A., & Wahyuni, R. S. (2021). Pengaruh Vaksinasi Outer Membran Protein 52 kDa Terhadap Perubahan Indeks Eritrosit Ikan Nila (*Oreochromis niloticus*) Yang Diinfeksi *Aeromonas hydrophila* Effect of Vaccination Outer Membrane Protein 52 kDa on Changes in Erythrocyte Index of Tilapia (*Oreochromis niloticus*) Infected by *Aeromonas hydrophila*. *Journal of Basic Medical Veterinary*, 10(1), 7-14.
 434. Odeyemi, O.A. (2013). Seafood bacterial pathogens and sustainability of aquaculture. *Asian J. Microbiol. Biotechnol. Environ. Sci*, 15(4), 1-5.
 435. Pereira, Carla, et al. Bacteriophages in the Control of *Aeromonas* sp. in Aquaculture Systems: An Integrative View. *Antibiotics* 11.2 (2022): 163.
 436. Roy, R.P. Investigation of pathogenic bacteria of a resident fish, *Lepidocephalichthys guntea* (Hamilton Buchanan), in relation to limnochemistry of a Terai river Lotchka in the Darjeeling foothills of West Bengal, India. Diss. University of North Bengal, 2014.
 437. Shahzad Ali, S. A., Shamim Akhter, S. A., Ali Muhammad, A. M., Iahtasham Khan, I. K., Khan, W. A., Iqbal, M. N., ... & Qurban Ali, Q. A. (2016). Identification, characterization and antibiotic sensitivity of *Aeromonas hydrophila*, a causative agent of epizootic ulcerative syndrome in wild and farmed fish from Potohar, Pakistan. *Pakistan Journal of Zoology*, 2016, 48, 3, 899-901.
 438. Sharma, A., et al. *Asparagus racemosus* aqueous root extract induced effects on cellular immune reaction of *Labeo rohita* (Hamilton). *Indian J. Animal Sci.*, 88.2 (2018): 251-258.
 439. Shulgina, L.V., Yakush, E.V., Shulgin, Y.P., Shenderyuk, V.V., Chukalova, N.N., & Baholdina, L.P. (2015). Antibiotics in aquaculture and their ecological significance. A review. *Izvestiya TINRO*, 181(2), 216-230.
 440. Srividya, R. (2020). Surveillance on bacterial diseases of cultured indian major carps in Andhra Pradesh, India (Doctoral dissertation, Sri Venkateswara Veterinary University Tirupati-517 502.(AP) India).
 441. Sruthi, P., Neeraja, T., Balasubramanian, A., Meshram, S., Prasad, G. S., Lavanya, C., ... & Singh, S. M. (2021). Phenotypic Characterization and Antimicrobial Resistance of *Aeromonas* Species Isolated from *Catla Catla* and *Labeo Rohita* Collected in Two

- Districts of Andhra Pradesh, India. J. Exp. Zool. India, 24(2).
442. Stojanov, I., Radulović, J. P., Kapetanov, M., & Petrović, J. (2015). Potential spread of antimicrobial resistance via drinking water in livestock husbandry. "One Health - New Challenges", First International Symposium of Veterinary Medicine (ISVM2015), 21-23 May 2015, Vrdnik, Serbia. Proceedings, 2015, 446-453 ref. 30.
 443. Zdanowicz, Marta, Zbigniew Jan Mudryk, and Piotr Perliński. Abundance and antibiotic resistance of *Aeromonas* isolated from the water of three carp ponds. Vet. Res. Commun., 44.1 (2020): 9-18.
 444. Улитко, В. Е. Продуктивные и биологические показатели карпа при использовании в комбикорме сорбционно-пробиотической добавки с эфирными маслами" биокоретрон. Диссертация, Ульяновск, 2018.
 445. Романова, Н. Н., Мышкин, А. В., Щелкунова, Ю. П., Токарева, С. Б., Сехина, О. В., & Кудинов, П. В. (2023). Оценка эффективности применения комплекса антибактериальных препаратов для лечения бактериальной геморрагической септицемии у рыб. Труды ВНИРО, 194, 165-175.
 446. Шульгина, Лидия Васильевна, et al. Антибиотики в объектах аквакультуры и их экологическая значимость. Обзор. Известия ТИНРО (Тихоокеанского научно-исследовательского рыбохозяйственного центра) 181 (2015).
- 11). Kostadinova, E., K. Alipieva, M. Stefova, D. Antonova, L. Evstatieva, G. Stefkov, I. Tsvetkova, H. NAYDENSKI, V. Bankova. Influence of cultivation on the chemical composition and antimicrobial activity of *Sideritis* spp. Phcog Mag. 4(14), 102 – 106 (2008)**
447. Adejumo, O., Williams, I., Ojewale, K., Igbokwe, C., & Ajayi, I. Phytochemistry and In vitro Antimicrobial Activity of Five Plant Species against Nine Common Human Pathogens. Dhaka Univ. J. Pharm. Sci. 20(2): 139-148, 2021.
 448. Ahmed, K. M. New challenges in the new year for Phcog Mag.: 5 years of quality publication. Pharmacognosy Mag., 2011, 7(25), 1.
 449. Aneva, I., Zhelev, P., Evstatieva, L., & Dimitrov, D. (2013). The ecological and floristic characteristics of populations of *Sideritis scardica* Griseb. in Slavyanka mountain. Bulg. J. Agricult. Sci., 19(2), 211-217.
 450. Aneva, I., Zhelev, P., Kozuharova, E., Danova, K., Nabavi, S. F., & Behzad, S. (2019). Genus *Sideritis*, section *Empedoclia* in southeastern Europe and Turkey—studies in ethnopharmacology and recent progress of biological activities. DARU J. Pharm. Sci., 27(1), 407-421.
 451. Antonina, V. A., Ljuba, E. N., & Dimitar, P. R. (2012). In situ and ex situ conservation of rare high-mountain medicinal plants in Bulgaria. Proceedings of the Seventh Conference on Medicinal and Aromatic Plants of Southeast European Countries, (Proceedings of the 7th CMAPSEEC), Subotica, Serbia, 27-31 May, 2012, 14-22 ref. 30
 452. Axiotis, Evangelos, et al. "Phytochemical Profile and Biological Activity of Endemic *Sideritis sipylea* Boiss. in North Aegean Greek Islands. Molecules 2020, 25.9:2022.
 453. Azizullah, Hasina. Isolering og karakterisering av polyfenoler fra gresk fjellte, *Sideritis scardica*. MS thesis. 2017.
 454. Balli-Neslihan, T.B.S.M., Genc D.T. *Sideritis trojana* Bornm. ve *Sideritis athoa* Papan. & Kokkini Metanol Ekstraktlarının Antimikrobiyal Aktivitelerinin Belirlenmesi. In

Proceedings & Abstracts. 2013, p. 65.

455. Cerdáa, X., van Oudenhovea, L., Bernsteinb, C., & Boulaya, R. R. (2014). A List of and Some Comments about the Trail Pheromones of Ants. NPC Natural Product Communications, 9, 8, 1115-1122.
456. Coğuplugil, A. (2019). Ratlarda Sideritis akmaniiden elde edilen ekstraktların lipid profili ve bazı biyokimyasal parametreler üzerine etkisinin araştırılması (Master's thesis, Sağlık Bilimleri Enstitüsü).
457. Ganos, C. G., Gortzi, O., Chinou, E. B., Calapai, G., & Chinou, I. B. (2022). Antimicrobial properties of selected native greek aromatic plants: An ethnopharmacological overview. Promising antimicrobials from natural products, 91-117.
458. González-Burgos, E., Carretero, M. E., & Gómez-Serranillos, M. P. Sideritis spp.: uses, chemical composition and pharmacological activities—a review. J. Ethnopharmacol., 2011, 135(2), 209-225.
459. Heiner, F., Feistel, B., J. Reichling. Sideritis scardica Griseb. DrugBase, 2018, 1-6.
460. Ibraliu, A., Trendafilova, A., Anđelković, B., Qazimi, B., Gođevac, D., Bebeci, E., ... & Papajani-Toska, V. (2015). Comparative Study of Balkan Sideritis Species from Albania, Bulgaria and Macedonia. European Journal of Medicinal Plants, 5(4), 328-340.
461. Ignatov, I. Sideritis scardica Griseb.(Mursalski tea; pirinski tea) from Bulgaria, which is growing in zones with high percent of long living people. Plant Cell Biotechnol. Mol. Biol. (2021): 141-153
462. Iordache, A.M., M. Culea, O. Cozar. Characterization of some extracts for therapeutic use by GC/MS. Processes in Isotopes and Molecules, Journal of Physics: Conference Series (vol.182, 1 (2009) p. 012027 doi:10.1088/1742-6596/182/1/012027
463. Karapandzova, M., Qazimi, B., Stefkov, G., et al. Chemical characterization, mineral content and radical scavenging activity of *Sideritis scardica* and *S. raeseri* from R. Macedonia and R. Albania. Nat. Prod. Communicat., 5, 8, 2013, 639-644.
464. Krgović, N., Jovanović, M., Aradski, A. A., Janković, T., Stević, T., Zdunić, G., ... & Šavikin, K. (2022). Bioassay-guided skin-beneficial effects of fractionated *Sideritis raeseri* subsp. *raeseri* extract. Plants, 11(20), 2677.
465. Menković N., Godevac D., Šavikin K., Zdunić G., Milosavljević S., Bojadži A., Avramoski O. Bioactive compounds of endemic species *Sideritis raeseri* subsp. *raeseri* grown in National Park Gal. Rec. Nat. Prod., 3, 7, 2013, 161-168.
466. Negoşanu, G. A. Asanica, C. Vîñătoru, B. Muşat, C. Bratu, M. Popescu. Research on the phenotypic and biochemical expression of *Sideritis scardica* cultivars acclimated and bred at the Plant Genetic Resources Bank - for Vegetables, Floricultural, Aromatic and Medicinal Plants, Buzău, Romania. December 2023. Acta Horticulturae, 2023, (1384), 427-434.
467. Pljevljakusic, D., K. Savikin, T. Jankovic et al. Chemical properties of the cultivated *Sideritis raeseri* Boiss. & Heldr. subsp *raeseri*. Food Chem., 2011, 124, 1, 226-233.
468. Provopoulou, K. D. (2022). Effect of temperature and boiling time on the composition of polyphenols and antioxidant power of green tea and mountain tea extracts (Master's thesis).
469. Qazimi, B., Stafilov, T., Bačeva-Andonovska, K., Tašev, K., Dragusha, S., Koraqi, H., ... & Ejupi, V. Comparison of mineral content between spontaneous and cultivated flowering stems of *Sideritis scardica*. Macedonian Pharmaceutical Bulletin, 68 (Suppl 2)

77 - 78 (2022).

470. Qazimi B., Gjoshe Stefkov, M. Karapandzova, I. Cvetkovikj, S. Kulevanova Volatile aroma compounds in infusions of stems and rosette leaves of *Sideritis raeseri* Boiss. & Heldr. from R. Macedonia, Albania and Greece. *Macedonian Pharmaceutical Bulletin*, 60 (1) 27 - 33 (2014).
471. Qazimi, B., Stefkov, G., Karapandzova, M., Cvetkovikj, I., & Kulevanova, S. (2014). Aroma compounds of mountain tea (*Sideritis scardica* and *S. raeseri*) from western Balkan. *Nat. Prod. Communicat.*, 9(9), <https://doi.org/10.1177/1934578X1400900937>.
472. Shtereva, L. A., R.D. Vassilevska-Ivanova, B.V. Kraptchev. In vitro cultures for micropropagation, mass multiplication and preservation of an endangered medicinal plant *Sideritis scardica* Griseb. *Botanica Serbica* 39.2 (2015).
473. Siafis, K. S. (2017). Application of irrigation to mountain tea (*Sideritis raeseri*) (2nd Cultivation season) (Master's thesis).
474. Solomou, A. D., Skoufogianni, E., Mylonas, C., Germani, R., & Danalatos, N. G. (2019) Cultivation and utilization of "Greek mountain tea" (*Sideritis* spp.): Current knowledge and future challenges. *AJAB. Asian J Agric & Biol*, 7(2), 289-299.
475. Stoumbou, S.X., 2023. Comparative Study of the Bioactivity and Chemical Composition of Plant Species of the *Sideritis* Genus.
476. Tadić, Vanja, et al. Chemical and antimicrobial analyses of *Sideritis romana* L. subsp. *purpurea* (Tal. ex Benth.) Heywood, an endemic of the Western Balkan. *Molecules* 22.9 (2017): 1395.
477. Todorova, M., & Trendafilova, A. *Sideritis scardica* Griseb., an endemic species of Balkan peninsula: Traditional uses, cultivation, chemical composition, biological activity. *J. Ethnopharmacol.*, 2014, 152(2), 256-265.
478. Tsibranska, I., B. Tytkowski, R. Kochanov, et al. Extraction of biologically active compounds from *Sideritis* ssp. L. *Food Bioprod. Proces.*, 2011, 89(4), 273-280.
479. Veli, A., Nikolova, R., Mustafa, Z., Rusev, G., & Gonsalvesh, L. (2023). Antioxidant Activity and Chemical Composition of Extracts from an Endemic Plant *Sideritis syriaca* 2. *Proceedings of University of Ruse*, 62.
480. Yurukova-Grancarova, D. P., & Yankova-Tsvetkova, P. E. (2012). On the embryology of *Sideritis scardica* Griseb.(Lamiaceae). *Proceedings of the 7th Conference on Medicinal and Aromatic Plants of Southeast European Countries*, (Proceedings of the 7th CMAPSEEC), Subotica, Serbia, 27-31 May, 2012, 2012, 34-39 ref. 21.
481. Zehiroğlu, C. (2017). Türkiye'de endemik olarak yetişen Kaz dağı çayının (*Sideritis trojana* bornm.) antioksidan, antimikrobiyal aktivitelerinin ve mineral içeriğinin araştırılması (Master's thesis, Fen Bilimleri Enstitüsü).
- 12). Georgiev, L., Chochkova, M., Ivanova, G., NAJDENSKI, H., Ninova, M., Milkova, T. Radical scavenging and antimicrobial activities of cinnamoyl amides of biogenic monoamines (2012) *Rivista Italiana delle Sostanze Grasse*, 89 (2), pp. 91-102.**
482. Ahlawat, Shruti, et al. "Bioevaluation and molecular docking analysis of novel phenylpropanoid derivatives as potent food preservative and anti-microbials." *3 Biotech* 11.2 (2021): 1-10.
483. Almoulah, N. Fadl, et al. Antibacterial, antiproliferative and antioxidant activity of leaf extracts of selected Solanaceae species. *South African J. Botany* 112 (2017): 368-374.

484. Aouadi, Kaïss, et al. Phytochemical Profiling, Antimicrobial and α -Glucosidase Inhibitory Potential of Phenolic-Enriched Extracts of the Aerial Parts from *Echium humile* Desf.: In Vitro Combined with In Silico Approach. *Plants* 11.9 (2022): 1131.
485. Andrade, M., Benfeito, S., Soares, P. et al. Fine-tuning of the hydrophobicity of caffeic acid: studies on the antimicrobial activity against *Staphylococcus aureus* and *Escherichia coli*. *RSC Advances* 2015, 5, 53915-53925.
486. Bułakowska A., Sławiński J., Hałasa R., Hering A., Gucwa M., Ochocka J.R., Stefanowicz-Hajduk J. An In Vitro Antimicrobial, Anticancer and Antioxidant Activity of N-[(2-Arylmethylthio)phenylsulfonyl]cinnamamide Derivatives. *Molecules*, 28(7), 2023.
487. Cao, P., Yang, J., Xia, L., Zhang, Z., Wu, Z., Hao, Y., ... & Wang, S. (2024). Two gene clusters and their positive regulator SIMYB13 that have undergone domestication-associated negative selection control phenolamide accumulation and drought tolerance in tomato. *Molecular Plant*, 17(4), 579-597.
488. Cho, J.-G., Huh, J., Jeong, R.-H., Cha, B.-J., Shrestha, S., Lee, D.-G., Kang, H.-C., Kim, J.-Y., Baek, N.-I Inhibition effect of phenyl compounds from the *Oryza sativa* roots on melanin production in murine B16-F10 melanoma cells. *Nat. Product Res.*, 2015, 29, 1052-1054.
489. Cho, M.H., & Lee, S.W. (2015). Phenolic Phytoalexins in Rice: Biological Functions and Biosynthesis. *Int. J. Mol. Sci.*, 16(12), 29120-29133.
490. Dastmalchi, Keyvan, et al. Temporal resistance of potato tubers: Antibacterial assays and metabolite profiling of wound-healing tissue extracts from contrasting cultivars. *Phytochemistry* 159 (2019): 75-89.
491. Firdaus, Soekamto NH, et al. Novel hydroxycinnamamide from morpholine and pyrrolidine: Synthesis, characterization, docking study, and anticancer activity against P388 leukemia murine cells. *J. Appl. Pharm. Sci.*, 11.01 (2021): 040-048.
492. Firdaus, F., Soekamto, N. H., Firdausiah, S., & Tahar, M. (2020). A Caffeic Acid Derivative Potential for Anticancer Drug: Synthesis of N-(piperidiny)l caffeamide and Its Activity against P388 Leukemia Murine Cells. *Jurnal Akta Kimia Indonesia (Indonesia Chimica Acta)*, 38-42.
493. França, Saraliny Bezerra, et al. Synthesis, applications and Structure-Activity Relationship (SAR) of cinnamic acid derivatives: a review. *Research, Society and Development* 10.1: e28010111691-e28010111691.
494. Fusini, G. (2015). Individuation and study of supported catalytic systems suitable for practically useful CC cross-coupling reactions performed in batch and/or flow conditions. Ph.D. Thesis in Chemical and Material Science, University of Pisa.
495. Guzman, J.D. Natural cinnamic acids, synthetic derivatives and hybrids with antimicrobial activity *Molecules* 2014, 19, 12, 19292-19349.
496. Machado, I. M. (2018). Synthesis of catecholic amino acids and peptides or conjugates of catechols with amino acids and peptides (Doctoral dissertation, Universidade do Minho (Portugal)).
497. Manaois, R.V., Zapater, J.E I., Morales, A.V., J.C.A. Cacerez. Phytochemical content, antioxidant capacity, and in vitro antibacterial activity of bran extracts of Philippine rice (*Oryza sativa*) cultivars. *SciEnggJ*, 2024, 17, 01, 8-17
498. Monteiro, L. S., et al. Synthesis and preliminary biological evaluation of new phenolic and catecholic dehydroamino acid derivatives. *Tetrahedron* 73.43 (2017):6199-6209.

499. Monteiro, L. S., & Paiva-Martins, F. (2022). Amino acids, amino acid derivatives and peptides as antioxidants. In *Lipid Oxidation in Food and Biological Systems: A Physical Chemistry Perspective* (pp. 381-404). Cham: Springer International Publishing.
500. Montes, R.C. Halogenated amides: coupling reactions and in silico investigation of antimicrobial activity. (2016). PhD thesis, State University of Paraiba.
501. Neelam, Ahlawat, S., Shankar, A., Lather, A., Khatkar, A., & Sharma, K. K. (2021). Bioevaluation and molecular docking analysis of novel phenylpropanoid derivatives as potent food preservative and anti-microbials. *3 Biotech*, 11, 1-10.
502. Nurlaida, N., Saadatul, H., & Aswar, A. (2023). Synthesis Optimization and Characterization of κ -Carrageenans from *Eucheuma cottonii*. *Jurnal Akta Kimia Indonesia (Indonesia Chimica Acta)*, 13-16.
503. Paramita, S. (2019). Synthesis of the compound 3-(3, 4-dihydroxyphenyl)-n-otolylacrylamide from 3-(3, 4-dihydroxyphenyl) acrylate and test of its bioactivity on HeLa cells (Doctoral dissertation, Hasanuddin University.).
504. Qi, Yanting, et al. Safety evaluation of FAD2 RNAi transgenic *Brassica napus* L. based on microbial diversity and metabonomic analysis. *Frontiers Plant Sci.*, 13 (2022).
505. Roumani, M. et al. Phenolamides in plants: An update on their function, regulation, and origin of their biosynthetic enzymes. *J. Exp. Botany* 72.7 (2021): 2334-2355.
506. Roumani, Marwa, et al. Phenolamides: Plant specialized metabolites with a wide range of promising pharmacological and health-promoting interests. *Biomed. & Pharmacother.* 131 (2020): 110762.
507. Santos, V. R. D. (2022). Antimicrobial and anti-inflammatory effect of phenolic acids isolated, combined and incorporated in chitosan hydrogen for endodontic application. PhD thesis – Paulista State University, Faculty of Dentistry of Araçatuba.
508. Silveira, Graziela Rangel, et al. In vitro anti-Toxoplasma gondii and antimicrobial activity of amides derived from cinnamic acid. *Molecules* 23.4 (2018): 774.
509. Soekamto, N. H., Firdausiah, S., Rasyid, H., Asmi, N., & Waelulu, M. (2021). Novel hydroxycinnamamide from morpholine and pyrrolidine: Synthesis, characterization, docking study, and anticancer activity against P388 leukemia murine cells. *J. Appl. Pharm. Sci.*, 11(1), 040-048.
510. Sun, J., Song, Y.-L. Zhang, J., Huang, Z., Huo, H.-X., Zheng, J., Zhang, Q., Zhao, Y.-F., Li, J., Tu, P.-F. Characterization and Quantitative Analysis of Phenylpropanoid Amides in Eggplant (*Solanum melongena* L.) by High Performance Liquid Chromatography Coupled with Diode Array Detection and Hybrid Ion Trap Time-of-Flight Mass Spectrometry. *J. Agricult.Food Chem.*, 2015, 63, 3426-3436.
511. Wei, G., Y. Liu, L. Huihui, L. Changqian, L. Shoujin, & H. Jiangmiao. (2015). Study on the chemical composition of *Dendrobium cylindrica*. *Modern Chinese Medicine*, (4), 311-314.
512. Yunjia, G., J. Yong, D. Fang, H. Zhiling, L. Haiyue, B. Zhong, ... & T. Pengfei. (2015). Research on the medicinal properties of *Cistanche deserticola* as a laxative. *Chinese Modern Medicine*, (4), 307-310.
513. Zenta, Firdaus, et al. Development trans-N-benzyl hydroxyl cinnamamide based compounds from cinnamic acids and characteristics anticancer potency. *J. Iranian Chem. Society* (2022): 1-9.

13). Bonovska, M., Tzvetkov, Y., NAJDENSKI, H., Bachvarova, Y. PCR for detection of

Mycobacterium tuberculosis in experimentally infected dogs (2005) Journal of Veterinary Medicine Series B: Infectious Diseases and Veterinary Public Health, 52 (4), pp. 165-170.

514. Al-Galebi, Ahlam Ali Soghi, Mithal Kareem Abass Al-Hassani, and Ebtesam Thamer Jeaz. Serodetection and Molecular Confirmation of *Mycobacterium tuberculosis* Infections in Dogs. J. Educ. Pure Science-University of Thi-Qar, 7.3 (2017): 109-119.
515. Atasever, A. and Yağlı, K., 2023. Tuberculosis in Domestic Animals. Cumhuriyet University Health Sciences Institute Journal, 8(2), 271-279.
516. Bagińska, M., & Rzewuska, M. (2010). Occurrence of mycobacteria from the *Mycobacterium tuberculosis* complex in animals - transmission of selected species between humans and animals. Veterinary Life, 85(09).
517. Barker, E. N., O'Halloran, C., & Gunn-Moore, D. A. (2024). Review Canine Tuberculosis—An Emerging Concern. The Veterinary Journal, 106111.
518. Barta, J. L. (2008). Man's Best Friend: Implications of Tuberculosis in a 16th Century Neutral Iroquois Dog from Canada. Multiplying and Dividing, 22.
519. Barua, Acheenta G., et al. Detection of *Mycobacterium tuberculosis* in Dog of Assam. Int. J. Curr. Microbiol. App. Sci 8.5 (2019): 1283-1288.
520. Cho, H.-S., Choi, U.-S., Oh, Y. Isolation of multidrug-resistant (MDR) *Mycobacterium bovis* from a dog in Korea. J. Vet. Med. Sci., 2022, 84(10), 1358-1362.
521. Deppenmeier, S., Schieszler, A., Nolte, I., Moser, I., Hewicker-Trautwein, M. Pulmonary tuberculosis with evidence of *Mycobacterium tuberculosis* in a Golden Retriever (2007) Tierärztliche Praxis Ausgabe K: Kleintiere - Heimtiere, 35 (2), 111-115.
522. Ghielmetti, Giovanni, and Urs Giger. *Mycobacterium avium*: an emerging pathogen for dog breeds with hereditary immunodeficiencies. Curr. Clin. Microbiol. Reports (2020):1-14.
523. Gong, Wenping, et al. An alert of *Mycobacterium tuberculosis* infection of rhesus macaques in a wild zoo in China. Experimental Animals 66.4 (2017): 357-365.
524. Greene, Craig E. Infectious diseases of the dog and cat. Vol. 310. Philadelphia: WB Saunders, 1998.
525. Hassan, N., Randhawa, C. S., & Zadon, S. (2005). *Mycobacterium bovis* Infection in an adult dog. Veterinarni Medicina, 50, 291-299.
526. Haydock, L.A.J., Abrams-Ogg, A.C.G., Weese, J.S., et al. Diagnostic and public health investigation of *Mycobacterium tuberculosis* infection in a dog in Ontario, Canada. J. Vet. Diagn. Invest., 2022, 34(2), 292-297.
527. Huchzermeyer, F. W., Groenewald, H. B., Myburgh, J. G., Steyl, J. C., & Crole, M. R. (2013). Osteoarthropathy of unknown aetiology in the long bones of farmed and wild Nile crocodiles (*Crocodylus niloticus*). J. South African Vet. Assoc., 84(1), 1-5.
528. Kim, S. (2022). Elucidation of canine immune response to *Mycobacterium avium* complex infection (Doctoral dissertation, Seoul National University Graduate School).
529. Krajewska-Wedzina, M., et al. Positive tuberculin test result in a dog - case report. Veterinary Life, 92.09 (2017).
530. Littleton, J., J. Park, A. Herring, T. Farmer. Multiplying and dividing: tuberculosis in Canada and Aotearoa New Zealand, 2008, 3, <http://hdl.handle.net/2292/2558>
531. Marrinhas, C., Malhão, F., Lopes, C., Sampaio, F., Moreira, R., Caniatti, M., ... & Marcos, R. (2022). Doing more with less: multiple uses of a single slide in veterinary

- cytology. A practical approach. Vet. Res. Commun., 46(3), 641-654. 2022.
532. Martinho, A.P.V., Franco, M.M.J., Ribeiro, M.G., et al. Case report: disseminated *Mycobacterium tuberculosis* infection in a dog. Am. J. Trop. Med. Hyg., 3, 88, 2013, 596-600.
 533. Moravkova, M., M. Slany, I. Trcka, M. Et al. Human-to-human and human-to-dog *Mycobacterium tuberculosis* transmission studied by IS6110 RFLP analysis: a case report. Veterinarni Medicina, 2011, 56, 6, 314-317.
 534. Parsons, S.D.C. Natural animal model systems to study tuberculosis. PhD Thesis, Stellenbosch University, 2010.
 535. Parsons, S.D.C., Gous, T.A., Warren, R.M., van Helden, P.D. Pulmonary *Mycobacterium tuberculosis* (Beijing strain) infection in a stray dog (2008) J. South African Vet. Assoc., 79 (2), pp. 95-98.
 536. Parsons, S.D.C., Warren, R.M., Ottenhoff, T.H.M., et al. Detection of *Mycobacterium tuberculosis* infection in dogs in a high-risk setting. Res. Vet. Sci., 92 (3), 2012, 414-419.
 537. Pesciaroli, M., Alvarez, J., Boniotti, M. B., Cagiola, M., Di Marco, V., , ... & Pasquali, P. (2014). Tuberculosis in domestic animal species. Res. Vet. Sci., 97, S78-S85.
 538. Radulski, L., Krajewska-Wedzina, M., & Lipiec, M. (2022). Occurrence of *Mycobacterium tuberculosis* complex infections in animals. Part II. Companion animals. Veterinary Life, 97(09).
 539. Ribeiro, M. G., et al. Pre-Multidrug-Resistant *Mycobacterium tuberculosis* Infection Causing Fatal Enteric Disease in a Dog from a Family with History of Human Tuberculosis. Transboundary and emerging diseases 64.5 (2017): e4-e7.
 540. Romero, A. M. (2006). Production of Reactive Nitrogen Intermediates in culture of mononuclear cells from goats exposed to BCG from *M. bovis*, supplemented with 1, 25 Dihydroxyvitamin D3 (Doctoral dissertation, Universidad Autónoma Agraria Antonio Narro).
 541. Szaluś-Jordanow, O., Augustynowicz-Kopeć, E., Czopowicz, M., Olkowski, A., Łobaczewski, A., Rzewuska, M., ... & Frymus, T. Intracardiac tuberculomas caused by *Mycobacterium tuberculosis* in a dog. BMC Vet. Res., (2016, 12, 1-6.
 542. Vangone, L., Cardillo, L., Riccardi, M. G., Borriello, G., Cerrone, A., Coppa, P., ... & Fusco, G. (2021). *Mycobacterium tuberculosis* SIT42 Infection in an Abused Dog in Southern Italy. Front. Vet. Sci., 8, 653360.
 543. Verma, R., R.Sangram, S. Kumar Singh. Canine tuberculosis: Mini Review. Indian J. Vet. Res., 22.2 (2013): 1-9.
 544. Warren, R. M., Gous, T. A., Van Helden, P. D., & Parsons, S. D. C. (2008). Pulmonary *Mycobacterium tuberculosis* (Beijing strain) infection in a stray dog: clinical communication. Journal of the South African Veterinary Association, 79(2), 95-98.
- 2008 Изтриване
545. Калмыков, В. М., А. Х. Найманов, and М. С. Калмыкова. Диагностика туберкулёза собак и кошек с использованием полимеразной цепной реакции. Молекулярная Диагностика 2017, 406-407.
 546. Утанова, Г. Х., and А. П. Силаев. Молекулярно-генетическая диагностика вируса бычьего иммунодефицита. Молекулярная Диагностика 2017, 407-408.
- 14). Zheleva-Dimitrova D, Gevrenova R, Zaharieva MM, NAJDENSKI H. et al. HPLC-UV and LC-MS Analyses of Acylquinic Acids in *Geigeria alata* (DC) Oliv. &**

Hiern. and their Contribution to Antioxidant and Antimicrobial Capacity. Phytochemical Analysis 2017, 28 (3), 176-184.

547. Alcázar, M. A., Kamimura, N., Soumyanath, A., Stevens, J. F., & Maier, C. S. (2021). Caffeoylquinic acids: Chemistry, biosynthesis, occurrence, analytical challenges, and bioactivity. *The Plant Journal*, 107(5), 1299-1319.
548. El Bashir, I. B., Kordofani, M. Y., Yagi, S., Al-Atar, A. A., Qahtan, A. A., Abdel-Salam, E. M., ... & Zengin, G. (2022). Antimalarial, cytotoxic and antioxidant activities of 14 medicinal plants from Sudan. *Nat. Resources Human Health*, 1-7.
549. Bernard, G. et al. MeJA elicitation of chicory hairy roots promotes efficient increase of 3, 5-diCQA accumulation, a potent antioxidant and antibacterial molecule. *Antibiotics* 9.10 (2020): 659.
550. Birsa M.L., Sarbu L.G. Health Benefits of Key Constituents in *Cichorium intybus* L. (2023) *Nutrients*, 15 (6), 1322. DOI: 10.3390/nu15061322.
551. Chew, J., S.-C. Peh, T.S. Yeang. Non-microbial natural products that inhibit drug-resistant *Staphylococcus aureus*. *Staphylococcus Aureus*. IntechOpen, 2018.
552. Clifford, M. N. "Some notes on the chlorogenic acids. Part 4. Botanical distribution of the chlorogenic acids." (2017).
553. Elbashir, S.M.Idris, et al. Free radical scavenging, α -glucosidase inhibitory and lipase inhibitory activities of eighteen Sudanese medicinal plants." *BMC Compl.Altern. Med.* 18.1 (2018): 1-12.
554. Fadul, E. et al. Anti-glycating and anti-oxidant compounds from traditionally used anti-diabetic plant *Geigeria alata* (DC) Oliv. & Hiern. *Nat. Product Res.*, 34.17 (2020): 2456-2464.
555. Ganzera, M., S. Sturm. Recent advances on HPLC/MS in medicinal plant analysis-An update covering 2011–2016. *J. Pharm. Biomed. Analysis* 147 (2018): 211-233.
556. Gronbach, M. (2020). Characterization of the ingredients of plant extracts, their investigation for their ER stress-relieving activity and application of sublimation to fruit powder for the identification of marker substances and isolation of natural products (Doctoral dissertation, Rostock, University of Rostock, 2021).
557. Heena, K.S., Kaur, V., Panwar, H., Sharma, P., & Jangra, R. (2024). Isolation of quinic acid from dropped *Citrus reticulata* Blanco fruits: its derivatization, antibacterial potential, docking studies, and ADMET profiling. *Front. Chemistry*, 12, 1372560.
558. Ibrahim, S., Osman, W., Maaz, M. A., Ali, A., Fadul, E., Arbab, A. H., ... & Mohamed, M. S. *Geigeria alata*-a potential source for anti-Alzheimer's constituents: In vitro and computational investigations. *Indonesian Journal of Pharmacy*, 34, 4, 2023.
559. Islam, S., Adam, Z., & Akanda, J. H. (2024). Quinic and caffeic acids derivatives: Affecting antioxidant capacities and phenolics contents of certain therapeutic and specialty crops employing water and ethanolic extracts. *Food Chem. Adv.*, 4, 100693.
560. Kłeczek, N. et al. *Carpesium divaricatum* Sieb. & Zucc. Revisited: newly identified constituents from aerial parts of the plant and their possible contribution to the biological activity of the plant. *Molecules* 24.8 (2019): 1614.
561. Lee C.-D., Cho H., Shim J., Tran G.H., Lee H.-D., Ahn K.H., Yoo E., Chung M.J., Lee S. Characteristics of phenolic compounds in *Peucedanum japonicum* according to various stem and seed colors. (2023) *Molecules*, 28 (17), 6266.
562. Li K., Yao Q., Zhang M., Li Q., Guo L., Li J., Yang J., Cai W. Exploring the effective

- components and potential mechanisms of Zukamu granules against acute upper respiratory tract infections by UHPLC-Q-Exactive Orbitrap-MS and network pharmacology analysis. (2023) *Arabian J. Chem.*, 16 (8), 104875.
563. Liu Y., Wang C., Wu J., Tan L., Gao P., Wu S., Tang D., Wang Q., Wang C., Li P., Liu J. Study on the comprehensive phytochemicals and the anti-ulcerative colitis effect of *Saussurea pulchella*. (2023) *Molecules*, 28 (4), 1526.
 564. Milutinović, V. et al. Methanol extracts of 28 Hieracium species from the Balkan Peninsula-Comparative LC–MS analysis, chemosystematic evaluation of their flavonoid and phenolic acid profiles and antioxidant potentials. *Phytochem. Anal.*, 29.1, 2018:30-47.
 565. Mohammed, S.I.H., EL-Kamali, H.H., EL-Tahir, A.S. (2022). Haematological effects of aqueous extracts of *Geigeria alata* in male albino rats. *Journal of The Faculty of Science and Technology*, (9 (2)), 125-133.
 566. Mohamed, M.A. M. (2021). Phytochemical, Antioxidant and Anti-acetylcholinesterase Screening of Some Selected Plants from Sudan (Doctoral dissertation, University of Khartoum).
 567. Nowak, Jadwiga, et al. Phytochemical Analysis of Polyphenols in Leaf Extract from *Vernonia amygdalina* Delile Plant Growing in Uganda. *Appl. Sci.*, 12.2 (2022): 912.
 568. Orlando, G., Zengin, G., Ferrante, C., Ronci, M., Recinella, L., Senkardes, I., ... & Menghini, L. (2019). Comprehensive chemical profiling and multidirectional biological investigation of two wild Anthemis species (*Anthemis tinctoria* var. *pallida* and *A. cretica* subsp. *tenuiloba*): focus on neuroprotective effects. *Molecules*, 24(14), 2582.
 569. Osman, W., Maaz, M. A., Ali, A., Fadul, E., Arbab, A. H., Al-Nour, M. Y., ... & Mohamed, M. S. (2023). *Geigeria alata*-A potential source for anti-alzheimer's constituents: in vitro and computational investigations. *Indonesian J. Pharmacy/Majalah Farmasi Indonesia*, 34(4).
 570. Pereira, I.S.P.et al. Phytochemical and biological studies on *Piptocarpha axillaris* (Less.) Baker (Asteraceae). *Biochem. Systematics Ecol.*, 85 (2019): 24-30.
 571. Quinty V., Nasreddine R., Colas C., Launay A., Nehmé R., El-Khiraoui A., Piot C., Draye M., Destandau E., Da Silva D., Chatel G. Antioxidant and anti-lipase capacities from the extracts obtained from two invasive plants: *Ambrosia artemisiifolia* and *Solidago canadensis*. (2023) *Food Biosci.*, 55, art. no. 103069. DOI: 10.1016/j.fbio.2023.103069
 572. Sinan, K. I., Zengin, G., Zheleva-Dimitrova, D., Gevrenova, R., Picot-Allain, M. C. N., Dall'Acqua, S., ... & Mahomoodally, M. F. (2021). Exploring the chemical profiles and biological values of two Spondias species (*S. dulcis* and *S. mombin*): Valuable Sources of Bioactive Natural Products. *Antioxidants*, 10(11), 1771.
 573. Singla, R., V. Jaitak. Recent advances in plant metabolites analysis, isolation, and characterization. *Rec. Trends Techniq. Plant Metabol. Eng.*, (2018): 75-115.
 574. Varvouni, E. F., Graikou, K., Gortzi, O., Cheilari, A., Aligiannis, N., & Chinou, I. (2021). Chemical and biological evaluation of the oil and seedcake from seeds of a Greek cardoon cultivar as potential functional vegetable oil. Comparison with sesame, flaxseed and extra virgin olive oils. *Foods*, 10(11), 2665.
 575. Xu, Z., et al. Lonicerin, an anti-algE flavonoid against *Pseudomonas aeruginosa* virulence screened from Shuanghuanglian formula by molecule docking based strategy. *J. Ethnopharm.*, 239 (2019): 111909.
 576. Yagi S., Yagi A. Medicinal plants used traditionally in Sudan to treat viral infections

- related to the signs and symptoms of COVID-19: A systematic review. (2023) Curr. Tradit. Med., 9 (6), e060223213450.
577. Zengin, G. et al. Chemical profiling and pharmaco-toxicological activity of *Origanum sipyleum* extracts: Exploring for novel sources for potential therapeutic agents. J. Food Biochem., 43.11 (2019): e13003.
 578. Zengin, G. et al. Multidirectional biological investigation and phytochemical profile of *Rubus sanctus* and *Rubus ibericus*. Food Chem. Toxicol., 127 (2019): 237-250.
 579. Zengin, G. et al. Shedding light on the biological and chemical fingerprints of three Achillea species (*A. biebersteinii*, *A. millefolium* and *A. teretifolia*)." Food & Function 8.3 (2017): 1152-1165.
- 15). NAJDENSKI, H., M. Heyndrickx, L. Herman, H. Werbrouck, E. Van Coillie. Quantification of *Yersinia enterocolitica* in raw milk using qPCR. J. Vet. Med., 2012, 160, 428-434 (IF-3.327)**
580. Ajauskaitė, A. *Yersinia* spp. prevalence of bacteria in pork products. (2017), PhD thesis (Lithuanian University of Health Sciences)..
 581. Bancercz-Kisiel, A. Szweda, W. Yersiniosis – a zoonotic foodborne disease of relevance to public health. Annals of Agricultural and Environmental Medicine 2015, 22, 397-402.
 582. Chapela, M.-J., A. Garrido-Maestu, A.G. Cabado. Detection of foodborne pathogens by qPCR: a practical approach for food industry applications. Cogent Food & Agriculture 1.1 (2015): 1013771.
 583. EFSA Panel on Biological Hazards (BIOHAZ). (2015). Scientific opinion on the public health risks related to the consumption of raw drinking milk. EFSA J., 13(1), 3940.
 584. Godic Torkar, Karmen, et al. The microbiological quality of Slovenian raw milk from vending machines and their hygienic-technical conditions. British Food J., 119.2 (2017): 377-389.
 585. Grigorenko, E., Fisher, C., Patel, S., Chancey, C., Rios, M., Nakhasi, H. L., & Duncan, R. C. Multiplex screening for blood-borne viral, bacterial, and protozoan parasites using an OpenArray platform. J. Mol. Diagnostics, 2014, 16(1), 136-144.
 586. Hongwei, Z., S.Z. Wenjie, W. Shuo. (2018). Detection of *Yersinia enterocolitica* in food by nucleic acid test strip method. Food Res. & Development, 39(24).
 587. Petsios, Stefanos, et al. Conventional and molecular methods used in the detection and subtyping of *Yersinia enterocolitica* in food. Intern. J. Food Microbiol., 237 (2016):55-72.
 588. Rina, S., Z. Liangjuan, P. Lu, & Z. Hongwei. (2018). Detection of *Yersinia pseudotuberculosis* in food by PCR-nucleic acid test strip method. J. Food Safety & Quality, 9(23).
 589. Raghianti, F., E.A. dos Santos, O.A. Martins. *Yersinia enterocolitica* in milk and dairy products: a review. Brazilian J. Hyg. Animal Sanity, 12.4 (2018):420-427.
 590. Rusak, L.A. Estudo molecular dos genótipos de *Yersinia enterocolitica* circulantes no Brasil. Doctoral Dissertation. 2017.
 591. Sha RiNa, S. R., Zhao LiangJuan, Z. L., Pang Lu, P. L., Zhang HongWei, Z. H. (2018). Detection of *Yersinia pseudotuberculosis* in food by PCR-nucleic acid strips method. J. Food. Safety & Quality.9,23, 6107-6111.
 592. Shoaib, M. et al. A comprehensive review on the prevalence, pathogenesis and

- detection of *Yersinia enterocolitica*. RSC Advances 9.70 (2019): 41010-41021.
593. Stachelska, M. A. Quantitative assessment of *Yersinia enterocolitica* in raw pork meat using real time PCR (qPCR) technique. J. Animal Feed Sci., 26.2 (2017): 141-147.
 594. Tavares, A. B., et al. Contamination sources of *Yersinia enterocolitica* during milk production. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 69.2 (2017): 483-490.
 595. Torkar, Karmen Godic, et al. "The microbiological quality of Slovenian raw milk from vending machines and their hygienic-technical conditions." British Food Journal (2017).
 596. Vanantwerpen, Gerty. Prevalence and risk factors of enteropathogenic *Yersinia* spp. in pigs at slaughter age. Diss. Ghent University, 2014.
 597. Van Damme, I. (2013). Isolation and spread of enteropathogenic *Yersinia* spp. throughout the pork production chain (Doctoral dissertation, Ghent University).
 598. Wei, J. Approaches to detect foodborne pathogenic microorganisms. Proceed. Fourth Int. Conf. Biolog. Inform. Biomed. Engineering. 2020.
 599. Zhou, C., Sun, C., Ruan, J., Zou, H., Li, Y. Determination of *Yersinia enterocolitica* in food by capillary electrophoresis with laser induced fluorescence detection. Analyt. Letters 48, 2015, 1988-2001.
 600. Киров, Б., В. Гълъбов, В. Карлова-Сергиева. Интегрирана автоматизирана система за детекция на патогени в течни проби от хранителни продукти—етапи на развитие и области на приложение. (2014).
- 16). De Rosa, S., Kamenarska, Z., Bankova, V., Stefanov, K., Dimitrova-Konaklieva, S., NAJDENSKI, H., Tzevtkova, I., Popov, S. Chemical composition and biological activities of the Black Sea algae *Polysiphonia denudata* (Dillw.) Kutz. and *Polysiphonia denudata* f. *fragilis* (Sperk) woronich (2001). Zeitschrift fur Naturforschung - Section C Journal of Biosciences, 56 (11-12), pp. 1008-1014.**
601. Antypenko, Lyudmyla, et al. "Monomethyl Suberate Screening for Antifungal Activity, Molecular Docking and Drug-Like Properties." Acta Chimica Slovenica 65.4 (2018): 836-841.
 602. Blasi, M. F., Rotini, A., Bacci, T., Targusi, M., Ferraro, G. B., Vecchioni, L., ... & Migliore, L. (2021). On Caretta caretta's shell: first spatial analysis of micro-and macro-epibionts on the Mediterranean loggerhead sea turtle carapace. Marine Biology Research, 17(7-8), 762-774.
 603. Celenk, F., A. Sukatar. Macroalgae of Izmir Gulf: *Cystoseira barbata*, *Cystoseira compressa* and *Cystoseira crinita* species have high α -glucosidase and moderate pancreatic lipase inhibition activities. Iranian J. Pharmaceutical Res., 19.2(2020):391-402.
 604. Chen, S. & Y. Haibin. Screening of 4 kinds of active ingredients of marine algae to inhibit pathogens. Wenzhou Medical College, 2012, 41 6, 583-585.
 605. Cruz-Rivera, E., Villareal, T.A. Macroalgal palatability and the flux of ciguatera toxins through marine food webs (2006) Harmful Algae, 5 (5), 497-525.
 606. De Oliveira, A.L., Avaliação química e biológica de espécimens de *Bostrychia radicans* (Rhodomelaceae), Dissertação, Universidade de São Paulo, Faculdade de Ciências Farmacêuticas de Ribeirão Preto, 2009.
 607. Gaubert, Julie, et al. Metabolomic variability of four macroalgal species of the genus *Lobophora* using diverse approaches. Phytochemistry 162 (2019): 165-172.
 608. Gołębiowski, M., Cerkowniak, M., Urbanek, A., et al. Identification and antifungal

- activity of novel organic compounds found in cuticular and internal lipids of medically important flies. *Microbiol. Res.*, 2015, 170, 213-222.
609. Güven, K. C., B. Coban, H. Erdugan. A chemical research on three red algae *Gracilaria bursa-pastoris*, *Phyllophora crispa* and *Laurencia obtusa* var. *pyramidata*. *Asian J. Chem.*, 26.18 (2014): 6118-6120.
 610. Januário, A. P., Félix, R., Félix, C., Reboleira, J., Valentão, P., & Lemos, M. F. (2021). Red seaweed-derived compounds as a potential new approach for acne vulgaris care. *Pharmaceutics*, 13(11), 1930.
 611. Kawaroe, M., et al. *Mikroalga potensi dan pemanfaatannya untuk produksi bio bahan bakar*. PT Penerbit IPB University Press, 2019.
 612. Linlin, C., X. Zhixin, W. Donghong, P. Fei, Z. Wenzhou, & X. Rong. (2017). Comparison of antioxidant activities of different extracts of three types of macroalgae from Quanzhou Bay. *Food Research & Development*, 38(23).
 613. Oliveira, A.L.L. *Algas e micro-organismos marinhos como fonte de substâncias bioativas: química e biologia de Bostrychia radicans e fungos endofíticos associados*. PhD thesis, Universidade de São Paulo, 2013.
 614. Permeh, P., Gohari, A., Saeidnia, S., Jamili, S., Permeh, S., & Mostafavi, P. G. (2013). Alpha amylase inhibitory and Antioxidant activity of *Padina boergesenii* (Allander and Kraft) from Persian Gulf. *Planta Medica*, 79(13), PA31.
 615. Rickert, E. et al. Seasonal variations in surface metabolite composition of *Fucus vesiculosus* and *Fucus serratus* from the Baltic Sea. *PLoS One* 11.12 (2016): e0168196.
 616. Rontani, J.F., M.A. Galeron. Autoxidation of chlorophyll phytyl side chain in senescent phototrophic organisms: A potential source of isophytol in the environment. *Organic Geochemistry* 97 (2016): 35-40.
 617. Saeidnia, S., Permeh, P., Nasiri, M., Mollazadeh, K., & Farahani, F. (2009). Biological activity of two red algae, *Gracilaria salicornia* and *Hypnea flagelliformis* from Persian Gulf. *Pharmacognosy Research*, 1(6).
 618. Shaobo, C., N. Haihan, Y. Haibin. (2011). Screening of active ingredients of four kinds of seaweed that inhibit marine pathogenic bacteria. *J. Wenzhou Medical College*, 41(6), 583-585.
 619. Takeara, R., Jimenez, P.C., Wilke, D.V., Odorico de Moraes, M., Pessoa, C., Peporine Lopes, N., Lopes, J.L.C., Monteiro da Cruz Lotufo, T., Costa-Lotufo, L.V. Antileukemic effects of *Didemnum psammatoedes* (Tunicata: Ascidiacea) constituents. (2008) *Comp. Biochem. Physiol. - A Molecular and Integrative Physiology*, 151 (3), 363-369.
 620. Xiongping, L., Z. Yi, C. Xiaoqing. (2008). Research progress on seaweed antibacterial active substances (review). *Subtropical Plant Science*, 37(3), 76-80.
 621. Yakimchyk, V.S., et al. Enantioselective catalytic approach to the C23–C28 subunit of 24 α -methyl steroids. *Steroids* 148 (2019): 82-90.
 622. Yu, Z., Hong, Y., Xie, K., Fan, Q. Research progresses on the physiological and pharmacological benefits of microalgae-derived biomolecules. *Foods*, 2022, 11(18), 2806.
 - 17). Konakchiev A, Todorova M, Mikhova B, Vitkova A, NAJDENSKI H. Composition and antimicrobial activity of *Achillea distans* essential oil. *Natural Product Communications*, 6(6), 2011, 905-906, ISSN 1934-578X IF 1.242**
 623. Abad, MJ., et al. The *Artemisia* L. genus: a review of bioactive essential oils.

- Molecules 17.3 (2012): 2542-2566.
624. Abad, M.J., L.M. Bedoya, P. Bermejo. Essential oils from the Asteraceae family active against multidrug-resistant bacteria. Fighting multidrug resistance with herbal extracts, essential oils and their components. Academic Press, 2013. 205-221.
 625. Andrushko, V., N. Andrushko. Principles, concepts, and strategies of stereoselective synthesis. Stereoselective Synthesis of Drugs and Natural Products (2013): 3-44.
 626. Benedec, D., et al. Polyphenolic composition, antioxidant and antibacterial activities for two Romanian subspecies of *Achillea distans* Waldst. et Kit. ex Willd. Molecules 18.8 (2013): 8725-8739.
 627. Benedec, Daniela, et al. Comparative HPLC-MS analysis of phenolics from *Achillea distans* and *Achillea millefolium* and their bioactivity. Studia Universitatis Babes-Bolyai. Chemia, vol. 60, no. 4, Dec. 2015, pp. 257+.
 628. Enna, S.J., S. Norton. Herbal supplements and the brain: understanding their health benefits and hazards. FT Press, 2012.
 629. Gavanji, S., Mohammadi, E., Larki, B., & Bakhtari, A. Antimicrobial and cytotoxic evaluation of some herbal essential oils in comparison with common antibiotics in bioassay condition. Integrative Med. Res., 2014, 3, 3, 142–152.
 630. Issabeagloo, E., B. Abri. Antimicrobial effects of yarrow (*Achillea millefolium*) essential oils against *Staphylococcus* species. African J.Pharm. Pharmacol., 6.41 (2012): 2895-2899.
 631. Issabeagloo, E. M. Taghizadieh. Inhibitory effect of *Ziziphus zizyphus* L. extract on *Staphylococcus* genera. Advanc. Biores., 3.3 (2012).
 632. Jeremić, Jovana Stanković, et al. "Antifungal activity of the essential oil from *Artemisia santonicum* and its constituent isogeranic acid." Lekovite sirovine 40 (2020): 62-65.
 633. Mohammadhosseini, M., S.D. Sarker, A.Akbarzadeh. Chemical composition of the essential oils and extracts of *Achillea* species and their biological activities: A review. J. Ethnopharmacol., 2017, 199, 257-315.
 634. Nekoei, M., M. Mohammadhosseini. Chemical compositions of the essential oils from the aerial parts of *Achillea wilhelmsii* using traditional Hydrodistillation, microwave assisted hydro-distillation and solvent-free microwave extraction methods: comparison with the volatile compounds obtained by headspace solid-phase microextraction. J. Essent. Oil Bearing Plants 19.1 (2016): 59-75.
 635. Popovici, M., Coldea, G., Coste, A., Pop, C., & TĂMAȘ, M. (2021). Comparative phytochemical evaluation in several *Achillea* species from Romania. Farmacia, 69(5).
 636. Özek, G., et al. Chemical compositions of *Achillea sivasica*: different plant part volatiles, enantiomers and fatty acids. Records of Natural Products, 120.2 (2018): 1.
 637. Razavi, S.M., Enferadi, M. (2022). Essential oil composition and metabolites changes of *Artemisia melanolepis* Boiss at different phenological stages. J. Plant Physiol. Breeding, 12(1), 39-49.
 638. Stanković J., Novaković M., Tešević V. et al., Antifungal activity of the essential oil from *Artemisia santonicum* and its constituent isogeranic acid. Lekovite Sirovine, 2020, 40, 62-65.
 639. Tatar, Özgür, et al. Plant-soil water status-induced changes in physiological and biochemical properties of yarrow. J. Essent. Oil Bearing Plants 19.7 (2016): 1776-1787.

640. Zhou, F., et al. In-vitro cardiovascular protective activity of a new achillinoside from *Achillea alpina*. *Revista Brasileira de Farmacognosia* 29.4 (2019): 445-448.
641. Романова, Ирина Борисовна. Клинико-лабораторное обоснование применения противовоспалительных препаратов на растительной основе в комплексном лечении воспалительных заболеваний пародонта у пациентов со скученным положением зубов. Diss. Первый Московский государственный медицинский университет им. ИМ Сеченова, 2017.
- 18). Veljanov, D., Vesselinova, A., Nikolova, S., NAJDENSKI, H., Kussovski, V., Markova, N. Experimental melioidosis in inbred mouse strains (1996) Zentralblatt fur Bakteriologie, 283 (3), pp. 351-359.**
642. Amemiya, Kei, et al. Animal Models for Melioidosis. *Curr. Trop. Med. Reports* (2017): 1-15.
643. Bottex, C., Gauthier, Y.P., Hagen, R.M., Finke, E.J., Splettstösser, W.D., Thibault, F.M., Neubauer, H., Vidal, D.R. Attempted passive prophylaxis with a monoclonal anti-*Burkholderia pseudomallei* exopolysaccharide antibody in a murine model of melioidosis (2005) *Immunopharmacology and Immunotoxicology*, 27 (4), pp. 565-583.
644. Cheng, A.C., Currie, B.J. Melioidosis: Epidemiology, pathophysiology, and management (2005) *Clin. Microbiol. Rev.*, 18 (2), 383-416.
645. Dance, D.A.B. (2009). Melioidosis and glanders as possible biological weapons. *Bioterrorism and infectious agents: a new dilemma for the 21st century*, 99-145.
646. Gauthier, Y. P., et al. "Protease production by *Burkholderia pseudomallei* and virulence in mice." *Acta tropica* 74.2-3 (2000): 215-220.
647. Gauthier, Yves P., et al. "Study on the pathophysiology of experimental *Burkholderia pseudomallei* infection in mice." *FEMS Immunology & Medical Microbiology* 30.1 (2001): 53-63.
648. Gelhaus, H. Carl, et al. Efficacy of post exposure administration of doxycycline in a murine model of inhalational melioidosis. *Scientific Reports* 3.1 (2013): 1-5.
649. Hampton, V., Kaestli, M., Mayo, M., Choy, J. L., Harrington, G., Richardson, L., ... & Currie, B. J. (2011). Melioidosis in birds and *Burkholderia pseudomallei* dispersal, Australia. *Emerging Inf. Dis.*, 17(7), 1310.
650. Herron, I. C., Laws, T. R., & Nelson, M. (2024). Marmosets as models of infectious diseases. *Frontiers Cell. Inf. Microbiol.*, 14, 1340017.
651. Jimma, T.K Vaccine development against plague, glanders and melioidosis in the former Soviet Union in comparison to the current state of global knowledge. Institute of Animal Hygiene and Veterinary Public Health Faculty of Veterinary Medicine, University of Leipzig, Dissertation, 2010.
652. Laws, T.R., Clark, G.C., D'Elia, R.V. Immune profiling of the progression of a BALB/c mouse aerosol infection by *Burkholderia pseudomallei* and the therapeutic implications of targeting HMGB1. *Int. J. Infect. Dis.*, 2015, 40, 1-8.
653. Lever, M.S., Nelson, M., Stagg, A.J., Beedham, R.J., Simpson, A.J.H. Experimental acute respiratory *Burkholderia pseudomallei* infection in BALB/c mice (2009) *International Journal of Experimental Pathology*, 90 (1), 16-25.
654. Mahfouz, M. E., Grayson, T. H., Dance, D. A., & Gilpin, M. L. (2006). Characterization of the mrgRS locus of the opportunistic pathogen *Burkholderia*

- pseudomallei*: temperature regulates the expression of a two-component signal transduction system. BMC Microbiology, 6, 1-16.
655. Santanirand, P., et al. "Obligatory role of gamma interferon for host survival in a murine model of infection with *Burkholderia pseudomallei*." Infection and immunity 67.7 (1999): 3593-3600.
 656. Soffler, Carl. Development and characterization of caprine infection models of melioidosis, The. Diss. Colorado State University, 2007.
 657. Sprague, L. D., & Neubauer, H. (2004). Melioidosis in animals: a review on epizootiology, diagnosis and clinical presentation. J. Vet. Med., Series B, 51(7), 305-320.
 658. Warner, J. M. melioidosis in Papua New Guinea. PhD thesis, James Cook University.
 659. West, T.E., Myers, N.D., Liggitt, H.D., et al. Murine pulmonary infection and inflammation induced by inhalation of *Burkholderia pseudomallei*. Int. J. Exp.Pathol., 2012, 93(6), 421-428.
 - 19). Dimitrova, L., Zaharieva, M. M., Popova, M., Kostadinova, N., Tsvetkova, I., Bankova, V., NAJDENSKI, H. Antimicrobial and antioxidant potential of different solvent extracts of the medicinal plant *Geum urbanum* L. Chem Cent J., 11, 1, 2017, DOI:10.1186/s13065-017-0343-8, 1-11. JCR-IF (Web of Science):2.284**
 660. Adams, S.J., et al. Foliar endophytic fungi from the endangered eastern mountain avens (*Geum peckii*, Rosaceae) in Canada. Plants 10.5 (2021):1026.
 661. Bunse, M, Mailänder, L, Lorenz, P, Stintzing, F, Kammerer D. "Evaluation of *Geum urbanum* L. Extracts with Respect to Their Antimicrobial Potential". Chemistry & Biodiversity, 19. 10.1002/cbdv.202100850., 2021.
 662. Bvumbi, C. et al. The effects of tormentic acid and extracts from *Callistemon citrinus* on *Candida albicans* and *Candida tropicalis* growth and inhibition of ergosterol biosynthesis in *Candida albicans*. The Scientific World Journal 2021.
 663. Bunse, M., L.K. Mailänder, P. Lorenz, F.C. Stintzing, D.R. Kammerer. Evaluation of *Geum urbanum* L. extracts with respect to their antimicrobial potential. Chemistry & Biodiversity, 2022, 19, 2, e202100850.
 664. Dakwa, R., Mozirandi, W. and Mukanganyama, S., 2023. In vitro antibiofilm activity of tormentic acid against *Pseudomonas aeruginosa*. Res. Square, 1-19.
 665. Farzaneh, A., et al. In vitro anti-diabetic and anti-oxidant activities of *Geum* species from Iran. Res. J. Pharmacognosy, 9.2 (2022): 37-44.
 666. Felix-Cuencas, L., et al. Bioactivity characterization of herbal molecules. Herbal Biomolecules in Healthcare Applications. Academic Press, 2022. 145-183.
 667. Garcia-Oliveira, P., et al. Scientific basis for the industrialization of traditionally used plants of the Rosaceae family. Food Chem., (2020):127197.
 668. Ham, S.H., et al. Plant growth-promoting microorganism *Pseudarthrobacter* sp. NIBRBAC000502770 enhances the growth and flavonoid content of *Geum aleppicum*. Microorganisms 10.6 (2022):1241.
 669. Kannin, Markku. Tropical Forestry Reports. Ed. Markku Kannin. University of Helsinki, Finland: Viikki Tropical Resources Institute (VITRI), 2022.
 670. Kashchenko, N.I., Olennikov, D.N. and Chirikova, N.K., 2023. Metabolites of *Geum aleppicum* and *Sibbaldianthe bifurca*: diversity and α -glucosidase inhibitory potential. Metabolites, 13(6), 689.

671. Kozłowska, M., Ścibisz, I., Przybył, J. L., Laudy, A. E., Majewska, E., Tarnowska, K., Małajowicz, J., Ziarno, M. Antioxidant and antibacterial activity of extracts from selected plant material. *Applied Sciences (Switzerland)*, 2022, 12, 19 C7-9871.
672. Ma, J. et al. The potential of pyrolyzates from rapeseed meal for food additives. *Caribbean J. Sci.*, 51.3 (2018): 619-632.
673. Madić, V. et al. Genotoxic and antigenotoxic potential of herbal mixture and five medicinal plants used in ethnopharmacology. *South African J. Botany*, 125 (2019):290-297.
674. Madić, V. Procena biološke aktivnosti biljne mešavine koja se primenjuje u etnofarmakološkom tretmanu dijabetesa. PhD thesis, University of Nic, Serbia
675. Olech, M., W. Ziemichód, N. Nowacka-Jechalke. The occurrence and biological activity of tormentic acid - a review. *Molecules* 26.13 (2021): 3797.
676. Özcan, K., T. Acet. Antimicrobial and antioxidant screening, synergy studies of *Helichrysum chionophilum* extracts against to resistant microbial strains. *Fresenius Env. Bull.*, 27.7 (2018): 5045-5052.
677. Palanisamy, C. P., et al. Antioxidant and antimicrobial activities of (6E, 10E)-2, 6, 24-trimethyl pentacos-2, 6, 10-triene from *Euclea crispa* leaves. *South African J. Botany* 124 (2019): 311-319.
678. Pooja, M., Poonam, R., Savan, D., & Sumitra, C. Chapter-5 Green synthesis, characterization and antimicrobial activity of copper oxide nanoparticles synthesized using delonix regia leaf. Chief Editor, 87.
679. Pozdnyakova, Y. et al. Biodiversity of wild spice plants of the Central Kazakhstan region and their medicinal potential. *Biodiversitas J. Biol. Diversity* 23.9 (2022).
680. Sadiki, F.Z., et al. Chemical composition and antibacterial activity of essential oil of *Tetraclinis articulata* (Vahl) masters branches of eastern Morocco. *Chem. Biol. Technol. Agricult.*, 5.1 (2018):24.
681. Salih, E.Y.A. Ethnobotany, phytochemistry and antimicrobial activity of *Combretum*, *Terminalia* and *Anogeissus* species (Combretaceae) growing naturally in Sudan. PhD thesis, Iniversity of Helsinki (2019).
682. Schmitt, M. et al. Investigation of antioxidant and elastase inhibitory activities of Geum urbanum aerial parts, Chemical characterization of extracts guided by chemical and biological assays. *Nat. Product Communicat.*, 15(3), 2020:1934578X20915307.
683. Schmitt, M. CHAVIC-Valorisation de la flore Champardennaise dans le domaine du vieillissement cutané. Doctoral Dissertation, Reims, 2019.
684. Telagathoti, Anusha et al. High-throughput volatilome fingerprint using PTR–ToF–MS shows species-specific patterns in Mortierella and closely related genera. *J. Fungi* 7.1 (2021):66.
685. Wadhvaniya, M., 2023. Unveiling Indianness: Exploring Hinduism Through The Eyes of Poets in Indian Literature. *Vidya-A Journal of Gujarat University*, 2(2), pp.69-75.
686. Zhang, B., He, Z.L., Jiang, L., Kuang, W.M., Liu, J., Sun, X., Li, Y.J. and Li, Y., 2023. Chemical constituents from *Potentilla kleiniana* and their chemotaxonomic significance. *Biochem. System. Ecol.*, 111, 104740.
687. Zhao, R. et al. Protective effects of aqueous extract from Gei Herba on blood-deficiency mice: insights gained by a metabolomic approach. *RSC Advances*, 10(17) (2020):10167-10177.

20). Kroumov, Alexander D, Scheufele, Fabiano B, Trigueros, Daniela E G, Modenes, Aparecido N, Zaharieva, Maya M, NAJDENSKI, H. Modeling and Techno-Economic Analysis of Algae for Bio-Energy and Co-Products. Algal Green Chemistry: Recent Progress in Biotechnology, 1, Elsevier, 2017, ISBN:9780444637840, DOI:10.1016/B978-0-444-63784-0.00011-4, 201-241

688. Aggarwal, M., Remya, N. The State-of-the-Art Production of Biofuel from Microalgae with Simultaneous Wastewater Treatment: Influence of Process Variables on Biofuel Yield and Production Cost. // Bioenergy Research, 2022, Vol. 15, Issue 1, p. 62-76, ISSN 19391234 (ISSN). doi:10.1007/s12155-021-10277-1.
689. Anwar, M. et al. Recent advancement and strategy on bio-hydrogen production from photosynthetic microalgae. Bioresource Technol., (2019):121972.
690. Bala, R., Mondal, M. K. Opportunities and challenges in industrial production of biofuels. // Biofuels and Bioenergy: Opportunities and Challenges. Elsevier, 2022, p. 3-21. ISBN 978-032385269-2 (ISBN); 978-032385270-8 (ISBN).
691. Choudhary, P. and R. K. Srivastava (2020). "Techno-economic case study: Bio-fixation of industrial emissions at an Indian oil and gas plant." Journal of Cleaner Production 266.10.1016/j.jclepro.2020.121820.
692. Costa, J.A.V., Freitas, B.C.B., Moraes, L., Zapparoli, M., Morais, M.G. Progress in the physicochemical treatment of microalgae biomass for value-added product recovery. Bioresource Technology, 301,122727.
693. Del Mondo, A. et al. Challenging microalgal vitamins for human health. Microbial Cell Factories 19.1 (2020): 1-23.
694. Khadim, S.R., Singh, P., Singh, A.K., Tiwari, A., Mohanta, A., & Asthana, R.K. (2018). Mass cultivation of *Dunaliella salina* in a flat plate photobioreactor and its effective harvesting. Bioresource Technol., 270, 20-29.
695. Khetkorn, W., Rastogi, R.P., Incharoensakdi, A., Lindblad, P., Madamwar, D., Pandey, A. and Larroche, C., (2017): Microalgal hydrogen production—A review. Bioresource Technol., 243, 1194-1206.
696. Masoumi, S., Borugadda, V. B., & Dalai, A. K. (2021). Techno-economic analysis of algal biorefineries. In Algal Biorefinery (pp. 265-292). Routledge.
697. Musa, M. et al. Microalgae dewatering for biofuels: A comparative techno-economic assessment using single and two-stage technologies. J. Cleaner Product., 229, 2019:325-336.
698. Ng, W. Z., Obon, A. A., Lee, C. L., Ong, Y. H., Gourich, W., Maran, K., Tang, D. B. Y., Song, C. P., Chan, E. S. Techno-economic analysis of enzymatic biodiesel co-produced in palm oil mills from sludge palm oil for improving renewable energy access in rural areas. Energy, 2022, 243 C7 – 122745.
699. Pereira, A.C.E. (2020). Arthrospira sp. cultivation from laboratory scale to pilot-scale. Master dissertation, University of Porto.
700. Pradhan, N., et al. Emerging trends in the pretreatment of microalgal biomass and recovery of value-added products: A review. Bioresource Technol. 369 C7-128395 (2022).
701. Razu, M.H., F. Hossain, M. Khan. Advancement of bio-hydrogen production from microalgae. Microalgae Biotechnology for Development of Biofuel and Wastewater Treatment. Springer, Singapore, 2019. 423-462.
702. Ramirez, G. (2020). Aislamiento y caracterización de una microalga verde para la fijación de CO2 (Master's thesis, Garcia Ramirez, Magly).

703. Sansone, C., Pistelli, L., Del Mondo, A., Calabrone, L., Fontana, A., Noonan, D. M., Albini, A., Brunet, C. The microalgal diatoxanthin inflects the cytokine storm in SARS-CoV-2 stimulated ACE2 overexpressing lung cells. *Antioxidants*, 2022, 11, 8 C7 - 1515,
704. Sansone, C., C. Brunet. Marine algal antioxidants. *Antioxidants* 2020, 9(3), 206.
705. Singh, R.P., Yadav, P., Kumar, I., Kumar, A. and Gupta, R.K., 2023. Bioprospecting and mechanisms of cyanobacterial hydrogen production and recent development for its enhancement as a clean energy. In *Cyanobacterial Biotechnology in the 21st Century* (pp. 107-131). Singapore: Springer Nature Singapore.
706. Xie, T., et al. "Flow Field, Flame Structure and Emissions Quantifications of Oxygenated Glycerol in a Swirl Flame Combustor." *Fuel* 321 C7 - 124052 (2022).
707. Бозиева, А.М., Заднепровская, Е. В., & Аллахвердиев, С. И. О. (2022). Получение биоводорода: последние достижения и современное состояние. *Глобальная энергия*, 28(4), 59-78.
- 21). NAJDENSKI, H., M. Heyndrickx, L. Herman, W. Messens. Fla-DGGE analysis of *Campylobacter jejuni* and *Campylobacter coli* in cecal samples of broilers without cultivation. *Vet. Microbiology*, 2008, 127, 196-202.**
708. Ahmed, M.U., Dunn, L., Ivanova, E.P. Evaluation of current molecular approaches for genotyping of *Campylobacter jejuni* strains. *Foodborne Pathog. Dis.*, 2012, 9(5), 375-385.
709. Ahmed, M. U. (2014). Investigation of a framework for epidemiological studies of *Campylobacter jejuni* based on the combination of genetic markers and MALDI-TOF MS typing schemes (Doctoral dissertation, Swinburne University of Technology Melbourne, Australia).
710. Belák, Á. (2009). Élelmiszer-biztonsági szempontból jelentős baktériumok kimutatása, PCR-alapú molekuláris azonosítása és tipizálása (Doctoral dissertation, Budapesti Corvinus Egyetem).
711. Bi, S., C. Miao, Z. Zhi, et al. Detection and genotyping of *Campylobacter jejuni* and *Campylobacter coli* in food samples by Fla-DGGE. *Modern Food Sci. Technol.*, 2010, 26, 10, 1148-1152.
712. Chinivasagam, H. N., Estella, W., Finn, D., Mayer, D. G., Rodrigues, H., & Diallo, I. (2024). Broiler farming practices using new or re-used bedding, inclusive of free-range, have no impact on *Campylobacter* levels, species diversity, *Campylobacter* community profiles and *Campylobacter* bacteriophages. *AIMS Microbiology*, 10(1), 12-40..
713. Daczowska-Kozon, E. G., Sawicki, W., & Skotarczak, K. The caeca-niche supporting survival of *Campylobacter* spp. in commercially reared broiler chickens. *Polish J. Food Nutr. Sci.*, 2010, 60(3), 265-271.
714. Elshagmani, E. (2015). The role of *Campylobacter concisus* in enteric infections (Doctoral dissertation, RMIT University).
715. Huq, M., & Istivan, T. (2021). Conventional and molecular detection methods of the opportunistic bacterial pathogen *Campylobacter concisus*. In *Campylobacter*. IntechOpen.
716. Lei S, et al. *J. South China University of Technol. (Nat. Sci.)*, 40 (5), 2012, 12-20.
717. Liping, X., X. Yongchun, Y. Yueqing. (2012). Research progress on *Campylobacter jejuni* typing technology. *Public Health and Preventive Medicine*, 23(6), 58-60.
718. Meng H-C, Bi S-L, Yan H, Shi L. Isolation of campylobacter strains in poultry products and genotyping identification of strains by means of PFGE and DGGE. *Huanan*

- Ligong Daxue Xuebao/J. South China Univ. Technol. (Nat. Sci.), 40 (5), 2012, 149-154.
719. Vanmarsenille, C, et al. Nanobodies targeting conserved epitopes on the major outer membrane protein of *Campylobacter* as potential tools for control of *Campylobacter* colonization. *Vet. Res.*, 48.1 (2017):86.
 720. Vanmarsenille, C. (2018). Protecting chickens against *Campylobacter jejuni* by Camelidae-derived nanobodies (Doctoral dissertation, Ghent University).
 721. Xing, L., X. Yongchun, & Y. Yueqing. Development of genotyping technologies of *Campylobacter jejuni*. *Publ. Health Prev. Med.*, 2012, 23 (006), 58-60.
 722. Yao, G. F., Hu, Y. L., Kong, N. Q., Liu, J. H., Luo, Y. W., Li, C. Y., & Bi, S. L. (2023). Rapid genotyping of *Campylobacter coli* strains from poultry meat by PFGE, Sau-PCR, and fla-DGGE. *Curr. Microbiol.*, 80(12), 402.
 723. Въшин, И.Т. Проучвания върху разпространението, видовия състав и някои особености на *Campylobacter* spp. при заклани животни. Дисертация „Доктор на ветеринарномедицинските науки” Стара Загора, 2009
- 22). NAJDENSKI, H., Kussovski, V., Vesselinova, A. Experimental *Burkholderia pseudomallei* infection of pigs. J. Vet. Med. Series B: Infectious Diseases and Veterinary Public Health, 2004, 51 (5), pp. 225-230.**
724. Amemiya, K., Bozue, J.A., Cote, C.K., Deshazer, D., Soffler, C., Welkos, S.L., & Worsham, P.L. (2017). Animal models for melioidosis. *Curr. Trop. Med. Reports*, 4, 208-222.
 725. Boddey, J.A. Molecular and cellular characterisation of *Burkholderia pseudomallei* interactions with host cells. PhD thesis, School of Medical Science, Griffith University, Gold Coast, Queensland, 2005
 726. Bondi, S.K., Goldberg, J.B. Strategies toward vaccines against *Burkholderia mallei* and *Burkholderia pseudomallei* (2008) *Expert Review of Vaccines*, 7 (9), 1357-1365.
 727. Broes, A., Taylor, D.J., & Martineau, G.P. (2019). Miscellaneous bacterial infections. *Diseases of Swine*, 981-1001.
 728. Conejero, L., N. Patel, M. Reynal, S. Oberdorf, J. Prior, P. Felgner, R. Titball, F. Salguero, G. Bancroft. Low-dose exposure of C57BL/6 mice to *Burkholderia pseudomallei* mimics chronic human melioidosis. *Am J Pathol.* 2011, 179 (1):270-280.
 729. Herron, I. C., Laws, T. R., & Nelson, M. (2024). Marmosets as models of infectious diseases. *Frontiers in Cellular and Infection Microbiology*, 14, 1340017.
 730. Koh, G.C.K.W. The effect of glibenclamide on the pathogenesis of melioidosis. PhD Thesis, Oxford University Press, 2012.
 731. Keluangkhot, V., Pethsouvanh, R., Strobel, M. Melioidosis [Mélioidose] (2005) *Medecine et Maladies Infectieuses*, 35 (10), 469-475.
 732. Koh, G. C. K. W. (2012). The effect of glibenclamide on the pathogenesis of melioidosis. PhD thesis, university of Cambridge.
 733. Kwanhian, Wiyada, et al. Investigation of melioidosis outbreak in pig farms in Southern Thailand. *Vet. Sci.*, 7.1 (2020): 9.
 734. Sprague, L.D., H. Neubauer. Melioidosis in animals: a review on epizootiology, diagnosis and clinical presentations. *J. Vet. Med.*, 2004, B51, 305-320.
 735. Soffler, C., Bosco-Lauth, A.M., Aboellail, T.A., et al. Pathogenesis of percutaneous infection of goats with *Burkholderia pseudomallei*: clinical, pathologic, and immunological responses in chronic melioidosis. *Int. J. Exp. Pathol.*, 2014, 95, 2, 101-119.

736. Soffler, C. The development and characterization of caprine infection models of melioidosis. PhD thesis, 2012, Colorado State University.
737. Tengfei, M., F. Yao, & M. Xuhu. (2016). Research progress on animal models of melioidosis. *Progress Microbiol. Immunol.*, 44(4), 65-69.
738. Van Schaik, E., Tom, M., DeVinney, R., Woods, D.E. Development of novel animal infection models for the study of acute and chronic *Burkholderia pseudomallei* pulmonary infections (2008) *Microb. Infect.*, 10 (12-13), 1291-1299.
739. Warawa, J. M. Evaluation of surrogate animal models of melioidosis. *Front. Microbiol. Cell. Infect. Microbiol.*, 2011, 1, 141, 1-12
- 23). NAJDENSKI, H., Iteman, I., Carniel, E. The genome of *Yersinia enterocolitica* is the most stable of the three pathogenic species. (1995) *Contributions to Microbiology and Immunology*, 13, pp. 281-284.**
740. Cebula, T. A., & LeClerc, J. E. (1997). Response from Cebula and LeClerc. *Trends in Microbiology*, 5(11), 428-429.
741. Dey, P. and Ray Chaudhuri, S., 2023. The opportunistic nature of gut commensal microbiota. *Crit. Rev. Microbiol.*, 49(6), 739-763.
742. Fredriksson-Ahomaa, M., Stolle, A., Stephan, R. Prevalence of pathogenic *Yersinia enterocolitica* in pigs slaughtered at a Swiss abattoir (2007). *Int. J. Food Microbiol.*, 119 (3), 207-212.
743. Fredriksson-Ahomaa, M., Stolle, A., Korkeala, H. Molecular epidemiology of *Yersinia enterocolitica* infections (2006) *FEMS Immunol. Med. Microbiol.*, 47 (3), 315-329.
744. Garzetti, D. (2015). Genome-based characterization of *Yersinia enterocolitica*: patho-evolution and adaptation of a versatile bacterium (Doctoral dissertation, München, Ludwig-Maximilians-Universität).
745. Gupta, U. and Dey, P., 2023. Rise of the guardians: gut microbial maneuvers in bacterial infections. *Life Sci.*, 121993.
746. Hallanvuo, S. Foodborne *Yersinia*: Identification and Molecular Epidemiology of Isolates from Human Infections. Academic Dissertation. University of Helsinki, Faculty of Agriculture and Forestry, 2009.
747. Huang XZ, Chu MC, Engelthaler DM, et al. Genotyping of a homogeneous group of *Yersinia pestis* strains isolated in the United States. *J. Clin. Microbiol.*, 2002, 40, 4, 1164-1173.
748. Kasimir, S. Verlaufsuntersuchungen zum Vorkommen potentiell humanpathogener *Yersinia enterocolitica* und *Campylobacter* spp. in Schweinebeständen von der Geburt bis zur Schlachtung sowie genotypisierung ausgewählter isolate. Doctoral dissertation, Veterinärmedizinischen Fakultät, Universität Leipzig, 2005.
749. Leon-Velarde, C.G. The application of bacteriophage host recognition binding proteins for the isolation of *Yersinia enterocolitica* in foods. Doctoral Dissertation, 2017, University of Guelph.
750. Lindler, L.E. *Yersinia pestis* as an emerged pathogen. What lessons can be learned? In: Biological weapons defense, Infectious diseases and counterbioterrorism (L.E. Lindler, F.J. Lebeda and G.W. Korch, eds.), 2005.
751. Lyte M, Nguyen KT. Alteration of *Escherichia coli* O157:H7 growth and molecular fingerprint by the neuroendocrine hormone noradrenaline. *Microbios*, 1997, 89, 360-61,

197-213.

752. Matic I, Radman M, Taddei F, et al. Highly variable mutation rates in commensal and pathogenic *Escherichia coli*. *Science*, 1997, 277, 5333, 1833-1834.
753. Metzgar D., Wills C. Evidence for the adaptive evolution of mutation rates. *Cell*, 2000, 101, 6, 581-584.
754. Metzgar D, Wills C. Evolutionary changes in mutation rates and spectra and their influence on the adaptation of pathogens. *Microb. Infection*, 2000, 2, 12, 1513-1522.
755. Pang T. Genetic dynamics of *Salmonella typhi* - diversity in clonality. *Trends Microbiol.*, 1998, 9, 339-342.
756. Taddei F, Matic I, Godelle B, et al. To be a mutator, or how pathogenic and commensal bacteria can evolve rapidly. *Trends Microbiol.*, 1997, 5, 11, 427-428.
757. Toutounian-Mashhad, K. (2007). Molekularbiologische Feintypisierung von *Yersinia* spp.-Stämmen aus Mastgeflügel. PhD thesis, Technical University of Berlin.
- 24). Voynikov, Y., Zheleva-Dimitrova, D., Gevrenova, R., Lozanov, V., Zaharieva, M. M., Tsvetkova, I., NAJDENSKI, H., Yagi, S., Almoulah, N. F., Momekov, G.. Hydroxycinnamic acid amide profile of *Solanum schimperianum* Hochst by UPLC-HRMS. *International Journal of Mass Spectrometry*, 408, Elsevier, 2016, ISSN:1387-3806, DOI:<http://dx.doi.org/10.1016/j.ijms.2016.08.008>, 42-50. JCR-IF (Web of Science):1.702**
758. Alajmi MF, Alam P, Rehman M, Husain FM, Khan AA, Siddiqui NA, Hussain A, Kalam M, Parvez MK. Interspecies Anticancer and Antimicrobial Activities of Genus *Solanum* and Estimation of Rutin by Validated UPLC-PDA Method. *Evidence-Based Complementary and Alternative Medicine, eCAM*, 2018.
759. Buitimea-Cantua NE, Gutierrez-Urbe JA, Serna-Saldivar SO. Phenolic-protein interactions: Effects on food properties and health benefits. *J. Med. Food*, 2018, 21 (2):188-198.
760. Chen, X. H., et al. "Identification of compounds in *Lycii* Cortex by UPLC-LTQ-Orbitrap-MS." *Zhongguo Zhong yao za zhi= Zhongguo zhongyao zazhi= China journal of Chinese materia medica* 44.20 (2019): 4486-4494.
761. Chen, Chen, et al. Multivariate statistical analysis of metabolites in *Anisodus tanguticus* (Maxim.) Pascher to determine geographical origins and network pharmacology. *Frontiers Plant Sci.*, 13 (2022): 927336-927336.
762. Dastmalchi, K. et al. Temporal resistance of potato tubers: Antibacterial assays and metabolite profiling of wound-healing tissue extracts from contrasting cultivars. *Phytochemistry* 159 (2019): 75-89.
763. Du, Nana, et al. Discovery of new muscarinic acetylcholine receptor antagonists from *Scopolia tangutica*. *Scientific Reports* 7 (2017).
764. Fan, A., Hou, B. L., Tang, Z., Wang, T., Zhang, D., Liang, Y., & Wang, Z. (2023). Liquid Chromatography-Tandem Mass Spectrometry-Based Metabolomics Analysis of Indigo Naturalis Treatment of Ulcerative Colitis in Mice. *J. Med. Food*, 26(12), 877-889.
765. Favre, Laurie, et al. Discrimination of four marine biofilm-forming bacteria by LC-MS metabolomics and influence of culture parameters. *J. Proteome Res.*, 16.5 (2017): 1962-1975.
766. Li, Z., Zhao, C., Zhao, X., Xia, Y., Sun, X., Xie, W., ... & Xu, G. (2018). Deep

- annotation of hydroxycinnamic acid amides in plants based on ultra-high-performance liquid chromatography–high-resolution mass spectrometry and its in silico database. *Analyt. Chem.*, 90(24), 14321–14330..
767. Milutinović, M., Nakarada, Đ., Božunović, J., Todorović, M., Gašić, U., Živković, S., Skorić, M., Ivković, Đ., Savić, J., Devrnja, N. and Aničić, N., 2023. *Solanum dulcamara* L. Berries: A convenient model system to study redox processes in relation to fruit ripening. *Antioxidants*, 12(2), 346.
768. Ozturk, Refiye Beyza, et al. Which extraction solvents and methods are more effective in terms of chemical composition and biological activity of *Alcea fasciculiflora* from Turkey?. *Molecules* 27.15 (2022): 5011.
769. Paul, K., et al. A Combined phenotypic and metabolomic approach for elucidating the biostimulant action of a plant-derived protein hydrolysate on tomato grown under limited water availability. *Frontiers Plant Sci.*, 10 (2019):493.
770. Roumani, M. et al. Phenolamides: Plant specialized metabolites with a wide range of promising pharmacological and health-promoting interests. *Biomed.& Pharmacother.*, 131 (2020):110762.
771. Roumani, M. (2021). Les phénolamides de la tomate: développement d’une approche d’ingénierie métabolique pour l’étude de leurs fonctions in planta, et évaluation de leurs activités biologiques (Doctoral dissertation, Université de Lorraine).
772. Samulski, G.B. et al. Dereplication of *Palicourea sessilis* ethanol extracts by UPLC-DAD-ESI-MS/MS discloses the presence of hydroxycinnamic acid amides and the absence of monoterpene indole alkaloids. *Biochem. Systemat. Ecol.*, 92 (2020): 104114.
773. Wang, W., H.D. Snooks, S. Sang. The chemistry and health benefits of dietary phenolamides. *J. Agric. Food Chem.* 2020, 68, 23, 6248–6267.
774. Zhang, W., X. Sun, W. Sui, X. Jiang, S. Cang, Q. Wang, R. Liu, H. Xu, Q. Li, W. Bi and Y. Cui (2020). Quality control of Xiebai San standard decoction assisted by network pharmacology strategy. *Chromatographia* 83(7): 873–884.
775. Zheng, B., Ding, Y., Johnson, J. B., Xiang, J., Li, Z., Zhang, Y., Luo, D. Enrichment and bioactivities of polyphenols of crude extract by deep eutectic solvent extraction from foxtail millet bran. *Int. J. Food Sci. Technol.*, 2022, 57, 12, 7974–7983.
776. Zheng, B., et al. Green extraction of phenolic compounds from foxtail millet bran by ultrasonic-assisted deep eutectic solvent extraction: Optimization, comparison and bioactivities. *LWT* 154 (2022): 112740.
- 25). Tsvetkova, I., NAYDENSKI H., Petrova A. et al. Antibacterial activity of some bulgarian higher basidiomycetes. *Int. J. Med. Mushr.*, 2006, 8, 63—66.**
777. Abdel-Azeem, A.M., M.A. Abdel-Azeem, W.F. Khalil. Endophytic fungi as a new source of antirheumatoid metabolites. *Bioactive food as dietary interventions for arthritis and related inflammatory diseases*. Academic Press, 2019. 355–384.
778. Badalyan, S. Medicinal aspects of edible ectomycorrhizal mushrooms. In *Edible Ectomycorrhizal Mushrooms* (pp. 317–334). Springer Berlin Heidelberg, 2012.
779. Elsayed, E. A., El Enshasy, H., Wadaan, M. A., & Aziz, R. Mushrooms: a potential natural source of anti-inflammatory compounds for medical applications. *Mediators of Inflammation*, 2014, Article ID 805841, pp.15.
780. Krupodorova, T., V. Barshteyn, E. Pokas. Antibacterial activity of *Fomitopsis betulina*

- cultural liquid. EUREKA: Life Sciences 6 (2019): 10-16.
781. Reyes, Renato G., et al. A new record of the mycoparasitic habit of *Collybia reinakeana* RGR-FE-NSC strain against *Aspergillus flavus*, *Fusarium oxysporum* and *Cladosporium sphaerospermum*. Int. J. Pharmac. Res. Allied Sci., 2017, 6(3), 29-32.
 782. Sajon, S.R. et al. Mushrooms: Natural factory of anti-oxidant, anti-inflammatory, analgesic and nutrition. J. Pharmacog. Phytochem., 7.1 (2018):464-475.
 783. Sánchez, C. Bioactives from mushroom and their application. Food Bioactives. Springer International Publishing, 2017. 23-57.
 784. Teplyakova, T.V., Psurtseva, N.V., Kosogova, T.A., et al. Antiviral activity of polyporoid mushrooms (higher Basidiomycetes) from Altai Mountains (Russia). Int. J. Med. Mushrooms, 2012, 14(1).
 785. Zafar, S. et al. Mushroom Species and Classification: Bioactives in Poisonous and Edible Mushrooms. Poisonous Plants & Phytochemicals in Drug Discovery, 2020:163-188.
 786. Zaichenko, T., Krupodorova, T., Barshteyn, V., & Dekhtiarenko, N. (2017). Антибактеріальні властивості деяких макроміцетів. Research Bulletin of the National Technical University of Ukraine "Kyiv Politechnic Institute", (3), 19-28..
 787. Круподьорова, Т.А., Н.А. Бісько, Н.Л. Поєдинок, и сотр. Антимікробна активність штамів *Ganoderma applanatum* (Pers.: Wallr.) pat. та *G. lucidum* (curt.: fr.) p. karst. в умовах глибинного культивування. Укр. Ботан. Журн., 2008, 65, 4, 590-595.
 788. Зайченко, Т. О., Круподьорова, Т. А., Барштейн, В. Ю., Дехтяренко, Н. В., Зайченко, Т. А., Круподерова, Т. А., ... & Дехтяренко, Н. В. (2017). Антибактеріальні властивості деяких макроміцетів.
 789. Сенюк, О.Ф., Горовой, Л.Ф., Паламар, Л.А., & Круль, Н.И. Влияние меланин-глюканового комплекса, выделенного из грибов трутовика, на продолжительность жизни самок мышей линии ICR. Пробл. старения и долголетия, 2014, 23, 1, 11-27
- 26). NAJDENSKI, H., Vesselinova, A., Golkocheva, E., Garbom, S., Wolf-Watz, H. Experimental infections with wild and mutant *Yersinia pseudotuberculosis* strains in rabbits (2003) Journal of Veterinary Medicine, Series B, 50 (6), pp. 280-288.**
790. Carolin A. Wiedig, Uwe Kramer, Sara Garbom, Hans Wolf-Watz, Ingo B. Autenrieth, Induction of CD8+ T cell responses by *Yersinia* vaccine carrier strains, Vaccine, 2005, 23, 42, 4984.
 791. Chassang, L. et al. Antemortem diagnosis and surgical management of splenitis due to *Yersinia pseudotuberculosis* infection in a pet rabbit (*Oryctolagus cuniculus*). J. Eexotic Pet Med., 29 (2019): 182-187.
 792. Fisher, M.L., Castillo, C., Mecsas, J. Intranasal inoculation of mice with *Yersinia pseudotuberculosis* causes a lethal lung infection that is dependent on *Yersinia* outer proteins and PhoP. Infect. Immun., 2007, 75 (1), 429-442.
 793. Fisher, M. L. (2008). The role of bacterial virulence factors in a newly defined model of murine pneumonic yersiniosis (Doctoral dissertation, Tufts University-Graduate School of Biomedical Sciences).
 794. Huismans, M., Hermans, K., Stock, E. Ultrasonographic diagnosis of hepatic coccidiosis in rabbits. Vlaams Diergeneeskundig Tijdschrift, 2022, 91(2), 55-61.
 795. Matthias Ullrich. Bacterial polysaccharides. Current innovations and future trends

(2009) Horizon Scientific Press

796. Pitkälä, A., Virtanen, T., Joutsen, S., Leimi, A., Tuominen, P. *Yersinia enterocolitica* ja *Yersinia pseudotuberculosis* suomalaisissa elintarvikkeissa – riskiprofiili. *Eviran Tutkimuksia*, 2009, 2, pp 75.
797. Sidiropoulou, A. (2004). Contribution to the control of infectious (bacterial) and parasitic diseases in systemically bred rabbits, which are also used as laboratory animals (Doctoral dissertation, Aristotle University of Thessaloniki).
798. Vassilakos D., A. Natoli, M. Dahlheim, A. R. Hoelzel, Balancing and Directional Selection at Exon-2 of the MHC DQB1 Locus among Populations of Odontocete Cetaceans, *Molecular Biology and Evolution*, 2008, 26, 3, 681.
799. Wanford, J.J., Hachani, A., Odendall, C. Reprogramming of cell death pathways by bacterial effectors as a widespread virulence strategy. *Infect. Immun.*, 2022, 90(5), e-00614-21.
800. Zwack, Erin E., et al. Guanylate binding proteins regulate inflammasome activation in response to hyperinjected *Yersinia* translocon components." *Infect. Immun.*, 85.10 (2017): e00778-16.
801. Воскресенская, Е. А. (2014). Новые подходы к микробиологическому мониторингу популяций *Yersinia pseudotuberculosis* и лабораторной диагностике псевдотуберкулеза (Doctoral dissertation, Российский научно-исследовательский противочумный институт "Микроб").
802. Кокорина, Г. И. (2012). Генотипы штаммов *Yersinia pseudotuberculosis* и их клиническое и диагностическое значение (Doctoral dissertation, Военно-медицинская академия им. СМ Кирова).
803. Кокорина, Г. И., О. А. Шендерович, and Г. Я. Ценева. Применение иммуноблота в диагностике затяжных форм иерсиниоза и изучении вопросов патогенеза (обзор литературы). *Клиническая лабораторная диагностика* 11 (2006): 47-49.
- 27). Popova, M., V. Bankova, I. Tsvetkova, C. NAYDENSKI, M. V. Silva. The first glycosides isolated from propolis: Diterpene rhamnosides. Z. Naturforsch. 56c, 1108 – 1111 (2001).**
804. Alday, E. M. Navarro-Navarro, A. Garibay-Escobar et al. Advances in pharmacological activities and chemical composition of propolis produced in Americas. In *Beekeeping and bee conservation: advances in research*, Chapter 5, 2016
805. Aminimoghadamfarouj, N., A. Nematollahi. Propolis diterpenes as a remarkable bio-source for drug discovery development: A review. *Int. J. Mol. Sci.*, 18.6 (2017):1290.
806. Aminimoghadamfarouj N, Nematollahi A. Structure elucidation and botanical characterization of diterpenes from a specific type of bee glue. *Molecules*. 2017; 22(7):1185.
807. Beltrán Herrera, E.A., & Figueroa Deras, M. N. (2021). Evaluación de la actividad antimicrobiana in vitro de extractos de propoleos de diferentes zonas de El Salvador (Doctoral dissertation, Universidad de El Salvador).
808. Chinou, I. Labdanes of natural origin-biological activities (1981-2004). *Current Medicinal Chemistry* 12(11), 1295-1317 (2005).
809. HA El-Mossalami, H A.N.A.A., & Abdel-Hakeim, Y. A. (2013). Using of propolis extract as a trial to extend the shelf-life and improving the quality criteria of fresh

- Egyptian sausage. Assiut Vet. Med. J., 59(139), 23-33.
810. King, D.I. (2017). Kangaroo island propolis: improved characterisation and assessment of chemistry and botanical origins through metabolomics (Doctoral dissertation).
 811. Méndez Ramírez, G. R. (2020). Evaluación de las características físicoquímicas y perfil cromatográfico por TLC de propoleo procedentes de cinco localidades de El Salvador (Doctoral dissertation, Universidad de El Salvador).
 812. Ohsaki, A., Yokoyama, R., Miyatake, H., & Fukuyama, Y. (2006). Two diterpene rhamnosides, Mimosasides B and C, from *Mimosa hostilis*. Chem. Pharmaceut. Bull., 54(12), 1728-1729.
 813. Salatino, A., Fernandes-Silva, C.C., Righi, A.A., & Salatino, M.L.F. Propolis research and the chemistry of plant products. Nat. Prod. Rep., 2011, 28(5), 925-936.
 814. Seidel, V., Peyfoon, E., Watson, D.G., Fearnley, J. Phytother. Res., 22 (9), 1256-1263 (2008).
 815. Shuzhen, Y., P. Litao, Y. Xiaolin, & P. Siyi. (2009). Research progress on the antifungal effect of propolis. Food Industry Science and Technology, (11), 349-352.
 816. Šturm, L NP Ulrih. Advances in the propolis chemical composition between 2013 and 2018: A review. eFood, 2020, 1(1), 24–37
 817. Temiz, A., Şener, A., Tüylü, A. et al. Antibacterial activity of bee propolis samples from different geographical regions of Turkey against two foodborne pathogens, *Salmonella enteritidis* and *Listeria monocytogenes*. Turkish J. Biol., 2011, 35(4), 503-511.
 818. Tran, Trong D., et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020): 4988, 1-35
 819. Кайгородов, Р. В., and Е. И. Попова. Эколого-биохимические факторы состава и свойств продуктов пчеловодства на примере меда и прополиса. История и методология физиолого-биохимических и почвенных исследований. 2017, 19-22.
- 28). Vesselinova, A., NAJDENSKI, H., Nikolova, S., Kussovski, V. Experimental melioidosis in hens (1996) Journal of Veterinary Medicine, Series B, 43 (6), pp. 371-378.**
820. Amemiya, Kei, et al. Animal models for melioidosis. Curr. Trop. Med. Reports 4.4 (2017): 208-222.
 821. Cheng, A.C., Currie, B.J. Melioidosis: Epidemiology, pathophysiology, and management (2005) Clin. Microbiol. Rev., 18 (2), pp. 383-416.
 822. Dance, D.A.B. 2010, Melioidosis and glanders as possible biological weapons. In Bioterrorism and Infectious Agents: A New Dilemma for the 21st Century (pp. 99-145). Springer New York.
 823. Galperin, M.Y. A census of membrane-bound and intracellular signal transduction proteins in bacteria: Bacterial IQ, extroverts and introverts. BMC Microbiol., 2005, 5, 35.
 824. Hampton, Vanya, et al. Melioidosis in birds and *Burkholderia pseudomallei* dispersal, Australia. Emerg. Inf. Dis., 17.7 (2011): 1310-1312.
 825. Herron, I.C., Laws, T.R., & Nelson, M. (2024). Marmosets as models of infectious diseases. Frontiers Cell. Inf. Microbiol., 14, 1340017.
 826. Jimma, T.K Vaccine development against plague, glanders and melioidosis in the former Soviet Union in comparison to the current state of global knowledge. Institute of Animal Hygiene and Veterinary Public Health Faculty of Veterinary Medicine, University

- of Leipzig, Doctoral Dissertation, 2010.
827. Mahfouz, M.E., et al. Characterization of the mrgRS locus of the opportunistic pathogen *Burkholderia pseudomallei*: temperature regulates the expression of a two-component signal transduction system. BMC Microbiology 6.1 (2006): 1-16.
 828. Soffler, C. Development and characterization of caprine infection models of melioidosis, Doctoral Dissertation, Colorado State University, 2007.
 829. Soltan Dallal, M.M., S. Hidarzadeh, M. Azarsa, et al. Synergistic effect polymyxin B sulphate and trimethoprim on *Yersinia enterocolitica* and closely related species. J. Zanzan Univ. Med. Sci. Health Serv. 2012, 20(79).
 830. Sonne, L., D.L. Raymundo, F.M. Boabaid, M.R. Borba, G.G.M. Snel, M.J.P. Gomes and D. Driemeier. Systemic infection by *Yersinia enterocolitica* in chinchillas (*Chinchilla laniger*). Pesquisa Veterinaria Brasileira. 2012, 32(5), 379-382. ISSN:0100-736X.
 831. Sprague, L.D., and H. Neubauer. Melioidosis in animals: a review on epizootiology, diagnosis and clinical presentations. J. Vet. Med., 2004, B51, 305-320.
 832. Taye Kissi, J. (2010). Vaccine development against plague, glanders and melioidosis in the former Soviet Union in comparsion to the current state of global knowledge (Doctoral dissertation, Leipzig, Univ., Diss., 2009).
 833. Warner, Jeffrey Mitchell. The epidemiology of melioidosis in Papua New Guinea. (Doctoral Dissertation, James Cook University), 2004.
- 29). Iteman, I., NAJDENSKI, H., Carniel, E. High genomic polymorphism in *Yersinia pseudotuberculosis*. (1995) Contributions to microbiology and immunology, 13, 106-111.**
834. Carnoy C, Floquet S, Marceau M, et al. The superantigen gene ypm is located in an unstable chromosomal locus of *Yersinia pseudotuberculosis*. J. Bacteriol., 2002, 184, 16, 4489-4499.
 835. Fredriksson-Ahomaa, M. Epidemiology of human *Yersinia pseudotuberculosis* infection [Epidemiologic von *Yersinia pseudotuberculosis* Infektionen beim Menschen] (2009) Archiv fur Lebensmittelhygiene, 60 (2), pp. 82-87.
 836. Fredriksson-Ahomaa, M., Tracing of enteropathogenic *Yersinia*. *Yersinia: Systems Biology and Control*. Caister Academic Press, 2012. 201-216.
 837. Hallanvuori, S. Foodborne *Yersinia*: identification and molecular epidemiology of isolates from human infections. Doctoral dissertation. University of Helsinki, 2009.
 838. Hallanvuori, S. et al. Molecular epidemiology of the five recent outbreaks of *Yersinia pseudotuberculosis* in Finland. The Genus *Yersinia*. Springer, Boston, MA, 2004. 309-312.
 839. Laukkanen-Ninios, R., M. Fredriksson-Ahomaa, H. Korkeala. Enteropathogenic *Yersinia* in foods. Food Associated Pathogens. CRC Press, Taylor & Francis Group, 2013. 316-338.
 840. Matic I, Radman M, Taddei F, et al. Highly variable mutation rates in commensal and pathogenic *Escherichia coli*. Science, 1997, 277, 5333, 1833-1834.
 841. Metzgar D, Wills C. Evolutionary changes in mutation rates and spectra and their influence on the adaptation of pathogens. Microb. Infection, 2000, 2, 12, 1513-1522
 842. Niskanen T, Laukkanen R, Murros A, et al. Characterisation of non-pathogenic *Yersinia pseudotuberculosis*-like strains isolated from food and environmental samples. Intern. J. Food Microbiol., 2009, 129, 2, 150-156.
 843. Niskanen, T. Diagnostics and epidemiology of *Yersinia pseudotuberculosis*.

- Academic Dissertation, Department of Food and Environmental Hygiene, Faculty of Veterinary Medicine, University of Helsinki, Finland, 2010.
844. Niskanen, T., M. Fredriksson-Ahomaa, and H. Korkeala. *Yersinia pseudotuberculosis* with limited genetic diversity is a common finding in tonsils of fattening pigs. J. Food Protection, 2002, 65, 3, 540-545.
 845. Odaert M, Berche P, Simonet M. Molecular typing of *Yersinia pseudotuberculosis* by using an IS200-like element. J. Clin. Microbiol., 1996, 34, 9, 2231-2235.
 846. Sebbane, Florent, et al. "The Superantigen Gene."
 847. Voskresenskaya, E., et al. Typing and clustering of *Yersinia pseudotuberculosis* isolates by restriction fragment length polymorphism analysis using insertion sequences. J. Clin. Microbiol., 52.6 (2014): 1978-1989.
 848. Voskresenskaya E, Leclercq A, Tseneva G, et al. Evaluation of ribotyping as a tool for molecular typing of *Yersinia pseudotuberculosis* strains of worldwide origin. J. Clin. Microbiol., 2005, 43, 12, 6155-6160.
- 30). NAJDENSKI, H, Dimova, T, Zaharieva, MM, Nikolov, B, Petrova-Dinkova, G, Dalakchieva, S, Popov, K, Hristova-Nikolova, I, Zehtindjiev, P, Peev, S, Trifonova-Hristova, A, Carniel, E, Panferova, YA, Tokarevich, NK. Migratory birds along the mediterranean Black sea flyway as carriers of zoonotic pathogens. Canadian Journal of Microbiology, 64, 12, 2018, ISSN:0008-4166, DOI:10.1139/cjm-2017-0763, 915-924. SJR (Scopus):0.613, JCR-IF (Web of Science):1.55**
849. Ahrabi, S.Z., Akyildiz, G., Kar, S. and Keles, A.G., 2023. Detection of the Crimean–Congo Hemorrhagic Fever Virus genome in questing *Ixodes* spp. and *Haemaphysalis* spp. in the periurban forestry areas of Istanbul: has a new biorisk emerged?. Vector-Borne and Zoonotic Diseases, 23, 10, 528-536.
 850. Akyildiz, G. et al. High prevalence and different genotypes of Crimean-Congo Hemorrhagic Fever Virus genome in Questing unfed adult *Hyalomma marginatum* in Thrace, Turkey. Ticks and Tick-borne Diseases, (2020):101622.
 851. Akyıldız, G. (2022). İstanbul'da *Ixodes ricinus* türü kenelerde lyme borreliozis varlığı, yayılışları ve biyogüvenlik riskleri (Doctoral dissertation, Marmara Üniversitesi (Turkey)).
 852. Al-Humam, N.A., & SaadEldin, W. F. (2021). The little white egret (*Egretta garzetta*) as a potential source of multidrug-resistant avian pathogenic *E. coli*. Slovenian Veterinary Research/Slovenski Veterinarski Zbornik, 58.
 853. Bacak, E., Ozsemir, A.C., Akyildiz, G. et al. (2024). Bidirectional tick transport by migratory birds of the African-Western Palearctic flyway over Turkish Thrace: observation of the current situation and future projection. Parasitol. Res., 123(1), 37.
 854. Berti, S. (2024). Changes in the waterbirds migratory patterns and wintering areas in Europe in relation to climate change. Master thesis, Università Ca' Foscari Venezia.
 855. Gan, L., Cao, X., Wang, Y., et al., (2019). Carriage and potential long distance transmission of *Listeria monocytogenes* by migratory black-headed gulls in Dianchi Lake, Kunming. Emerging Microbes & Infections, 8(1), 1195-1204.
 856. Chen, T. et al. Characterization of *Listeria monocytogenes* isolated from wildlife in central New York." Vet. Med. Sci. (2022), 8, 3, 1319-1320.
 857. Ebani, V.V. et al. Molecular survey on the occurrence of avian haemosporidia,

- Coxiella burnetii* and *Francisella tularensis* in waterfowl from central Italy. *Int. J. Parasitol.: Parasites and Wildlife* 10 (2019): 87-92.
858. Eisenberg, Tobias, et al. Expanding the host range: infection of a reptilian host (*Furcifer pardalis*) by an atypical *Brucella* strain." *Antonie van Leeuwenhoek* 113.10 (2020): 1531-1537.
 859. Fair, J. M., Al-Hmoud, N., Alrwashdeh, M.M., et al. (2024). Transboundary determinants of avian zoonotic infectious diseases: challenges for strengthening research capacity and connecting surveillance networks. *Frontiers in Microbiology*, 15, 1341842.
 860. Gan, Lin, et al. Carriage and potential long distance transmission of *Listeria monocytogenes* by migratory black-headed gulls in Dianchi Lake, Kunming. *Emerging Microbes & Infections* 8.1 (2019):1195-1204.
 861. Gayle, A.A. Ai for early warning of seasonal infectious disease: shapely additive explanations improves prediction of extraordinary West Nile virus events in Europe. *medRxiv* (2020).
 862. Gayle, A.A. Artificial intelligence predicts and explains West Nile Virus risks across Europe: extraordinary outbreaks determined by climate and local factors. *MedRxiv*, 2020-08.
 863. Gayle, A.A. Explainable AI Unravels Local Factors Driving Extraordinary Outbreaks of West Nile Virus in Europe." *medRxiv* (2020).
 864. Islam, M.S. et al. Virulence determinants and multidrug resistance of *Escherichia coli* isolated from migratory birds. *Antibiotics* 10.2 (2021):190.
 865. Islam, M.S., Paul, A., Talukder, M. et al. (2021). Migratory birds travelling to Bangladesh are potential carriers of multi-drug resistant *Enterococcus* spp., *Salmonella* spp., and *Vibrio* spp. *Saudi J. Biol. Sci.*, 28(10), 5963-5970.
 866. Jamil, T., Akar, K., Erdenlig, S., et al.. Spatio-temporal distribution of Brucellosis in European terrestrial and marine wildlife species and its regional implications. *Microorganisms*, 2022, 10, 10 C7 – 1970.
 867. Kar, S., & Keles, A. G. (2021). eo17 Possible direct and human-mediated impact of climate change on tick populations in Turkey. In: *Climate, Ticks and Disease*, 115-124.
 868. Kar, S., Akyildiz, G., Guven, E. et al. (2021). Monthly infestation characteristics of ticks on cattle in Thrace, a Crimean Congo hemorrhagic fever-endemic area of Turkey. *Parasitol. Res.*, 120, 3395-3404.
 869. Kar, S., & Ergünay, K. (2021). eo66 The strange case of tick-borne. In: *Climate, Ticks and Disease*, 455-459..
 870. Kobuszevska, A., & Wysok, B. (2024). Pathogenic bacteria in free-living birds, and its public health significance. *Animals*, 14(6), 968.
 871. Kochanova, E., Nair, A., Sukhikh, N. et al. (2021). Patterns of cryptic diversity and phylogeography in four freshwater copepod crustaceans in European lakes. *Diversity*, 13(9), 448.
 872. Malik, Y. S., Arun Prince Milton, A., Ghatak, S. et al. (2021). Migratory birds and public health risks. *Role of Birds in Transmitting Zoonotic Pathogens*, 15-22.
 873. Melo, A.M., et al. Aspergillosis, avian species and the one health perspective: the possible importance of birds in azole resistance. *Microorganisms* 8.12 (2020): 2037.
 874. Olvera-Ramírez, A.M., McEwan, N.R., Stanley, K., Nava-Díaz, R. and Aguilar-Tipacamú, G., 2023. A systematic review on the role of wildlife as carriers and spreaders of *Campylobacter* spp. *Animals*, 13(8), p.1334.

875. Orf, G.S., Ahouidi, A.D., Mata, M., et al. (2024). Next-generation sequencing survey of acute febrile illness in Senegal (2020–2022). *Frontiers in Microbiology*, 15, 1362714.
876. Sauvala, M. et al. Hunted game birds—carriers of foodborne pathogens. *Food Microbiol.*, 98 (2021): 103768.
877. Schoder, D., Guldemann, C., Märklbauer, E. Asymptomatic carriage of *Listeria monocytogenes* by animals and humans and its impact on the food chain. *Foods*, 2022, 11, 21, 3472,
878. Smith, H.G., et al. Presence and antimicrobial resistance profiles of *Escherichia coli*, *Enterococcus* spp. and *Salmonella* sp. in 12 species of Australian shorebirds and terns. *Zoonoses and Public Health* (2022).
879. Smith, M.M. et al. A serologic survey of *Francisella tularensis* exposure in wildlife on the Arctic Coastal Plain of Alaska, USA. *J. Wildlife Diseases* 58.4 (2022): 746-55.
880. Smith, H. (2020). Wild Australian shorebirds as reservoirs of pathogenic bacteria and antimicrobial resistance. PhD thesis, Federation University Australia.
881. Smoglica, C., Graziosi, G., De Angelis, D., et al. (2024). Wild birds as drivers of *Salmonella* Braenderup and multidrug resistant bacteria in Wetlands of Northern Italy. *Transboundary and Emerging Diseases*, 2024.
882. Song, C. and Li, N., 2024. Dynamic analysis and bifurcation control of a delayed fractional-order eco-epidemiological migratory bird model with fear effect. *Int. J. Biomathemat.*, 17, 03, p.2350022.
883. Wareth, Gamal, et al. Susceptibility of avian species to *Brucella* infection: A hypothesis-driven study. *Pathogens* 9.2 (2020):77.
884. Wareth, G. et al. Susceptibility of avian species to *Brucella* infection: A hypothesis-driven study. *Pathogens*, 9.2 (2020):77.
885. Доронин, И. В., В. Р. Алексеев, and А. А. Котов. Морфологическое и молекулярно-генетическое разнообразие пресноводных гарпактицид (Crustacea: Соперода: Harpacticoida) Северо-Запада Евразии. 2020, PhD thesis, Saint Petersburg.
886. Панин, А.Л., Микробиологический мониторинг возбудителей сапрозоонозов в полярных регионах. (Doctoral Dissertation), Москва, 2023.
- 31). NAJDENSKI, H., Golkocheva, E., Kussovski, V., Ivanova, E., Manov, V., Iliev, M., Vesselinova, A., Skurnik, M. Experimental pig yersiniosis to assess attenuation of *Yersinia enterocolitica* O:8 mutant strains (2006) *FEMS Immunol. Med. Microbiol.*, 47 (3), 425-435.**
887. Axler-DiPerte, G. L. (2007). Identification and characterization of the *Yersinia enterocolitica* YtxR regulon (Doctoral dissertation, New York University).
888. Bartra, S.C.S. Outer membrane proteins of *Yersinia pestis* Ail and OmpA (2010) Institute of Microbiology, Umea University, Umea, Sweden, PhD Thesis.
889. Leibiger, R., Niedung, K., Geginat, G. et al. *Yersinia enterocolitica* Yop mutants as oral live carrier vaccines. (2008), *Vaccine*, 26 (51), 6664-6670.
890. Schaake, J., Kronshage, M., Uliczka, F., et al. Human and animal isolates of *Yersinia enterocolitica* show significant serotype-specific colonization and host-specific immune defense properties. *Infect. Immun.*, 2013, 11, 81, 4013-4025.
891. Schaake, J., Drees, A., Grüning, P., et al. Essential role of invasins for colonization and persistence of *Yersinia enterocolitica* in its natural reservoir host, the pig. *Inf. Immun.*,

- 2014, 82(3), 960-969.
892. Schesser Bartra, S. C. (2010). Outer membrane proteins of *Yersinia pestis*: Ail and OmpA (Doctoral dissertation, Institutionen för molekylärbiologi, Umeå Universitet).
893. Valentin-Weigand, P., Heesemann, J., & Dersch, P. Unique virulence properties of *Yersinia enterocolitica* O: 3—An emerging zoonotic pathogen using pigs as preferred reservoir host. *Int. J. Med. Microbiol.*, 304(7), 2014, 824-834.
894. Axler-DiPerte, Grace L. Identification and characterization of the *Yersinia enterocolitica* YtxR regulon. Diss. New York University, 2007.
- 32). De Rosa, S., Kamenarska, Z., Stefanov, K., Dimitrova-Konaklieva, S., NAJDENSKI, C., Tzevtkova, I., Ninova, V., Popov, S. Chemical composition of *Corallina mediterranea* Areschoug and *Corallina granifera* Ell. et Soland. *Zeitschrift fur Naturforschung - Section C Journal of Biosciences*, 2003, 58 (5-6), 325-332.**
895. Abd El-malek, Fady, et al. Polyhydroxyalkanoate nanoparticles produced by marine bacteria cultivated on cost effective Mediterranean algal hydrolysate media. *J. Biotechnol.*, 328 (2021): 95-105.
896. Borik, R.M. Volatile compounds extraction, fractionation and identification from the red alga *Corallina officinalis*. *World Appl. Sci. J.*, 2014, 30 (6), 741-746.
897. De Oliveira, A.L., Avaliação química e biológica de espécimens de *Bostrychia radicans* (Rhodomelaceae), Dissertação, Universidade de São Paulo, Faculdade de Ciências Farmacêuticas de Ribeirão Preto, 2009.
898. El Zawawy, Nessma A., et al. A novel study on the inhibitory effect of marine macroalgal extracts on hyphal growth and biofilm formation of candidemia isolates. *Sci. Reports* 10.1 (2020): 1-10.
899. Erickson, A.A., V.J. Paul, van Alstyne, K.L., L. M. Kwiatkowski. Palatability of macroalgae that use different types of chemical defenses. *J. Chem. Ecol.*, 2006, 32, 9.
900. Kumar, J., Dhar, P., Tayade, A.B. et al. Chemical composition and biological activities of trans-Himalayan alga *Spirogyra porticalis* (Muell.) Cleve *PLoS ONE* 2015, 10, e0118255
901. Labib, W., S. Hosny. Nitrogen and protein contents of *Ulva fasciata* and *Corallina officinalis* under environmental variations. *Egypt. J. Aquat. Biol. Fisheries* 24.3 (2020): 425-438.
902. Martínez, S., & Laurenis, Del V. Evaluación de los metabolitos secundarios y la actividad biológica del alga invasora *Caulerpa racemosa*. (Doctoral Dissertation, Universidad de Oriente Núcleo de Sucre. (2013).
903. Rauter, A.P., M.M. Filipe, C. Prata, et al. A new dihydroxysterol from the marine phytoplankton *Diacronema* sp. *Fitoterapia*, 2005, 76, 5, 433-438.
904. Takeara, R. et al. Antileukemic effects of *Didemnum psammatoedes* (Tunicata: Ascidiacea) constituents. *Compar. Biochem. Physiol. Part A: Molecular & Integrative Physiology* 151.3 (2008): 363-369.
905. Zivanovic A, Skropeta D. c-AMP dependent protein kinase A inhibitory activity of six algal extracts from South Eastern Australia and their fatty acid composition. *Nat. Prod. Commun.*, 2012, 7 (7), 2012, 923-926.
- 33). NAJDENSKI, H., S. Nikolova, A. Vesselinova, P. Nejkov. Studies on *Yersinia enterocolitica* O:3 experimental infection in pigs. *J. Vet. Med., B*, 45, 1998, 59-64.**

906. Arnold, T. Nachweis von *Salmonella* und *Yersinia enterocolitica* im persistent infizierten Schwein, (2002), Doctoral Dissertation, University of Leipzig.
907. Bartling C, Truyen U, Isa G, et al. Investigations on the prevalence of *Yersinia*-specific antibodies in cattle in Bavaria. Meeting of the Bacteriology and Mycology Working Group of the German-Vet.-Med.-Society, JUN 09-12, 2004 Berlin, Germany. Berliner und Munchener Tierärztliche Wochenschrift, 2004, 117, 11-12, 499-507.
908. Brugmann, M., M. Peters, J. Mumme. *Yersinia enterocolitica* – septicemia in an American Minipig. Deutsche Tierärztliche Wochenschrift, 2001, 108, 6, 257.
909. Caspari, K. Untersuchungen zum Vorkommen von humanpathogenen *Yersinia enterocolitica* bei Schlachtschweinen aus verschiedenen Haltungsformen. Dissertation, Aus dem Institut für Lebensmittelqualität und -sicherheit des Zentrums für Lebensmittelwissenschaften der Tierärztlichen Hochschule Hannover, 2005.
910. Esnault E., Rouaud A., Labbé A. et al. Controlled experimental infection in pigs with a strain of *Yersinia enterocolitica* harboring genetic markers for human pathogenicity: colonization and stability. Infect. Immun., 91 (7), 2023, e00157-23..
911. Fredriksson-Ahomaa, M., Molecular epidemiology of yadA-positive *Yersinia enterocolitica*. (2001), Doctoral Dissertation, Helsinki University.
912. Garthoff LH, Sobotka TJ. From farm to table to brain: Foodborne pathogen infection and the potential role of the neuro-immune-endocrine system in neurotoxic sequelae. Nutrit. Neuroscie., 2001, 4, 5, 333-374.
913. Hensel A, Nikolaou K, Bartling C, et al. On the prevalence of anti-yersinia outer protein antibodies in bavarian slaughter pigs: Berliner und Munchener Tierärztliche Wochenschrift. 2004, 117, 1-2, 30-38.
914. Louis, A.L. Untersuchungen zum Nachweis von *Yersinia enterocolitica* im Kot von Mastschweinen mittels Immunfluoreszenztest. Dissertation, Aus der Klinik für kleine Klauentiere und forensische Medizin und Ambulatorischen Klinik der Tierärztlichen Hochschule Hannover, 2005.
915. Neubauer H, Sprague LD, Scholz H, et al. *Yersinia enterocolitica* infections: Impact on animal health. Berliner und Munchener Tierärztliche Wochenschrift, 2001, 114, 1-2, 8-12.
916. Голкочева, Е. Мембранно свързаните протеини – фактор на специфичност в серологични изследвания при йерсиниозата. Дисертация за присъждане на образователната и научна степен “доктор”. София, 2003.
- 34). Georgieva, K., Popova, M., Dimitrova, L., Trusheva, B., Phuong, D. T. L., Lien, N. T. P., Najdenski H., Bankova, V.. Phytochemical analysis of Vietnamese propolis produced by the stingless bee *Lisotrigona cacciae*. 14, 4, PloS one, 2019, ISSN:19326203, DOI:10.1371/journal.pone.0216074, 1-13. ISI IF:2.776**
917. Abbas, M., S. Sukarsa. Type and size of pollen collected by *Tetragonula laeviceps* at various altitudes. Biodiversitas J. Biolog. Diversity 23.3 (2022).
918. Abdullah, N.A. et al. Phytochemicals, mineral contents, antioxidants, and antimicrobial activities of propolis produced by Brunei stingless bees *Geniotrigona thoracica*, *Heterotrigona itama*, and *Tetrigona binghami*. Saudi J. Biol. Sci., 27.11 (2020): 2902-2911.
919. Afata T. N., Dekebo A. Chemical composition and antimicrobial effect of Western Ethiopian propolis. 2023, Chemistry & Biodiversity, 20, 2, e202200922..

920. Afata, T.N. et al. Phytochemical investigation, physicochemical characterization, and antimicrobial activities of Ethiopian propolis. *Arabian J. Chem.*, 15.7 (2022): 103931.
921. Alawode, T.T., Lajide, L., Olaleye, M. et al. 2023. *Crinum jagus*: antiproliferative studies of extracts on HepG2 cell line and in silico assessment of phytoconstituents as potential inhibitors of p53–mortalin interaction. *Future J. Pharmaceut. Sci.*, 9(1), 1-15.
922. Alawode, T.T., Lajide, L., Olalye T., & Owolabi, B.J. (2022). Metabolite profiling of ethyl acetate stem extract of *solanum erianthum* for potential anticancer compounds. *Afr. J. Pharmacet. Res. Dev.*, 14, 60-66.
923. Alrashada, Y.N., Hassanien, H.A., Abbas, A.O. et al. (2023). Dietary propolis improves the growth performance, redox status, and immune response of Nile tilapia upon a cold-stress challenge. *Plos one*, 18(11), e0293727.
924. Asma, S.T., Acaroz, U., Bobiş, O. et al. (2024). 9 Propolis. In: *Honey Bees, Beekeeping and Bee Products*, CRC Press, 147-169.
925. Balderrama-Carmona, A.P., Ramos-García, V., Ruiz-López, L., & Felipe-Ortega-Fonseca, X. (2024). Propolis as a bioindicator of contamination with toxic metals. In: *Heavy Metal Remediation: Sustainable Nexus Approach* (pp. 289-317). Cham: Springer Nature Switzerland.
926. Batistuta, M. A., Aulia, A., & Kustiawan, P. M. (2021). Potensi aktivitas anti virus dari produk alami lebah kelulut. *Jurnal Farmasi Udayana*, 10(2), 144-148.
927. Belmehdi, O., El Menyiy, N., Bouyahya, A. et al., 2023. Recent advances in the chemical composition and biological activities of propolis. *Food Rev. Int.*, 39(9), 6078-6128.
928. Belmehdi, Omar, et al. "Recent advances in the chemical composition and biological activities of propolis. *Food Reviews International* (2022): 1-51.
929. Chuttong, B., Lim, K., Praphawilai, P. et al. (2023). Exploring the functional properties of propolis, geopropolis, and cerumen, with a special emphasis on their antimicrobial effects. *Foods*, 12(21), 3909.
930. de Almeida-Junior S., Ferraz M.V.F., de Oliveira A.R. et al. Advances in the phytochemical screening and biological potential of propolis. *Fundam Clin. Pharmacol.* 2023, 10.
931. Ebrahimzadeh, M.A. et al. Enhanced catalytic and antibacterial efficiency of biosynthesized *Convolvulus fruticosus* extract capped gold nanoparticles (CFE@ AuNPs). *J. Photochem. Photobiol. B: Biology* 209 (2020): 111949.
932. Hamouda, Sayed Mohammed, et al. "Apitherapy of Septic Metacarpal and Metatarsal Wounds (An Experimental Study on Donkeys)." *Clin. Med. Res.*, 8.4 (2019):77.
933. Hamzah, S.A., N. Zawawi, S Sabri. A Review on the association of bacteria with stingless bees. *Sains Malaysiana* 49.8 (2020):1853-1863.
934. Isidorov, V.A. et al. Chemical composition and biological activity of argentinian propolis of four species of stingless bees. *Molecules* 27.22 (2022): 7686.
935. Kegode, T.M., Ndungu, N. and Kiatoko, N., 2023. Determination of total flavonoids, phenolics, and antioxidant activity of propolis from six stingless bee species in Kenya. *JSFA Reports*, 3(9), 441-446.
936. Kegode, T.M. et al. Phytochemical composition and bio-functional properties of *Apis mellifera* propolis from Kenya. *Royal Society open science* 9.7 (2022):211214.
937. Kegode, T.M., & Lattorff, H.M.G. (2022). The effects of hive materials on phytochemical and biological properties of honeybee propolis. *JSFA Reports*, 2(10), 466-471.

938. Kitamura, H. Effects of propolis extract and propolis-derived compounds on obesity and diabetes: knowledge from cellular and animal models. *Molecules* 24.23 (2019):4394.
939. Kung, Y.C. et al. Alkaline hydrolysis and discrimination of propolis at different pH values using high throughput 2D IR spectroscopy and LC-MS/MS." *Biores. Technol., Rep.*, 18 (2022):101120.
940. Le Nguyen T., Nguyen A. T., Vu Thi K. O., Vu Thi H., Nguyen Tu M., Nguyen Phan N. M., Truong Vinh X., Diep Thi L. P., Nguyen Thi P. L. Chemical constituents and cytotoxicity of *Lepidotrigona ventralis* propolis. *Vietnam Journal of Chemistry*, vol. 61, 1, 47-79. 2023 Feb.
941. Li, M.M., J. Lu, Y. Deng. Dracaenone, a novel type of homoisoflavone: natural source, biological activity and chemical synthesis. *Curr. Org. Chem.*, 26.9 (2022):887-897.
942. Llave, A. L. A. R. Evaluation of the anticarcinogenic potential of propolis ethanolic extract (PEE) from Philippine stingless bee (*Tetragonula biroi* Friese). *Int. J. Sci. Eng.Sci.*, 2024, 8, 2, 93-104.
943. Maragou, N.C., Strati, I.F., Gialouris, P.L. et al. (2023). Honey and bee products. In: *Emerging Food Authentication Methodologies Using GC/MS* (pp. 137-213). Cham: Springer International Publishing.
944. Mohiuddin I, Kumar TR, Zargar MI, et al. GC-MS analysis, phytochemical screening, and antibacterial activity of *Cerana indica* propolis from Kashmir Region. *Separations*. 2022; 9(11):363.
945. Negri, G., Salatino, A., Pereira, L.L.R. et al. 2022, A highly complex stingless bee propolis: Composition and influence of the period of collection. *JSFA Reports*, 2(2), 64-80.
946. Ngaini, Z., Hussain, H., Kelabo, E. et al. (2023). Chemical profiling, biological properties and environmental contaminants of stingless bee honey and propolis. *J. Apicult. Res.*. 62, 1, 131-147.
947. Nguyen, H.X., Nguyen, M.T.T., & Nguyen, N.T. (2023). Some epoxy lignans from the Vietnamese propolis of stingless bee *Trigona minor* and their bioactivities. *Vietnam J. Chem.*, 61, 94-100.
948. Phuong Thao, N., Minh, N.T., Huyen, T.K. et al. (2021). Chemical constituents and antimicrobial activity of *Lisotrigona cacciae* propolis collected in Hoa Binh Province. *Vietnam J. Sci. Technol. Eng.*, 63(2), 70-73..
949. Prashanthi, G., & Sujatha, K. (2023). GC MS-analysis of bio active compounds from propolis and antibacterial activity against *B. cereus* isolated from Tasar Silkworm Cadavers. *Bulletin of Pure & Applied Sciences-Botany*, 42, 2..
950. Purwaningsih Y., Masduqi, A.F., Indriyanti, E. et al. Photoprotective and antioxidant potential of Indonesia's klanceng honey beehive waste. *Sci. Technol.Indonesia*. 8, 1, 137 - 143, 2023.
951. Rivera-Yañez N., Ruiz-Hurtado P.A., Rivera-Yañez C.R. et al. The role of propolis as a natural product with potential gastric cancer treatment properties: a systematic review. *Foods*. 2023, 16;12(2):415.
952. Rozman, A.S. et al. A comprehensive review of stingless bee products: phytochemical composition and beneficial properties of honey, propolis, and pollen. *Appl. Sci.*, 12.13 (2022): 6370.
953. Ruiz Ruiz, J.C., Pacheco López, N.A., Rejón Méndez, E.G. et al. Phenolic content and bioactivity as geographical classifiers of propolis from stingless bees in Southeastern

Mexico. Foods 2023, 12, 1434.

954. Saenab, A.. Chemical and Biological Characteristics of Anacardic acid as a Bioactive Compound and Its Application on Ruminant. WARTAZOA Indonesian Bulletin of Animal and Veterinary Sciences, 2022, 32, 3
955. Salleh, S.N.A.S. et al. A comprehensive review on chemical compounds, biological actions and potential health benefits of stingless bee propolis. Sains Malaysiana 51.3 (2022): 733-745.
956. Salleh, S.N.A.S., Hanapiah, N.A.M., Ahmad, H. et al. (2021). Determination of total phenolics, flavonoids, and antioxidant activity and GC-MS analysis of Malaysian stingless bee propolis water extracts. Scientifica, 2021..
957. Sankaran, S., Dubey, R., Lohidasan, S. Optimization of extraction conditions using response surface methodology and HPTLC fingerprinting analysis of Indian propolis. J. Biol. Active Products from Nature, 13, 2023.
958. Shirzadi-Ahodshti, M. et al. Biogenic synthesis of spherical-shaped noble metal nanoparticles using *Vicia faba* extract (X@ VF, X = Au, Ag) for photocatalytic degradation of organic hazardous dye and their in vitro antifungal, antibacterial and anticancer activities. Inorganic Chem. Communicat., 146 (2022):110042.
959. Shirzadi-Ahodshti, M. et al. Facile and eco-benign synthesis of a novel MnFe₂O₄@ SiO₂@ Au magnetic nanocomposite with antibacterial properties and enhanced photocatalytic activity under UV and visible-light irradiations. Appl.Organomet. Chem., 34.5 (2020): e5614.
960. Shirzadi-Ahodshti, M., S. Mortazavi-Derazkola, M.A. Ebrahimzadeh. Biosynthesis of noble metal nanoparticles using crataegus monogyna leaf extract (CML@ X-NPs, X= Ag, Au): Antibacterial and cytotoxic activities against breast and gastric cancer cell lines. Surfaces and Interfaces 21 (2020):100697.
961. Syahariza, Z.A., & Kee, L. S. (2022). Antioxidant activity of stingless bee propolis using different extraction method. Int. J. Eng. Advanc. Res., 4(4), 1-15.
962. Syaifie, P.H. et al. Computational study of asian propolis compounds as potential anti-type 2 diabetes mellitus agents by using inverse virtual screening with the DIA-DB web server, tanimoto similarity analysis, and molecular dynamic simulation." Molecules 27.13 (2022):3972.
963. Thai, P.H., Ngoc, P.T., & Le Thuong, H.T. Studying the biological characteristics of the stingless bee *Tetragonilla colina* Smith 1857 (Apidae:Melipona) in Hanoi, Vietnam. Vietnam J. Agricult. Sci., 2021, 4, 2, 1067-1076..
964. Tran, T.D. et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020):4988.
965. Zin, N.B.M., et al., HPTLS fingerprinting coupled with chemometric analysis for evaluation of different extraction methods on stingless bee's propolis. Bioscience Research, 2022, 19, 211-223..
966. Zulhendri, F. et al. Propolis of stingless bees for the development of novel functional food and nutraceutical ingredients: A systematic scoping review of the experimental evidence. J. Funct. Foods, 88 (2022): 104902.
967. Wina, E., dan A Saenab. Chemical and biological characteristics of anacardic acid as a bioactive compound and its application on ruminant. WARTAZOA, 32, 3 2022, 165-176
- 35). Chochkova, M. G., Chorbadzhiyska, E. Y., Ivanova, G. I., Najdenski, H., Ninova,**

M., Milkova, T. S. Antimicrobial and Radical Scavenging Activities of N-Hydroxycinnamoyl – L-Cysteine and - L-Proline Ethyl Esters. 2, The Natural Products Journal, 2012, 50-54. ISI IF:0.7

968. Herrera-R,A., Castrillón, W., Pastrana, M. et al. (2021). Promising hybrids derived from S-allylcysteine and NSAIDs fragments against colorectal cancer: Synthesis, in-vitro evaluation, drug-likeness and in-silico adme/tox studies. *Iranian J. Pharmaceut. Res.*, 20(3), 351.
 969. Kwak, S.Y. et al. Chemical modulation of bioactive compounds via oligopeptide or amino acid conjugation. *Peptide Sci.*, 100.6 (2013):584-591.
 970. Machado, I.M. Synthesis of catecholic aminoacids and peptides or conjugates of catechols with amino acids. Doctoral dissertation, 2018, University of Minho, Portugal.
 971. Monteiro, L.S. et al. An efficient one-pot synthesis of polyphenolic amino acids and evaluation of their radical-scavenging activity. *Bioorg. Chem.*, 89 (2019): 102983.
 972. Monteiro, L.S. et al. Synthesis and preliminary biological evaluation of new phenolic and catecholic dehydroamino acid derivatives. *Tetrahedron*, 73.43 (2017):6199-6209.
 973. Monteiro, L.S., & Paiva-Martins, F. (2022). Amino acids, amino acid derivatives and peptides as antioxidants. In: *Lipid Oxidation in Food and Biological Systems: A Physical Chemistry Perspective* (pp. 381-404). Cham: Springer International Publishing.
 974. Neelam, Ahlawat, S., Shankar, A. et al. Bioevaluation and molecular docking analysis of novel phenylpropanoid derivatives as potent food preservative and anti-microbials. *3 Biotech* 11, 70 (2021).
 975. Noumi, E. G.A.T. Tiam. Floristic inventory of woody species of the Oku Sacred forest in the North-West Cameroon, theoretical and philosophical approach. *Int. J. Curr. Res. Biosci. Plant Biol* 3.1 (2016): 66-91.
 976. Ahlawat, Shruti, et al. "Bioevaluation and molecular docking analysis of novel phenylpropanoid derivatives as potent food preservative and anti-microbials." *3 Biotech* 11.2 (2021): 1-10.
 977. Machado, Inês Marques. Synthesis of catecholic amino acids and peptides or conjugates of catechols with amino acids and peptides. Diss. 2018.
 978. Zenta, F. et al. Development trans-N-benzyl hydroxyl cinnamamide based compounds from cinnamic acids and characteristics anticancer potency. *J. Iranian Chem. Soc.*, (2022):1-9.
- 36). Popova, M., Lazarova, H., Trusheva, B., Popova, M., Bankova, V., Mihály, J., Najdenski H.,, Tsvetkova, I., Szegedi, Á.. Nanostructured silver silica materials as potential propolis carriers. 263, 2018, 28-33. ISI IF:3.649**
979. Bernardo, M.P., et al. Innovations in antimicrobial engineered nanomaterials. *Adv. Nanostruct. Materials Environm. Remediation*. Springer, Cham, 2019. 253-277.
 980. Ceylan, O., .H Karakus, H. Cicek. Design and in vitro antibiofilm activity of propolis diffusion-controlled biopolymers. *Biotechnol. Appl. Biochem.*, 2021, 68, 4, 789-800.
 981. Chien, K.Z., Kavin, T., Jia, H. et al. Drug delivery approaches to improve the efficiency of phytoderivatives against UV induced damage - A review. *J. Drug Deliv. Sci. Technol.*, 87, 2023, 104793.
 982. Correa-González, Y.X., M.A. Rojas-Cardozo, C.E. Mora-Huertas. Potentialities of the Colombian propolis in pharmaceuticals and cosmetics: A standpoint from the quality control. *Revista Colombiana de Ciencias Químico-Farmacéuticas* 48.3 (2019): 762-788.

983. Das T.K., Ganguly S., Bhawal P. et al. A facile green synthesis of silver nanoparticles decorated silica nanocomposites using mussel inspired polydopamine chemistry and assessment its catalytic activity. *J. Environ. Chem. Eng.*, 6 (6), 6989-7001, 2018.
984. Dias, R.T.A. Desenvolvimento de sistemas visando o tratamento de lesões cutâneas à base de fibras de PLA/PEG e própolis vermelha produzidas por solution blow spinning. 2019, Doctoral dissertation, Universidade Federal da Paraíba, Brasil.
985. Ikeda, N.Y. Evaluation of the residues from the ethanolic extraction of organic propolis as a source of biological compounds. PhD thesis, Universidade de São Paulo.
986. Melendez-Rodriguez, B, et al. Electrospun antimicrobial films of poly (3-hydroxybutyrate-co-3-hydroxyvalerate) containing eugenol essential oil encapsulated in mesoporous silica nanoparticles. *Nanomaterials*, 9.2 (2019):227.
987. Mendez-Pfeiffer, P., Juarez, J., Hernandez, J. et al. (2021). Nanocarriers as drug delivery systems for propolis: A therapeutic approach. *J. Drug Deliv. Sci. Technol.*, 65, 102762.
988. Oliveira, L.F.A.D.M. (2021). Nanopartículas de sílica mesoporosa carregadas com extrato de própolis vermelha: síntese, caracterização e avaliação in vitro da atividade antioxidante, antimicrobiana e antiviral. Doctoral Dissertation, Universidade Federal de Alagoas, Brasil.
989. Remedio, L. N. (2022). Desenvolvimento de filmes de desintegração oral como veículo de extrato etanólico de própolis verde utilizando-se a técnica de impressão (Doctoral dissertation, Universidade de São Paulo).
990. Remedio L.N., Garcia V.A.D.S., Rochetti A.L. et al. Hydroxypropyl methylcellulose orally disintegration films produced by tape casting with the incorporation of green propolis ethanolic extract using the printing technique. *Food Hydrocoll.*, 135, 2023, 108176.
991. Syukri, Y.A.N.D.I., et al. Fabrication of propolis self-nano emulsifying using virgin coconut oil as lipid-based vehicle. *Adv. Mater. Res.*, 2021, 1162, 119-128..
992. Гребенникова, Т.А., В.В. Трошина, Ж.Е. Белая. Маркеры и генетические предикторы остеопороза в рутинной клинической практике. *Consilium Medicum* 21.4 (2019).
- 37). Gacheva G., Gigova L., Ivanova N., Iliev I., Toshkova R., Gardeva E., Kussovski V., NAJDENSKI H. Suboptimal growth temperatures enhance the biological activity of cultured cyanobacterium *Gloeocapsa* sp. *Journal of Applied Phycology*, 1, 25, 2013, 183-194, ISSN: 0921-8971. IF 2.326**
993. Adar, O., Kaplan-Levy, R.N., Schönhözl, M. et al. Optimal temperature and salinity for growth of two Cyanobacteria from Hammat Gader hot springs. *Nedev, Dead Sea and Arava Studies*, 2017, 9, 14, 128-136..
994. Balu, S. et al. Polycyclic aromatic hydrocarbon sequestration by intertidal phototrophic biofilms cultivated in hydrophobic and hydrophilic biofilm-promoting culture vessels. *J. Hazard. Mater.*, (2022): 129318.
995. Deutsch, Y. et al. Endophytes from algae, a potential source for new biologically active metabolites for disease management in aquaculture. *Front. Marine Sci.*, 8 (2021): 333.
996. do Amaral S.C., Xavier L.P., Vasconcelos V. et al. A promising source of antifungal metabolites. *Marine Drugs*, 21 (6), art. no. 359, 2023.
997. Emalya, N., Yunardi, Y., Munawar, E. et al. (2024). Synergistic removal of organic and nutrients from landfill leachate using photobioreactor-cultivated microalgae-bacteria consortium. *Global J. Environm. Sci. Management*, 10(2), 683-698.

998. Georgiev, Y.N., T.G. Batsalova, B.M. Dzhambazov. Immunomodulating polysaccharide complexes and antioxidant metabolites from *Anabaena laxa*, *Oscillatoria limosa* and *Phormidesmis molle*. *Algal Research*, 60, 2021, 102538
999. Gong, W. et al. Extracellular polymeric substances produced by the thermophilic cyanobacterium *Gloeocapsa gelatinosa*: Characterization and assessment of their antioxidant and metal-chelating activities. *Marine Drugs* 20.4 (2022): 227.
1000. Heydarizadeh, P., Poirier, I., Loizeau, D., et al. Plastids of marine phytoplankton produce bioactive pigments and lipids. *Marine Drugs*, 2013, 9, 11, 3425-3471.
1001. Leão, P.N., Ramos, V., Gonçalves, P.B., et al. Chemoecological screening reveals high bioactivity in diverse culturable Portuguese marine cyanobacteria. *Marine Drugs*, 4, 11, 2013, 1316-1335.
1002. Lu, Y.-Z., et al. Research progress of pharmacological active substances from diazotrophic cyanobacteria, *Chinese Tradit. Herbal Drugs*, 49.18, (2018): 4453-4460.
1003. Maruthanayagam, V., et al. Effects of surface material on growth pattern and bioactive exopolymers production of intertidal cyanobacteria *Phormidium* sp. *Indian J. Geo-Marine Sci.*, 49(10), (2020), 1669-1677.
1004. Montero, X. Á. (2017). Modulación de la producción y caracterización estructural de los exopolisacáridos en cianobacterias diazotróficas, y estudio de su utilización para el tratamiento del digestato líquido de la digestión anaeróbica de efluentes de una procesadora de pescado (Doctoral dissertation, Universidade de Santiago de Compostela).
1005. Mota, R., Lima, R. T., Flores, C. et al. Assessing the antitumor potential of variants of the extracellular carbohydrate polymer from *Synechocystis* Δ sigF mutant. *Polymers*, 15(6), 1382.
1006. Mudimu, O., N. Rybalka, T. Bauersachs, et al. Biotechnological screening of microalgal and cyanobacterial strains for biogas production and antibacterial and antifungal effects. *Metabolites*, 2014, 4, 2, 373-393.
1007. Ördög, V. Mikroalgák biotechnológiai alkalmazása a növénytermesztésben és növényvédelemben. Doctoral Dissertation, Nyugat-magyarországi Egyetem, 2015.
1008. Stirk, W.A., J. van Staden. Bioprospecting for bioactive compounds in microalgae: Antimicrobial compounds. *Biotechnol. Advances* (2022): 107977.
1009. Veerabhadran, M., et al. Effects of flask configuration on biofilm growth and metabolites of intertidal Cyanobacteria isolated from a mangrove forest." *Journal of applied microbiology* 125.1 (2018): 190-202.
- 38). Vesselinova A., NAJDENSKI H., Nikolova S., Wesselinova D.. Arthritis after Experimental Infection with *Yersinia enterocolitica* 0:3 in Rabbits. *Journal of Veterinary Medicine, Series B*, 48, Wiley-Blackwell Publishing Ltd, 2001, DOI:<https://doi.org/10.1111/j.1439-0450.2001.00422.x>, 43-53**
1010. Dalal, M.M.S., M.T. Akhi, A.R.P. Azar. Study of *Yersinia enterocolitica* in acute children diarrhea under fourteen years old. *Med. J. Tabriz Univ. Med. Sci. Health Services* 30.4 (2009): 49-52.
1011. Sonne, L et al. Systemic infection by *Yersinia enterocolitica* in chinchillas (*Chinchilla laniger*). *Pesquisa Veterinária Brasileira* 32.5 (2012): 379-382.
1012. Wang, X. et al. *Yersinia*. In: *Laboratory Models for Foodborne Infections*. CRC Press, 2017. 427-437.

1013. Голкочева, Е. Мембраносвързаните протеини- фактор на специфичност в серологични изследвания при йерсиниозата. Дисертация за присъждане на образователната и научна степен "доктор". София, 2003.

39). Veljanov, D., A. Vesselinova, S. Nikolova, V. Kussovski, H. NAJDENSKI. Experimental infection with *Yersinia pseudotuberculosis* of ground squirrels (*Citellus Citellus*). J. Vet. Med. B 40, 1993, 589-596.

1014. Голкочева, Е. Мембраносвързаните протеини- фактор на специфичност в серологични изследвания при йерсиниозата. Дисертация за присъждане на образователната и научна степен "доктор". София, 2003.

1015. Шурыгина, И.А. (2003). Морфологические проявления псевдотуберкулеза. In: Псевдотуберкулез (pp. 81-108).

1016. Шурыгина, И.А. Патогенетические механизмы формирования псевдотуберкулеза, вызванного возбудителями с различным плазмидным спектром (экспериментально-клиническое исследование). Doctoral Dissertation, Восточно-Сибирский научный центр СО РАМН, 2004.

40). Golkocheva-Markova E, Christova I, Stoilov R, NAJDENSKI H. Cross-reaction between *Yersinia* outer membrane proteins and anti-*Borrelia* antibodies in sera of patients with Lyme disease. Clin Microbiol Infect 2008; 14:873– 875.

1017. Barbour, V., J. Clark, L. Peiperl, E. Veitch. Tropical and travel-associated diseases. Med Hyg, 2008, 8, 533.

1018. Carter, J.D. Bacterial agents in spondyloarthritis: a destiny from diversity?. Best Practice & Research Clinical Rheumatology 24.5 (2010):701-714.

1019. Duránte Fernánde, C. Desarrollo y utilidad de las técnicas de ELISA y quimioluminiscencia para el diagnóstico de la tularemia humana. 2019, PhD thesis, Universidad de Valladolid.

1020. Fang X., Kang L., Qiu Y.-F. et al. *Yersinia enterocolitica* in Crohn's disease. Frontiers in Cellular and Infection Microbiology, 13, art. no. 1129996, 2023.

1021. Grażewska, W. Holec-Gąsior, L., 2023. Antibody cross-reactivity in serodiagnosis of Lyme disease. antibodies, 12(4), p.63.

1022. Joyner, G., Mavin, S., Milner, R., Lim, C. Introduction of IgM testing for the diagnosis of acute Lyme borreliosis: a study of the benefits, limitations and costs, Europ. J. Clin. Microbiol. Inf. Dis., 2022, 41(4), 671-675.

1023. Ringwood, T., et al. Current evidence for human yersiniosis in Ireland. Eur. J. Clin. Microbiol. & Inf. Dis., 31.11 (2012): 2969-2981.

1024. Ringwood, T. Epidemiology and comparative analysis of *Yersinia* in Ireland. Doctoral Dissertation. University College Cork, 2013.

1025. Silva, Â.A. O. et al. Performance of *Treponema pallidum* recombinant proteins in the serological diagnosis of syphilis. PloS one 15.6 (2020):e0234043.

1026. Wielkoszynski, T. et al. Novel diagnostic ELISA test for discrimination between infections with *Yersinia enterocolitica* and *Yersinia pseudotuberculosis*. Europ. J. Clin. Microbiol. & Inf. Dis., 37.12 (2018): 2301-2306.

1027. Паунова-Кръстева, Ц.С. Фенотипни вариации свързани с полизахаридните антигени при *Escherichia coli* O157. Дисертация за присъждане на образователната и научна степен „доктор”. София 2015.

1028. Симакова, ДИ. Конструирование видоспецифического антигенного полимерного препарата для серологической диагностики псевдотуберкулеза. Doctoral Dissertation. Ростовский-на-Дону научно-исследовательский противочумный институт Роспотребнадзора, 2019.

41). Nikolova, S., NAJDENSKI, H., Wesselinova, D., Vesselinova, A., Kazatchka, D., Neikov, P. Immunologic and electronmicroscopic studies on *Yersinia enterocolitica* O:3 infected pigs. Zbl. Bact., 1997, 286, 503-510.

1029. Brüggmann, M., M. Peters, and J. Mumme. "Case report: *Yersinia enterocolitica* septicemia in an American minipig." DTW. Deutsche Tierärztliche Wochenschrift 108.6 (2001): 257-260.

1030. Carniel, E., I Autenrieth, G. Cornelis, et al. *Y. enterocolitica* and *Y. pseudotuberculosis*. In: Prokaryotes, Springer, 2006, 270-398.

1031. Perry, M., A. Whyte. Immunology of the tonsils. Immunol. Today, 1998,9, 414-421.

1032. Rakin, A., Garzetti, D., Bouabe, H., & Sprague, L. D. (2015). *Yersinia enterocolitica*. In: Molecular medical microbiology (pp. 1319-1344). Academic Press.

1033. Robins-Browne, E.L. Hartland. *Yersinia* species. In: International Handbook of Foodborne Pathogens. 2003, CRC Press.

1034. Бениова, С.Н. Органные поражения у детей, больных псевдотуберкулезом, в динамике болезни: клиника, диагностика, патогенез, исходы. Doctoral Dissertation, Дальневосточный государственный медицинский университет, 2003.

1035. Голкочева, Е. Мембранносвързаните протеини – фактор на специфичност в серологични изследвания при йерсиниозата. Дисертация за присъждане на образователната и научна степен “доктор”. София, 2003.

1036. Железникова, Г.Ф. et al. Иммунный ответ при остром псевдотуберкулезе у детей." Медицинская иммунология 4.1, 2002, 45-58.

42). Scheufele, FB, Hinterholz, CL, Zaharieva, MM, NAJDENSKI, H, Módenes, AN, Trigueros, DEG, Borba, C, Espinoza-Quiñones, FR, Kroumov, AD. Complex mathematical analysis of photobioreactor system. Engineering in Life Sciences, John Wiley & Sons Ltd., 2019, ISSN:1618-2863, DOI:10.1002/elsc.201800044, 1-14. SJR (Scopus):0.56, JCR-IF (Web of Science):1.934

1037. Ahmad, D., Ghani, I.F.A., Yaacob, N.S. et al. Experiment and kinetic modeling of soil extract effects on *Dunaliella primolecta* growth. Res. Square, 2021.

1038. Alonso-Bastida, A., Franco-Nava, M.A., Adam-Medina, M. et al. Mathematical modeling of thermal interactions in a self-cooling pilot-scale photobioreactor. Case Studies in Thermal Engineering, 2022, 31 C7 – 101825.

1039. Chaouki, J., R. Sotudeh-Gharebagh. Scale-Up Processes: Iterative Methods for the Chemical, Mineral and Biological Industries. 2021, pp.350.

1040. Gonçalves, V.D. et al. Combination of light emitting diodes (LEDs) for photostimulation of carotenoids and chlorophylls synthesis in *Tetradismus* sp." Algal Res., 43 (2019):101649.

1041. Hinterholz, C. L. (2018). Desenvolvimento de um fotobiorreator de placas planas para cultivo de microalgas com alta densidade celular. PhD thesis, University of Toledo.

1042. Karam, A.L. et al. Chlorophyll a and non-pigmented biomass are sufficient predictors for estimating light attenuation during cultivation of *Dunaliella viridis*. Algal Res., 55

- (2021):102283.
1043. Nastiti, K.D., A. Rahman. Multiobjective optimization of synechocytis culture in flat-plate photobioreactor toward optimal growth and exergy. *Journal of Physics: Conference Series*. Vol. 1858. No. 1. IOP Publishing, 2021.
 1044. Rakhmanov, Sh. et al. 2021 IOP Conf. Ser.: Earth Environ. Sci., 939, 012054
 1045. Rodríguez-Zuñiga, D., Méndez-Zavala, A., Solís-Quiroz, O. et al. 2023. Microalgae as cell factories for biofuel and bioenergetic precursor molecules. *Plants as Bioreactors for Industrial Molecules*, pp.299-316.
 1046. Schediwy, K. et al. Microalgal kinetics - a guideline for photobioreactor design and process development. *Engineer. Life Sci.*, 2019, 19(12), 830-843.
 1047. Ugya, A.Y., Meguellati, K. Microalgae biomass modelling and optimisation for sustainable biotechnology - a concise review. *J. Ecol. Eng.*, 2022, 23, 9, p. 309-318,
 1048. Vasile, N.S., et al. Computational analysis of dynamic light exposure of unicellular algal cells in a flat-panel photobioreactor to support light-induced CO₂ bioprocess development. *Frontiers in Microbiology* 12 (2021).
 1049. Yuvraj, Padmanabhan, P. Improvements in conventional modeling practices for effective simulation and understanding of microalgal growth in photobioreactors: an experimental study. *Biotechnol. Bioproc.*, E 26, 483–500 (2021).
 1050. . Zhang Ting, Feng Aiguo, Jiang Nan, & Liu Chunjiang. (2023). Experimental study on the growth characteristics of *Chlorella* under flow conditions. *Experimental Fluid Mechanics*, 38, 1-9.
- 43). Tokarevich, NK, Panferova, YA, Freylikhman, OA, Blinova, OV, Medvedev, SG, Mironov, SV, Grigoryeva, LA, Tretyakov, KA, Dimova, T, Zaharieva, MM, Nikolov, B, Zehtindjiev, P, NAJDENSKI, H. *Coxiella burnetii* in ticks and wild birds. *Ticks and Tick-borne Diseases*, 10, 2, Elsevier, 2019, ISSN:1877959X, DOI:10.1016/j.ttbdis.2018.11.020, 377-385. SJR (Scopus):1.182, JCR-IF (Web of Science):2.749**
1051. Abreu, Daniel Paiva Barros de. Frugal innovation applied on the development of molecular tools for the bioecological study of ixodid ticks and associated *Rickettsia* spp. bacteria in Brazilian territor. 2019, Doctoral Dissertation, Universidade Federal Rural do Rio de Janeiro.
 1052. Balti, G., Galon, C., Derghal, M. et al. (2021). *Atelerix algirus*, the North African Hedgehog: suitable wild host for infected ticks and fleas and reservoir of vector-borne pathogens in Tunisia. *Pathogens*, 10(8), 953.
 1053. Borawski, K. et al. *Coxiella burnetii* and Q fever-a review. *Przegląd Epidemiologiczny* (2020): 43.
 1054. Cardona-Romero, M. et al. Seroprevalence and detection of *Rickettsia* spp. in wild birds of Arauca, Orinoquia region, Colombia. *Vet. Parasitol.: Regional Studies and Reports* 30 (2022):100720.
 1055. Celina, S. S., Cerný, J. *Coxiella burnetii* in ticks, livestock, pets and wildlife: A mini-review. *Front. Vet. Sci.*, 2022, 9 C7 – 1068129.
 1056. Devaux, Christian A., et al. "*Coxiella burnetii* in dromedary camels (*Camelus dromedarius*): a possible threat for humans and livestock in North Africa and the Near and Middle East?." *Frontiers in Veterinary Science* 7 (2020).

1057. Ebani, V.V., Mancianti, F. Potential role of birds in the epidemiology of *Coxiella burnetii*, *Coxiella*-like agents and *Hepatozoon* spp. *Pathogens*, 2022, 11, 3 C7 – 298.
1058. Ebani, V.V., Guardone, L., Bertelloni, F. et al. (2021). Survey on the presence of bacterial and parasitic zoonotic agents in the feces of wild birds. *Vet. Sci.*, 8(9), 171.
1059. Ebani, V. V. (2023). *Coxiella burnetii* Infection in Cats. *Pathogens*, 12(12), 1415.
1060. Epelboin, L., De Souza Ribeiro Mioni, M., Couesnon, A., et al. 2023. *Coxiella burnetii* infection in livestock, pets, wildlife, and ticks in Latin America and the Caribbean: a comprehensive review of the literature. *Curr. Trop. Med. Reports*, pp.1-44.
1061. Frangoulidis, D., Kahlhofer, C., Said, A.S. et al. (2021). High prevalence and new genotype of *Coxiella burnetii* in ticks infesting camels in Somalia. *Pathogens*, 10(6), 741.
1062. Fu, M., He, P., OuYang, X. et al. Novel genotypes of *Coxiella burnetii* circulating in rats in Yunnan Province, China. *BMC Vet. Res.*, 2022, 18, 1 C7 – 204.
1063. Gardner, B.R., Hufschmid, J., Stenos, J. et al., 2023. Pacific Gulls (*Larus pacificus*) as potential vectors of *Coxiella burnetii* in an Australian Fur Seal breeding colony. *Pathogens*, 12(1), p.122.
1064. Grostieta, E., Zazueta-Islas, H.M., Cruz-Valdez, T. et al. Molecular detection of *Coxiella*-like endosymbionts and absence of *Coxiella burnetii* in *Amblyomma mixtum* from Veracruz, Mexico. *Exp. Appl. Acarology*, 2022, 88, 1, 113-125.
1065. He Peisheng, Yuan Qinghong, Zhang Shan, Qin Qingqing, Wan Weiqiang, Li Danny, ... & Xiong Xiaolu. (2023). Genotyping study on Yunnan Yulong rodents carrying *Coxiella burnetii*. *Parasites and Medicine Acta Entomologica Sinica*, 30(01), 10-17.
1066. Keve, G., A. Sándor, S. Hornok. Hard ticks (Acari: Ixodidae) associated with birds in Europe: Review of literature data. *Front. Vet. Sci.*, 9 (2022).
1067. Körner, S. et al. The prevalence of *Coxiella burnetii* in hard ticks in europe and their role in q fever transmission revisited - A systematic review. *Front. Vet. Sci.*, 8 (2021).
1068. Labbé Sandelin, L. *Neoehrlichia mikurensis* in Sweden : An emerging tick-borne human pathogen. 1853. *Acta Universitatis Upsaliensis*, 2022.
1069. Loureiro, F., Cardoso, L., Matos, A., & Matos, M. (2024). *Coxiella burnetii* in wild birds from Europe. *J. Advanc. Vet. Res.*, 14(4), 760-766.
1070. Lu, M., Tian, J., Zhao, H., et al.. Molecular survey of vector-borne pathogens in ticks, sheep keds, and domestic animals from Ngawa, Southwest China. *Pathogens*, 2022, 11, 5 C7 – 606.
1071. Lu, M. et al. Ehrlichia, Coxiella and Bartonella infections in rodents from Guizhou Province, Southwest China. *Ticks and Tick-borne Diseases* (2022):101974.
1072. Mandel, C.G., Sanchez, S.E., Monahan, C.C. et al. (2024). Metabolism and physiology of pathogenic bacterial obligate intracellular parasites. *Front. Cell. Inf. Microbiol.*, 14, 1284701.
1073. Orkun, Ö. Description of a novel Babesia sp. genotype from a naturally infected Eurasian lynx (*Lynx lynx*) in Anatolia, Turkey, with remarks on its morphology and phylogenetic relation to other piroplasmid species. *Ticks and Tick-borne Dis.* 13.6 (2022): 102026.
1074. Ouarti, B. et al. Molecular detection of microorganisms in lice collected from farm animals in Northeastern Algeria. *Comp. Immunol. Microbiol. Inf. Dis.*, 74 (2020): 101569-101569.
1075. Sellens, E. (2021). Q fever and Australia's veterinary workforce: Research to inform vaccine policy (Doctoral dissertation).
1076. Tarasiuk, K., Weiner, M., 2023. Ocena ryzyka zakażeń bakteryjnych przenoszonych

- przez kleszcze u funkcjonariuszy Nadbużańskiego Oddziału Straży Granicznej. Akademia Bialska Nauk Stosowanych im. Jana Pawła II.
1077. Truong, A., et al. *Toxoplasma gondii* and *Rickettsia* spp. in ticks collected from migratory birds in the Republic of Korea. *Sci. Reports* 12.1 (2022):1-9.
 1078. Wilhelmsson, P. et al. Migratory birds as disseminators of ticks and the tick-borne pathogens *Borrelia* bacteria and tick-borne encephalitis (TBE) virus: a seasonal study at Ottenby Bird Observatory in South-eastern Sweden. *Parasites & Vectors* 13.1 (2020): 1-17.
 1079. Yessinou, R.E. et al. Prevalence of *Coxiella*-infections in ticks-review and meta-analysis. *Ticks and Tick-borne Diseases* (2022): 101926.
 1080. Zendoia, I.I. et al. Stable prevalence of *Coxiella burnetii* in wildlife after a decade of surveillance in northern Spain. *Vet. Microbiol.*, 268 (2022):109422.
 1081. Бондаренко, Е.И., Филимонова, Е.С., Краснова, Е.И. et al. (2021). Случаи заболевания Ку-лихорадкой, выявленные у жителей Новосибирской области, госпитализированных с подозрением на инфекции, передаваемые клещами. *Клиническая лабораторная диагностика*, 66(4), 229-236.
 1082. Григорьева, Л. А., et al. "Многолетний мониторинг численности опасных для человека иксодовых клещей *Ixodes persulcatus* и *I. Ricinus* (Acari: ixodinae) на территории Санкт-Петербурга и Ленинградской области." *Паразитология* 54.1 (2020): 13-24.
 1083. Панин, А. Л. Микробиологический мониторинг возбудителей сапрозоонозов в полярных регионах. Doctoral dissertatin, Moscow, 2023.
 - 44). Zaharieva, MM, Kroumov, AD, Dimitrova, L, Tsvetkova, I, Trochopoulos, A, Konstantinov, SM, Berger, MR, Momchilova, M, Yoncheva, K, NAJDENSKI, HM. Micellar curcumin improves the antibacterial activity of the alkylphosphocholines erufosine and miltefosine against pathogenic *Staphylococcus aureus* strains. *Biotechnology & Biotechnological Equipment*, 33, 1, Fransis & Taylor, 2019, ISSN:1310-2818, DOI:10.1080/13102818.2018.1533792, 38-53. SJR (Scopus):0.38, JCR-IF (Web of Science):1.186**
 1084. Alavi, Mehran, and Mahendra Rai. "Antibacterial and wound healing activities of micro/nanocarriers based on carboxymethyl and quaternized chitosan derivatives." *Biopolymer-Based Nano Films*. Elsevier, 2021. 191-201.
 1085. Blake M, Page E, Smith M, Calhoun T. Miltefosine Impacts Small Molecule Transport in Gram-Positive Bacteria. *ChemRxiv*. 2024.
 1086. Buliga, D-I. et al. Enhancing the light fastness of natural dyes by encapsulation in silica matrix. *J. Photochem. Photobiol. A: Chemistry* 432 (2022): 114085.
 1087. Karthikeyan, A., N. Senthil, T. Min. Nanocurcumin: A promising candidate for therapeutic applications. *Front. Pharmacol.*, 11 (2020), 529594.
 1088. Khatri, P., Rani, A., Hameed, S. et al. 2023. Current understanding of the molecular basis of spices for the development of potential antimicrobial medicine. *Antibiotics*, 12(2), 270.
 1089. Lawson, Becki, and Dick Best. *Passerines and other small birds*. BSAVA manual of wildlife casualties. BSAVA Library, 2016. 421-438.
 1090. Makgoo, L., & Mbita, Z. (2022). Germicidal and antineoplastic activities of curcumin and curcumin-derived nanoparticles. *Ginger-Cultivation and Use*.
 1091. Mukherjee, S. et al. *Nano Curcumin: Making it useful for Human Therapy*. (2020).

1092. Pawelski, D., Walewska, A., Ksiezak, S. et al. (2021). Monocarbonyl analogs of curcumin based on the Pseudopelletierine scaffold: synthesis and anti-inflammatory activity. *Int. J. Mol. Sci.*, 22(21), 11384.
1093. Pourhajibagher, M. et al. Sonodynamic excitation of nanomicelle curcumin for eradication of *Streptococcus mutans* under sonodynamic antimicrobial chemotherapy: Enhanced anti-caries activity of nanomicelle curcumin. *Photodiagn. Photodyn. Therapy* (2020): 101780.
1094. Pourhajibagher, M. et al. Quorum quenching of *Streptococcus mutans* via the nano-quercetin-based antimicrobial photodynamic therapy as a potential target for cariogenic biofilm. *BMC Microbiology* 22.1 (2022):1-18.
1095. Pourhajibagher, M., Bahrami, R., Bazarjani, F., Bahador, A., 2023. Anti-multispecies microbial biofilms and anti-inflammatory effects of antimicrobial photo-sonodynamic therapy based on acrylic resin containing nano-resveratrol. *Photodiagn. Photodyn. Therapy*, 43, 103669.
1096. Prajapati, S.K. et al. Antimicrobial application potential of phytoconstituents from turmeric and garlic. *Bioact. Nat. Prod. Pharmaceut. Appl.*. Springer, Cham 409-435.
1097. Sankhwar, R. et al. Application of nano-curcumin as a natural antimicrobial agent against Gram-positive pathogens. *J. Appl. Nat. Sci.*, 13.1 (2021): 110-126.
1098. Torrontegi, Olalla, et al. "Naturally Avian Influenza Virus–Infected Wild Birds Are More Likely to Test Positive for Mycobacterium spp. and Salmonella spp." *Avian diseases* 63.1 (2018): 131-137.
1099. Tsokana, Constantina N., et al. "European Brown hare (*Lepus europaeus*) as a source of emerging and re-emerging pathogens of Public Health importance: A review." *0 Veterinary Medicine and Science* 6(3), (2020), 550-564.
1100. Varga, Molly. "Deer." *BSAVA Manual of Wildlife Casualties*. BSAVA Library, 2016. 275-298.
1101. Venkatas, J., Daniels, A., Singh, M. The potential of curcumin-capped nanoparticle synthesis in cancer therapy: a green synthesis approach. *Nanomaterials*, 2022, 12, 18 C7- 3201.
1102. Гончарук, М., et al. Анализ рисков развития заболеваний для программы реинтродукции амурских (дальневосточных) леопардов (*Panthera pardus orientalis*).
- 45). NAJDENSKI H., Golkocheva E., Kussovski V., Vesselinova A., Garbom S., Walf-Watz H.. Attenuation and Preserved Immunogenic Potential of *Yersinia pseudotuberculosis* Mutant Strains Evidenced in Oral Pig Model. *Zoonoses & Public Health*, 56, 4, Wiley-Blackwell Publishing Ltd, 2009, ISSN:18631959, 18632378, DOI:10.1111/j.1863-2378.2008.01153.x, 157-168. JCR-IF (Web of Science):1.906**
1103. Jiménez V., S. del Carmen. Identificación de proteínas de secreción con capacidad inmunogénica de aislamientos de *Yersinia pseudotuberculosis* provenientes de planteles cuyícolas del departamento de Nariño. PhD thesis, 2011.
1104. Martínez, P.O. Prevalence of enteropathogenic *Yersinia* in pigs from different European countries and contamination in the pork. Doctoral Dissertation, Faculty of Veterinary Medicine, University of Helsinki, 2010.
1105. Moura, C.AA, et al. Evidence of improved reporting of swine vaccination trials in the post-REFLECT statement publication period. *J. Swine Health Product*. 27.5 (2019): 265.
1106. Булгакова, Н. Ф. Оценка степени аттенуации и сохранения иммуногенности

мутантных штаммов *Yersinia pseudotuberculosis* при экспериментальном пероральном введении пороссятам. (Болгария. Швеция), Ветеринария, 2011, 3, 757.

46). NAJDENSKI H., Golkocheva E., Vesselinova A., Rüssmann, H.. Comparison of the course of infection of virulent *Yersinia enterocolitica* serotype O:8 with an isogenic soda mutant in the peroral rabbit model. International Journal of Medical Microbiology, 6, Urban und Fischer Verlag GmbH und Co. KG, 2004, ISSN:14384221, 16180607, 383-393. JCR-IF (Web of Science):2.611

1107. Bai, G., Pata, J., McDonough, K.A., Golubov, A., Smith, E. Differential gene regulation in *Yersinia pestis* versus *Yersinia pseudotuberculosis*: Effects of hypoxia and potential role of a plasmid regulator (2007) *Advances in Experimental Medicine and Biology*, 603, pp. 131-144.

1108. Champion, O.L., Karlyshev, A., Cooper, I.A.M. et al. *Yersinia pseudotuberculosis* mntH functions in intracellular manganese accumulation, which is essential for virulence and survival in cells expressing functional Nramp1. *Microbiology*, 2011, 4, 1115-1122.

1109. Papp-Wallace, K.M., A.S. Moomaw, , M.E. Maguire. Manganese: uptake, biological function, and role of virulence. 236-256. In: *Molecular Microbiology of Heavy Metals*, (D.H. Nies, S.Silver, eds.), Springer, 2007.

1110. Papp-Wallace, K.M., Maguire, M.E. Manganese transport and the role of manganese in virulence (2006) *Annual Review of Microbiology*, 60, pp. 187-209.

1111. Wang, Xin, et al. *Yersinia*. *Laboratory Models for Foodborne Infections*. CRC Press, 2017, 427-437.

47). Stoykova B., Chochkova M., Ivanova G., Markova N., Enchev V., Tsvetkova I., Najdenski H., Štícha M., Milkova T.. Ultrasound-assisted green bromination of N-cinnamoyl amino acid amides–Structural characterization and antimicrobial evaluation. 1135, Journal of Molecular Structure, 2017, DOI:<https://doi.org/10.1016/j.molstruc.2017.01.056>, 144-152. ISI IF:1.753

1112. Bărbălan, G. et al. The effect of some physical-chemical factors on *Yersinia enterocolitica* strains, *Lucrări Științifice Medicină Veterinară* 2021, LIV(1), TIMIȘOARA.

1113. Cumpănășoiu, C., C.E. Cumpănășoiu, R. Trif, et al. Behavior of some strains of *Yersinia enterocolitica* at freezing and salt. *Lucrări Științifice Medicină Veterinară*, XLIII (1), 2009, 224-229.

1114. Jităreanu A., I.C. Caba, L. Agoroaei. Halogenation – a versatile tool for drug synthesis - the importance of developing effective and eco-friendly reaction protocols. *Curr. Analys. Biotechnol.*, 2:2019, 11-25.

1115. Jităreanu A., et al. Bromination - a versatile tool for drugs optimization. *The Medical Surgical L.*, 2018, 122, 3, 614-626..

1116. Lin Jun, Hu Xiaolong, Li Junyan et al. Research progress in N-phenylpropenoyl-L-amino acids. *Advances in Pharmacy*. 2018, 42(1):52-59.

1117. Sabuzi, F. et al. Sustainable bromination of organic compounds: A critical review." *Coordination Chemistry Reviews*, 385 (2019):100-136.

1118. Van Kerrebroeck, R., T. Horsten, C.V. Stevens. Bromide oxidation: a safe strategy for electrophilic brominations. *Eur. J. Org. Chem.*, 2022.35 (2022): e202200310.

48). Iliev, M., NAJDENSKI, H. Monitoring of plasmid dissociation and pathogenic

potential among *Yersinia enterocolitica* and *Yersinia pseudotuberculosis* during storage of refrigerated pork meat. 58, 4, Annals of Microbiology, 2008, 623-632. JCR-IF (Web of Science):0.5

1119. Bărbălan G., I. Nichita, N. Mederle, et al. The effect of some physical-chemical factors on *Yersinia enterocolitica* strains. *Medicina Veterinara*, 2021, 54, 1, 5-10
1120. Cumpănașoiu C., C.E. Cumpănașoiu, R. Trif et al. Behavior of some strains of *Yersinia enterocolitica* at freezing and salt. *Lucrury Științifice Medicina Veterinara*, XLIII (1), 2009, 224-229, Timișoara.
1121. Li, C. et al. Isolate specific cold response of *Yersinia enterocolitica* in transcriptional, proteomic, and membrane physiological changes. *Front. Microbiol.*, 10 (2020): 3037.
1122. Sabuzi, Federica, et al. "Sustainable bromination of organic compounds: A critical review." *Coordination Chemistry Reviews* 385 (2019): 100-136.
1123. Stoykova, B., et al. Anti-influenza drug derivatives with potential biological activity. The PhD Student Scientific Session of the FMNS–2016: 6.
1124. Thabet, S.S., Thabet, M. (2021). Viability of the isolated *Yersinia enterocolitica* strains from Damietta Cheese and ice cream at different refrigeration and freezing temperatures. *J. Appl. Vet. Sci.*, 6(2), 1-14
1125. Zadernowska, A., Chajęcka-Wierzchowska, W., & Łaniewska-Trokenheim, Ł. *Yersinia enterocolitica*: a dangerous, but often ignored, foodborne pathogen. *Food Rev. Intern.*, 2014, 30(1), 53-70.

49). Konakchiev A., Mikhova B., Todorova M., NAJDENSKI H., Tzvetkova I., Vitkova A., Duddeck H. Composition of the Essential Oil of *Achillea Asplenifolia* Vent. from Bulgaria. *Journal of Essential Oil Bearing Plants*, 8, 3, Taylor and Francis Ltd., 2005, ISSN:0972060X, 318-323. JCR-IF (Web of Science):0.27

1126. Boris, T.U.R. K., Baričević, D., & Batič, F. (2021). Essential oil content, chamazulene content and antioxidative properties of *Achillea millefolium* agg. extracts from Slovenia. *Acta Agricult. Slovenica*, 117(2), 1-10.
1127. Kindlovits, S., É. Németh. Sources of variability of yarrow (*Achillea* spp..) essential oil. *Acta Aliment.*, 41.Suppl.-1 (2012):92-103.
1128. Öğretmen, N.G.. Civanperçemi (*Achillea asplenifolia* ve *Achillea collina*) popülasyonlarının verim ve bazı kalite özellikleri üzerine farklı kültürel uygulamaların etkisi. Master thesis. Adnan Menderes Üniversitesi, Fen Bilimleri Enstitüsü, 2014.

50). Ivanova, V., Graefe, U., Schlegel, B., Kolarova, M., Aleksieva, K., NAJDENSKI, H., Tzvetkova, I., Chipeva, V. Usnic acid, metabolite from *Neuropogon* sp., an antarctic lichen isolation, structure elucidation and biological activity. *Biotechnology and Biotechnological Equipment*, 18, 1, 2004, ISSN:1310-2818, 66-71. ISI IF:0.622

1129. Frenák, R., Vilková, M., Garberová, M. et al. (2023). Isolation and identification of lichen substances for biological and ecological roles. In: *Plant Specialized Metabolites: Phytochemistry, Ecology and Biotechnology* (pp.1-66). Cham: Springer Nature Switzerland.
1130. Galanty, A., P. Paško, I. Podolak. Enantioselective activity of usnic acid: a comprehensive review and future perspectives. *Phytochem. Rev.*, 18.2 (2019): 527-548.
1131. Kosanić, M., & Ranković, B. (2015). Lichen secondary metabolites as potential antibiotic agents. In *Lichen Secondary Metabolites* (pp. 81-104). Springer Int. Publishing.

1132. Luzina, O.A., N.F. Salakhutdinov. Biological activity of usnic acid and its derivatives: Part 1. Activity against unicellular organisms. Russian J. Bioorg. Chem., 42.2 (2016): 115-132.
1133. Maulidiyah, M., Darmawan, A., Ahmad, E. et al. (2021). Antioxidant activity-guided isolation of usnic acid and diffractaic acid compounds from lichen genus *Usnea* sp. J. Appl. Pharm. Sci., 11(2), 075-083.
1134. Stoica A.E., Albuleț D., Bîrcă A.C. et al.. Electrospun nanofibrous mesh based on PVA, chitosan, and usnic acid for applications in wound healing. Int. J. Mol. Sci., 24 (13), 11037, 2023.
1135. Лузина, О.А., Н.Ф. Салахутдинов. Биологическая активность усниновой кислоты и ее производных. Часть 2. Действие усниновой кислоты и ее производных на высшие организмы, молекулярные и физико-химические аспекты биологической активности (обзорная статья). Биоорганическая химия 42.3 (2016): 276-276.

51). Raducheva, T., J. Kurteva, N. Markova, D. Veljanov, H. NAJDENSKI. Behavior of *Salmonella dublin* in mice and rats upon intraperitoneal infection.. Zbl. Bact., 280, 1994, 520-525. JCR-IF (Web of Science):0.729

1136. Aydin, Merve, Derya Arslan Danacioğlu, and Selman Türker. "Propolisin genel özellikleri ve kullanımı." Gıda 46.1: 69-81.
1137. dos Santos, Cíntia Maria, et al. "Red propolis as a source of antimicrobial phytochemicals: extraction using high-performance alternative solvents." Frontiers in Microbiology 12 (2021): 1166.
1138. Farooq, Muhammad Qamar, Nabeel Mujtaba Abbasi, and Jared L. Anderson. "Deep eutectic solvents in separations: Methods of preparation, polarity, and applications in extractions and capillary electrochromatography." Journal of Chromatography A 1633 (2020): 461613.
1139. Fedorka-Cray, P.J., et al. Alternate routes of invasion may affect pathogenesis of *Salmonella typhimurium* in swine. Inf. Immun., 63.7 (1995):2658.
1140. Havelaar, A.H., et al. A rat model for dose–response relationships of *Salmonella enteritidis* infection. J. Appl. Microbiol., 91.3 (2001):442-452.
1141. Jiang L., Li W., Hou X. et al. Nitric oxide is a host cue for *Salmonella typhimurium* systemic infection in mice. Communicat. Biol., 6 (1), 501, 2023.
1142. Naughton, P.J., G. Grant. Modelling of salmonellosis. Biology of Growing Animals. 2, Elsevier, 2005. 235-257.
1143. Naughton, P.J., et al. A rat model of infection by *Salmonella typhimurium* or *Salm. enteritidis*. J. Appl. Bacteriol., 81.6 (1996):651-656.
1144. Naughton, P.J. et al. *Salmonella typhimurium* and *Salmonella enteritidis* induce gut growth and increase the polyamine content of the rat small intestine in vivo. FEMS Immunol. & Med. Microbiol., 12.3-4 (1995):251-257.
1145. Nie Jingjing, Li Chuanbao, Pian Yaya, et al. Establishment of a rat model of *Salmonella enteritidis* infection and study of disease outcome. Medical Animal Control, 2021

52). Zaharieva, MM, Genova-Kalou, P, Dincheva, I, Badjakov, I, Krumova, S, Enchev, V, NAJDENSKI, H, Markova, N . Anti-Herpes Simplex virus and antibacterial activities of *Graptopetalum paraguayense* E. Walther leaf extract: a pilot study. Biotechnology & Biotechnological Equipment, 33, 1, Taylor&Francis, 2019, ISSN:1310-2818, DOI:10.1080/13102818.2019.1656108, 1251-1259. SJR (Scopus):0.38, JCR-IF (Web of

1146. Al-Mamoori1, F., D.M. Al-Tawalbe, M.Alnaqeeb. Medicinal plants for the treatment and management of oral infections: A review. Trop. J. Nat. Prod. Res., 2021; 5(9):1528-1536
1147. Garber, A, L. Barnard, C. Pickrell. Review of whole plant extracts with activity against Herpes Simplex Viruses in vitro and in vivo. J. Evid.-Based Integr. Med., 26 (2021):2515690X20978394.
1148. Medina Mendoza, R.A. Composición química y actividades biológicas de tres especies de Graptopetalum nativas de Sinaloa. Master thesis, Universidad Autonoma de Sinaloa.
1149. Mohan, S. et al. Bioactive natural antivirals: An updated review of the available plants and isolated molecules. Molecules 25.21 (2020):4878.
1150. Nandi, S.S. et al. (2023). Medicinal plants against Herpes Simplex Virus (HSV) Type 1 infections: ethnopharmacology, chemistry, and clinical and preclinical studies. In: Pal, D. (eds) Anti-Viral Metabolites from Medicinal Plants. Reference Series in Phytochemistry, Springer, Cham.
1151. Todorova N, Rangelov M, Dincheva I, Badjakov I, Enchev V, Markova N Potential of hydroxybenzoic acids from Graptopetalum paraguayense for inhibiting of herpes simplex virus DNA polymerase - metabolome profiling, molecular docking and quantum-chemical analysis (2022) Pharmacia 69(1): 113-123. <https://doi.org/10.3897/pharmacia.69.e79467>.
1152. Sitarek, P. et al. Potential synergistic action of bioactive compounds from plant extracts against skin infecting microorganisms. Int. J. Mol. Sci., 21.14 (2020):5105.
- 53). Konakchiev, A., M. Todorova, B. Mikhova, A. Vitkova, H. NAJDENSKI, H. Duddeck. Chemical composition and antimicrobial activity of the essential oil from two *Achillea collina* Becker.. Compt. Rend. Acad. Bulg. Sci, 59, 5, BAS, 2006, ISSN:1310-1331 (Print), 505-510. SJR (Scopus):0.22**
1153. Abad, M.J., L.M. Bedoya, P. Bermejo. Essential oils from the Asteraceae family active against multidrug-resistant bacteria. In. Fighting Multidrug Resistance with Herbal Extracts, Essential Oils and their Components. Elsevier Inc., 2013, 205-221.
1154. Andrushko, N., & Andrushko, V. Asymmetric hydrogenation of C=O and C=N Bonds in stereoselective synthesis. Stereoselective synthesis of drugs and natural products, John Wiley & Sons, Inc. 2013, DOI: 10.1002/9781118596784.ssd030
1155. Abad, M.J., Bedoya, L.M., Luis Apaza, L, et al. The Artemisia L. genus: a review of bioactive essential oils. Molecules, 2012, 17 (3), 2542-2566.
1156. Benedec, D., Vlase, L., Oniga, I., et al. Polyphenolic composition, antioxidant and antibacterial activities for two Romanian subspecies of *Achillea distans* Waldst. et Kit. ex Willd. Molecules, 8, 18, 2013 , 8725-8739.
1157. Chizzola, R. Volatile compounds in the aerial parts of *Achillea collina* collected in the urban area of Vienna (Austria). Nat. Prod. Commun.,12, 2017, 12, 1933-1936.
1158. Doğan, H., Uskutoğlu, T., Fidan, H. et al. (2021). Chemical compositions, antioxidant activities, and mineral matter contents of *Achillea collina* Becker ex Rchb from the flora of Bulgaria. Commagene J. Biol., 5(2), 143-149.
1159. Enna, S. J., & Norton, S. Herbal supplements and the brain: understanding their health benefits and hazards. FT Press, Pearson Education, Inc. 2012. ISBN-10:0-13-282497-3
1160. Hussin, W.A., & El-Sayed, W.M. Synergic interactions between selected botanical extracts and tetracycline against Gram positive and Gram negative bacteria. J. Biol. Sci.,

2011, 11, 7, 433-441.

1161. Issabeagloo, E., & Taghizadieh, M. Inhibitory Effect of *Ziziphus zizyphus* L. Extract on *Staphylococcus* Genera. *Adv. Biores.*, 2012, 3(3).

1162. Issabeagloo, E., & Abri, B. Antimicrobial effects of yarrow (*Achillea millefolium*) essential oils against *Staphylococcus* species. *African J. Pharm. Pharmacol.*, 2012, 6(41), 2895-2899.

1163. Jemia, MB., Rouis, Z., Maggio, A., et al. Chemical composition and free radical scavenging activity of the essential oil of *achillea ligustica* growing wild in lipari (aeolian islands, sicily). *Nat. Prod. Communicat.*, 11, 8, 2013, 1629-1632.

1164. Schlecker, A. Metall-katalysierte Enin-Zyklisierungen. Max– Planck-Institut für Kohlenforschung in Mülheim an der Ruhr, Universität Dortmund, 2008.

54). Zaharieva, M. M., Trochopoulos, A., Dimitrova, L., Berger, M. R., NAJDENSKI, H., Konstantinov, S., Kroumov, A. D.. New Insights in Routine Procedure for Mathematical Evaluation of in vitro Cytotoxicity Data from Cancer Cell Lines. 22, 2, International Journal Bioautomation, 2018, DOI:10.7546/ijba.2018.22.2.87-106, 87-106. SJR (Scopus):0.231

1165. Sudha, D., Revathi, A., Arunadevi, N., & Kirubavathy, S. J. (2024). Structural characterization, DNA interaction and pharmacological evaluation of Cu (II), Co (II), Ni (II), Fe (II) dihydrate complexes derived from pyridine-2, 5-dicarboxylic acid and 2-methylimidazole as organic moieties. *J. Mol. Str.*, 138632.

1166. Uota, S.T. (2020). Microalgae as a new source of neuroprotective compounds (Doctoral dissertation, Universidade do Algarve (Portugal)).

55). NAJDENSKI H., Vesselinova A., Golkocheva E., Garbom S., Wolf-Watz H.. Characterization of Infections with Wild and Mutant *Yersinia pseudotuberculosis* Strains in Rabbit Oral Model. The Genus *Yersinia*, 2003, ISBN:978-0-306-48416-2, ISSN:0065-2598, DOI:10.1007/b100541, 117-120

1167. Ковширина Ю.В. Клинические и иммунопатологические особенности псевдотуберкулеза у детей. (2008) Doctoral dissertation, Новосибирск, с.206.

1168. Разгулин С.А. Научное обоснование нового подхода к профилактике кишечных антропонозов у военнослужащих в эндемичных районах. (2006) Doctoral dissertation, Пермь, с.265.

1169. Седулина, О.Ф. (2002). Клинико-морфологическая характеристика поражений желудка при иерсиниозах у детей (Doctoral dissertation, Владивостокский государственный медицинский университет)..

56). NAJDENSKI H., Kussovski V., Michailov Y., Vesselinova A. Protective effect of Oxadin on experimental *Yersinia enterocolitica* infection in rats.. Die Pharmazie, 57, 2002, 337-339

1170. Cao, X., S. Ke. Updated report on synthesis and biological properties of oxadiazine-based heterocyclic derivatives. *Rec. Adv. Med. Chem.*, 2 (2015):107.

1171. Ke, S., X. Cao, Y. Liang, et al. Synthesis and biological properties of dihydro-oxadiazine-based heterocyclic derivatives. *Mini Rev Med Chem.* 2011, 11 (8):642-57.

1172. Qiao, X., Han, Y., Huang, S. et al. (2024). Visible-light-induced [3+ 3] cycloaddition reaction of phenol and hydrazone to access 1, 3, 4-oxadiazines scaffolds. *Molecular*

Catalysis, 561, 114156.

1173. Schmidt, E.Y., Semenova, N.V., Ivanova, E.V. et al. (2019). Acetylene-based two-step diastereoselective synthesis of bridgehead dihydro-oxadiazines using ketones and hydrazine as the only reactants. Chem. Commun., 55(18), 2632-2635.

57). Le Guern, A.-S., C. Savin, H. Angermeier, S. Brémont, D. Clermont, E. Mühle, P. Orozova, H. NAJDENSKI, J. Pizarro-Cerdá. *Yersinia artesiana* sp. nov., *Yersinia proxima* sp. nov., *Yersinia alsatica* sp. nov., *Yersinia vastinensis* sp. nov., *Yersinia thracica* sp. nov. and *Yersinia occitanica* sp. nov., isolated from humans and animals.. Int. J. Syst. Evolut. Microbiol., 7, 10, 2020, ISSN:1466-5026, DOI:10.1099/ijsem.0.004417, 1-10. SJR (Scopus):1.02

1174. Burbick CR, Munson E, Lawhon SD, et al. An update on novel taxa and revised taxonomic status of bacteria (including members of the phylum Planctomycetota) isolated from aquatic host species described in 2018 to 2021. J Clin Microbiol. 2023, 22;61(2):e0142622.

1175. Chen, G. et al. Obtaining specific sequence tags for *Yersinia pestis* and visually detecting them using the CRISPR-Cas12a system. Pathogens 10.5 (2021): 562.

1176. Lawhon SD, Burbick CR, Munson E, et al. Update on novel taxa and revised taxonomic status of bacteria isolated from nondomestic animals described in 2018 to 2021. J Clin Microbiol. 2023, 22;61(2):e0142522.

1177. Lim SJ, Jithpratuck W, Wasyluk K, et al. Associations of microbial diversity with age and other clinical variables among pediatric chronic rhinosinusitis (CRS) patients. Microorganisms. 2023; 11(2):422.

1178. Munson E, Carroll KC. Update on accepted novel bacterial isolates derived from human clinical specimens and taxonomic revisions published in 2020 and 2021. J Clin Microbiol. 2023, 26;61(1):e0028222.

1179. Munson E, Lawhon SD, Burbick CR, et al. 2023. An update on novel taxa and revised taxonomic status of bacteria isolated from domestic animals described in 2018 to 2021. J Clin Microbiol 61:e00281-22..

1180. Nguyen, Scott Van, et al. "Yersinia occitanica is a later heterotypic synonym of *Yersinia kristensenii* subsp. rochesterensis and elevation of *Yersinia kristensenii* subsp. rochesterensis to species status." International Journal of Systematic and Evolutionary Microbiology (2021): 004626.

1181. Špačková, M., et al. Overview of basic epidemiological characteristics and descriptive analysis of the incidence of human yersiniosis in the Czech Republic in 2018-2020. Epidemiologie, Mikrobiologie, Imunologie: Casopis Společnosti pro Epidemiologii a Mikrobiologii Ceske Lekarske Společnosti JE Purkyne 71.1 (2022):32-39.

58). Popova, MP, Trusheva, B.S., Nedialkov, P.T., Tsvetkova, I., Pardo-Mora, D.P., NAJDENSKI, H., Torres-García, O.A., Sforcin, J.M., Bankova, V.S. New Δ -tocotrienol derivatives from Colombian propolis. Natural Product Research, 34, 19, Taylor and Francis Ltd., 2020, ISSN:14786427, 14786419, 2779-2786. SJR (Scopus):0.46, JCR-IF (Web of Science):2.158

1182. Asma, S.T., Acaroz, U., Bobiş, O. et al. (2024). Propolis. In: Honey Bees, Beekeeping and Bee Products (pp. 147-169). CRC Press..

1183. Belmehdi, O., El Meniyi, N., Bouyahya, A. et al. 2023. Recent advances in the chemical

- composition and biological activities of propolis. Food Rev. Int., 39(9), 6078-6128.
1184. Belmehdi, O., Bouyahya, A., Jekő, J. et al. (2021). Chemical analysis, antibacterial, and antioxidant activities of flavonoid-rich extracts from four Moroccan propolis. J. Food Proc. Preserv., 45, e15816.
1185. Gallego, M.G.S. Determinación de la actividad anti-Candida albicans de propóleos sonorenses colectados en las diferentes estaciones del Año, y sus principales constituyentes. Master thesis, Universidad de Sonora..
1186. Hossain, R. et al. Propolis: An update on its chemistry and pharmacological applications. Chinese Medicine, 17.1 (2022): 1-60.
1187. Pardo-Mora, D.P., Murillo, O.J., Rey-Buitrago, M. et al. (2021). Apoptosis-related gene expression induced by Colombian propolis samples in canine osteosarcoma cell line. Vet. World, 14(4), 964.
1188. Tran, T.D. et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020):4988.
1189. Zeutso, J.F. et al. Antioxidant and cytotoxicity activities of δ -tocotrienol from the seeds of *Allophylus africanus*. Nat. Prod. Res., (2021): 1-11.
- 59). Gigova L., G. Gacheva, R. Toshkova, E. Gardeva, N. Ivanova, I. Iliev, V. Kusssovski, H. NAJDENSKI. Effects of temperature on Synechocystis sp. R10 (Cyanoprocarvota) at two irradiance levels. Genetics and Plant Physiology, 2, 1-2, BAS, 2012, ISSN:1314-6394 (Print), 38-49**
1190. Montalvão, S, et al. Large-scale bioprospecting of cyanobacteria, micro-and macroalgae from the Aegean Sea. New biotechnol., 33.3 (2016):399-406.
1191. Stirk, W. A., & van Staden, J. (2022). Bioprospecting for bioactive compounds in microalgae: Antimicrobial compounds. Biotechnology advances, 59, 107977.
- 60). Iliev, M., H. NAJDENSKI, A. Stals, H. Werbrouck, L. Herman, E. van Coille. Optimization of Real-Time PCR protocol for detection of pathogenic Yersinia enterocolitica strains.. Bulg. J. Vet. Med., 11, 3, 2008, 179-184. JCR-IF (Web of Science):0.28**
1192. Mangal, M. et al. Molecular detection of foodborne pathogens: A rapid and accurate answer to food safety. Crit. Rev. Food Sci. Nutrition 56.9 (2016): 1568-1584.
1193. Abbas, K.H. Rapid detection of Yersinia enterocolitica by using Real-Time PCR technique in some types of foods in Al-Qadisiyah province. Al-Qadisiyah J. Vet. Med. Sci., 14.1 (2015): 34-38.
- 61). NAJDENSKI H., Nikolova S., Wesselinova D., Kazatchca D., Vesselinova A. Experimental Mixed Infection With Yersinia Enterocolitica and Listeria Monocytogenes in Guinea Pigs. 45, 1-10, Wiley-Blackwell Publishing Ltd, 1998, ISSN:0931-1793, DOI:10.1111/j.1439-0450.1998.tb00834.x, 611-620. JCR-IF (Web of Science):0.573**
1194. Бакулов, Игорь Алексеевич, et al. Листерии и листериоз. (2016).
- 62). Toshkova, R., E. Ivanova, H. NAJDENSKI, J. Gumpert. Antitumour immunization of hamsters by allogenic myeloid tumour cells. Compt. rend. Acad. bulg. Sci., 50, 9-10, BAS, 1997, ISSN:1310-1331 (Print), 71-74**
1195. Allan, E.J., C. Hoischen, J. Gumpert. Bacterial L-forms. Adv. Appl. Microbiol., 68,

63). Mileva M, Ilieva Y, Jovtchev G, Gateva S, Zaharieva, MM, Georgieva A, Dimitrova L, Dobрева A, Angelova Ts, Vilhelmova-Ilieva N, Valcheva V, NAJDENSKI H. Rose flowers—A Delicate Perfume or a Natural Healer?. Biomolecules, 11, 1, MDPI, 2021, ISSN:2218-273X, DOI:<https://doi.org/10.3390/biom11010127>, 127-159. SJR (Scopus):1.614, JCR-IF (Web of Science):4.694

1196. Aituarova, A., Zhusupova, G.E., Zhussupova, A. and Ross, S.A., 2023. Study of the Chemical Composition of *Rosa beggeriana* Schrenk's Fruits and Leaves. *Plants*, 12(18), p.3297.
1197. Alborzi SS, Roosta A. The effect of different solvents on the production of rose concrete and rose absolute, experimental study, and thermodynamic aspects using the UNIFAC model. *Chem. Eng. Res. Design*. 2022, 13.
1198. Ali, E.F., Al-Yasi, H.M., Majrashi, A.M. et al. 2022. Chemical and nutritional characterization of the different organs of taif's rose (*Rosa damascena* Mill. var. *trigintipetala*) and possible recycling of the solid distillation wastes in Taif City, Saudi Arabia. *Agriculture*, 12(11), 1925.
1199. Aituarova, A., Zhusupova, G.E., Zhussupova, A., & Ross, S. A. (2023). Study of the chemical composition of *Rosa beggeriana* Schrenk's fruits and leaves. *Plants*, 12(18), 3297.
1200. Antoniadou, M., Rozos, G., Vaiou, N. et al. (2023). The in vitro assessment of antibacterial and antioxidant efficacy in *Rosa damascena* and *Hypericum perforatum* extracts against pathogenic strains in the interplay of dental caries, oral health, and food microbiota. *Microorganisms*, 12(1), 60.
1201. Bao, M., Xi, Y., Wang, R. et al. (2024). Trehalose signaling regulates metabolites associated with the quality of rose flowers under drought stress. *Env. Exp. Botany*, 105813.
1202. Bashir, A., Ahmad, T., Farooq, S. et al. 2023. A secondary metabolite of *Cercospora* sp., associated with *Rosa damascena* Mill., inhibits proliferation, biofilm production, ergosterol synthesis and other virulence factors in *Candida albicans*. *Microb. Ecol.*, 85(4), 1276-1287.
1203. Bashir, A., Manzoor, M.M., Ahmad, T. et al. 2023. Endophytic fungal community of *Rosa damascena* Mill. as a promising source of indigenous biostimulants: Elucidating its spatial distribution, chemical diversity, and ecological functions. *Microbiol. Res.*, 276, 127479.
1204. Bentley, P.R., Fisher, J.C., Dallimer, M. et al. 2023. Nature, smells, and human wellbeing. *Ambio*, 52(1), 1-14.
1205. Bottoni M, Milani F, Galimberti PM, Vignati L, Romanini PL, Lavezzo L, Martinetti L, Giuliani C, Fico G. Ca' Granda, Hortus simplicium: Restoring an Ancient Medicinal Garden of XV–XIX Century in Milan (Italy). *Molecules*. 2021; 26(22):6933.
1206. Bottoni, M., Milani, F., Galimberti, P. M. Et al. (2021). Ca'Granda, *Hortus simplicium*: Restoring an ancient medicinal garden of XV–XIX century in Milan (Italy). *Molecules*, 26(22), 6933..
1207. Charoimek, N., Phusuwan, S., Petcharak, C. et al., 2023. Do abiotic stresses affect the aroma of damask roses?. *Plants*, 12(19), 3428.
1208. Choi, Y.G., Choi, W.S., Song, J.Y. et al. 2023. Antiinflammatory effect of the ethanolic extract of Korean native herb *Potentilla rugulosa* Nakai in Bisphenol-a-stimulated A549 cells. *J. Toxicol. Env. Health, Part A*, 86(20), 758-773.
1209. Demirel, S., 2022. Medical evaluation of the antimicrobial activity of rose oil on some

- standard bacteria strains and clinical isolates. *Altern. Ther. Health & Medicine*, 28(6).
1210. Ekhtiari, Z., Hadjzadeh, M. A. R., & Gholamnezhad, Z. (2023). Medicinal plants and foods with metaphorical concepts in Rumi's "Masnavi Manavi": The psychosomatic approach to human health. *Avicenna J. Phytomed.*, 13(6).
 1211. Fang, R., Zweig, M., Li, J., et al. 2023. Diversity of volatile organic compounds in 14 rose cultivars. *J. Essent. Oil Res.*, 35(3), 220-237.
 1212. Festini Mikulec, Z., 2022. Izolacija i identifikacija eteričnog ulja stolisne ruže (*Rosa x centifolia* L.'Fantin-Latour') (Doctoral dissertation, University of Zagreb).
 1213. Flórez González, S.J., Stashenko, E.E., Ocazonez, R.E., et al. (2024). In vitro safety assessment of extracts and compounds from plants as sunscreen ingredients. *Int. J. Toxicol.*, 10915818231225661.
 1214. Fuentes, J.L., Pedraza Barrera, C.A., Villamizar Mantilla, D.A. et al. 2022. Flower extracts from ornamental plants as sources of sunscreen ingredients: determination by in vitro methods of photoprotective efficacy, antigenotoxicity and safety. *Molecules*, 27(17), 5525.
 1215. Galal, T.M., Al-Yasi, H.M., Fawzy, M.A. et al. 2022. Evaluation of the phytochemical and pharmacological potential of taif's rose (*Rosa damascena* Mill var. *trigintipetala*) for possible recycling of pruning wastes. *Life*, 12(2), 273.
 1216. González-Minero, F.J., Bravo-Díaz, L. and Moreno-Toral, E., 2023. Pharmacy and fragrances: traditional and current use of plants and their extracts. *Cosmetics*, 10(6), 157.
 1217. Harismah, K., Mirzaei, M. and Beser, N., 2023, Multi-purpose Plants of Essential Oils in Residential Gardens. In *Proceedings of the International Conference of Contemporary Affairs in Architecture and Urbanism-ICCAUA* (Vol. 6, No. 1, pp. 1114-1122).
 1218. Hegde, A.S., Gupta, S., Sharma, S. et al. 2022. Edible rose flowers: a doorway to gastronomic and nutraceutical research. *Food Res. Int.*, 162, 111977.
 1219. Kalembe-Drożdż M., Grzywacz-Kisiełewska A., Cierniak A. Surowce polifenolowe Za Stosowania i perspektywy.
 1220. Kant, K., Gupta, S., Kaur, N. et al. 2023. Novel foliar approaches enhancing active constituents, flower yield and essential oil content in Damask rose (*Rosa damascena* Mill.): A review. *J. Plant Nutrit.*, 46(18), 4532-4558.
 1221. Kapler A. 2021. Biologia, ekologia i możliwości uprawy mniej znanych roślin bogatych w polifenole. w: Małgorzata Kalembe-Drożdż Agata Grzywacz-Kisiełewska Agnieszka Cierniak (red.) Surowce polifenolowe Zastosowania i perspektywy. Conference: Surowce polifenolowe. Zastosowania i perspektywyAt: Krakowska Akademia im. Andrzeja Frycza Modrzewskiego, Kraków.
 1222. Karageorgiou, E.M., 2023. Development of a high-performance liquid chromatography coupled to a photodiode detector (HPLC-DAD) method for the determination of selected phenolic compounds found in plant infusions. PhD thesis, University of West Attika.
 1223. Katekar VP, Rao AB, Sardeshpande VR. A cleaner and ecological rosewater production technology based on solar energy for rural livelihood. *Cleaner Circ. Bioecon.*, 2022, 16:100022.
 1224. Katekar, V.P., Rao, A.B. and Sardeshpande, V.R., 2022. Review of the rose essential oil extraction by hydrodistillation: An investigation for the optimum operating condition for maximum yield. *Sustain. Chem. Pharm.*, 29, 100783.
 1225. Kędzierska, M., Sala, K., Bańkosz, M. et al. 2023. Investigation of physicochemical

- properties and surface morphology of hydrogel materials incorporating rosehip extract. *Materials*, 16(17), 6037.
1226. Khoei, S., Madadi, M., 2023. Medicinal plant-based terpenoids in nanoparticles synthesis, characterization, and their applications. In: *Secondary Metabolites from Medicinal Plants* (pp. 53-86). CRC Press.
 1227. Koljančić, N., Vyvirska, O. Španik, I., 2023. Aroma compounds in essential oils: analyzing chemical composition using two-dimensional gas chromatography–high resolution time-of-flight mass spectrometry combined with chemometrics. *Plants*, 12(12), 2362.
 1228. Koraqi, H., Aydar, A.Y., Khalid, W. et al. (2024). Ultrasound-assisted extraction with natural deep eutectic solvent for phenolic compounds recovery from *Rosa damascena* Mill.: Experimental design optimization using central composite design. *Microchem. J.*, 196, 109585.
 1229. Koyama S, Heinbockel T. Chemical constituents of essential oils used in olfactory training: focus on COVID-19 induced olfactory dysfunction. *Front. Pharmacol.*, 2022, 2:1550.
 1230. Lekshmi, S. G., Sethi, S., Pooja, B. K. et al. (2023). Ornamental plant extracts: Application in food colouration and packaging, antioxidant, antimicrobial and pharmacological potential—A concise review. *Food Chem. Adv.*, 3, 100529.
 1231. Kunc, N., Hudina, M., Osterc, G., & Grohar, M. C. (2024). Determination of volatile compounds in blossoms of *Rosa spinosissima*, *Rosa pendulina*, *Rosa gallica*, and their cultivars. *Agriculture*, 14(2), 253.
 1232. Lee, Y., E. Park, B Jang, et al. Antifungal activity of Bulgarian rose damascena oil against vaginitis-causing opportunistic fungi. *Evidence-Based Compl. Altern. Med.*, 2023, 1-9
 1233. Li, J., Yao, A., Yao, J. et al. 2022. Dynamic profiles of rose jam metabolomes reveal sugar-pickling impacts on their nutrient content. *Food Biosci.*, 101947.
 1234. Li, M., Zhang, Y., Xi, H. et al. 2022. Characterization of rose essential oils by double-region atmospheric pressure chemical ionization mass spectrometry (DRAPCI-MS) with principal component analysis (PCA), hierarchical cluster analysis (HCA), and heatmap analysis. *Anal. Lett.*, 55(15), 2382-2393.
 1235. Liu, X., Wang, S., Cui, L. et al. 2023. Flowers: precious food and medicine resources. *Food Science and Human Wellness*, 12(4), 1020-1052.
 1236. Malek, M.A., B Gowda, S.G., Gowda, D., & Hui, S. P. (2024). Analysis of lipid composition and characterization of acyl steryl glycosides in rose petals by using nontargeted LC/MS. *J. Food Measur. Charact.*, 1-12.
 1237. Martínez Mena, M. (2023). *La Botánica dentro del Perfume*. (Master thesis, Universidad de Sevilla, Sevilla).
 1238. Musolino, V., Marrelli, M., Perri, M.R. et al. (2022). *Centranthus ruber* (L.) DC. and *Tropaeolum majus* L.: phytochemical profile, in vitro anti-denaturation effects and lipase inhibitory activity of two ornamental plants traditionally used as herbal remedies. *Molecules*, 28(1), 32.
 1239. Oargă, D. P., Cornea-Cipcigan, M., & Cordea, M. I. (2024). Unveiling the mechanisms for the development of rosehip-based dermatological products: an updated review. *Frontiers in Pharmacology*, 15, 1390419.
 1240. Noh, Y. M., Ait Hida, A., Raymond, O. et al. (2024). The scent of roses, a bouquet of fragrance diversity. *J. Exp. Botani*, 2024, 75, 5, 1252-1264.
 1241. Parra-Pacheco, B., Cruz-Moreno, B. A., Aguirre-Becerra, H. et al. (2024). Bioactive compounds from organic waste. *Molecules*, 29(10), 2243..

1242. Pirker, T., Pferschy-Wenzig, E., Bauer, R. Inhibition of COX-2 mRNA expression by damask rose flowers. In: eBook of Abstracts (for personal use and information of participants throughout the conference, not for citation; abstracts will be published in a special issue of *Planta Medica* depending on agreement of authors). GA – 69th Annual Meeting 2021, Bonn, Germany Virtual conference, 5 – 8 Sept., Society for Medical Plant and Natural Product Research 2021, PC 9 – 44, 191.
1243. Pirker, T., Pferschy-Wenzig, E., & Bauer, R. (2021). Inhibition of COX-2 mRNA expression by damask rose flowers. *Planta Medica*, 87(15), PC9-44.
1244. Prants, K., 2022. *Rosa damascena* Mill roosi kroonlehtede mõju õunasiidri fermentatsioonile (Master's thesis, Eesti Maaülikool).
1245. Rajabzadeh, S., Pirbalouti, A. G., Yadegari, M., & Rahimi, T. (2024). Physiological and phytochemical responses of *Rosa damascena* Mill. to the foliar application of different elicitors. *J. Essent. Oil Bear. Plants*, 27(1), 272–284.
1246. Rani, M.L., Indumathy, V., Vinodhini, M. et al, 2022. Therapeutic value of selected traditional flowers - a review. *J. Pharm. Negative Results*, 6496-6511.
1247. Ren, H., Yang, W., Jing, W. et al. (2024). Multi-omics analysis reveals key regulatory defense pathways and genes involved in salt tolerance of rose plants. *Horticult. Res.*, 11, 5, uhae068.
1248. Shang, Y. F., Hao, W. D., Zhang, W. et al., Non-targeted LC-MS metabolite profiling: contrasting water distillation and methanol extraction of *Rose Damascens* Mill., 2024, 1-19.
1249. Simin, N., Živanović, N., Božanić Tanjga, B. et al. (2024). New garden rose (*Rosa* × *hybrida*) genotypes with intensely colored flowers as rich sources of bioactive compounds. *Plants*, 13(3), 424.
1250. Slaga, T.J., Snyder, P.W. Safety assessment of *Rosa centifolia*-derived Ingredients as used in cosmetics. Scientific Literature Review for Public Comment, 2021, 13-14.
1251. Sofiya Karunanithi G., Bharath Kumar. Study on the Effect of Dual Solvent Proportions on Composition of Rosa x Damascena Concrete Oil Obtained using Soxhlet Extraction Method. *Asian J. Chem.* / 2022 / 34(1) / pp 78-84.
1252. Sofiya, K. and Kumar, G.B., 2022. Study on the Effect of Dual Solvent Proportions on Composition of Rosa x damascena Concrete Oil Obtained using Soxhlet Extraction Method. *Asian Journal of Chemistry*, (1), 78-84.
1253. Sumbul, S.A, Heyat MBB, Rahman K, et al. (2024), Efficacy and classification of *Sesamum indicum* linn seeds with *Rosa damascena* mill oil in uncomplicated pelvic inflammatory disease using machine learning. *Front. Chem.* 12:1361980.
1254. Syukri, D., & Haiyee, Z. A. (2024). Optimum Condition for the Formation of β-ionone by Thermal Decomposition of Carotenoids Extract from Orange Sweet Potato (L.). *The Open Agricult. J.*, 18(1).
1255. T Pirker, E Pferschy-Wenzig, R Bauer. Inhibition of COX-2 mRNA expression by damask rose flowers. *Planta Med* 2021; 87(15): 1309 DOI: 10.1055/s-0041-1736970.
1256. Tanjga, B.B., Lončar, B., Aćimović, M. et al. 2022. Volatile profile of garden rose (*Rosa hybrida*) hydrosol and evaluation of its biological activity in vitro. *Horticulturae*, 8(10), 895.
1257. Tsygankova V.A., Oliynyk O.O., Kvasko O.Y. et al. Effect of plant growth regulators ivin, methyur and kamethur on the organogenesis of miniature rose (*Rosa mini* L.) in Vitro. *Int J Med Biotechnol Genetics*. 2022;S1:02:001:1-8.
1258. Wang, C., Kim, I.J., Seong, H.R. et al. 2023. Antioxidative and Anti-Inflammatory

- Activities of Rosebud Extracts of Newly Crossbred Roses. *Nutrients*, 15(10), 2376.
1259. Wang, L., Yu, M., Ding, S. et al. (2023). Zebrafish models for the evaluation of essential oils (EOs): A comprehensive review. *Qual. Assur. Safety Crops & Foods*, 15(4), 156-178.
1260. Xu, X., Wang, N., Feng, L. and Wang, J., 2023. Simple sequence repeat fingerprint identification of essential-oil-bearing *Rosa rugosa* via high-resolution melting (HRM) analysis. *Biomolecules*, 13(10), 1468.
1261. Younis, I.Y., El-Hawary, S.S., Eldahshan, O.A. et al. Green synthesis of magnesium nanoparticles mediated from *Rosa floribunda* charisma extract and its antioxidant, antiaging, and antibiofilm activities. *Sci. Rep.*, 11, 16868 (2021).
1262. Yousefi, B., Shahbazi, K., Khamis-Abady, H., and Gheitury, M., 2021. Assessment of variability of essential oil components in different accessions of Damask rose (*Rosa damascena* Mill.) by multivariate analysis. *Trends Phytochem. Res.*, 5(4), 199-208.
1263. Zhai, D., Hu, Y., Liu, L. Et al. (2023). Exploring the molecular mechanism of Licorice rose beverage anti-melasma based on network pharmacology, molecular docking technology and in vivo and in vitro experimental verification. *Heliyon*, 9(12).
1264. Zhang, X., D. Chen, M. Long, et al. Study on the quality stability of rose effervescent tablets and evaluation of antioxidant activity." *CONVERTER* 2021, 7 (2021):933-946.
1265. Zumsteg, J., Bossard, E., Gourguillon, L. Et al. (2023). Comparison of nocturnal and diurnal metabolomes of rose flowers and leaves. *Metabolomics*, 20(1), 4.
- 64). Grozdanova, T., Trusheva, B., Alipieva, K., Popova, M., Dimitrova, L., NAJDENSKI, H., Zaharieva, M. M., Ilieva, Y., Vasileva, B., Miloshev, G., Georgieva, M., Bankova, V. Extracts of medicinal plants with natural deep eutectic solvents: enhanced antimicrobial activity and low genotoxicity. *BMC Chemistry*, 14, 73, Springer Nature, 2020, ISSN:2661-801X, DOI:10.1186/s13065-020-00726-x, 1-9. SJR (Scopus):0.426, JCR-IF (Web of Science):2.493**
1266. Abdallah, M. (2022). Extraction of bioactives from marine by-products using deep eutectic solvents for dry eye disease treatment. PhD thesis, Universidade Nova de Lisboa.
1267. Afonso, A. C., et al. Phytochemicals against drug-resistant bacterial biofilms and use of green extraction solvents to increase their bioactivity. *Adv. Exp. Med. Biol.*, Cham: Springer International Publishing, 2022. 1-18.
1268. Akbar, N., Khan, N.A., Ibrahim, T. et al. 2023. Antimicrobial activity of novel deep eutectic solvents. *Scientia Pharmaceutica*, 91(1), p.9.
1269. Alioui, O. et al. Theoretical and experimental evidence for the use of natural deep eutectic solvents to increase the solubility and extractability of curcumin. *J. Mol. Liq.*, 359 (2022):119149.
1270. Baby, J.N., Akila, B., Chiu, T.W. et al. 2023. Deep eutectic solvent-assisted synthesis of a strontium tungstate bifunctional catalyst: investigation on the electrocatalytic determination and photocatalytic degradation of acetaminophen and metformin drugs. *Inorg. Chem.*, 62(21), 8249-8260.
1271. Bagheri, H., S. Ghader, S. AbdulAmeer, N. Ahmad Comprehensive study on deep eutectic solvent density based on various EoSs: SRK, PT, VTSRK, sPC-SAFT. *J. Mol. Liq.*, 393, 2024, 123627.
1272. Brahmi-Chendouh, N., Piccolella, S., Gravina, C. et al. Ready-to-use nutraceutical formulations from edible and waste organs of algerian artichokes. *Foods*, 2022, 11, 24 C7-3955.

1273. Bragagnolo, F.S., M.M. Strieder, R.S. Pizani, et al. Revisiting natural deep eutectic solvents (NADES) as extraction media and ready-to-use purposes. *TrAC Trends Anal. Chem.*, 175, 2024, 117726.
1274. Chanioti, S., Siamandoura, P. and Tzia, C., 2023. Green technologies in food processing. In: *Green Chemistry in Agriculture and Food Production* (pp. 150-197). CRC Press.
1275. de Oliveira, F.L. et al. Fruit by-products as potential prebiotics and promising functional ingredients to produce fermented milk. *Food Res. Int.* (2022):111841.
1276. Emsen, B., Surmen, B. and Karapinar, H.S., 2023. In vitro antioxidant and cytotoxic effects of three endemic plants from Turkey based on their phenolic profile. *Plant Biosystems-An International Journal Dealing with all Aspects of Plant Biology*, 157(2), 346-356.
1277. Fuad F.M., M.M. Nadzir, A. Harun Kamaruddin. Hydrophilic natural deep eutectic solvent : A review on physicochemical properties and extractability of bioactive compounds. *J. Mol. Et al.* 339 (2021) 116923.
1278. García, C.B., Concha, J., Culleré, L. et al. 2023. Has the toxicity of therapeutic deep eutectic systems been assessed? *Appl. Sci.*, 13(10), 5980.
1279. García-Valcarcel, A. I., & Martín-Esteban, A. (2024). Ultrasound-assisted extraction of sulfonamides from soil samples using natural deep eutectic solvents and their determination by liquid chromatography tandem mass spectrometry. *Microchem. J.*, 110850.
1280. Guo, X., Ren, T., Li, H., Lu, Q., & Di, X. (2024). Antioxidant activity and mechanism exploration for microwave-assisted extraction of flavonoids from *Scutellariae Radix* using natural deep eutectic solvent. *Microchem. J.*, 200, 110300.
1281. Hemaiswarya, Shanmugam, Pranav Kumar Prabhakar, and Mukesh Doble. Synergistic Herb Interactions with Anticancer Drugs. *Herb-Drug Combinations: A New Complementary Therapeutic Strategy*. Singapore: Springer Nature Singapore, 2022. 145-73.
1282. Hikmawanti NPE, Ramadon D, Jantan I, Mun'im A. Natural deep eutectic solvents (NADES): phytochemical extraction performance enhancer for pharmaceutical and nutraceutical product development. *Plants*. 2021; 10(10):2091.
1283. Houël, E. et al. Treating leishmaniasis in Amazonia, part 2: Multi-target evaluation of widely used plants to understand medicinal practices. *J. Ethnopharmacol.* 289 (2022): 115054.
1284. Joarder, S., Bansal, D., Meena, H. et al. 2023. Bioinspired green deep eutectic solvents: preparation, catalytic activity, and biocompatibility. *J. Mol. Liq.*, 121355.
1285. Koh, Q.Q., Kua, Y.L., Gan, S. et al. 2023. Sugar-based natural deep eutectic solvent (NADES): Physicochemical properties, antimicrobial activity, toxicity, biodegradability and potential use as green extraction media for phytonutrients. *Sust. Chem. Pharm.*, 35, 101218.
1286. Lai, Z.Y., Yiin, C.L., Lock, S.S.M. et al. 2023. A review on natural based deep eutectic solvents (NADESs): fundamentals and potential applications in removing heavy metals from soil. *Env. Sci. Pollut. Res.*, 30(55), 116878-116905.
1287. Lazović, M., Ivković, Đ., Jankov, M., et al. (2024). Enhancement of propolis food preservation and functional ingredient characteristics by natural eutectic solvents extraction of phytochemicals. *Food Biosci.*, 57, 103467.
1288. Leal, M., Moreno, M.A., Albornoz, P.L. et al. 2023. Morphological characterization of *nicotiana tabacum* inflorescences and chemical-functional analysis of extracts obtained from its powder by using green solvents (NaDESs). *Plants*, 12(7), 1554.
1289. Li, Y., Sun, M., Cao, Y. et al. (2024). Designing low toxic deep eutectic solvents for the green recycle of lithium-ion batteries cathodes. *ChemSusChem*, e202301953.

1290. Molnar, M., Gašo-Sokač, D., Komar, M. et al. (2024). Potential of deep eutectic solvents in the extraction of organic compounds from food industry by-products and agro-industrial waste. *Separations*, 11(1), 35.
1291. Nikbazzm, R., Rahimi, Z., Moradi, Y. et al. The effect of cranberry supplementation on *Helicobacter pylori* eradication in *H. pylori* positive subjects: A systematic review and meta-analysis of randomised controlled trials. *British J. Nutrit.* 2022, 128, 1090-1099.
1292. Nguyen, T.T., Nguyen, D.V., Tran, Q.H., et. al. (2024). Choline chloride based natural deep eutectic solvents coupling with ultra-high performance liquid chromatography-tandem mass spectroscopy for effective extraction and rapid detection of adenosine and cordycepin in *Cordyceps militaris*. *J. Mol. Liq.*, 397, 124107.
1293. Ozkan, G. (2023). Valorization of artichoke outer petals by using ultrasound-assisted extraction and natural deep eutectic solvents (NADES) for the recovery of phenolic compounds. *J. Sci. Food Agricult.*, 104, 5, 2744-2749.
1294. Panossian, A. (2023). *Herba Sideritis*: a putative adaptogen for reducing the risk of age-related cognitive decline and neurodegenerative disorders. *Phytomed. Plus*, 100519.
1295. Ramon, M. R., Durand, E., Garcia-Sosa, K., & Peña-Rodríguez, L. M. (2023). Exploring the potential of deep eutectic solvents (DES) in bioactive natural product research: from DES to NaDES, THEDES, and beyond. *PeerJ Analyt. Chem.*, 5, e28.
1296. Sailau, Z. et al. Studying the formation of choline chloride-and glucose-based natural deep eutectic solvent at the molecular level. *J. Mol. Model.*, 28.8 (2022):1-8.
1297. Saputri, F. A., Mun'im, A., Putri, C. R., & Aryani, D. (2022). Validasi metode analisis kurkuminoid dan xantorizol pada rimpang temulawak (*Curcuma xanthorrhiza*) dengan KLT-densitometri. *MPI (Media Pharmaceutica Indonesiana)*, 4(2), 147-156.
1298. Schneider, H. (2023). Reactive eutectic media based on ammonium formate for the valorization of bio-sourced materials (Doctoral dissertation, Universität Potsdam).
1299. Schuh, L., Reginato, M., Florêncio, I. et al. (2023). From nature to innovation: the uncharted potential of natural deep eutectic solvents. *Molecules*, 28(22), 7653.
1300. Sherlin V.A., Joseph, X.B., Wang, S.F. et al. Natural deep eutectic solvent assisted synthesis of FeMnO₃ entrapped functionalized carbon nanofiber composite: An electrochemical detection of nimesulide. *J. Mol. Liq.*, 2022, 367 C7 – 120421.
1301. Syafira, W. A. (2023). Pengaruh natural deep eutectic solvents (NADES) terhadap aktivitas antioksidan ekstrak bekatul (*Oryza sativa* L.) (Doctoral dissertation, Universitas Islam Negeri Maulana Malik Ibrahim).
1302. Siddiqui, R., Khodja, A., Ibrahim, T. et al. 2023. The increasing importance of novel deep eutectic solvents as potential effective antimicrobials and other medicinal properties. *World J. Microbiol. Biotechnol.*, 39(12), 330.
1303. Susawaengsup, C., Choengpanya, K., Sornsakdanuphap, J. et al 2023. Phytochemical and pharmacological properties of a traditional herb, *Strobilanthes Cusia* (Nees) Kuntze. *Mol. Biotechnol.*, 1-12.
1304. Thakur, M., Modi, V. K. Biocolorants in food: Sources, extraction, applications and future prospects. *Crit. Rev. Food Sci. Nutr.*, 2022, 6, 2024, 14, 4674-4713.
1305. Torres-Vega, J. et al. Polyphenolic compounds extracted and purified from *Buddleja globosa* Hope (Buddlejaceae) leaves using natural deep eutectic solvents and centrifugal partition chromatography. *Molecules* 26.8 (2021): 2192.
1306. Triasih, R. (2022). Aktivitas antijamur ekstrak kulit biji kakao (*Theobroma cacao* L.)

- dengan natural deep eutectic solvent terhadap *Candida albicans* (Doctoral dissertation, Universitas dr. Soebandi).
1307. Tzani, A., Karadendrou, M. A., Tsiaka, T. et al. Exploring the role of natural deep eutectic solvents (NADES) towards the valorization of food processing industry waste. Deep eutectic solvents: properties, applications and toxicity. Nova Sci. Publishers, Inc., 2022, 19-52.
 1308. Wong, Y.S., R. Yusoff, G.C. Ngoh. Phenolic compounds extraction by assistive technologies and natural deep eutectic solvents. Rev. Chem. Eng., 2024, 40, 2, 24, 229-246.
 1309. Zengin, G., Cádiz-Gurrea, M.D.L.L., Fernández-Ochoa, Á. et al. Selectivity tuning by natural deep eutectic solvents (NADESs) for extraction of bioactive compounds from *Cytinus hypocistis*—studies of antioxidative, enzyme-inhibitive properties and LC-MS rofiles. Molecules, 2022, 27, 18 C7 – 5788.
 1310. Zulhendri F, Perera CO, Tandean S. Can propolis be a useful adjuvant in brain and neurological disorders and injuries? A systematic scoping review of the latest experimental evidence. Biomedicines. 2021; 9(9):1227.
- 65). Valcheva V., Savova-Lalkovska T., Vyazovaya A., Dimitrova A., Bonovska M., NAJDENSKI H. First insight into phylogeography of *Mycobacterium bovis* and *M.caprae* from cattle in Bulgaria,. Infection, Genetics and Evolution, 81, Elsevier, 2020, ISSN:1567-1348, DOI:10.1016/j.meegid.2020.104240, JCR-IF (Web of Science): 2.611**
1311. Krajewska-Wędzina, Monika, et al. *Mycobacterium bovis* transmission between cattle and a farmer in Central Poland. Pathogens 11.10 (2022):1170.
 1312. Krajewska-Wędzina, M., Didkowska, A., & Radulski, Ł. (2022). Występowanie zakażeń *Mycobacterium tuberculosis* complex u zwierząt. Część I. Małe przeżuwacze. Życie Weterynaryjne, 97(08), 546-549.
 1313. Lorente Leal, V. (2023). Nuevas metodologías para el diagnóstico de la tuberculosis bovina. PhD thesis, Universidad Complutense, Madrid.
 1314. Martínez-Lirola, M., Herranz, M., Serrano, S.B. et al. 2023. A One Health approach revealed the long-term role of *Mycobacterium caprae* as the hidden cause of human tuberculosis in a region of Spain, 2003 to 2022. Eurosurveillance, 28(12), 2200852.
 1315. Papaventsis, Dimitrios, et al. Occupational exposure to zoonotic tuberculosis caused by *Mycobacterium caprae*, Northern Greece, 2019. Emerg. Inf. Dis., 27.7 (2021):1997-1999.
 1316. Rîmbu, C.M., Horhoge, C.E., Carp-Cărare, C. et al. (2023). Aspecte privind epidemiologia speciilor *Mycobacterium bovis* și *Mycobacterium caprae* în județul Vaslui, Romania. In Gestionarea fondului genetic animalier–probleme, soluții, perspective (pp. 425-433).
 1317. Romano, G. E. (2021). Achados em genômica comparativa de *Mycobacterium caprae* e seus impactos no metabolismo e crescimento in vitro (Doctoral dissertation, Universidade de São Paulo).
 1318. Seña Acosta, R. (2022). Actividad antituberculosa in vitro de nuevos híbridos de chalconas, sulfonamidas y pirazolinas, frente a *Mycobacterium bovis* y *Mycobacterium tuberculosis*. Doctoral dissertation.
 1319. Zhang, S., Chen, C., Rao, Z. et al. 2020. Survey of human and bovine tuberculosis infection on dairy farms in southwestern China. Res. Square, <https://doi.org/10.21203/rs.2.16233/v3>
 1320. Тимошенко, В., Музыка, А., Кирикович, С. Et al. (2023). Биоэнергетическая оценка технологических процессов на молочно-товарных фермах и комплексах

различной мощности. In Gestionarea fondului genetic animalier–probleme, soluții, perspective (pp. 352-360).

66). Gevrenova, R, Zaharieva, MM, Kroumov, AD, Voutquenne-Nazabadioko, L, Zheleva-Dimitrova, D, Balabanova, V, NAJDENSKI, H, Konstantinov, S. Gypsophila saponins enhance the cytotoxicity of etoposide in HD-MY-Z lymphoma cells. Food and Chemical Toxicology, 133, Elsevier, 2019, ISSN:0278-6915, DOI:10.1016/j.fct.2019.110777, 110777. SJR (Scopus):0.9, JCR-IF (Web of Science):4.679

1321. Bechkri, S., Magid, A.A., Voutquenne-Nazabadioko, L. et al. 2023. Triterpenoid saponins from *Silene coeli-rosa*. Phytochem. Lett., 54, 50-56.

1322. Bechkri, Sara, et al. "Antioxidant Activity-Guided Isolation of Flavonoids from *Silene Gallica* Aerial Parts. Phytochem. Lett., 50 (2022):61-66.

1323. Cui, X., Ma, X., Li, C. et al. 2023. A review: structure–activity relationship between saponins and cellular immunity. Mol. Biol. Rep., 50(3), 2779-2793.

1324. Cui, X., Ma, X., Li, C., Meng, H., Han, C. A review: structure–activity relationship between saponins and cellular immunity. Mol. Biol. Rep., 2022.

1325. Hemaiswarya, S., P.K. Prabhakar, M. Doble. Synergistic herb interactions with anticancer drugs. Herb-drug combinations: a new complementary therapeutic strategy. Singapore: Springer Nature Singapore, 2022, 145-73.

1326. Kocacık, A. (2021). Çöven Ekstraktının Kurutulması ve Dondurmada Emülgatör Olarak Kullanılmasının Araştırılması (Master's thesis, Fen Bilimleri Enstitüsü).

1327. Liu, Bo, et al. Spirostane saponins with a rearranged A/B ring system isolated from the rhizomes of *Ophiopogon japonicus*. Phytochem., 193 (2022):112975.

1328. Tedeschi, L.O., et al. Nutritional aspects of ecologically relevant phytochemicals in ruminant production. Front. Vet. Sci., 8 (2021):155.

1329. Wang, Xuanbin, et al. Flavonoids and saponins: what have we got or missed? Phytomedicine 109 (2022):154580.

1330. Wei, J., Yang, Y., Peng, Y. et al. 2023. Biosynthesis and the transcriptional regulation of terpenoids in tea plants (*Camellia sinensis*). Int. J. Mol. Sci., 24(8), 6937.

67). Koev K., Donkov N., Stankova N., NAJDENSKI H., Nurgaliev T., Nikov R., Avramov L.. Application of silver antibacterial nanolayers for hard contact lenses coating. IOP Conference Series: Materials Science and Engineering, 618, 1, IOP Publishing, 2019, ISSN:1757-8981, DOI:10.1088/1757-899X/618/1/012028, JCR-IF (Web of Science):0.55

1331. Negut, I., Albu, C., Bitu, B. (2024). Advances in antimicrobial coatings for preventing infections of head-related implantable medical devices. Coatings, 14(3), 256.

1332. Shaker, L., Abdulhadi, S., Al-azzawi, W. et al. (2023). Colorless poly(vinyl pyrrolidone) hydrogel contact lenses synergized with silver nanoparticles. J. Optics. 53(1).

1333. Willcox, Mark, et al. CLEAR-Contact lens wettability, cleaning, disinfection and interactions with tears. Cont. Lens Anter. Eye 44.2 (2021):157-191.

1334. Zhao, L., Zhao, Y., Wei, J. et al. (2021). Antibacterial mechanism of dihydrotanshinone i. Nat. Prod. Comm. 16(2).

68). Mincheva, I, Zaharieva, MM, Batovska, D, NAJDENSKI, H, Ionkova, I,

Kozuharova, E. Antibacterial activity of extracts from *Potentilla reptans* L. Pharmacia, 66, 1, Meditsinski Universitet - Sofia, 2019, ISSN:0428-0296, DOI:10.3897/pharmacia.66.e35293, 7-11. SJR (Scopus):0.158

1335. Copenheaver, C.A., Peterson, J.A., DeBose, K.G. and Barney, J.N., 2023. Non-native plants observed in North America by 18th century naturalists. *Ecoscience*, 30(1), 39-51.

1336. Maha, H.L., Fidrianny, I. and Suciati, T., 2023. An updated review of *Typhonium flagelliforme*: phytochemical compound, pharmacological activities and the use of vitexin and isovitexin as flavonoid compound in cosmetics development. *Pharmacia*, 70(3), 673-680.

1337. Suhendy, H., Fidrianny, I., Insanu, M. (2023). Phytochemical compounds and pharmacological activities of *Ipomoea batatas* L.: An updated review. *Pharmacia*, 70(4), 1283-1294.

1338. Özgünseven, A., A. Özgünseven. *Potentilla speciosa* Willd. var. *speciosa* Willd. ve sekonder metabolitlerinin α -glukozidaz ve tirozinaz inhibitör etkilerinin araştırılması. 2021.

69). Magdalena Bonovska, Tanya Savova, Reneta Petrova, Violeta Valcheva, HRISTO NAJDENSKI. Cases of paratuberculosis in deer in Bulgaria. Comptes rendus de l'Academie Bulgare des Sciences, 72, 3, BAS, 2019, ISSN:2367-5535, DOI:10.7546/CRABS.2019.03.18, 422-428. JCR-IF (Web of Science):0.321

1339. Ekundayo, T. C., & Okoh, A. I. (2020). Systematic assessment of mycobacterium avium subspecies paratuberculosis infections from 1911–2019: a growth analysis of association with human autoimmune diseases. *microorganisms*, 8(8), 1212.

70). Balabanova, V, Voynikov, Y, Zheleva-Dimitrova, D, Gevrenova, R, Zaharieva, MM, NAJDENSKI, H. Preliminary study on bioactive fractions from sudanese plant *solanum schimperianum* hochst. Comptes Rendus de L'Academie Bulgare des Sciences, 71, 5, Bulgarian Academy of Sciences, 2018, ISSN:1310-1331, DOI:10.7546/CRABS.2018.05.07, 633-639. SJR (Scopus):0.205, JCR-IF (Web of Science):0.321

1340. Li J., Liu D., Tian X. et al. Novel antibacterial modalities against methicillin resistant *Staphylococcus aureus* derived from plants. *Crit. Rev.Food Sci. Nutrit.*: 1-9, 2018.

71). Poehlein, Anja, NAJDENSKI, HRISTO, Simeonova, D, Diliana D.. Draft genome sequence of *Klebsiella pneumoniae* subsp. *pneumoniae* ATCC 9621. Genome announcements, 5, 12, ASM, 2017, DOI:10.1128/genomeA.01718-16, SJR (Scopus):0.217

1341. Carraturo, F., Del Giudice, C., Compagnone, M. et al. (2021). Evaluation of microbial communities of bottled mineral waters and preliminary traceability analysis using NGS microbial fingerprints. *Water*, 13(20), 2824.

1342. Kumru, S., Tekedar, H. C., Blom, J. et al. (2020). Genomic diversity in flavobacterial pathogens of aquatic origin. *Microb.Pathogen.*, 142, 104053.

1343. Leigh, S. C., Catabay, C., German, D. P. (2022). Sustained changes in digestive physiology and microbiome across sequential generations of zebrafish fed different diets. *Comp. Biochem. Physiol. Part A: Molec. & Integr. Physiol.*, 273, 111285.

1344. Mitrea, Laura, et al. *Klebsiella pneumonia* - a useful pathogenic strain for biotechnological purposes: 1, 3-propanediol biosynthesis." (2019).

1345. Sverdrup, A. E., Frolova, L. L., Sagdeeva, S. (2019). The monitoring of Sredniy Kaban lake by 16S rRNA gene amplicon data set-based bacterial diversity. *Journal of*

Environmental Treatment Techniques, 7(Special Issue), 1016-1025.

1346. Фролова, Л.Л., Свердруп, А.Э., Маланин, С.Ю et al. Метагеном гидробионтов озер Кабан города Казани: анализ видового разнообразия гидробионтов по маркерным генам. 2019, 218.

72). Gotova I., Dimitrov Z., NAJDENSKI H. Selected *Lactobacillus bulgaricus* and *Streptococcus thermophilus* Strains from Bulgarian Yogurt Demonstrate Significant Anti-Inflammatory Potential. Acta Microbiologica Bulgarica, 33, 3, Bulgarska Akademiia Na Naukite, 2017, ISSN:0204-8809, 137-142.

1347. Bharindwal, S., Goswami, N., Jha, P. et al. R. Prospective use of probiotics to maintain astronaut health during spaceflight. Life 2023, 13, 727.

1348. Palanivelu, J., et al. Probiotics in functional foods: survival assessment and approaches for improved viability. Appl. Sci., 12.1 (2022):455.

1349. Petrova, P. et al. Traditional bulgarian dairy products: ethnic foods with health benefits. Microorganisms 9.3 (2021):480.

73). Golkocheva-Markova, E., R. Nenova, R. Stoilov, I. Christova, H. NAJDENSKI. Cross-reactivity between *Yersinia* outer membrane proteins and anti-Francisella and anti-Borrelia antibodies in serodiagnosis of *Yersinia*-triggered reactive arthritis.. Compt. rend. Acad. bulg. Sci., 64, 1, BAS, 2011, 61-66. JCR-IF (Web of Science):0.206

1350. Djoumerska-Alexieva, Iglia, and Tchavdar Vassilev. "Enhanced Binding Polyspecificity of Human IgG after Acid Treatment." Comptes rendus de l'Académie bulgare des Sciences 64.8 (2011).

1351. Grażlewska, W. and Holec-Gąsior, L., 2023. Antibody Cross-Reactivity in Serodiagnosis of Lyme Disease. Antibodies, 12(4), p.63.

74). Batovska, D., Todorova, I., Parushev, S., Tsvetkova, I., NAJDENSKI, H., Ubukata, M.. Evaluation of antibacterial activity of synthetic aliphatic and aromatic monoacylglycerols. 57, 3, Polskie Towarzystwo Mikrobiologów Polish Society of Microbiologists, 2008, 261-265. ISI IF:0.99

1352. Chagas, F.O. et al. Expanding the chemical repertoire of the endophyte streptomyces albospinus RLe7 reveals amphotericin B as an inducer of a fungal phenotype. J. Nat. Prod. (2017), 80(5), 1302-1309

1353. Вензель, С.В., Каннингем, К.Т., Кишник, Э.Р., & Чаудхари, В. Антибактериальная композиция, содержащая сложный эфир бензойной кислоты, и способы подавления бактериального роста посредством ее применения. Патент на изобретение, RU 2729079 C1, 2020.

75). Iliev, M., H. NAJDENSKI. Optimisation of PCR protocol for detection and differentiation of pathogenic serotypes of *Yersinia enterocolitica* in milk. Compt. rend. Acad. bulg. Sci., 61, 10, BAS, 2008, ISSN:1310-1331 (Print), 1271-1278. SJR (Scopus):0.22

1354. Rowe M, J. Donaghy. Microbiological aspects of dairy ingredients. Dairy Ingredients for Food Processing (2011):59-101.

76). Yordanov M., Golkocheva E., NAJDENSKI H.. Modulation of complement activity in Vitro and in Vivo by *Yersinia* wild and mutant strains. Folia Microbiologica,

51, Springer Netherlands, 2006, ISSN:00155632, 18749356, 27-32. JCR-IF (Web of Science):1.144

1355. Железникова, Г.Ф., М.К. Бехтерева. Энтеропатогенные иерсинии и иммунная система хозяина (часть 1). Российский иммунологический журнал 4.2 (2010):111-122.

77). Bengoechea J. A., NAJDENSKI H., Skurnik M. Essential role of the lipopolysaccharide O-antigen status of *Yersinia enterocolitica* O:8 in the complex outer membrane regulatory network including outer membrane components. Molecular Microbiology, 52, Wiley-Blackwell Publishing Ltd, 2004, ISSN:0950-382X, 451-469. SJR (Scopus):2.25, JCR-IF (Web of Science):6.108

1356. Паунова-Кръстева, Ц.С. Фенотипни вариации свързани с полизахаридните антигени при *Escherichia coli* O157. Дисертация за присъждане на образователната и научна степен „доктор”. София 2015

78). NAJDENSKI H., Vesselinova A. Experimental mixed infection of rabbits with *Yersinia enterocolitica* and *Listeria monocytogenes*.. Journal of veterinary medicine. B, Infectious diseases and veterinary public health, 49, 2002, ISSN:0931-1793, DOI:10.1046/j.1439-0450.2002.00514.x, 97-104

1357. Bartling, C. et al. Prevalence of anti-*Yersinia* antibodies in European brown hares in North-Rhine Westphalia, Germany. Deutsche Tierärztliche Wochenschrift, 111(6), 259–264, 259.

1358. El-Ghany W.A.A. Listeriosis in rabbits (*Oryctolagus cuniculus*): a significant bacterial disease with an emerging zoonosis. J. Adv. Vet. Res., 13 (1), 150-156. 2023.

1359. Nikolaou, K., Hensel, A., Bartling, C. et al. Prevalence of anti-*Yersinia* outer protein antibodies in goats in lower Saxony. (2005) J. Vet. Med. Series B: Inf. Dis. Vet. Public Health, 52(1), 17-24.

79). Ivanova, E, Yanchev, I, NAJDENSKI, H, Toshkova, R, Dimitrova, P, Manov, V. Studies on the interactions of immunostimulated macrophages and *Yersinia enterocolitica* O:8. Canadian Journal of Microbiology, 46, 3, NRC Research Press, 2000, ISSN:0008-4166, DOI:10.1139, 218-228. SJR (Scopus):0.667, JCR-IF (Web of Science):1.327

1360. Allan EJ, Hoischen C, & Gumpert J. Bacterial L-forms. Adv. Appl. Microbiol., 2009, 68, 1-39.

80). Ivanova, E., R. Toshkova, H. NAJDENSKI, V. Ivanova, M. Kolarova, V. Yanchev. Effect of *Ulva Lactuca* heteropolysaccharide on the immune response of *Yersinia pseudotuberculosis* infected mice.. Compt. rend. Acad. bulg. Sci., 49, 4, BAS, 1996, ISSN:1310-1331 (Print), 85-89. SJR (Scopus):0.22

1361. Siddhanta, A. K., et al. Water soluble polysaccharides of marine algal species of *Ulva* (Ulvales, Chlorophyta) of Indian waters. (2001).

81). Veljanov D., Vesselinova A., Nikolova S., Markova N., NAJDENSKI H.. Macrophage damage in vitro by *Listeria monocytogenes* and its listeriolysin: morphological and cytochemical criteria. *Listeria* 1992: 11. International Symposium, Copenhagen (Denmark), 11-14 May 1992, SSI, 1992

1362. Карпова, М. Р. Инфекция и гемопоэз. Федеральное государственное автономное образовательное учреждение высшего образования" Национальный исследовательский Томский государственный университет", 1999.

82). Trusheva B., Todorov I., Ninova M., NAJDENSKI H., Daneshmand A., Bankova V. Antibacterial mono- and sesquiterpene esters of benzoic acids from Iranian propolis. Chemistry Central Journal, 2010, 8, 1-4. ISSN: 1752-153X

1363. Abubakar, M.B., Abdullah, W.Z., Sulaiman, S.A., & Ang, B.S. Polyphenols as key players for the antileukaemic effects of propolis. Evidence-Based Compl. Altern. Med., 2014, 371730, 11.

1364. Adanero Jorge, F. Caracterización de propóleos de Castilla y León= Characterization of Castilla y León propolis. (Doctoral dissertation, Univerisdad de Leon).

1365. Afrouzan, H., Zakeri, S., Mehrizi, A.A. et al. (2017). Anti-plasmodial assessment of four different Iranian propolis extracts. Arch. Iranian Med., (AIM), 20(5).

1366. Ali, I. H., Daoud, A. S., & Shareef, A. Y. (2012). Physical properties and chemical analysis of Iraqi propolis. Tikrit J. Pure Sci., 17(2), 26-31.

1367. Altabbal, S., Athamnah, K., Rahma, A., Wali, A. F., Eid, A. H., Iratni, R., & Al Dhaheri, Y. (2023). Propolis: A detailed insight of its anticancer molecular mechanisms. Pharmaceuticals, 16(3), 450.

1368. Al-Khalifa, K.S., et al. Influence of propolis extract (caffeic acid phenethyl ester) addition on the *Candida albicans* adhesion and surface properties of autopolymerized acrylic resin. Int. J. Dent., 2022, 6118660.

1369. Álvarez L.R.G. Determinación del poder antibiótico in vitro del extracto etanólico del propóleo sobre *Staphylococcus aureus* y *Escherichia coli* presentes en metritis puerperal bovina. 2013, Universidad de Cuenca, Ecuador

1370. Álvarez, P. F. C. (2013). Facultad de ciencias económicas y administrativas escuela de administración de empresas (Doctoral Dissertation, Universidad De Cuenca).

1371. Amarante, J.F., Ribeiro, M.F., Costa, M.M. et al. (2019). Chemical composition and antimicrobial activity of two extract of propolis against isolates of *Staphylococcus* spp. and multiresistant bacterials. Pesquisa Vet. Brasileira, 39, 734-743.

1372. Antón, A.R.S. Atividade de extratos de própolis e de nanopartículas de prata sobre *Candida* spp. isoladas de casos de estomatite protética. 2019, (Doctoral dissertation, Universidade Federal da Bahia, Savador).

1373. Aschenbrenner, A. K., Kwon, M., Conrad, J. et al. (2016). Identification and characterization of two bisabolene synthases from linear glandular trichomes of sunflower (*Helianthus annuus* L., Asteraceae). Phytochemistry, 124, 29-37.

1374. Baudel, M. (2017). L'apithérapie. (Doctoral dissertation, Université Picardie Jules Verne, 123)

1375. Bayram, N., Sorkun, K., Öz, G. et al. (2018). Chemical characterization of 64 propolis samples from Hakkari, Turkey. Records Nat. Prod., 12(6).

1376. Benhanifia, M., Mohamed, W.M. Phenolics constituents of different types of propolis and their antimicrobial activities Anti-Infective Agents. 2015, 13, 1, 17-27.

1377. Benhanifia, M., Soltani, A. 2022. Biological activity of propolis: An update. 2022030009.

1378. Betoloum, S. M., Mbaïogaoun, A., Mbaïhougadobe, S. et al. (2022). Phytochemical tests, assessment of antioxidant properties and isolation of two compounds of ethyl acetate extract of

- Chadian propolis: case of Bebotho (Southern Chad). *Am. J. Anal. Chem.*, 13(7), 241-254.
1379. Bogdanov, V.L. (2012). Influence of initial stresses on the stressed state of a composite with a periodic system of parallel coaxial normal tensile cracks. *J. Mathem. Sci.*, 186, 1-13.
 1380. Bogdanova P.B., Karapetkovska-Hristova, V., Presilski, S. et al. (2017). Assessment of heavy metals in propolis and soil from Pelagonia region, Republic of Macedonia. *Macedonian J. Chem. Chem. Eng.*, 36(1), 23-33.
 1381. Bouaroura ép Redjem, A., & Segueni, N. (2020). Etude comparative du profil chimique et de l'activité antioxydante de plusieurs propolis de l'Est algérien et investigation phytochimique de la propolis la plus active (Doctoral dissertation, Université Frères Mentouri-Constantine 1).
 1382. Cao Xueping, Shen Xiaoge, & Hu Fuliang. (2015). Active substances of propolis against pathogenic microorganisms and their mechanism of action. *Chinese Apiculture*, 66(12), 41-44.
 1383. Chacon-Morales, Pablo A., et al. Hemisynthesis and bactericidal activity of several substituted benzoic acid esters of 13 (S)-labdan-8 α , 15-diol, a diterpene from *Oxylobus glanduliferus*. *Chem. Nat. Comp.*, 55.4 (2019): 677-684.
 1384. Chacon-Morales, P.A., Amaro-Luis, J.M., Rojas Fermin, L.B. et al. (2019). Hemisynthesis and bactericidal activity of several substituted benzoic acid esters of 13 (S)-labdan-8 α , 15-diol, a diterpene from *Oxylobus glanduliferus*. *Chem. Nat. Comp.*, 55, 677-684.
 1385. Chuttong, B., Lim, K., Praphawilai, P. Et al. (2023). Exploring the functional properties of propolis, geopropolis, and cerumen, with a special emphasis on their antimicrobial effects. *Foods*, 12(21), 3909.
 1386. Dahiya, R., & Mourya, R. Synthetic studies on novel nitroquinazolinone analogs with antimicrobial potential. *Bull. Pharm. Res*, 2013, 3(2), 51-7.
 1387. de Lima Silva, W.E., Junior, W.D.F., da Rosa, P.R. et al. 2015. In vitro activity of propolis: synergism in combination with antibiotics against *Staphylococcus* spp. *African J. Microbiol. Res.*, 9(1), 1-5.
 1388. El-Ghany W.A.A. Listeriosis in Rabbits (*Oryctolagus cuniculus*): A Significant Bacterial Disease with an Emerging Zoonosis. *Journal of Advanced Veterinary Research*, 13 (1), pp. 150 – 156. 2023.
 1389. Farooq, S., Azarpira, A., Iqbal Choudhary, M. (2016). Reversal of multi-drug resistance in *Staphylococcus aureus* by natural product-way forward. *Letters in Drug Design & Discovery*, 13(7), 668-675.
 1390. Ferreira, A.R. (2016). Avaliação da atividade antifúngica de ésteres benzoicos estruturalmente relacionados frente a espécies de *Candida*. (Doctoral dissertation, Universidade Federal Da Paraíba).
 1391. Feyzi, N., Ebadi, A., & Dastan, D. (2023). Chingin from *Ferula haussknechtii* as AChE inhibitor and confirmation of the absolute configuration. *J. Biomol. Struct. Dyn.*, 1-10.
 1392. Gajek, G. et al. Antagonistic effects of CAPE (a component of propolis) on the cytotoxicity and genotoxicity of irinotecan and SN38 in human gastrointestinal cancer cells in vitro. *Molecules*, 25.3 (2020):658.
 1393. Gajek, G. (2023). Modulowanie biologicznej aktywności irinotekanu i jego metabolitu (SN38) przez ester feniloetylowy kwasu kawowego (CAPE) w badaniach in

- vitro.(Doctoral dissertation,University of Lodzki).
1394. Gallego, M.G.S. Determinación de la actividad anti-candida albicans de propóleos sonorenses colectados en las diferentes estaciones del año, y sus principales constituyentes, 2021,(Master's thesis, Universidad de Sonora)
 1395. Ghosh, S., McArthur, R., Guo, Z.C. et al. (2017). Evidence for anti-pseudogymnoascus destructans (Pd) activity of propolis. *Antibiotics*, 7(1), 2.
 1396. Ghosh, S., Roy, K., & Pal, C. (2019). Terpenoids against infectious diseases. In *Terpenoids against human diseases* (187-208). CRC Press.
 1397. Gilbert-Girard, S., Reigada, I., Savijoki, K. et al. (2021). Screening of natural compounds identifies ferutinin as an antibacterial and anti-biofilm compound. *Biofouling*, 37(7), 791-807.
 1398. Gilbert-Girard, S. (2021). Biomolecular screening and characterization of antimicrobials acting on biofilms and planktonic bacteria. (Doctoral dissertation, University of Helsinki).
 1399. Giampieri, F. et al. Bee products: an emblematic example of underutilized sources of bioactive compounds. *J. Agr. Food Chem.*, 70.23 (2022):6833-6848.
 1400. Grajales-Conesa, J., Elías-Chirino, J., Lozano-Guzmán, E. et al. (2021). Stingless bees propolis antimicrobial activity in combination with garlic, *Allium sativum* (Amaryllidaceae). *Revista Biol. Tropical*, 69(1), 22-35.
 1401. Guzman, E.L., Guzman, O.D.L.O., Luis, A.C., et al. Interaction between propoleum extracts and ciprofloxacin and levofloxacin for the in vitro inhibition of methicillin-resistant *Staphylococcus aureus* isolates. *African J. Microbiol. Res.*, 2014, 8(10), 1089-1097.
 1402. Guzman, E.D. et al. Efecto antagonica de propoleo sobre ciprofloxacino y levofloxacino. *Quimica Medicinal*, 2016, 91-96. ISSN 2448-914X.
 1403. Hadi, W. (2019). Ekstrak etanol propolis trigona sp. malang indonesia sebagai quorum sensing inhibitor isolat biofilm *Staphylococcus aureus* dari rinosinusitis kronis (Doctoral dissertation, Universitas Brawijaya).
 1404. Hasan, A.E.Z., Ambarsari, L., Widjaja, W.K. et al. Potency of nanopropolis stinglessbee *Trigona* spp Indonesia as antibacterial agent. *IOSR J. Pharm.* 2014, 4, 12,01-09.
 1405. Hibi, T., Ohtsuka, H., Shimasaki, T. et al. (2018). Tschimganine and its derivatives extend the chronological life span of yeast via activation of the Sty1 pathway. *Genes to Cells*, 23(8), 620-637.
 1406. Hochheim, S. et al. A bioguided approach for the screening of antibacterial compounds isolated from the hydroalcoholic extract of the native brazilian bee's propolis using mollicutes as a model. *Front. Microbiol.*, 11 (2020):558.
 1407. Hossain, R. et al. Propolis: An update on its chemistry and pharmacological applications. *Chinese Med.*, 17.1 (2022):1-60.
 1408. Huang, S., Zhang, C.P., Wang, K. et al. (2014). Recent advances in the chemical composition of propolis. *Molecules*, 19(12), 19610-19632.
 1409. Huang, Z. C., Hu, F. Progress of study on the chemical composition of propolis (2008-2012). *Nat. Prod. Res. Developm.*, 2013, 25 (8), 1146-1153.
 1410. Kehinde, A. Z. (2021). Development of novel bioproducts from breadfruit (Doctoral dissertation, University of British Columbia).
 1411. Kim, Y. J., Yong, Y. K., & Aslam, M. S. (2019). Narrative and meta-analytic study of in vivo efficiency of the bioactive compounds of propolis in tooth decay. *Pharmacogn.*

Rev., 13(26).

1412. Kuete, V., Wiench, B., Hegazy, M.E.F., et al. Antibacterial activity and cytotoxicity of selected Egyptian medicinal plants. *Planta Medica-Natural Products and Medicinal Plant Research*, (2012), 78(2), 193.
1413. Lacerda, R. C. C. (2012). Avaliação da composição química e atividade antioxidante da própolis orgânica de *Apis mellifera* visando à preservação ambiental do ecossistema envolvido (Doctoral dissertation, Universidade de São Paulo).
1414. Li, J. et al. Isotschimgine alleviates nonalcoholic steatohepatitis and fibrosis via FXR agonism in mice. *Phytother. Res.*, 2021, 35(6), 3351-3364..
1415. Majdi, M., Liu, Q., Karimzadeh, G., et al. Biosynthesis and localization of parthenolide in glandular trichomes of feverfew (*Tanacetum parthenium* L. Schulz Bip.). *Phytochemistry*, 2011, 72(14), 1739-1750.
1416. Makhlof M.E.M., El-Sheekh M.M., EL-Sayed A.I.M. In vitro antibiofilm, antibacterial, antioxidant, and antitumor activities of the brown alga *Padina pavonica* biomass extract. *International Journal of Environmental Health Research* 2023. DOI: 10.1080/09603123.2023.2165045
1417. Memari, H.R., Pazouki, L., & Niinemets, Ü. (2013). The biochemistry and molecular biology of volatile messengers in trees. In: *Biology, controls and models of tree volatile organic compound emissions* (pp. 47-93). Springer Netherlands.
1418. Miguel, M.G., & Antunes, M.D. Is propolis safe as an alternative medicine?. *J. Pharm. & Bioallied Sci.*, 2011, 3(4), 479-495.
1419. Moniruzzaman, M., Amrah Sulaiman, S., & Gan, S. H. (2017). Phenolic acid and flavonoid composition of Malaysian honeys. *J. Food Biochem.*, 41(2), e12282.
1420. Morales Muñoz, H. (2015). Perfil de HPLC, RMN y actividad biológica de extractos etanólicos de geopropóleos de abejas meliponas de la Región Centro de Veracruz. (Universidad Veracruzana).
1421. Murase, H., Shimazawa, M., Kakino, M., et al. The effects of Brazilian green propolis against excessive light-induced cell damage in retina and fibroblast cells. *Evidence-Based Compl. Altern. Med.*, 2013, 238279.
1422. Naco, J., Borche, M., Stefce, P. et al. (2017). Heavy metal levels of propolis in R. Macedonia. *Научни трудове на Съюза на учените–Пловдив. Серия В: Техника и технологии*, 15, 138-141.
1423. Ohtsuka, H. et al. Tschimganine has different targets for chronological lifespan extension and growth inhibition in fission yeast. *Bioscie. Biotechnol. Biochem.* 86.6 (2022):775-779.
1424. Okhale, S. E., Nkwegu, C., Ugbabe, G.E. et al. (2021). Bee propolis: Production optimization and applications in Nigeria. *J. Pharm. Phytother.*, 13(1), 33-45.
1425. Paul, S., Emmanuel, T., Matchawe, C., et al. Pentacyclic triterpenes and crude extracts with antimicrobial activity from Cameroonian brown propolis sample. *J. Appl. Pharm. Sci.*, 2014, 4, 7, 1-9.
1426. Pereira R.C., Paradas W.C., de Carvalho R.T., de Lima Moreira D., Kelecom A., Passos R.M.F., Atella G.C., Salgado L.T. Chemical Defense against Herbivory in the Brown Marine Macroalga *Padina gymnospora* Could Be Attributed to a New Hydrocarbon Compound. *Plants*, 12 (5), art. no. 1073, 2023. DOI: 10.3390/plants12051073.

1427. Robles-Zepeda, R.E., Martínez, J.H., Escobar, A.G. et al. (2012). Botanical origin and biological activity of propolis. Medicinal plants: Biodiversity and drugs. 1st ed. New York: CRC Press, Taylor & Francis Group, 570-597.
1428. Safari, M., Badban, L., Rashidipour, A. Comparison the protective effects of aqueous extract of iranian propolis in 6-hydroxydopamine-induced model of parkinsonism in male rat with L-DOPA: A behavioral and histological evaluation, Koomesh, 2014, 15, 4, 584-591
1429. Salatino, A., Salatino, M. L. F., & Negri, G. (2021). How diverse is the chemistry and plant origin of Brazilian propolis?. Apidologie, 1-23.
1430. Sajjadi, S.E., Eskandarian, A.A., Shokoohinia, Y. et al. (2016). Antileishmanial activity of prenylated coumarins isolated from *Ferulago angulata* and *Prangos asperula*. Res. Pharm. Sci., 11(4), 324-331.
1431. Santos-Buelga, C., & González-Paramás, A. M. (2017). Phenolic composition of propolis. Bee products-chemical and biological properties, 99-111.
1432. Sarker LS., Demissie ZA., Mahmoud SS. Cloning of a sesquiterpene synthase from *Lavandula x intermedia* glandular trichomes. Planta, 5, 238, 2013, 983-989.
1433. Sarkez, N.H. (2014). Antimicrobial properties of Libyan propolis against *Staphylococcus aureus*. Libyan J. Med. Res, 8, 50-55.
1434. Sawicka, D., Car, H., Borawska, M.H., & Nikliński, J. The anticancer activity of propolis. Folia Histochem. Cytobiol., 2012, 50(1), 25-37.
1435. Shang, H. et al. Effect of propolis supplementation on C-reactive protein levels and other inflammatory factors: A systematic review and meta-analysis of randomized controlled trials. J. King Saud University-Science 32.2 (2020):1694-1701.
1436. Sirma, E. Determination of the chemical content and botanical origin of propolis of Pervari (Siirt) Region (Çemikari, Kovanağzi, Sariyaprak) (Master's thesis, Institute of Science and Technology, Fen Bilimleri Enstitüsü).
1437. Soltani, A., & Benhanifia, M. (2023). Propolis, plant sources and antimicrobial activity: An overview. Anti-Infective Agents, 21(5), 20-39.
1438. Sorimachi, K., & Nakamoto, T. Alternative medicine safety: *Agaricus blazei* and propolis. Combin. Chem. & High Throughput Screen., 2011, 14(7), 616-621.
1439. Suleman, T. (2016). The antimicrobial and chemical properties of South African propolis (Doctoral dissertation, University of the Witwatersrand).
1440. Sultana F., Wahab M.A., Nahiduzzaman M., Mohiuddin M., Iqbal M.Z., Shakil A., Mamun A.-A., Khan M.S.R., Wong L., Asaduzzaman M. Seaweed farming for food and nutritional security, climate change mitigation and adaptation, and women empowerment: A review. Aquaculture and Fisheries, 8 (5), pp. 463 – 480, 2023.
1441. Syaifie, P.H. et al. Computational study of asian propolis compounds as potential anti-type 2 diabetes mellitus agents by using inverse virtual screening with the DIA-DB web server, tanimoto similarity analysis, and molecular dynamic simulation. Molecules 27.13 (2022):3972.
1442. Tamfu, A.N., Munvera, A.M., Botezatu, A.V.D. et al. (2022). Synthesis of benzoyl esters of β -amyrin and lupeol and evaluation of their antibiofilm and antidiabetic activities. Results Chem., 4, 100322.
1443. Tatli Seven, P., Seven, I., Gul Baykalir, B. et al. (2018). Nanotechnology and nano-propolis in animal production and health: An overview. Italian J. Animal Sci., 17(4), 921-930.
1444. Tamfu, A.N. et al. Synthesis of benzoyl esters of β -amyrin and lupeol and evaluation

- of their antibiofilm and antidiabetic activities. *Results Chem.*, 4 (2022):100322.
1445. Tran, T.D. et al. Lessons from exploring chemical space and chemical diversity of propolis components. *Int. J. Mol. Sci.*, 21.14 (2020):4988.
1446. Trinh, X.-T. et al. A Comprehensive review of natural compounds for wound healing: targeting bioactivity perspective. *Int. J. Mol. Sci.*, 23.17 (2022):9573.
1447. Usman, U.Z., Mohamed, M. Analysis of phytochemical compounds in water and ethanol extracts of Malaysian propolis. *Int. J. Pharma Bio Sci.*, 2015, 6(2), P374-P380.
1448. Usman, U.Z., Bakar, A.B.A., Zin, A.A.M., & Mohamed, M. (2017). LC-MS analysis and effects of Malaysian propolis on insulin, glucagon, pancreas and oxidative stress status in streptozotocin-induced diabetic rats. *J. Med. Biomed. Res.*, 16(1), 15-27..
1449. Usman, U. Z., Bakar, A. B. A., & Mohamed, M. (2016). Phytochemical composition and activity against hyperglycaemia of Malaysian propolis in diabetic rats. *Biomed Res*, 27(1), 46-51.
1450. Vranješ, M. (2023). *Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa* (Doctoral dissertation, University of Zagreb).
1451. Wang, Z.Q., Huang, C., Huang, J., et al. The stereochemistry of two monoterpenoid diastereomers from *Ferula dissecta*. *RSC Advances*, 2014, 4(28), 14373-14377.
1452. Yalcin, G., & Lee, C.K. (2019). Recent studies on anti-aging compounds with *Saccharomyces cerevisiae* as a model organism. *Transl. Med. Aging*, 3, 109-115.
1453. Yilmaz, M.A., O. Cakir. Phytochemical constituents of a precious honeybee product: propolis. In: *Biological Activities of Honeybee Products*, 2023, 18-46.
1454. Zaccaria, V. (2018). *Chemical composition and nutraceutical properties of beehive products: focus on propolis*. (Doctoral dissertation, Università Degli Studi Di Pavia)
1455. Zakerkish, M. et al. The effect of Iranian propolis on glucose metabolism, lipid profile, insulin resistance, renal function and inflammatory biomarkers in patients with type 2 diabetes mellitus: a randomized double-blind clinical trial. *Sci. Rep.* 9.1 (2019):7289.
1456. Zhang, C., & Hu, F. Propolis terpenoids. *Nat. Prod. Res. Developm.*, 2012, 24 (7), 976-984.
1457. Zhang, C., & Hu, F. Study of propolis (2010) - overview. *Bee J.*, 2011, 31 (7), 5-8.
1458. Zhang, C., Shun, P. H. & Hu, F. Study of the chemical composition of propolis from different geographical and plant origin. *Chinese Pharm. J.*, 2013, 48 (22), 1889-1892.
1459. Zhou, Y., Wang, C., Xin, F. et al. (2018). Synthesis, insecticidal, fungicidal activities and structure–activity relationships of tschimganin analogs. *Molecules*, 23(6), 1473.
1460. Zingue, S., Nde, C.B.M., Michel, T. et al. (2017). Ethanol-extracted Cameroonian propolis exerts estrogenic effects and alleviates hot flushes in ovariectomized Wistar rats. *BMC Compl. Altern. Med.*, 17, 1-17.
- 83). Trusheva, B., M. Popova, H. NAYDENSKI, I. Tsvetkova, J.G. Rodriguez, V. Bankova. New polyisoprenylated benzophenones from Venezuelan propolis. *Fitoterapia*, 75(7-8), 683-689 (2004)**
1461. Abdel-Kader, M.S., Hefnawy, M.M., & Al-Majed, A.R.A. Natural phenolic compounds: planar chromatography separation, In: *Encyclopedia of Chromatography*, 3rd Edition, 2011. DOI: 10.1081/E-ECHR3-120043819
1462. Abe, M., M. Nakada. New construction of the bicyclo [3.3. 1] nonane system via Lewis acid promoted regioselective ring-opening reaction of the tricyclo [4.4. 0.05, 7]

- dec-2-ene derivative. *Tetrahedron Lett.*, 47.36 (2006): 6347-6351.
1463. Ahangari Z., Naseri M., Vatandoost F. (2018). Propolis: chemical composition and its applications in endodontics. *Iran. Endod. J.*, 13 (3), 285-292, 2018.
 1464. Al Marghitas, Liviu, Daniel S. Dezmirean, and Otilia Bobis. "Important developments in Romanian propolis research." *Evidence-Based Complementary and Alternative Medicine* (2013).
 1465. Alday, Efrain, et al. Advances in pharmacological activities and chemical composition of propolis produced in Americas. *Beekeeping and Bee Conservation - Advances in Research* (2016).
 1466. Adanero Jorge, F. Caracterización de propóleos de Castilla y León= Characterization of Castilla y León propolis. Doctoral dissertation, Universidade de Leon.
 1467. Antón, A.R.S. Atividade de extratos de própolis e de nanopartículas de prata sobre *Candida* spp. isoladas de casos de estomatite protética. 2019, (Doctoral dissertation, Universidade Federal da Bahia, Savador).
 1468. Arce Calvario, K.R. (2011). Evaluación in vitro de la actividad antibacteriana de propóleos sonorenses en aislados clínicos de *Staphylococcus aureus* y *Enterococcus* spp. Doctoral dissertation, Universidad de Sonora.
 1469. Calvario, A., K. Rocío. Evaluación in vitro de la actividad antibacteriana de propóleos sonorenses en aislados clínicos de *Staphylococcus aureus* y *Enterococcus* spp. (2011).
 1470. Baghdad, Hicham. Etude de la composition chimique et evaluation biologique de la propolis de plusieurs regions de Tlemcen (Algerie, Doctoral Dissertation, 13-03-2018.
 1471. Begum, A.B, Khanum, N.F., Ranganatha, V. L., et al. Evaluation of benzophenone-N-ethyl morpholine ethers as antibacterial and antifungal activities. *J. Chem.*, 2014, Article ID 941074, 6 pages.
 1472. Begum, A.B., Khanum, N.F., Naveen, P., et al. Efficacy of 5-(2-aryloxy methyl-2-phenyl-1, 3, 4-oxadiazoles as antibacterial and antifungal agents. *J. Appl. Pharm. Sci.*, 2013, 3(11), 105-109.
 1473. Benhanifia, M., Soltani, A. Biological activity of propolis: an update. *Preprints* 2022, 2022030009.
 1474. Benhanifia, M., Mohamed, W.M. Phenolics constituents of different types of propolis and their antimicrobial activities *Anti-Infective Agents* 2015, 13, 17-27.
 1475. Bogdanov, V.L. (2012). Influence of initial stresses on the stressed state of a composite with a periodic system of parallel coaxial normal tensile cracks. *J. Math. Sci.*, 186, 1-13.
 1476. Boisard, S. et al. Unusual chemical composition of a Mexican propolis collected in Quintana Roo, Mexico. *J. Apicult. Res.*, 54.4 (2022): 350-357.
 1477. Bonsignore, G., S. Martinotti, E. Ranzato. Propolis: A multifaceted approach for wound healing. *Gums, Resins and Latexes of Plant Origin: Chemistry, Biological Activities and Uses*. Cham: Springer International Publishing, 2022. 689-697.
 1478. Bruschi, M.L., et al. Nanostructured Propolis as Therapeutic Systems With Antimicrobial Activity. *Nano-and Microscale Drug Delivery Systems*. 2017. 377-391.
 1479. Bruschi, M.L. Desenvolvimento e caracterização de sistemas de liberação de própolis intrabolsa periodontal. Doctoral Dissertation. Universidade de São Paulo, 2006.
 1480. Bushra Begum A., et al. Evaluation of benzophenone-N-ethyl morpholine ethers as antibacterial and antifungal activities. *J. Chem.*, 2014 (2014).

1481. Cabral, I.S.R. (2008). Isolamento e identificação de compostos com atividade antibacteriana da própolis vermelha brasileira (Doctoral dissertation, Universidade de São Paulo).
1482. Cardinault, N., Cayeux, M.O., & du Sert, P.P. La propolis: origine, composition et propriétés. *Phytothérapie*, 2012, 10(5), 298-304.
1483. Costabile G., Gasteyer K.I., Nadithe V. et al. Physicochemical and in vitro evaluation of drug delivery of an antibacterial synthetic benzophenone in biodegradable PLGA nanoparticles. *AAPS Pharm. Sci. Tech.*, 1-10, 2018.
1484. Cuesta-Rubio, O., Piccinelli, A. L. Tropical propolis: recent advances in chemical components and botanical origin. In: *Med. Plants: Biodiv. Drugs*, 2012, 209.
1485. Cuesta-Rubio O., Monzote L., Fernández-Acosta R. et al. A review of nemorosone: Chemistry and biological properties. *Phytochemistry*, 210, 113674, 2023.
1486. Cuesta-Rubio, O., A.L. Piccinelli, M. Campo Fernandez, et al. Chemical characterization of Cuban propolis by HPLC– PDA, HPLC– MS, and NMR: The brown, red, and yellow Cuban varieties of propolis *J. Agric. Food Chem.* 55, 7502-7509 (2007)
1487. da Silva Nascimento, R. Desenvolvimento de métodos analíticos para a análise de própolis utilizando técnicas espectrométricas e análise multivariada. Doctoral dissertation, Universidade Federal de Minas Gerais, (2013).
1488. da Silva, R.O., Andrade, V.M., Rêgo, E.S.B. et al (2015). Acute and sub-acute oral toxicity of Brazilian red propolis in rats. *J. Ethnopharm.*, 170, 66-71.
1489. de Castro Ishida, V.F., Negri, G., Salatino, A., & Bandeira, M.F.C. A new type of Brazilian propolis: prenylated benzophenones in propolis from Amazon and effects against cariogenic bacteria. *Food Chem.*, 2011, 125(3), 966-972.
1490. de L. Paula, L.A., Cândido, A.C., Santos, M.F. et al. (2021). Antiparasitic properties of propolis extracts and their compounds. *Chem. & Biodiv.*, 18(9), e2100310.
1491. de Macêdo, P.C.D. Etiologia da candidíase esofágica e avaliação do efeito antifúngico do extrato de própolis in vitro e in vivo. PhD Thesis, Universidade Federal de Pernambuco, 2011.
1492. Değirmencioglu, H.T. (2018). Evaluation of phenolic profile, botanical origin, antioxidant and antimicrobial activities of Turkish propolis (Master's thesis, Sağlık Bilimleri Enstitüsü).
1493. Duke, C.C., et al. A sedge plant as the source of Kangaroo Island propolis rich in prenylated p-coumarate ester and stilbenes. *Phytochemistry* 134 (2017):87-97.
1494. El-Khamsa, M.S. (2017). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation, Université Ferhat Abbas–Setif).
1495. Falcão, S.I.D.M. Chemical composition of Portuguese propolis. *Bioactive Properties*. (2013). Doctoral dissertation, Universidade do Porto, Portugal.
1496. Ferraz, C.G. (2011). Derivados poliprenilados de benzofenonas, triterpenos, esteróides, bifenila e xantona de *clusia burllemarxii* e atividade citotóxica contra células GL-15, de glioblastoma humano. Doctoral dissertation, Universidade Federal da Bahia.
1497. Fernández Acosta, R., Piñeros, O., Pardo Andreu, G.L. (2019). The mitochondrial uncoupling as a promising pharmacological target against cancer. *J. Pharm.& Pharmacogn. Res.*, 7(2).
1498. Ferreira, J.M., G. Negri. Composição química e atividade biológica das própolis brasileiras: verde e vermelha. *Acta Apicola Brasilica* 6.1 (2018):06-15.
1499. Freires, I.A., S.M. de Alencar, P.L. Rosalen. A pharmacological perspective on the use

- of Brazilian red propolis and its isolated compounds against human diseases. Eur. J. Med. Chem., 110 (2016): 267-279.
1500. Freires, I.A., Pinguero, J M., Miranda, S.L. et al. (2018). Red propolis: Phenolics, polyphenolics, and applications to microbiological health and disease. In Polyphenols: Prevention and Treatment of Human Disease (pp. 293-300). Academic Press.
 1501. Galan, J.P.M.. Caracterización físico-química y evaluación de la actividad antifúngica de propóleos recolectados en el suroeste antioqueño. Master's thesis, Universidad Nacional de Colombia Sede Medellín.
 1502. Gallego, M. G. S. Determinación de la actividad anti-*Candida albicans* de propóleos sonorenses colectados en las diferentes estaciones del año, y sus principales constituyentes. 2021, PhD thesis, Universidad de Sonora.
 1503. García, L.R.P., Galán, J.P.M., Pajón, C.M.G., et al. Physicochemical characterization and antimicrobial activity of propolis from municipality of La Union (Antioquia, Colombia). Rev. Fac. Nal. Agr. Medellín, 2010, 63(1), 5373-5383.
 1504. Gil-Gonzales, J., Durango-Restrepo, D.L., Rojano, B.A., Martin-Loaiza, G. Antioxidant activity and Chemical Composition of Colombian Propolis. In: Natural Antioxidants and Biocides from Wild Medicinal Plants Edited by C. Cespedes, D. Sampietro, D. Seigler, M. Rai. CABI, 2013, 92-116
 1505. Gómez, P.A.M., Jon, L.Y.T.C., Torres, D.J.M. et al. (2021). Antibacterial, antibiofilm, and cytotoxic activities and chemical compositions of Peruvian propolis in an in vitro oral biofilm. F1000Research, 10.
 1506. Grosso, G.S. (2017). Origen, naturaleza, propiedades fisicoquímicas y valor terapéutico del propóleo (pp. 75-78). Sello Editorial Universidad del Tolima.
 1507. Hassani A.R. et al. In vitro reduction in colonization of *Streptococcus mutans* by honey beeswax ethyl acetate extract. J. Arak Univ. Med. Sci.-Rahavard Danesh. 2009; 11 (4): 87-95.
 1508. Hegazi, A.G., Abd El Hady, F.K., Shalaby, H.A. Pakistan Journal of Biological Sciences 2007, 10(19), 3295-3305.
 1509. Herrera-López, M.G., et al. Botanical origin of triterpenoids from Yucatecan propolis. Phytochem. Lett., 29, (2019), 25-29.
 1510. Hongzhuan, X, H. Fuliang. (2009). Chemical components of Brazilian red propolis. Chinese Apiculture, (6), 35-36.
 1511. Hongzhuan, X., H. Fuliang. (2009). Pharmacological effects of Brazilian red propolis. Chinese Apiculture, (8), 31-32.
 1512. Hosni, S., H. Kasri, & Ghaemi. (2008). The effect of bee wax ethyl acetate extract on *Streptococcus mutans* colonization. Journal of Arak Univ. Med. Sci., 11(4), 87-95.
 1513. Huang, S., Zhang, C. P., Wang, K., Li, G. Q., & Hu, F. L. Recent advances in the chemical composition of propolis. Molecules, 19(12), 2014, 19610-19632.
 1514. Huang S., Zhang C., & Hu F. (2013). Research progress on chemical components of propolis from 2008 to 2012. Nat. Product Res. Develop., 25(8), 1146-1153.
 1515. Ibrahim, R.S., A.M. Metwally. Plant source of Egyptian propolis using multivariate-assisted digitally-enhanced TLC data analysis. J. Nat. Prod. Plant Resour., 2017, 73, 20-25.
 1516. Isla, M., Vit, P., Brito, R. et al. (2005). Caramelos a base de propóleos y su posible aceptación en la ciudad de Mérida. Revista del Instituto Nacional de Higiene Rafael Rangel, 36(1), 6-12.
 1517. Isla, M. et al. Caramelos a base de propóleos y su posible aceptación en la ciudad de

- Mérida. Revista del Instituto Nacional de Higiene Rafael Rangel 36.1 (2005): 6-12.
1518. Juanes, C.C. Red propolis and L-Lysine inhibiting angiogenesis in Walker Tumor in hamster cheek pouch model. Doctoral dissertation, Universidade Federal do Ceará, Fortaleza, 2018.
 1519. Kardar, M.N., et al. Characterisation of triterpenes and new phenolic lipids in Cameroonian propolis. *Phytochemistry* 106 (2014):156-163.
 1520. Kaya, G., Keskin, M. (2020). Biochemical properties and urease, α -amylase inhibitory effects of *Ocimum basilicum* L.(Reyhan). *J. Med. Herbs Ethnomed.*, 6, 52-55.
 1521. Khanum, S.A., Shashikanth, S., Umesha, S. et al. Synthesis and antimicrobial study of novel heterocyclic compounds from hydroxybenzophenones *Eur. J. Med. Chem.*, 40(11), 1156-1162 (2005).
 1522. Kim, Y. J., Yong, Y. K., Aslam, M. S. (2019). Narrative and meta-analytic study of in vivo efficiency of the bioactive compounds of propolis in tooth decay. *Pharmacogn. Rev.*, 13(26).
 1523. Kumar S., Sharma S., Chattopadhyay SK. The potential health benefit of polyisoprenylated benzophenones from *Garcinia* and related genera: Ethnobotanical and therapeutic importance. *Fitoterapia*, 1, 89, 2013, 86-125.
 1524. Kustiawan, P.M. et al. In vitro cytotoxicity of Indonesian stingless bee products against cancer cell lines. *Asian Pacific J. Trop. Biomed.*, 2014, 4(7), 549-556.
 1525. La Propolis, C.I.G.S I. 7. Activités biologiques de la propolis. *Dédicace*, 10.
 1526. Lipovka Y., Alday E., Hernandez J., Velazquez C. Molecular mechanisms of biologically active compounds from propolis in breast cancer: state of the art and future directions. *Food Rev. Int.*, 39 (5), pp. 2931 – 2968, 2023.
 1527. Liu, H. et al. Acylphloroglucinol and tocotrienol derivatives from the fruits of *Garcinia multiflora*. *RSC Advances* 7.47 (2017):29295-29301.
 1528. López BG-C., Schmidt EM., Eberlin MN., Sawaya ACHF. Phytochemical markers of different types of red propolis. *Food Chem.*, 146, 2014, 174-180.
 1529. López, M.G.H., Irabien, L.M.C. (2019). Caracterización química y actividad biológica de propóleos producidos en el estado de Yucatán (Doctoral dissertation, Centro de Investigación Científica de Yucatán).
 1530. López, B.G.C., Sawaya, A.C.H.F. (2012). A review of the plant origins, composition and biological activity of red propolis. *Natural products: Structure, bioactivity and applications*, 83-96.
 1531. Machado, G.M.D., Leon, L.L., De Castro, S.L. Activity of Brazilian and Bulgarian propolis against different species of *Leishmania*. *Memorias do Instituto Oswaldo Cruz*, 2007, 102, 1, 73-77.
 1532. Maleckaitė, M. (2020). Vandenių ir nevandenių propolio tirpalų poveikio pirminėje žiurkių smegenų ląstelių kultūroje išemijos sąlygomis tyrimas (Doctoral dissertation, Lietuvos sveikatos mokslų universitetas).
 1533. Mărghitaş LA., Dezmirean DS., Bobiş O. Important developments in Romanian propolis research. *Evid. Based Compl. Altern. Med.*, 2013, 159392, ISSN: 1741-427X.
 1534. Marín-Prida, J., Andreu, G.L.P., Rossignoli, C.P. et al. (2017). The cytotoxic effects of VE-3N, a novel 1, 4-dihydropyridine derivative, involve the mitochondrial bioenergetic disruption via uncoupling mechanisms. *Toxicology in vitro*, 42, 21-30.
 1535. Martínez Galán, J.P. Caracterización físico-química y evaluación de la actividad

- antifúngica de propóleos recolectados en el suroeste antioqueño. Doctoral Dissertation, Facultad de Ciencias Agropecuarias (2009).
1536. Massaro, C.F. Bee propolis: a potential anti-herpes drug. Doctoral Dissertation, Studi di Urbino, Italy, 2008.
 1537. Massaro, Carmelina Flavia. "Bee Propolis: A Potential Anti-Herpes Drug." (2007).
 1538. Mayworm, M.A.S., Fernandes-Silva, C.C., Salatino, M.L.F. et al. A simple and inexpensive procedure for detection of a marker of Brazilian alecrim propolis. J. Apicult. Res., 2015, 54, 36-39.
 1539. Medina Alvarado, A. L. (2018). Caracterización química y actividad antiproliferativa de propóleos de la región de Jerez, Zacatecas. PhD thesis, Universidad Veracruzana.
 1540. Meneses, E.A., D.L. Durango, C.M. García. Antifungal activity against postharvest fungi by extracts from Colombian propolis. Química Nova, 32.8 (2009):2011-2017.
 1541. Miguel, M.G., & Antunes, M.D. Is propolis safe as an alternative medicine? J. Pharmacy & Bioal. Sci., 2011, 3(4), 479.
 1542. Mingxia, L. (2009). Application of propolis in agronomy and plant protection. Chinese Bee Industry, (8), 32-32.
 1543. Miranda, S.L.F. et al. Brazilian red propolis reduces orange-complex periodontopathogens growing in multispecies biofilms. Biofouling 35,3, (2019), 308-319.
 1544. Mora, D.P. P. et al. The chemical composition and events related to the cytotoxic effects of propolis on osteosarcoma cells: A comparative assessment of Colombian samples. Phytother. Res., 33.3 (2019):591-601.
 1545. Naldoni, F. J. et al. Antimicrobial activity of benzophenones and extracts from the fruits of *Garcinia brasiliensis*. J. Med. Food, 12.2 (2009): 403-407.
 1546. Naveen, P., Al-Ghorbani, M., Asha, M.S., et al. Synthesis and inhibition of microbial growth by benzophenone analogues-a simplistic approach. Asian J. Biomed. Pharm. Sci., 2014, 4(29), 55-60.
 1547. Neto, MS Regueira, et al. Seasonal variation of Brazilian red propolis: Antibacterial activity, synergistic effect and phytochemical screening. Food Chem. Toxicol., 107 (2017):572-580.
 1548. Niken, P. Chemical constituents, botanical origin and biological activity of propolis of *Tetragonula sapiens*, a stingless bee, in Southeast Sulawesi, Indonesia. 2015, PhD thesis.
 1549. Nina, N, et al. Chemical profiling and antioxidant activity of Bolivian propolis. J. Sci. Food Agricult., 96.6 (2016):2142-2153.
 1550. Oliveira, L.F.A.D.M. (2021). Nanopartículas de sílica mesoporosa carregadas com extrato de própolis vermelha: síntese, caracterização e avaliação in vitro da atividade antioxidante, antimicrobiana e antiviral. Doctoral dissertation, Universidade Federal de Alagoas.
 1551. Okhale, S. E., et al. Bee propolis: Production optimization and applications in Nigeria. J. Pharmacogn. Phytother., 13.1 (2021): 33-45.
 1552. Patrícia Cerqueira de Macêdo, Danielle. "Etiologia da candidíase esofágica e avaliação do efeito antifúngico do extrato de própolis in vitro e in vivo." (2011).
 1553. Palomino García, L.R., Martínez Galán, J.P., García Pajón, C.M. et al. (2010). Caracterización fisicoquímica y actividad antimicrobiana del propóleos en el Municipio de La Unión (Antioquia, Colombia). Revista Facultad Nacional de Agronomía Medellín, 63(1), 5373-5383.
 1554. Patrícia Cerqueira de Macêdo, D. (2011). Etiologia da candidíase esofágica e

- avaliação do efeito antifúngico do extrato de própolis in vitro e in vivo. Doctoral dissertation, Universidade Federal de Pernambuco.
1555. Pereira, I.N. *Própolis: matéria-prima de potencial aplicação farmacêutica*. Master's Thesis, Universidade Federal do Rio Grande do Sul, 2011.
 1556. Prashanth, T., et al. Synthesis, characterization, docking study and antimicrobial activity of 2-(4-benzoylphenoxy)-1-[2-(1-methyl-1 H-indol-3-yl) methyl)-1 H-benzo [d] imidazol-1-yl] ethanone derivatives. *J. Iranian Chem. Soc.*, (2021):1-16.
 1557. Prashanth, T., Naveen, P., Al-Ghorbani, M. et al. (2014). Synthesis and Inhibition of Microbial Growth by Benzophenone Analogues-A Simplistic Approach. *Asian J. Biomed. Pharm. Sci.*, 4(29), 55.
 1558. Rastrelli, L. (2012). Tropical propolis: recent advances in chemical components and botanical origin. *Med. Plants*, 235-266.
 1559. Regueira, M. S., et al. Seasonal variation of Brazilian red propolis: Antibacterial activity, synergistic effect and phytochemical screening. *Food Chem. Toxicol.*, (2017).
 1560. Righi, A.A. *Extratos brutos e constituintes de própolis brasileiras: avaliação dos efeitos nos carrapatos Rhipicephalus sanguineus, Rhipicephalus microplus e Amblyomma cajennense*. PhD Thesis, Universidade de São Paulo, 2013.
 1561. Righi, A.A. *Perfil químico de amostras de própolis brasileiras*, Doctoral Dissertation, Universidade de São Paulo, 2008.
 1562. Rios, N., Yáñez, C., Rojas, L., et al. Chemical composition of essential oil of *Apis mellifera* propolis from Falcón State, Venezuela. *Emirates J. Food Agricult.*, 2014, 26(7), 639-642.
 1563. Rivera-Yañez, N. et al. Biomedical properties of propolis on diverse chronic diseases and its potential applications and health benefits. *Nutrients* 13.1 (2020):78.
 1564. Robles-Zepeda, R.E. et al. Botanical origin and biological activity of propolis. In: *Medicinal plants: Biodiversity and drugs*. 1st ed. New York: CRC Press, Taylor & Francis Group (2012):570-597.
 1565. Rosero, A.L.F. (2013). *Complicaciones y accidentes intraoperatorios y postoperatorios de la exodoncia simple y su tratamiento*. Facultad Piloto de Odontología (Doctoral dissertation, Universidad de Guayaquil).
 1566. Rusak G. In: *Scientific Evidence of the Use of Propolis in Ethnomedicine* (N. Orsolic, I. Basic, Editors), Transworld Research Network, Trivandrum, 2008, pp. 17 – 31.
 1567. Rusak G. In: *Scientific Evidence of the Use of Propolis in Ethnomedicine* (N. Orsolic, I. Basic, Editors), Transworld Research Network, Trivandrum, 2008, pp. 17 – 31.
 1568. Salatino, A., Fernandes-Silva, C.C., Righi, A.A., & Salatino, M.L.F. Propolis research and the chemistry of plant products. *Nat. Prod. Reports*, 2011, 28(5), 925-936.
 1569. Salatino, A., Salatino, M. L. F., & Negri, G. (2021). How diverse is the chemistry and plant origin of Brazilian propolis?. *Apidologie*, 1-23.
 1570. Salomão, K., P.R.S. Pereira, L.C. Campos et al. Brazilian propolis: correlation between chemical composition and antimicrobial activity *Evidence-based Compl. Altern. Med.*, 5(3), 317-324 (2008)
 1571. Santos, L.M., et al. Propolis: types, composition, biological activities and veterinary product patent prospecting. *J. Sci. Food Agr.*, 2020, 100, 4, 1369-1382.
 1572. Santos, Laerte M., et al. Propolis: types, composition, biological activities, and veterinary product patent prospecting. *J. Sci. Food Agr.* 100.4 (2020): 1369-1382.

1573. Sawaya, A.C.H.F. A Review of the plant origins, composition and biological activity of red propolis. *Natural Products: Structure, Bioactivity and Applications* (2012).
1574. Sawaya, A.C.H.F., Abdelnur, P.V., Eberlin, M.N., et al. Fingerprinting of propolis by easy ambient sonic-spray ionization mass spectrometry. *Talanta*, 2010, 81, 1-2, 100-108.
1575. Seidel, V. et al. Comparative study of the antibacterial activity of propolis from different geographical and climatic zones. *Phytother. Res.*, 22.9 (2008):1256-1263.
1576. Shen X., Z. Cuiping, H. Fuliang. (2015). Research progress on the chemical components of Brazilian propolis. *Nat. Product Res. Develop.*, 27(5), 915.
1577. Shoaie Hassani, A., K. Hamdi, A. Ghaemi. In vitro reduction in colonization of streptococcus mutans by honey beeswax ethyl acetate extract. *J. Arak University Med. Sci.*, 11.4 (2008):87-95.
1578. Soltani, E. K. (2018). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation).
1579. Silici, S. Turkish propolis: chemical constituents. 2010, *Mellifera*, 10(19), 24-33.
1580. Silici, S., Plant-honeybee interactions. *Rec. Adv. Plant Sci.*, 2020:313-340.
1581. Silva, A.C. Avaliação in vitro do potencial leishmanicida de derivados triazólicos e derivados de benzofenonas. (2017). Doctoral dissertation, Universidade Federal de Vicosa
1582. Silva-Carvalho, R., Baltazar, F., & Almeida-Aguiar, C. (2015). Propolis: a complex natural product with a plethora of biological activities that can be explored for drug development. *Evidence-Based Compl. Altern. Med.*, 2015, 1-29, 206439.
1583. Sirma, E. Pervari (siirt) bölgesi (çemikari, kovanagzi, sariyaprak) propolislerinin kimyasal içerikleri ve botanik orjininin belirlenmesi. (Master's thesis, Fen Bilimleri Enstitüsü).
1584. Soltani, El-khamsa. Caractérisation et activités biologiques de substances naturelles, cas de la propolis. Doctoral Dissertation), 2018.
1585. Soltani, A., M. Benhanifia. Propolis, plant sources and antimicrobial activity: an overview. *Anti-Infective Agents*, 2023, 21, 5, 20-39.
1586. Souza, J.H. (2023). Potencial antibacteriano e antibiofilme da própolis vermelha brasileira frente à bactérias periodontopatogênicas. Doctoral Dissertation, Universidade Federal de Uberlândia..
1587. Toreti VC., Sato HH., Pastore GM., Park YK. Recent progress of propolis for its biological and chemical compositions and its botanical origin. *Evidence-based Compl. Altern. Med.*, 2013, art. no. 697390.
1588. Torres, R.N.S. et al. The volatile constituents of propolis from Piauí. *Quimica Nova* 31.3 (2008):479-485.
1589. Tran, T.D., et al. Lessons from exploring chemical space and chemical diversity of propolis components. *Int. J. Mol. Sci.*, 21.14 (2020):4988,1-35.
1590. Trujillo Celi, Cristian Fabián. Composición química de cinco muestras de propóleo del cantón Yantzaza de la provincia de Zamora Chinchipe. BS thesis. Machala: Universidad Técnica de Machala, 2015.
1591. Yusoff, Y. M. (2018). The use of metabolomics tools to assess the therapeutic natural products of honey and propolis from Malaysia and New Zealand. Doctoral dissertation, University of Strathclyde.
1592. Кайгородов, Р., С.А. Суворова. Биохимические особенности растительных источников прополиса умеренной природной зоны. *Вестник Пермского*

Университета. Серия: Биология, 2013, (3), 65-68.

1593. Кайгородов, Р., О.А.Малькова, Ю.В. Кайгородова. Тестирование антиоксидантных свойств спиртовых экстрактов прополиса с использованием липосомной модели. *Educatio* 4 (11)-3 (2015).

84). Trusheva B., Popova M., Koendhori EB., Tsvetkova I., NAYDENSKI C., Bankova V. Indonesian propolis: Chemical composition, biological activity and botanical origin. *Natural Product Research*, 6, 25, 2011, 606-613, ISSN: 1478-6419. IF – 1.009.

1594. Abdullah, N.A. et al. Phytochemicals, mineral contents, antioxidants, and antimicrobial activities of propolis produced by Brunei stingless bees *Geniotrigona thoracica*, *Heterotrigona itama*, and *Tetrigona binghami*. *Saudi J. Biol. Sci.*, 27.11 (2020):2902-2911.

1595. Abdillah, R., Rachmaini, F., Fadhilah, D., & Almahdy, A. (2024). Fetal protective effect of Indonesian propolis from *Apis mellifera* against rifampicin-pyrazinamide induced impaired pregnancy in BALB/c mice. *J. Compl. Integr. Med.*, (0).

1596. Aim, T.U.N.A.N., Smail, T., Ulaiman, S.I.A.M.S. et al. (2018). Chemical constituents of Malaysian *Apis mellifera* propolis. *Sains Malaysiana*, 47(1), 117-122.

1597. Ali, S.I., S. Tasleem, S.B.S. Naqvi (2019). Microbial evaluation of infant milk formula. *Bioscience Research. World Health*, 16, 1, 639-647.

1598. Alsherbiny, M.A., Bhuyan, D.J., Radwan, I. et al. (2021). Metabolomic identification of anticancer metabolites of Australian propolis and proteomic elucidation of its synergistic mechanisms with doxorubicin in the MCF7 cells. *Int. J. Mol. Sci.*, 22(15), 7840.

1599. Anđelković, B.D. Primena rezultata NMR i FTIR spetroskopskih tehnika u multivarijantnoj analizi za klasifikaciju propolisa. Doctoral dissertation, Univerzitet u Beogradu-Hemijski fakultet, 2017.

1600. Avula, B. et al. Quantification and characterization of phenolic compounds from northern Indian propolis extracts and dietary supplements. *J. AOAC Int.* 103.5 (2020): 1378-1393.

1601. Azonwade F., Mabanza-Banza B.B., Le Ray A.-M. et al. Chemodiversity of propolis samples collected in various areas of Benin and Congo: Chromatographic profiling and chemical characterization guided by ¹³C NMR dereplication. *Phytochem. Anal.*, 2023, 34 (4), 461-475.

1602. Barrera, E., Gil, J., Restrepo, A. et al. (2015). A coating of chitosan and propolis extract for the postharvest treatment of papaya (*Carica papaya* L. cv. Hawaiiiana). *Revista Facultad Nacional de Agronomía Medellín*, 68(2), 7667-7678.

1603. Bhuyan, D.J., Alsherbiny, M.A., Low, M.N. et al. (2021). Broad-spectrum pharmacological activity of Australian propolis and metabolomic-driven identification of marker metabolites of propolis samples from three continents. *Food & Function*, 12(6), 2498-2519.

1604. Blicharska, N., & Seidel, V. (2019). Chemical diversity and biological activity of African propolis. *Progress Chem. Org. Nat. Products*, 109, 415-450.

1605. Cardinault, N., Cayeux, M.O., & du Sert, P.P. La propolis: origine, composition et propriétés. *Phytothérapie*, 2012, 10(5), 298-304.

1606. da Silva, C. C. F., Salatino, A., da Motta, L. B., Negri, G., & Salatino, M. L. F. (2019). Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from

- Ceará state. *Revista Brasileira de Farmacognosia*, 29(3), 309-318.
1607. Dawkar, V., & Chalwade, A. (2023). Ayurveda and environmental network: Antimicrobial, pharmacological and ecological potential of *Swietenia macrophylla* Meliaceae (L.) Jacq. *Biota: Biologi dan Pendidikan Biologi*, 16(2), 76-99.
 1608. De Groot AC. Propolis: A review of properties, applications, chemical composition, contact allergy, and other adverse effects. *Dermatitis*, 6, 24, 2013, 263-282.
 1609. De Souza, G.G., Pfenning, L.H., De Moura, F., et al. Isolation, identification and antimicrobial activity of propolis-associated fungi. *Nat. Prod. Res.* 18(27), 2013, 1705-1707.
 1610. Djati, G., Djati, I.N.S.G., 2023. Effect of application of propolis as feed supplement and preservation agent to pathogenic microbes contamination of local chicken meat. In: *Proc. 12th Int. Conf. Green Technol. (ICGT 2022)* (Vol. 221, p. 187). Springer Nature.
 1611. El-aziz, E.A.E.D.A., Elgayar, S.F., Mady, F.M. et al. (2021). The potential of optimized liposomes in enhancement of cytotoxicity and apoptosis of encapsulated Egyptian propolis on hep-2 cell line. *Pharmaceutics*, 13(12), 2184.
 1612. Ervina, M. The recent use of *Swietenia mahagoni* (L.) Jacq. as antidiabetes type 2 phytomedicine: A systematic review. *Heliyon* 6.3 (2020):e03536.
 1613. Fan Y., Ma L., Zhang W. et al. Microemulsion can improve the immune-enhancing activity of propolis flavonoid on immunosuppression and immune response. *Int. J. Biol. Macromol.*, 63, 2014, 126-132.
 1614. Fan, Y., Ma, L., Zhang, W., et al. The design of propolis flavone microemulsion and its effect on enhancing the immunity and antioxidant activity in mice. *Int. J. Biol. Macromol.*, 2014, 65, 200-207.
 1615. Farida, S. et al. The beneficial effect of Indonesian propolis wax from *Tetragonula* sp. as a therapy in limited vaginal candidiasis patients. *Saudi J. Biol. Sci.*, 27.1 (2020):142-146.
 1616. Fernández-León, K.J. et al. Comparison of in vitro anti-*Staphylococcus aureus* activity of eight antibiotics and four dilutions of propolis. *J. Selva Andina Res. Soc.*, 13.1 (2022): 35-48.
 1617. Gil-Gonzales, J., Durango-Restrepo, D.L., Rojano, B.A., Martin-Loaiza, G. Antioxidant activity and Chemical Composition of Colombian Propolis. In: *Natural Antioxidants and Biocides from Wild Medicinal Plants* Edited by C. Cespedes, D. Sampietro, D. Seigler, M. Rai. CABI, 2013, 92-116.
 1618. Hendraningrat, A. (2014). Efek Pemberian Ekstrak Propolis Terhadap Profil Sel Darah Putih Pada Tikus Putih (*Rattus norvegicus*) Yang Diberi Deksametason (Doctoral dissertation, Universitas Airlangga).
 1619. Herrera-López, M.G., Rubio-Hernández, E.I., Leyte-Lugo, M.A. et al. (2019). Botanical origin of triterpenoids from Yucatecan propolis., *Phytochem. Lett.*, 29, 25-29.
 1620. Herrera-López, Mercedes G., et al. Resorcinolic lipids from Yucatecan propolis. *J. Brazilian Chem. Soc.*, 31.1 (2020):186-192.
 1621. Hidayat, S.A. et al. Optimization of East Java propolis extraction as anti SARS-Cov-2 by molecular docking study. *J. Ilmu dan Teknologi Hasil Ternak (JITEK)* 17.2 (2022): 123-134.
 1622. Hossain, R. et al. Propolis: An update on its chemistry and pharmacological applications. *Chinese Medicine* 17.1 (2022): 1-60.
 1623. Hu, H. et al. Two novel markers to discriminate poplar-type propolis from poplar bud extracts: 9-oxo-ODE and 9-oxo-ODA. *J. Food Comp. Anal.*, 105 (2022):104196.
 1624. Huang, S., Zhang, C.P., Wang, K. et al. Recent advances in the chemical composition

- of propolis. *Molecules*, 19(12), 2014, 19610-19632.
1625. Huang, Z. C., & Hu, F. Progress of study on the chemical composition of propolis (2008-2012). *Nat. Prod. Res. Developm.*, 2013, 25 (8), 1146-1153.
 1626. Inui, S., Hosoya, T., Shimamura, Y., et al. Solophenols B–D and solomonin: new prenylated polyphenols isolated from propolis collected from the Solomon Islands and their antibacterial activity. *J. Agric. Food Chem.*, 2012, 60(47), 11765-11770.
 1627. Iqbal, M., Fan, T.P., Watson, D. et al. (2019). Preliminary studies: the potential anti-angiogenic activities of two Sulawesi Island (Indonesia) propolis and their chemical characterization. *Heliyon*, 5(7), e01978.
 1628. Isidorov, V.A., Maslowiecka, J., Szoka, L., et al. Chemical composition and biological activity of Argentinian propolis of four species of stingless bees, *Molecules*, 2022, 27(22), 7686.
 1629. Isidorov, V. A., Dallagnol, A. M., & Zalewski, A. (2024). Chemical Composition of Volatile and Extractive Components of Canary (Tenerife) Propolis. *Molecules*, 29(8), 1863.
 1630. Ismail T.N.N.T., Sulaiman S.A., Ponnuraj K.T. et al. Chemical constituents of Malaysian *Apis mellifera* propolis. *Sains Malays.*, 47 (1), 117-122, 2018.
 1631. Jaya, F. (2017). *Produk-produk lebah madu dan hasil olahannya*. Universitas Brawijaya Press. ISBN 978-602-432-145-1.
 1632. Jee, Y.S. (2012). First experiences with laparoscopic assisted distal gastrectomy: in the view of comparison with high volume centers. *J. Korean Surg. Soc.*, 83(3), 130-134.
 1633. Juwita, D.A., Ahmadin, A., Abdillah, R. et al. (2023). Hepatoprotective effect of Indonesian propolis from in carbon tetrachloride (CCl₄) induced liver injury in mice. *Curr. Issues Pharm. Med. Sci.*, 36,4, 189-193..
 1634. Lee, K.P., Sudjarwo, G.W., Kim, J.S. et al. (2014). The anti-inflammatory effect of Indonesian *Areca catechu* leaf extract in vitro and in vivo. *Nutr. Res. Practice*, 8(3), 267.
 1635. Kardar, M. N., Zhang, T., Coxon, G. D., et al. Characterisation of triterpenes and new phenolic lipids in Cameroonian propolis. *Phytochemistry*. 2014, 106, 156-163.
 1636. Keskin, M. Chemical characterization of arabic gum-chitosan-propolis beads and determination of alpha-amylase inhibition effect. *Progress in Nutrition* 22.2 (2020):562-567.
 1637. Keskin, M. Determination of chemical composition and α -amylase inhibitory effect of new propolis extracts. *Combinatorial chemistry & high throughput screening* (2020): 23(9):939-944.
 1638. Kim, J., & Yang, Y. J. (2014). Plain water intake of Korean adults according to life style, anthropometric and dietary characteristic: the Korea National Health and Nutrition Examination Surveys. *Nutr. Res. Pract.* 2014, 8(5): 580–588.
 1639. Kinasih, I., et al. Addition of black soldier fly larvae (*Hermetia illucens* L.) and propolis to broiler chicken performance. *IOP Conference Series: Earth and Environmental Science*. Vol. 187. No. 1. IOP Publishing, 2018.
 1640. Kinghorn, A. Douglas, et al., eds. *Progress in the chemistry of organic natural products*. Cham, Switzerland: Springer, 2017.
 1641. Kumazawa, S., Murase, M., Momose, N. et al. Analysis of antioxidant prenylflavonoids in different parts of *Macaranga tanarius*, the plant origin of Okinawan propolis. *Asian Pac. J. Trop. Med.*, 2014, 7(1), 16-20.
 1642. Kustiawan, P.M., Puthong, S., Arung, E. et al. In vitro cytotoxicity of Indonesian stingless bee products against human cancer cell lines. *Asian Pac. J. Trop. Biomed.*, 2014, 4, 7, 549-556.

1643. Kustiawan, P.M. et al. Molecular mechanism of cardol, isolated from *Trigona incisa* stingless bee propolis, induced apoptosis in the SW620 human colorectal cancer cell line. *BMC Pharmacol. Toxicol.*, 18.1 (2017):32.
1644. Lee, K.P., Sudjarwo, G.W., Kim, J.S., et al. The anti-inflammatory effect of Indonesian Areca catechu leaf extract in vitro and in vivo. *Nutr. Res. Practice*, 2014,8, 4,
1645. Lipovka Y., Alday E., Hernandez J. et al. Molecular mechanisms of biologically active compounds from propolis in breast cancer: state of the art and future directions. *Food Rev. Int.*, 39 (5), 2931-2968, 2023.
1646. Magnavacca, A. et al. The antiviral and immunomodulatory activities of propolis: An update and future perspectives for respiratory diseases. *Med. Res. Rev.*, 42.2 (2022): 897-945.
1647. Manginstar, C.O., Tallei, T.E., Niode, N.J. et al. (2024). Therapeutic potential of propolis in alleviating inflammatory response and promoting wound healing in skin burn. *Phyther. Res.*, 38(2), 856-879.
1648. Magnavacca, A. (2022). Brazilian propolis: a multifaceted natural product with healing properties - a comprehensive study on the ability to improve cutaneous wound healing through hif-1 modulation. Doctoral dissertation, Università degli Studi di Milano.
1649. Masadah, R., Ekawardhani, S., Putra, R.E. et al, M. (2023). Ethanolic extract of propolis from *Tetragonula laeviceps*: selective cytotoxicity for MCF-7 breast cancer cells. *Pharmacogn. J.*, 15(6).
1650. Massaro, C. F., Simpson, J. B., Powell, D., & Brooks, P. (2015). Chemical composition and antimicrobial activity of honeybee (*Apis mellifera ligustica*) propolis from subtropical eastern Australia. *The Science of Nature*, 102, 1-11.
1651. Massaro, C.F. Antibacterial natural products of propolis and honeys from Australian stingless bees (*Tetragonula carbonaria*) (Doctoral dissertation, University of the Sunshine Coast).
1652. Meemongkolkiat T., Puthong S., Khongkarat P. et al. Antiproliferative and anti-tyrosinase activities of 93 propolis from *Tetragonula laeviceps* and *Tetragonula pegdeni* in Thailand. *Sains Malaysiana*, 52 (4), 1145-1160, 2023.
1653. Meemongkolkiat, T., Puthong, S., Khongkarat, P. et al. (2024). In vitro cytotoxic activity on KATO-III cancer cell lines of mangiferolic acid purified from Thai *Tetragonula laeviceps* propolis. *Heliyon*, 10(9).
1654. Mohd Badiyazaman, A.A., Nazif Aziz, A., Mohamad, H. et al. (2024). Bioassay-guided isolation of cycloartane-type triterpenes, their cytotoxic properties and identification of potential chemical markers from *Geniotrigona thoracica* propolis. *Nat. Product Res.*, 1-6.
1655. Muscat M. Use of Propolis chemical and Asian tiger mosquito bites. Case report and review. *Malta Med. J.*, 2013, 1, 25, 58-61.
1656. Nasirli F., Daikh A., Doğan N.M. et al. In vitro evaluation of cytotoxic activity of Algerian propolis against human breast adenocarcinoma (MDA-MB-231) cells and investigation of its potential mechanism of action. *Curr. Bioact. Compounds*, 2023, 19 (1), 28-38.
1657. Negri, G., Silva, C.C.F., Coelho, G.R. et al. (2019). Cardanols detected in non-polar propolis extracts from *Scaptotrigona* aff. *postica* (Hymenoptera, Apidae, Meliponini). *Brazilian J. Food Technol.*, 22, e2018265.
1658. Nugraha, A.P., Koendhori, E.B., Fahmi, A. et al. (2023). Ethanolic extract of propolis from Indonesian stingless bee (*Tetragonula biroii*) is ineffective against ESBL-producing *Klebsiella pneumoniae*: An in vitro study and a brief review of propolis antibacterial and

- anti-inflammatory activity. *World J. Adv. Res. Rev.*, 20(3), 51-59.
1659. Özkök A., Karlıdağ S., Keskin M. et al. Palynological, chemical, antimicrobial, and enzyme inhibition properties of *Cannabis sativa* L. propolis. *Eur. Food Res. Technol.*, 2023, 249 (8), 2175-2187.
 1660. Pailee, P., Sangpetsiripan, S., Mahidol, C. et al. (2015). Cytotoxic and cancer chemopreventive properties of prenylated stilbenoids from *Macaranga siamensis*. *Tetrahedron*, 71(34), 5562-5571.
 1661. Pant, K., Sharma, A., Chopra, H.K. and Nanda, V., 2023. Impact of biodiversification on propolis composition, functionality, and application in foods as natural preservative: A review. *Food Control*, p.110097.
 1662. Pereira, Daniel Santiago, et al. Effect of extracts of amazonian bee propolis on *Xanthomonas axonopodis* pv. *passiflorae* in the State of Pará-Brazil. *Embrapa Amazônia Oriental-Artigo em periódico indexado (ALICE)* (2020), 9, 11, e3719119464.
 1663. Pohara, H. A. (2020). Aktivitas imunostimulan snedds propolis terhadap parameter jumlah leukosit, neutrofil, dan limfosit pada tikus putih jantan galur wistar. Doctoral dissertation, Universitas Islam Indonesia.
 1664. Pratami, D.K., et al. Formulation and determination of quality parameters of propolis extract microcapsule tablets from *Tetragonula sapiens*. *Int. J. Appl. Pharm.* (2022):47-52.
 1665. Pujirahayu, N., Suzuki, T., Katayama, T. (2019). Cycloartane-type triterpenes and botanical origin of propolis of stingless indonesian bee *Tetragonula sapiens*. *Plants*, 8(3), 57.
 1666. Pumerantz, S.A. PEGylated liposomal vancomycin: A glimmer of hope for improving treatment outcomes in MRSA pneumonia. *Rec. Patents Anti-Infect. Drug Discov.*, 2012, 7(3), 205-212.
 1667. Rahim, M.K.A. Structural and biochemical characterization of bees propolis. Doctoral dissertation. Pusat Pengajian Sains Perubatan, Universiti Sains Malaysia, 2020.
 1668. Renteria, I.P.D.G. (2018). Caracterización química y actividad biológica de extractos etanólicos de propóleo del estado de Sinaloa (Doctoral dissertation, Universidad Autónoma de Sinaloa).
 1669. Rivera-Yañez, N., Rivera-Yañez, C.R., Pozo-Molina, G. et al. (2020). Biomedical properties of propolis on diverse chronic diseases and its potential applications and health benefits. *Nutrients*, 13(1), 78.
 1670. Ruiz-Bustos P., Alday E., Garibay-Escobar A. et al. Propolis: antineoplastic activity, constituents, and mechanisms of action. *Curr Top Med Chem.*, 2023, 23(18):1753-1764.
 1671. Saeed, F., Ahmad, R.S., Arshad, M.U. et al. (2016). Propolis to curb lifestyle related disorders: an overview. *Int. J. Food Properties*, 19(2), 420-437.
 1672. Sahlan M., Supardi T. Encapsulation of Indonesian propolis by Casein micelle. *Int. J. Pharm.Biol. Sci.*, 2013, 1, 4, 297-305.
 1673. Sahlan, M., Akhmariadi, A., Pratami, D.K. et al. (2018). Scale up production indonesian liquid propolis from raw propolis and wild beehive using bubbling vacuum evaporator. In: *MATEC Web of Conferences* (Vol. 248, p. 04001). EDP Sciences.
 1674. Salawu, K. M., Oyerinde, A. A., Aliyu, A. et al. (2020). Growth inhibitory properties and antimicrobial evaluation of *Aloe schweinfurthii* (Baker) leaf rind extract. *J. Pharm.& Bioresources*, 17(2), 174-179.
 1675. Saleh, K.A.A. (2018). Profiling of propolis samples from UK, Indonesia and sub-Saharan Africa and isolation of some pure components. Doctoral dissertation, University

of Strathclyde..

1676. Santos, R. L., Pereira, D.S., Xavier Júnior, S.R. et al. (2017). Levantamento fitogeográfico de *Dalbergia Lf* (Leguminosae-papilionoideae) com potencial produtivo para própolis vermelha no Estado do Pará.. *Revista Verde de Agroecologia e Desenvolvimento Sustentável*, 12, 3, 590-595.
1677. Sauri-Duch, E., Gutiérrez-Canul, C., Cuevas-Glory, L.F. et al. (2021). Determination of quality characteristics, phenolic compounds and antioxidant activity of propolis from southeastern Mexico. *J. Apicult. Sci.*, 65(1), 109-122.
1678. Schneiderova K., Smejkal K. (2015). Phytochemical profile of *Paulownia tomentosa* (Thunb) Steud. *Phytochem. Rev.*, 14 (5), 799–833.
1679. Schneiderová, K., & Šmejkal, K. (2014). Phytochemical profile of *Paulownia tomentosa* (Thunb). Steud. *Phytochemistry Reviews*, 2014, 1-35.
1680. Sforcin, J.M. Biological properties and therapeutic applications of propolis. *Phytother. Res.*, 30.6 (2016):894-905.
1681. Shahinozzaman, M, D.N. Obanda, S. Tawata. Chemical composition and pharmacological properties of Macaranga-type Pacific propolis: A review. *Phytotherapy Res.*, 2021, 35.1, 207-222.
1682. Siheri, W., Ebiloma, G.U., Igoli, J.O. et al. (2019). Isolation of a novel flavanonol and an alkylresorcinol with highly potent anti-trypanosomal activity from Libyan propolis., *Molecules*, 24(6), 1041.
1683. Silva-Carvalho, R., Baltazar, F., Almeida-Aguiar, C. Propolis: a complex natural product with a plethora of biological activities that can be explored for drug development. *Evidence-based Comple. Altern. Med.* 2015, art. number 206439.
1684. Silva, C.C.F.D., Salatino, A., Motta, L.B.D. et al. (2019). Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from Ceará state. *Revista Brasileira de Farmacognosia*, 29, 309-318.
1685. Sofrenić, I., Ljujić, J., Simić, K. et al. (2021). Application of LC–MS/MS with ion mobility for chemical analysis of propolis extracts with antimicrobial potential. *J. Serbian Chem. Soc.*, 86(12), 1205-1218.
1686. Souza, E.A.D., Inoue, H.T., Fernandes Júnior, A. et al. Influence of seasonality and production method on the antibacterial activity of propolis. *Acta Scientiarum. Animal Sci.*, 2014, 36(1), 49-53.
1687. Spulber, R., Colta, T., Băbeanu, N., & Ovidiu, P.O.P.A. (2017). Chemical diversity of polyphenols from bee pollen and propolis. *AgroLife Scientific J.*, 6(2).
1688. Susilo, A., Cahyati, M., Pranowo, D. et al. (2024). Chrysin inhibits Indonesian serotype foot-and-mouth-disease virus replication: insights from DFT, molecular docking, and molecular dynamics analyses. *J. Trop. Biodiv. & Biotechnol.*, 9(1), 1-16.
1689. Suzuki, T., Katayama, T., Pujirahayu, N. (2019). Cycloartane-type triterpenes and botanical origin of propolis of stingless indonesian bee *Tetragonula sapiens*. *Plants* (2223-7747), 8(3).
1690. Svečnjak, L. et al. Mediterranean propolis from the Adriatic Sea islands as a source of natural antioxidants: comprehensive chemical biodiversity determined by GC-MS, FTIR-ATR, UHPLC-DAD-QqTOF-MS, DPPH and FRAP assay. *Antioxidants* 9.4 (2020):337.
1691. Toreti, V.C., Sato, H.H., Pastore, G.M., Park, Y.K. Recent progress of propolis for its biological and chemical compositions and its botanical origin. *Evid. Based Compl. Altern.*

- Med., art. no. 697390, 2013, ISSN: 1741-427X
1692. Tran, T.D., et al. Lessons from exploring chemical space and chemical diversity of propolis components. *Int. J. Mol. Sci.*, 21.14 (2020): 4988.
 1693. Vranješ, M., 2023. *Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa* (Doctoral dissertation, University of Zagreb. Faculty of Veterinary Medicine).
 1694. Wali, A.F., et al. Bee propolis (bee's glue): a phytochemistry review. *J. Crit. Rev.* 4.4 (2017).
 1695. Wibowo, I., Utami, N., Anggraeni, T. et al. (2021). Propolis can improve caudal fin regeneration in zebrafish (*Danio rerio*) induced by the combined administration of Alloxan and glucose. *Zebrafish*, 18(4), 274-281.
 1696. Yilmaz, M.A., O. Cakir. Phytochemical constituents of a precious honeybee product: propolis. In: *Biological Activity of Honeybee products*, 2014.
 1697. Yuan, J., Lu, Y., Abula, S., et al. Optimization on preparation condition of propolis flavonoids liposome by response surface methodology and research of its immunoenhancement activity. *Evid.-Based Compl. Alt. Med.*, art. no. 505703, 2013.
 1698. Yusoff, Y. M. (2018). The use of metabolomics tools to assess the therapeutic natural products of honey and propolis from Malaysia and New Zealand. Doctoral dissertation, University of Strathclyde.
 1699. Yuniasari, H. (2019). Pengaruh pemberian ekstrak propolis terhadap pertumbuhan *propionibacterium acnes* secara in vitro. Doctoral dissertation, Universitas Lampung.
 1700. Zng C., Hu F. (2012). Overview of propolis research at home and abroad in 2011. *Chinese Bee Industry*, (0Z1), 64-73.
 1701. Zhang, C., Shun, P. H. & Hu, F. Study of the chemical composition of propolis from different geographical and plant origin. *Chinese Pharm. J.*, 2013, 48 (022), 1889-1892.
 1702. Zhang Hongcheng, Zhao Liangliang, Hu Hao, & Dong Jie. (2014). Analysis of polyphenols in propolis and their antioxidant activity. *Food Science*, (13), 59-65.
 1703. Zulhendri, F., Chandrasekaran, K., Kowacz, M. et al. (2021). Antiviral, antibacterial, antifungal, and antiparasitic properties of propolis: A review. *Foods*, 10(6), 1360..
- 85). Popova, M., V. Bankova, S. Bogdanov, I. Tsvetkova, C. NAYDENSKI, G.- L. Marcazzan, A.-G. Sabatini. Chemical characteristics of poplar type propolis of different geographic origin. *Apidologie* 38 306– 311 (2007)**
1704. Abdulla, B., Rashed, R., Hamasalih, R. et al. Antibacterial and antifungal activities of *Apis mellifera* L. honey, propolis, royal jelly in Iraqi Kurdistan region. *Latin Am. J. Biotechnol. Life Sci.*, 2023.
 1705. Abdulah, T. (2016). Comparison between Iraqi and European ether extract of propolis on *Staphylococcus aureus* isolated from minced meat. *Assiut Vet. Med. J.*, 62(151), 1-7.
 1706. Aboulghazi, A., et al. Physicochemical characterization and in vitro evaluation of the antioxidant and anticandidal activities of Moroccan propolis. *Vet. World* 15.2 (2022):341.
 1707. Aguiar, T. et al. Residual polysaccharides from fungi reduce the bacterial spot in tomato plants. *Bragantia* 77.2 (2018):299-313.
 1708. Alafandy, A.S. Assessment of crude propolis as a direct pulp capping agent in primary and immature permanent teeth. *Endodontic Practice Today*, 2014, 8(3), 199-206.
 1709. Alafandy, A.S., S.S. Barakat. Histological evaluation of pulp response to Syrian crude propolis as a pulpotomy agent in primary and immature permanent teeth--In vivo study."

- Quintessence Int., 9.3 (2015):201-09.
1710. Alvarez, M.V., Moreira, M.R., & Ponce, A. Antiquorum sensing and antimicrobial activity of natural agents with potential use in food. *J. Food Safety*, 2012, 32(3), 379-387.
 1711. Alvarez, M.V., Ponce, A.G., Mazzucotelli, C.A. et al. The impact of biopreservatives and storage temperature in the quality and safety of minimally processed mixed vegetables for soup. *J. Sci. Food Agricult.* 2015, 95, 962-971.
 1712. Alvarez, M.V., Ponce, A.G., Moreira, M.R. combined effect of bioactive compounds and storage temperature on sensory quality and safety of minimally processed celery, leek and butternut squash. *J. Food Safety*, 2015, 35, 560-574.
 1713. Alvarez, M.V., et al. Physical treatments and propolis extract to enhance quality attributes of fresh-cut mixed vegetables. *J. Food Process. Preserv.*, 41.5 (2017):e13127.
 1714. Alvear, M., Santos, E., Cabezas, F. et al. (2021). Geographic area of collection determines the chemical composition and antimicrobial potential of three extracts of Chilean propolis. *Plants*, 10(8), 1543.
 1715. Anđelković, B.D. Primena rezultata NMR i FTIR spektroskopskih tehnika u multivarijantnoj analizi za klasifikaciju propolisa. Diss. Univerzitet u Beogradu-Hemijski fakultet, 2017.
 1716. Anđelković, B. et al. Metabolomics study of *Populus* type propolis. *J. Pharm. Biomed. Anal.*, 135 (2017):217-226.
 1717. Asawahame, C., Sutjarittangtham, K., Eitssayeam, S. et al. Antibacterial activity and inhibition of adherence of *Streptococcus mutans* by propolis electrospun fibers. *AAPS PharmSciTech*, 2015, 16, 182-191.
 1718. Ashry, E.S.H.E., & Ahmad, T.A. The use of propolis as vaccine's adjuvant. *Vaccine*, 2012, 31(1), 31-39
 1719. Bakchiche, B. et al. Chemical composition and biological activities of honeybee products from Algeria. *J. Appl. Biotechnol. Rep.* 7.2 (2020):93-103.
 1720. Bakchiche, B. et al. Total phenolic, flavonoid contents and antioxidant activities of honey and propolis collected from the region of Laghouat (South of Algeria). *World News of Natural Sciences. An International Scientific Journal* 11 (2017).
 1721. Bayram, N. et al. Chemical characterization of 64 propolis samples from Hakkari, Turkey. *Rec. Nat. Products*, (2018), 12(6).
 1722. Baysa, M. (2018). Propolisin antipiretik etkisinin belirlenmesi (Master's thesis, Fen Bilimleri Enstitüsü).
 1723. Bazine Nahla, K.W.K.M. (2023). Evaluation de l'effet protecteur de la propolis contre la cardiotoxicité chez la souris. Master's thesis.
 1724. Benhanifia M., Mohamed WM., Bellik Y., Benbarek H. Antimicrobial and antioxidant activities of different propolis samples from north-western Algeria. *Int. J. Food Sci. Technol.*, 12, 48, 2013, 2521-2527.
 1725. Benhanifia, M., Mohamed, W.M. Phenolics constituents of different types of propolis and their antimicrobial activities *Anti-Infective Agents* 2015, 13, 17-27.
 1726. Benhanifia, M., Soltani, A. Biological activity of propolis: An Update. *Preprints* 2022, 2022030009..
 1727. Benkovic, V., A.H. Knezevic, D. Dikic, et al. Radioprotective effects of propolis and quercetin in γ -irradiated mice evaluated by the alkaline comet assay. *Phytomedicine* 15(10), 851-858 (2008)

1728. Betances-Salcedo, E. et al. Flavonoid and antioxidant capacity of propolis prediction using near infrared spectroscopy. *Sensors* 17.7 (2017):1647.
1729. Bogdanov, S. (2007). Authenticity of honey and other bee products: state of the art. *Bull. Univ. Agricult. Sci. Vet. Med. Cluj-Napoca. Animal Sci. Biotechnol.*, 2007, Vol. 63/64, 1-8 ref. 54.
1730. Bogdanov, V. L. (2012). Influence of initial stresses on the stressed state of a composite with a periodic system of parallel coaxial normal tensile cracks. *J. Math. Sci.*, 186, 1-13.
1731. Boke Sarikahya, N. et al. Comparative study of antiviral, cytotoxic, antioxidant activities, total phenolic profile and chemical content of propolis samples in different colors from Turkiye. *Antioxidants* 11.10 (2022):2075.
1732. Bouchelaghem, S. Propolis characterization and antimicrobial activities against *Staphylococcus aureus* and *Candida albicans*: A review. *Saudi J. Biol. Sci.* (2021).
1733. Bremer, M. et al. Investigation on the potential of poplar bark from short-rotation coppices as bio-based fungicidal additives. *BioEnergy Res.*, (2021):1-10.
1734. Buriol, L. et al. Chemical composition and biological activity of oil propolis extract: an alternative to ethanolic extract. *Quimica Nova* (2009), 296 – 302.
1735. Caetano, A.R., Oliveira, R.D., Pereira, R.F.C. et al. (2023). Examination of raw samples and ethanol extracts of gerês propolis collected in different years. *Plants*, 12(22), 3909.
1736. Çakır, B., Güzel, N., 2023. Optimization of propolis extraction under thermosonication and quantification of bioactive compounds by LC-MS/MS. *Food Anal. Methods*, pp.1-13.
1737. Cardinault, N., Cayeux, M.O., du Sert, P.P. La propolis: origine, composition et propriétés. *Phytothérapie*, 2012, 10(5), 298-304.
1738. Çelemlı, Ö.G., M. Atakay, K. Sorkun. The correlation between botanical source and the biologically active compounds of propolis. *İstanbul J. Pharm.*, 49.2 (2019):81-87.
1739. Cerqueira, Patrícia, Ana Cunha, and Cristina Almeida-Aguiar. "Potential of propolis antifungal activity for clinical applications." *Journal of Applied Microbiology*.
1740. Cerqueira, P., A. Cunha, C. Almeida-Aguiar. Potential of propolis antifungal activity for clinical applications. *J. Appl. Microbiol.*, 133(3), 1207-1228.
1741. Chiappe, C. S., Iurlina, M. O., & Saiz, A. I. (2020). Anti-quorum sensing activity of Argentinean honey and effect of pH on main flavonoids. *Adv. J. Food Sci. Technol.* 18(1), 1-8.
1742. Choudhary P., Tushir S., Bala M. et al. Exploring the potential of bee-derived antioxidants for maintaining oral hygiene and dental health: a comprehensive review. *Antioxidants*, 12 (7), art. no. 1452, 2023.
1743. Coneac, G.H., Vlaia, L., Olariu, I., et al. Experimental researches for standardization of hidroalcoholic extracts of propolis from the west region of Romania. *Farmacia*, 2014, 62(2), 400-412.
1744. Curifuta, M., Vidal, J., Sánchez-Venegas, J., et al. The in vitro antifungal evaluation of a commercial extract of Chilean propolis against six fungi of agricultural importance. *Ciencia e Investigación Agraria*, 2012, 39(2), 347-359.
1745. Dadar, M., et al. Evaluation of in vitro anti-brucella activity and chemical composition of different geographically distinct propolis from Iran. *Arch. Razi Institute* 77.1 (2022):55-62.
1746. de Almeida, S.L., Schmidt, É.C., Pereira, D.T., et al. Effect of ultraviolet-B radiation in laboratory on morphological and ultrastructural characteristics and physiological

- parameters of selected cultivar of *Oryza sativa* L. *Protoplasma*, 6, 250, 2013, 1303-1313.
1747. de Castro Ishida, V.F., Negri, G., Salatino, A. et al. A new type of Brazilian propolis: Prenylated benzophenones in propolis from Amazon and effects against cariogenic bacteria. *Food Chem.*, 2011, 125(3), 966-972.
 1748. de Figueiredo, S.M., A. Nogueira-Machado, J., de M Almeida, B., et al. Immunomodulatory properties of green propolis. *Recent Patents on Endocrine, Metabolic & Immune Drug Discovery*, 2014, 8(2), 85-94.
 1749. De Groot AC. Propolis: a review of properties, applications, chemical composition, contact allergy, and other adverse effects. *Dermatitis*, 6, 24, 2013, 263-282.
 1750. de Oliveira, Lídia Procópio. "Caracterização química da geoprópolis de abelhas sem ferrão amazônicas." (2014).
 1751. de Oliveira, Lídia Procópio. "Estudo químico da geoprópolis de *Melipona seminigra* Merriale Cockerell, 1919." (2015).
 1752. Devequi-Nunes, D. et al. Chemical characterization and biological activity of six different extracts of propolis through conventional methods and supercritical extraction. *PLoS One* 13.12 (2018): e0207676.
 1753. Dezmiorean, D.S., et al. (2020), Plant sources responsible for the chemical composition and main bioactive properties of poplar-type propolis. *Plants* 2021, 10, 22.
 1754. Dezmiorean, D. S. et al. Influence of geographic origin, plant source and polyphenolic substances on antimicrobial properties of propolis against human and honey bee pathogens. *J. Apicult. Res.*, 56.5 (2017):588-597.
 1755. Diallo, M. (2020). Chemical characterization and bioactivity of poplar, green and red propolis: a screening study with a food preservation purpose (Doctoral dissertation, Instituto Politécnico de Bragança (Portugal)).
 1756. Dimkić, I. et al. Phenolic profiles and antimicrobial activity of various plant resins as potential botanical sources of Serbian propolis. *Indust. Crops Products* 94 (2016):856-871.
 1757. Ding, J. et al. Daily Brazilian green propolis intake elevates blood artemisin C levels in humans. *J. Sci. Food Agric.* (2021), 101, 11, 4855-4861..
 1758. Dolabella, L.M.P. Estudo eletroquímico sobre a corrosão de aço carbono e aço inoxidável em meio contendo cloreto na presença de extrato etanólico de própolis. (2016), Doctoral dissertation, Universidade Federal de Minas Gerais.
 1759. dos Santos, Thaíse Fernanda Santana, et al. "MSPD procedure for determining buprofezin, tetradifon, vinclozolin, and bifenthrin residues in propolis by gas chromatography–mass spectrometry." *Analytical and bioanalytical chemistry* 390.5 (2008): 1425-1430.
 1760. Dranca, F., F. Ursachi, M. Oroian. Bee bread: physicochemical characterization and phenolic content extraction optimization. *Foods* 9.10 (2020):1358.
 1761. Drescher, N., Wallace, H.M., Katouli, M. et al. Diversity matters: how bees benefit from different resin sources. *Oecologia*, 176(4), 2014, 943-953.
 1762. Duke, C.C., et al. A sedge plant as the source of Kangaroo Island propolis rich in prenylated p-coumarate ester and stilbenes. *Phytochemistry* 134 (2017):87-97.
 1763. El Sayed, H., T. A. Ahmad. The use of propolis as vaccine's adjuvant. *Vaccine* 31.1 (2012):31-39.
 1764. El-Shahawi, M.S., R. Al-Hindi. Physicochemical characteristics of Saudi Arabian locally produced raw and diluted honeys and their relations to antimicrobial activity. *J.*

1765. Escriche, I., M. Juan-Borrás. Standardizing the analysis of phenolic profile in propolis. Food Res. Int., 106 (2018):834-841.
1766. Evran, E., Durakli-Velioglu, S., Velioglu, H.M. et al. (2023). Effect of wax separation on macro-and micro-elements, phenolic compounds, pesticide residues, and toxic elements in propolis. Food Sci. & Nutrit., 12(3), 1736-1748..
1767. Falcão SI., Freire C., Vilas-Boas M. A proposal for physicochemical standards and antioxidant activity of Portuguese propolis. JAOCS, J. Am. Oil Chemists' Soc., 11, 90, 2013, 1729-1741.
1768. Falcao, S.I.D.M. (2013). Chemical composition of Portuguese propolis boactive properties (Doctoral dissertation, Universidade do Porto (Portugal)).
1769. Fedotova, V.V., D.A. Konovalov. Propolis research in Russia. Indian J. Pharm. Educ. Res 53 (2019):500-509.
1770. Fidan, M., Pinar, S.M., Erez, M.E. et al. (2022). Determination of botanical origin and mineral content of propolis samples from Balveren (Şırnak) beekeepers accommodation areas. Commagene J. Biol., 6(2), 165-171..
1771. Fokt, H., Pereira, A., Ferreira, A.M. et al. How do bees prevent hive infections? The antimicrobial properties of propolis. Curr. Res. Technol. Educ. Topics Appl. Microbiol. Microbial Biotechnol., 2010, 1, 481-493.
1772. Fortier, J. et al. Potential for hybrid poplar riparian buffers to provide ecosystem services in three watersheds with contrasting agricultural land use. Forests 7.2 (2016):37.
1773. Frazão, J. Desenvolvimento de formulações cosméticas utilizando produtos apícolas e voláteis de cogumelos silvestres: determinação da estabilidade e toxicidade.)Master's thesis, Instituto Politecnico de Braganca, Portugal). 2017.
1774. Gardini, S. et al. Chemical composition of Italian propolis of different ecoregional origin. J. Apicult. Res., 57.5 (2018):639-647.
1775. Garmani, M., Kohsari, H., Hadi, Seyed Elangi, & Seyed Zahra. (2019). Antibacterial and antioxidant activity of several types of bee propolis collected from different geographical regions of Golestan province. Sci. J. Vet. Microbiol., 14(2), 25-35.
1776. Gracić, D. (2016). Inhibicija korozije bakra pomoću propolisa (Doctoral dissertation, University of Split. Faculty of Chemistry and Technology. Division of Engineering and Chemistry).
1777. Hage, S., G.E. Morlock. Bioprofiling of Salicaceae bud extracts through high-performance thin-layer chromatography hyphenated to biochemical, microbiological and chemical detections. J. Chromatography A 1490 (2017):201-211.
1778. Hamilton, K.D. Evaluation of the anti-inflammatory, anti-oxidant and wound-healing potential of cerumen from the Australian native stingless bee, *Tetragonula carbonaria*. Doctoral dissertation, University of the Sunshine Coast, Queensland, 2015.
1779. Hasanah, K.U. Uji daya antifungi propolis terhadap *Candida albicans* dan *Pityrosporum ovale*. (2012).
1780. Hernández-Martínez, J.A., Zepeda-Bastida, A., Morales-Rodríguez, I. et al. (2024). Potential antidiabetic activity of *Apis mellifera* propolis extraction obtained with ultrasound. Foods, 13(2), 348.
1781. . Hernández Martínez, J.A. (2023). Compuestos bioactivos, actividad antioxidante y antidiabética de extractos de propóleo obtenidos por ultrasonido en condiciones de

- digestión simulada. Master's thesis.
1782. Herrera, C.L., Alvear, M., Barrientos, L., et al. The antifungal effect of six commercial extracts of Chilean propolis on *Candida* spp. *Ciencia e Investigacion Agraria*, 2010, 37, 1, 75-84.
 1783. Hu, H. et al. Two novel markers to discriminate poplar-type propolis from poplar bud extracts: 9-oxo-ODE and 9-oxo-ODA. *J. Food Comp. Analysis* 105 (2022):104196.
 1784. Ibrahim, N, et al. Chemical and biological analyses of Malaysian stingless bee propolis extracts. *Malaysian J. Anal. Sci.*, 20.2 (2016): 413-422.
 1785. Irigoiti, Y., D.K. Yamul, A.S. Navarro. Co-crystallized sucrose with propolis extract as a food ingredient: Powder characterization and antioxidant stability. *LWT* 143 (2021):111164.
 1786. Isidorov, V.A., Szczepaniak, L., & Bakier, S. Rapid GC/MS determination of botanical precursors of Eurasian propolis. *Food Chem.*, 2014, 142, 101-106.
 1787. Isidorov, Valery A. et al. Activity of selected plant extracts against honey bee pathogen *Paenibacillus* larvae. *Apidologie* 49.6 (2018):687-704.
 1788. Isidorov, V.A. et al. In vitro study of the antimicrobial activity of European propolis against *Paenibacillus larvae*. *Apidologie* 48.3 (2017):411-422.
 1789. Isidorov, V.A., Dallagnol, A.M., & Zalewski, A. (2024). Chemical composition of volatile and extractive components of Canary (Tenerife) propolis. *Molecules*, 29(8), 1863.
 1790. Islam M., Malakar S., Dwivedi U. et al. Impact of different drying techniques on grapefruit peels and subsequent optimization of ultrasonic extraction conditions for bioactive compounds. *J. Food Process Eng.*, 46 (6), e14331, 2023.
 1791. Jermalionok, Josif. "Jonažolių sirupo modeliavimas ir kokybės vertinimas." (2011). https://lsmuni.lt/cris/bitstream/20.500.12512/100458/1/Diplominis_darbas.pdf
 1792. Jiang, X. et al. A new propolis type from Changbai mountains in North-east China: chemical composition, botanical origin and biological activity. *Molecules* 24.7 (2019):1369.
 1793. Jiang, X. et al. Grouping, spectrum–effect relationship and antioxidant compounds of Chinese propolis from different regions using multivariate analyses and off-line anti-DPPH assay. *Molecules* 25.14 (2020):3243.
 1794. Junaković E.P., Šandor K., Vujnović A., Oršolić N., Andrišić M., Žarković I., Špigelski K.V., Fajdić D., Sinković S., Terzić S. Spectrophotometric determination of the main polyphenol groups in propolis samples from different regions of Croatia [Spektrofotometrijsko određivanje glavnih polifenolnih skupina u uzorcima propolisa iz različitih područja Republike Hrvatske.] *Veterinarski Arhiv*, 93 (2), 257 – 270, 2023.
 1795. Kadhim, M. J., et al. Propolis in livestock nutrition. *Entomol. Ornithol. Herpetol* 7 2018, 207, 7, 2161..
 1796. Kapare, H. et al. Standardization, anti-carcinogenic potential and biosafety of Indian propolis. *J. Ayurveda Integr. Med.*, 10.2 (2019):81-87.
 1797. Kasote, D.M., et al. HPLC, NMR based chemical profiling and biological characterisation of Indian propolis. *Fitoterapia* 122 (2017):52-60.
 1798. Katekhaye, S, et al. Gaps in propolis research: challenges posed to commercialization and the need for an holistic approach. *J. Apicult. Res.*, 58.4 (2019):604-616.
 1799. Kekeçoğlu, M., Bellici, A.E., Yildirim, İ. Et al. 2023. Arı Ürünlerinin Kullanımı ve Apiterapi Uygulamalarına Bağlı İstenmeyen Etkileri: Sistemik Derleme. *J. Trad. Med. Compl. Therapies*, 6(2).

1800. Kızıldaş, H. Farklı kovanların propolis üretimine ve içeriğine (fenolik bileşim) etkisi (Master's thesis, Fen Bilimleri Enstitüsü).
1801. Ken, M. & F. Shuichi. Okinawa production of POLYPLASDONE of plants origin. Chem.Biology, 2010, 48 (1), 35-42.
1802. Keskin, Ş., L. Yatanaslan, S. Karlidağ. Farklı illerden toplanan propolis örneklerinin kimyasal karakterizasyonu. Uludağ Arıcılık Dergisi 20.1 (2020):81-88.
1803. Kolaylı S., Birinci C., Kara Y. et al.. A melissopalynological and chemical characterization of Anatolian propolis and an assessment of its antioxidant potential. Eur. Food Res. Technol., 249 (5), 1213-1233, 2023.
1804. Komzáková, Karolína. Fytochemické studium vybraných genotypů bazalky pravé (*Ocimum basilicum* L.). <https://theses.cz/id/uhticf/Komzkov.Karolna.2019.pdf>
1805. Koru, Ö., Bedir, O., Vit, P. et al. (2024). Evaluation of antimicrobial efficacy and phenolic compound profiles in geopropolis samples from Bolivia and Venezuela. Kafkas Univ. Vet. Fak. Dergisi, 30, 2, 215.
1806. Kucukates, E. (2022). Antifungal activity of propolis against candida species: propolis and antifungal action. In: Candida and Candidiasis. IntechOpen..
1807. Kumar, R., Srivastava, P., 2023. Comparative analysis of physicochemical and antioxidant activity of propolis. J. Exp. Zool., India, 26(2).
1808. Kumazawa S., N. Jun, O.Toshiro et al. (2010). Plant origin of propolis produced in Okinawa. Chemistry and Biology, 48(1), 35-42.
1809. Kylymnyuk, O., Khimich, O. and Laptieyev, O., 2023. Propolis in the diet of broiler chickens as a natural source of biologically active substance complexes. Feeds and Feed Production, (95), 179-185.
1810. Lai, M., Zhou, Y., Xiaoming, F. et al. Volatile components analysis of ethanol extract of propolis. Modern Food Sci. Technol., 2012, 28 (4), 456-461.
1811. Lectong Cusme, Nieve Esther, and Neiva Maricela Quiñonez Becerra. Efecto de La E-Polilisina y propóleo como conservantes en la vida útil del yogurt. MS thesis. Calceta: ESPAM MFL, 2020.
1812. Leoni, V., et al. Variabilità all'origine e fattori di qualità: nuove esperienze analitiche perla caratterizzazione della propoli." CEC, Erboristeria Domani, 2020 (420), 60-67.
1813. Machado, B.A.S. et al. Chemical composition and biological activity of extracts obtained by supercritical extraction and ethanolic extraction of brown, green and red propolis derived from different geographic regions in Brazil. PloS one 11.1 (2016): e0145954.
1814. Maroof, K., S.H. Gan. A review on chemical compositions, biological activity and formulation techniques of Malaysian honey bee and meliponine propolis. J. Biol. Active Products from Nature 10.6 (2020):507-523.
1815. Miguel, M.G., Antunes, M.D. Is propolis safe as an alternative medicine?. J.Pharm. & Bioal. Sci., 2011, 3(4), 479.
1816. Mohd Suib, M.S. et al. Ethanolic extract of propolis from the Malaysian stingless bee *Geniotrigona thoracica* inhibits formation of THP-1 derived macrophage foam cells. J. Apicult. Res., (2020):1-13.
1817. Mohd, K.S. et al. Propolis: traditional uses, phytochemical composition and pharmacological properties. Int. J. Eng. & Technol., 7.4.43 (2018):78-82.
1818. Moise, A.R., O. Bobiş. *Baccharis dracunculifolia* and *Dalbergia ecastophyllum*, main plant sources for bioactive properties in green and red Brazilian propolis." Plants 9.11

- (2020):1619.
1819. Moritz, R.F.A., de Miranda J, Fries I, et al. Research strategies to improve honeybee health in Europe. *Apidologie*, 2010, 41, 3, 227-242.
 1820. Nabernik, U. (2019). Razvoj metode za določitev permeabilnosti sestavin propolisa skozi kožo prašičjega uhlja in vitro (Doctoral dissertation, Univerza v Ljubljani, Fakulteta za farmacijo).
 1821. Niculae, M. et al. In vitro synergistic antimicrobial activity of Romanian propolis and antibiotics against *Escherichia coli* isolated from bovine mastitis. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 43.2 (2015):327-334.
 1822. Nweze, N.E., et al. Effects of Nigerian red propolis in rats infected with *Trypanosoma brucei brucei*. *Comp. Clin. Pathol.*, 26.5 (2017):1129-1133.
 1823. Nyandwi, R. et al. Determination and quantification of gallic acid in raw propolis by high-performance liquid chromatography–diode array detector in Burundi. *East Africa Sci.*, 1.1 (2019):43-48.
 1824. Okińczyc, P. et al. Phytochemical profile, plant precursors and some properties of Georgian propolis. *Molecules* 27.22 (2022):7714.
 1825. Okur, Ö.D.. Determination of antioxidant activity and total phenolic contents in yogurt added with black cumin (*Nigella sativa* L.) honey. *Ovidius Univ. Annals Chem.* 32.1 (2021):1-5.
 1826. Oroian, M., F. Ursachi, F. Dranca. Influence of ultrasonic amplitude, temperature, time and solvent concentration on bioactive compounds extraction from propolis. *Ultrasonics Sonochem.* 64 (2020): 105021.
 1827. Oroian, M., F. Ursachi, F. Dranca. Ultrasound-assisted extraction of polyphenols from crude pollen. *Antioxidants* 9.4 (2020): 322.
 1828. Oroian, M., F. Dranca, F. Ursachi. Comparative evaluation of maceration, microwave and ultrasonic-assisted extraction of phenolic compounds from propolis. *J. Food Sci. Technol.*, 57.1 (2020):70-78.
 1829. Oruç, H.H., Çayci, M., Sorucu, A. et al, 2023. Characterization of commercially available propolis products in Turkey based on individual phenolic compounds. *J. Apicult. Res.*, 62(5), 1225-1232.
 1830. Oruç, Hasan Hüseyin, Ali Sorucu, and Levent Aydın. Propolisin sağlık açısından önemi, kalitesinin belirlenmesi ve Türkiye açısından irdelenmesi. (2013).
 1831. Oruç, H.H. (2014). Propolisin sağlık açısından önemi, kalitesinin belirlenmesi ve Türkiye açısından irdelenmesi. *Uludağ Arıcılık Dergisi*, 14(1), 35-43.
 1832. Oruç, H.H. et al. Effects of season and altitude on biological active certain phenolic compounds levels and partial standardization of propolis. *Ankara Üniversitesi Veteriner Fakültesi Dergisi* 64.1 (2017):13-20.
 1833. Osés, S.M., et al. Design of a food product composed of honey and propolis. *J. Apicult. Res.*, 54.5 (2015):461-467.
 1834. Ouriques, L.C. et al. Physiological, morphological and ultrastructural responses to exposure to ultraviolet radiation in the red alga *Aglaothamnion uruguayense* (WR Taylor). *Brazilian J. Botany* 40.3 (2017):783-791.
 1835. Özenirler, Ç. et al. A new record for propolis substitute: pruning sealer. *Mellifera* 18.1 (2018):36-39.
 1836. Papachroni, D. et al. Phytochemical analysis and biological evaluation of selected

- African propolis samples from Cameroon and Congo. Nat. Prod. Comm. 10.1 (2015): 1934578X1501000118.
1837. Papotti, G, Bertelli D, Plessi M, et al. Use of HR-NMR to classify propolis obtained using different harvesting methods. Intern. J. Food Sci. Technol., 2010, 45, 8,1610-1618.
 1838. Papotti, G., Bertelli, D., Bortolotti, L., et al. Chemical and functional characterization of Italian propolis obtained by different harvesting methods. J. Agricult. Food Chem., 2012, 60(11), 2852-2862.
 1839. Paula V.B., Estevinho L.M., Cardoso S.M. et al. Comparative methods to evaluate the antioxidant capacity of propolis: an attempt to explain the differences. Molecules, 28 (12), art. no. 4847, 2023.
 1840. Pavlovic, R. et al. Effectiveness of different analytical methods for the characterization of propolis: a case of study in Northern Italy. Molecules 25.3 (2020):504.
 1841. Peixoto, S.R. (2023). Extraction of bioactive compounds from propolis for application in different matrices in food industry (Doctoral dissertation).
 1842. Pelvan, E. et al. Development of propolis and essential oils containing oral/throat spray formulation against SARS-CoV-2 infection. J. Funct. Foods 97 (2022):105225.
 1843. Perak J.E., 2023. Određivanje biodostupnosti odabranih polifenolnih sastavnica propolisa u uvjetima in vitro simulirane probave (Doctoral dissertation, University of Zagreb. Faculty of Science. Department of Chemistry).
 1844. Perak Junaković, E., Šandor, K., Vujnović, A. et al. 2023. Spectrophotometric determination of the main polyphenol groups in propolis samples from different regions of Croatia. Veterinarski arhiv, 93(2), 257-270.
 1845. Pereira, D.T. et al. Effects of salinity on the physiology of the red macroalga, *Acanthophora spicifera* (Rhodophyta, Ceramiales). Acta Bot. Brasilica 31.4 (2017):555-565.
 1846. Pereira, L, A. Cunha, C. Almeida-Aguiar. Portuguese propolis from Caramulo as a biocontrol agent of the apple blue mold. Food Control 139 (2022):109071.
 1847. Ping, Z., C. Jianqing, H. Fuliang, H. Yuanqiang, S. Qiaoyun. HPLC fingerprint graphs identification of different geographical origin of propolis and its authenticity. Apiculture of China, 2009, 60, 10.
 1848. Pontis, J.A., da Costa, L.A.M.A., da Silva, S.J.R., Flach, A. Color, phenolic and flavonoid content, and antioxidant activity of honey from Roraima, Brazil, Food, Sci. Technol., 2014, 34, 1, 69-73.
 1849. Puc, M. et al. Maščobne kisline in olja kot glavna sestavina prehranskih dopolnil. Univerza v Ljubljani, Biotehniška fakulteta, 2018.
 1850. Qiao J., Wang Y., Zhang Y. et al. Botanical origins and antioxidant activities of two types of flavonoid-rich poplar-type propolis. Foods, 12 (12), 2304, 2023.
 1851. Quetglas-Llabrés, M.M., Quispe, C., Herrera-Bravo, J. et al. (2022). Pharmacological properties of bergapten: mechanistic and therapeutic aspects. Oxidative medicine and cellular longevity, 2022:8615242..
 1852. Reyes Méndez, L.M.. Produção, caracterização e estudo da estabilidade de filmes à base de gelatina e extrato de própolis vermelha enriquecidos com óleos essenciais de manjerição (*Ocimum basilicum*), cravo (*Syzygium aromaticum*) ou hortelã (*Mentha piperita*). Doctoral dissertation. Universidade de São Paulo.
 1853. Ristivojević, P. et al. Phenolic composition and biological activities of geographically different type of propolis and black cottonwood resins against oral streptococci, vaginal

- microbiota and phytopathogenic *Fusarium* species. *J. Appl. Microbiol.*, 129.2 (2020):296-310.
1854. Ristivojević, P. et al. Poplar-type propolis: chemical composition, botanical origin and biological activity. *Nat. Product Commun.*, 10.11 (2015):1934578X1501001117.
1855. Rivera, C.A.S. et al. Applied techniques for extraction, purification, and characterization of medicinal plants active compounds. *Nat. Products Chem.: Biomed. Pharm. Phytochem.*, (2020):155-189.
1856. Rivero-Cruz, B., Martínez-Chávez, A. Development and validation of a RP-HPLC method for the simultaneous quantification of flavonoids markers in Mexican propolis. *Food Anal. Methods*, 2015, 8, 413-418.
1857. Rivero-Cruz, B., A. Martínez-Chávez. Development and validation of a RP-HPLC method for the simultaneous quantification of flavonoids markers in Mexican propolis. *Food Anal. Methods*, 8.2 (2015):413-419.
1858. Roman, A., Popiela-Pleban, E., Kowalska-Górska, M. (2012). Stan toksykologiczny propolisu wykorzystywanego w przemyśle farmaceutycznym i kosmetycznym. *Przemysł Chemiczny*, 91, 937-940.
1859. Romas, K., L. Vyshnevskaya, T. Zubchenko. Study of phenolic composition of medicine in the form of androgenic action capsules. *Ukrainian Biopharm. J.* 2 (63) (2020): 66-70.
1860. Rončević, M., Radas, K., Ljubojević Hadžavdić, S. (2023). Hypersensitivity Reactions to Homemade Topical Preparations. *Acta Dermatovenereologica Croatica*, 31(3), 117-124.
1861. Rosero, A. L. F. (2013). Facultad Piloto de Odontología (Doctoral dissertation, Universidad de Guayaquil).
1862. Saavedra, N., Barrientos, L., Herrera, C. et al. Efecto de propóleos chilenos sobre la bacteria cariogénica *Lactobacillus fermentum*. *Ciencia Investig. Agraria*, 2011, 38(1), 117-125.
1863. Saeed, F. et al. Propolis to curb lifestyle related disorders: An overview. *Int. J. Food Properties* 19.2 (2016): 420-437.
1864. Sağdıç, O., S. Karasu, H. Goktas. Piyasada satılan ticari propolis örneklerinin biyoaktif bileşenlerinin belirlenmesi. *Avrupa Bilim ve Teknoloji Dergisi* 19 (2020): 19-31.
1865. Şahin, Y. Arı ürünlerinde farklı materyalden kaşık kullanımının etkileri (Master's thesis, Fen Bilimleri Enstitüsü).
1866. Salcido Gallego, Mónica Guadalupe. Determinación de la actividad antibacteriana y antioxidante en propóleos de dos regiones de Sonora. (2010). <http://repositorioinstitucional.uson.mx/bitstream/handle/unison/2157/salcidogallegomonicaguadalupel.pdf?sequence=1&isAllowed=y>
1867. Salvatori, C. et al. Effectiveness of a standardized propolis extract in non-surgical periodontal therapy. (2021). DOI: <https://doi.org/10.21203/rs.3.rs-418104/v1>
1868. Sankaran S., Dubey R., Lohidasan S. Optimization of extraction conditions using response surface methodology and HPTLC fingerprinting analysis of Indian propolis. *J. Biol. Active Products from Nature*, 13 (1), 76-93, 2023.
1869. Santana dos Santos, T.F., Aquino, A., Dórea, H.S. et al. (2008). MSPD procedure for determining buprofezin, tetradifon, vinclozolin, and bifenthrin residues in propolis by gas chromatography–mass spectrometry. *Analyt. Bioanalyt. Chem.*, 390, 1425-1430..
1870. Santos, L.M., et al. Propolis: types, composition, biological activities, and veterinary product patent prospecting. *J. Sci. Food Agr.* 100.4 (2020): 1369-1382.
1871. Sarra, B. (2022). Antimicrobial and antibiofilm activities of Hungarian propolis on *Candida albicans* and *Staphylococcus aureus* and its mechanism of action. PhD thesis,

University of Pecs..

1872. Schmidt, É.C. et al. Profiles of carotenoids and amino acids and total phenolic compounds of the red alga *Pterocladia capillacea* exposed to cadmium and different salinities. *J. Appl. Phycol.*, 28.3 (2016):1955-1963.
1873. Segueni, N., Keskin, Ş., Kadour, B. et al. (2021). Comparison between phenolic content, antioxidant, and antibacterial activity of Algerian and Turkish propolis. *Comb. Chem. & High Throughput Screening*, 24(10), 1679-1687.
1874. Selvaraju, G.D. et al. Fabrication and characterization of surgical sutures with propolis silver nano particles and analysis of its antimicrobial properties. *J. King Saud University-Science* 34.5 (2022):102082.
1875. Seven, P.T., Yilmaz, S., Seven, I. et al. The Effects of propolis in animals exposed oxidative stress. In: *Oxidative Stress - Environmental Induction and Dietary Antioxidants* (V. Lushchak, ed.), 2012, 267-288.
1876. Shafik, Ansam. "Assessment of crude propolis as a direct pulp capping agent in primary and immature permanent teeth." *ENDO (Lond Engl)* 8.3 (2014): 199-206.
1877. Shafriani, N.R. (2021). Pengaruh propolis terhadap il-6 pada diabetes melitus tipe 2: a literature review. *Herb-Medicine Journal: Terbitan Berkala Ilmiah Herbal, Kedokteran dan Kesehatan*, 4(3), 57-71.
1878. Shariatipour, N. al. Assessing the potential of native ecotypes of *Poa pratensis* L. for forage yield and phytochemical compositions under water deficit conditions. *Sci. Rep.* 12.1 (2022):1-14.
1879. Shen, H. et al. Radial variation law of wood properties for *Populus deltoides*, *Populus nigra* and their hybrid progenies. *Beijing Linze Daxue Xuebao/Journal of Beijing Forestry University* 42, 5, 2020: 50-58.
1880. Siheri, W. et al. The chemical and biological properties of propolis. *Bee products-chemical and biological properties*. Springer, Cham, 2017. 137-178.
1881. Silici, S. Plant-honeybee interactions (Book Chapter), *Recent Advances in Plant Science* 2020: 313-340.
1882. Silici, S. Honeybee products and apitherapy. *Turkish J. Agr.-Food Sci. Technol.* 7.9 (2019):1249-1262.
1883. Silici, S. Propolis üzerine ön klinik araştırmalar. *Erciyes Üniversitesi Fen Bilimleri Enstitüsü Fen Bilimleri Dergisi* 31.3 (2015):185-191.
1884. Silva, L.R.D.O. (2023). Desenvolvimento de sistemas microparticulados contendo extrato de própolis marrom da região de União dos Palmares (Alagoas). Doctoral dissertation, Universidade Federal de Alagoas..
1885. Simionatto, E., Facco, J.T., Morel, A.F., et al. Chiral analysis of monoterpenes in volatile oils from propolis. *J. Chilean Chem. Soc.*, 2012, 57(3), 1240-1243.
1886. Simone, M.D. Colony-level immunity benefits and behavioral mechanisms of resin collection by honey bees. PhD Thesis, University of Minnesota, 2010.
1887. Simone-Finstrom, M., M. Spivak. Propolis and bee health: the natural history and significance of resin use by honey bees. *Apidologie* 41.3 (2010): 295-311.
1888. Soltani, E.K. (2018). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation).
1889. Soltani, A., Benhanifia, M. (2023). Propolis, plant sources and antimicrobial activity: an overview. *Anti-Infective Agents*, 21(5), 20-39.

1890. Sorucu, A., H.H. Oruç. Determination of biologically active phenolic compounds in propolis by LC–MS/MS according to seasons and altitudes. *J. Food Meas. Charact.* 13.3 (2019):2461-2469.
1891. Spulber, R. et al. Chemical diversity of polyphenols from bee pollen and propolis. *AgroLife Sci. J.*, 6.2 (2017):183-194.
1892. Stanciauskaite, M. et al. Evaluation of chemical composition, sun protection factor and antioxidant activity of Lithuanian propolis and its plant precursors. *Plants* 11.24 (2022): 3558.
1893. Stanciauskaite, M. et al. Extracts of poplar buds (*Populus balsamifera* L., *Populus nigra* L.) and Lithuanian propolis: comparison of their composition and biological activities. *Plants* 10.5 (2021):828.
1894. Suarez, H., Jiménez, Á., & Díaz, C. Determination of microbiological and sensory parameters of fish fillets with propolis preserved under refrigeration. *Revista MVZ Córdoba*, 19(3), 2014, 4214-4225.
1895. Sudarmadi, S.D. (2017). Pengaruh ekstrak ethanol propolis terhadap peningkatan ekspresi protein bax dan penurunan proliferasi sel pada kultur sel kanker payudara (cell line mcf-7) (Doctoral dissertation, UNS (Sebelas Maret University)).
1896. Suleman, T. The antimicrobial and chemical properties of South African propolis. Doctoral dissertation, 2016.
1897. Sun, M. (2019). Commercial propolis liquid products: comparison of physicochemical properties and antioxidant and antimicrobial properties: a thesis presented in partial fulfilment of the requirements for the degree of Master of Food Technology at Massey University, Auckland, New Zealand (Doctoral dissertation, Massey University).
1898. Sunil, L. S., Vignesh, S., & Meenatchi, R. (2021). Physicochemical properties and antimicrobial activity of Indian propolis. *J. Pharm. Innov*, 10, 1660-1664.
1899. Tian W., Z. Yazhou, F. Xiaoming et al. (2012). Analysis of volatile components of propolis alcohol extract. *Modern Food Sci. Technol.*, 28(4), 456-461.
1900. Tran, CTN, et al. Quality assessment and chemical diversity of Australian propolis from *Apis mellifera* bees. *Sci. Rep.*, 12.1 (2022):1-14.
1901. Thang, T.T.N. (2022). Caractérisation de la propolis d'abeille provenant du Canada, de la Roumanie et du Brésil. Doctoral thesis.
1902. Truchado, P., López-Gálvez, F., Gil, M.I. et al. *Food Chem.* 115(4), 1337-1344 (2009)
1903. Trujillo Celi, Cristian Fabián. Composición química de cinco muestras de propóleo del cantón Yantzaza de la provincia de Zamora Chinchipe. BS thesis. Machala: Universidad Técnica de Machala, 2015.
1904. Türk, M.U., Şahinler, N., Dinler, H. (2022). Chemical structure and antifungal activity of Aegean Region of propolis in Türkiye. *Turkish J. Agric. Food Sci. Technol.*, 10(12), 2571-2582.
1905. Ugrina, Teo. Utjecaj temperature na inhibiciju korozije bakra pomoću propolisa. Doctoral dissertation, University of Split. Faculty of Chemistry and Technology, 2016.
1906. Ünal, H. H. (2017). Effects of season and altitude on biological active certain phenolic compounds levels and partial standardization of propolis. Doctoral dissertation, Ankara Üniversitesi
1907. Vargas Tapia, Eulalia. "Identificação da fonte botânica, caracterização química e avaliação das atividades biológicas das própolis coletadas no Peru." (2018). http://repositorio.unicamp.br/bitstream/REPOSIP/334302/1/Tapia_EulaliaVargas_D.pdf

1908. Varoni M.E., Lodi, G., Sardella, A. et al. Plant polyphenols and oral health: old phytochemicals for new fields. *Curr. Med. Chem.*, 2012, 19(11), 1706-1720.
1909. Veloz, J., Cabezas, F., Fernández, C. and Lespinasse, M., 2023. The structural diversity of flavonoid present in chilean propolis determines their biological activity.
1910. Veloz, J.J., Saavedra, N., Lillo, A. et al. Antibiofilm activity of chilean propolis on *Streptococcus mutans* is influenced by the year of collection. *BioMed Res. Int.*, 2015, Art.num. 291351
1911. Verdugo Torres, María Augusta, and Bruno Esteban Tola Álvarez. Capacidad antioxidante y composición química de varios extractos de propóleos de la zona sur del Ecuador. MS thesis. 2017.
1912. Vlaia, Lavinia Lia, et al. "Preparation and Characterization of Inclusions Complexes between Propolis Ethanolic Extracts and 2-hydroxypropyl- β -cyclodextrin." *Rev. Chim.(Bucharest)* 67 (2016): 378.
1913. Vranješ, M., 2023. Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa (Doctoral dissertation, University of Zagreb. Faculty of Veterinary Medicine).
1914. Vránová, T. Antimikrobiální účinky extraktů řasy *Chlorella sorokiniana* vůči bakterii *Paenibacillus larvae*. 2017, Doctoral dissertation.
1915. Vujić, M. (2022). Kvaliteta i sigurnost dodataka prehrani na bazi pčelinjih proizvoda (Doctoral dissertation, University of Zagreb, Fac. Food Technol. Biotechnol.).
1916. Walker R.D.E.E.N. (2014). Elaboración, validación y evaluación de una guía farmacoterapéutica para una unidad de cuidados intensivos de un centro de salud privado en la ciudad de Valdivia. (Doctoral dissertation, Universidad Austral de Chile).
1917. Wang, K., Ping, S., Huang, S., et al. Molecular mechanisms underlying the in vitro anti-inflammatory effects of a flavonoid-rich ethanol extract from chinese propolis (poplar type). *Evidence-Based Compl. Altern. Med.*, 2013, 127672, ISSN: 1741-427X.
1918. Widelski J., Okińczyc P., Suśniak K. et al. Phytochemical profile and antimicrobial potential of propolis samples from Kazakhstan. *Molecules*, 28 (7), art. no. 2984, 2023.
1919. Xue, F. et al. Physicochemical properties of chitosan/zein/essential oil emulsion-based active films functionalized by polyphenols. *Future Foods*, 3, 2021, 100033.
1920. Yang, R., Yang, Y., Hu, Y. et al. 2023. Comparison of bioactive compounds and antioxidant activities in differentially pigmented *Cerasus humilis* fruits. *Molecules*, 28(17), p.6272.
1921. Yuniarto, Haryono. Pengaruh Ekstrak Ethanol Propolis Terhadap Ekspresi Protein Bcl2, Cyclin D1 dan Apoptosis pada Kultur Sel Kanker Kolon. Diss. UNS (Sebelas Maret University), 2016.
1922. Yusop, S.A.T.W. et al. Antioxidant, antimicrobial and cytotoxicity activities of propolis from Beladin, Sarawak stingless bees *Trigona itama* extract. *Materials Today: Proceedings* 19 (2019):1752-1760.
1923. Zeng Y., Zhang S., Song H. Physiological responses of *Populus euphratica* to leaf rust infection. *Zhiwu Shengli Xuebao/Plant Physiology Journal*, 59 (2), 353-361, 2023.
1924. Zhao, Y., Tian, W., & Peng, W. Anti-proliferation and insulin resistance alleviation of hepatocellular carcinoma cells HepG2 in vitro by Chinese propolis. *J. Food Nutrition Res.*, 2014, 2(5), 228-235.
1925. Zhao Y., T. Wenli, G.Zhanbao, et al. (2012). Optimization of ethanol extraction process of propolis based on response surface methodology. *Chinese J. Agric. Sci.*

- Technol., 14(3), 85-93.
1926. Zhou Ping, Chen Jianqing, Hu Fuliang, Hu Yuanqiang, & Shao Qiaoyun. (2009). HPLC fingerprint determination and authenticity determination of propolis from different origins. Chinese Bee Industry, (10), 5-8.
 1927. Zhou, Y., M. Lai. Surface methodology based on the response to optimize propolis ethanol extraction process. Chinese Agricult. Sci. Technol., 2012, 14 (3), 85-93.
 1928. Кайгородов, Р. Биохимические особенности растительных источников прополиса умеренной природной зоны. Вестник Пермского университета. Серия: биология, 2013, (3), 65-68.
 1929. Кайгородов, Р.В., О.А. Малькова, Ю.В. Кайгородова. Суммарное содержание фенольных соединений в прополисе разного ботанико-географического происхождения. Евразийский союз ученых 5-7 (14) (2015).
 1930. Кайгородов, Р., О.А. Малькова, Ю.В. Кайгородова. Тестирование антиоксидантных свойств спиртовых экстрактов прополиса с использованием липосомной модели. Educatio 4 (11)-3 (2015).
 1931. Мухидинов, З.К., С.Р. Усманова, Ф.Ю. Насырова. Перспективы изучения прополиса в Таджикистане. Вестник Авиценны 19.3 (2017).
 1932. Ромась, К.П., Вишневецька, Л.І., Зубченко, Т.М. et al. (2020). Вивчення фенольного складу лікувально-профілактичного засобу у формі капсул андрогенної дії.
 - 86). Popova M., Dimitrova R., Al-Lawati H.T., Tsvetkova I., NAJDENSKI H., Bankova V. Omani propolis: chemical profiling, antibacterial activity and new propolis plant sources. Chemistry Central Journal 2013, 7:158 doi:10.1186/1752-153X-7-158 (IF - 1.31)**
 1933. Abd Rashid, N., Mohammed, S.N.F., Syed Abd Halim, S. A. et al. (2022). Therapeutic potential of honey and propolis on ocular disease. Pharmaceuticals, 15(11), 1419.
 1934. Ádám, K., Péter, C., Ákos, J. (2022). A propolisz baktériumellenes hatékonysága–1. rész: Irodalmi összefoglaló. Magyar Állatorvosok Lapja, 144(5).
 1935. Acar, B.O., Gülenadağ, E., 2023. Comparison of antimicrobial activities of ethanol-and water-based propolis extracts on various foodborne pathogens by agar-well diffusion method. Res. Agricult. Sci., 54(3), 130-136.
 1936. Albokhadaim, I. Influence of dietary supplementation of propolis on hematology, biochemistry and lipid profile of rats fed high cholesterol diet. J. Adv. Vet. Animal Res., 2.1 (2015):56-63.
 1937. Al-Ghamdi, A.A. et al. Chemical compositions and characteristics of organic compounds in propolis from Yemen. Saudi J. Biol. Sci., 24.5 (2017):1094-1103.
 1938. Al-Lawati, H.T., H.I. Salim Al-Ajmi, M.I. Waly. Antioxidant and health properties of beehive products against oxidative stress-mediated carcinogenesis. Bioactive Components, Diet and Medical Treatment in Cancer Prevention. Springer, Cham, 2018. 97-103.
 1939. Alvaro, M.R., H.H. Alejandra, D.C. Antonio. In vitro antibacterial activity of *Maclura tinctoria* and *Azadirachta indica* against *Streptococcus mutans* and *Porphyromonas gingivalis*. J. Pharm. Res. Int., (2015):291-298.
 1940. Atoum, D., Fernandez-Pastor, I., Young, L et al., 2023. Use of multivariate analysis to unravel the differences between two chamomile varieties and their anticancer and antioxidant activities. Plants, 12(12), p.2297.
 1941. Avula, B. et al. Quantification and characterization of phenolic compounds from northern

- Indian propolis extracts and dietary supplements. J. AOAC Int., 103.5 (2020): 1378-1393.
1942. Badiazaman, A.A.M. et al. Phytochemical screening and antioxidant properties of stingless bee *Geniotrigona thoracica* propolis. Malaysian J. Fund. Appl. Sci., 15.2-1 (2019):330-335.
 1943. Bhuyan, D.J. et al. Broad-spectrum pharmacological activity of Australian propolis and metabolomic-driven identification of marker metabolites of propolis samples from three continents. Food & Function 12.6 (2021):2498-2519.
 1944. Bouchelaghem, S. et al. Evaluation of total phenolic and flavonoid contents, antibacterial and antibiofilm activities of Hungarian propolis ethanolic extract against *Staphylococcus aureus*. Molecules 27.2 (2022):574.
 1945. Bouchelaghem, S. Propolis characterization and antimicrobial activities against *Staphylococcus aureus* and *Candida albicans*: A review. Saudi J. Biol. Sci., 2022, 29(4), 1936-1946.
 1946. Buahorm, S., Puthong, S., Palaga, T. et al. (2015). Cardanol isolated from Thai *Apis mellifera* propolis induces cell cycle arrest and apoptosis of BT-474 breast cancer cells via p21 upregulation. DARU J. Pharm. Sci., 23(1), 1.
 1947. Calimag, K.P.D. et al. Attenuation of carrageenan-induced hind paw edema and plasma TNF- α level by Philippine stingless bee (*Tetragonula biroi* Friese) propolis. Exp. Animals (2020):20-0118.
 1948. Carol, D.M.E. et al. GC-MS characterization and antiulcer properties of the triterpenoid fraction from propolis of the north west region of Cameroon. J. Sci. Res. Rep. 2017:1-18.
 1949. Chuttong, B., Lim, K., Praphawilai, P. et al. (2023). Exploring the functional properties of propolis, geopropolis, and cerumen, with a special emphasis on their antimicrobial effects. Foods, 12(21), 3909.
 1950. Cortes, M.E. In vitro antimicrobial activity and biocompatibility of propolis containing nanohydroxyapatite. Nanohydroxyapatite-based antibacterial surfaces to prevent biofilm associated biomaterials bone infection (2015): 113.
 1951. da Silva, Caroline Cristina Fernandes, et al. "Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from Ceará state." Brazilian Journal of Pharmacognosy 29.3(2019): 309-318.
 1952. de Figueiredo, S.M, A Nogueira-Machado, J., de M Almeida, B., et al. Immunomodulatory properties of green propolis. Recent Patents on Endocrine, Metabolic & Immune Drug Discovery, 2014, 8(2), 85-94.
 1953. de Riesgo Asociados, A.P. (2016). Comportamento epidemiologica de los factores de riesgo asociados a prediabetes. Facultad de Ciencias Médicas (Doctoral dissertation, Universidad de San Carlos de Guatemala).
 1954. Dezmirean, D.S. et al. Influence of geographic origin, plant source and polyphenolic substances on antimicrobial properties of propolis against human and honey bee pathogens. J. Apicult. Res., 56.5 (2017):588-597.
 1955. do Nascimento, T.G. et al. Comprehensive multivariate correlations between climatic effect, metabolite-profile, antioxidant capacity and antibacterial activity of Brazilian red propolis metabolites during seasonal study. Sci. Rep., 9.1 (2019):18293.
 1956. Dvykaliuk, R. et al. (2023). Development of safety and quality of propolis as a food raw material. Animal Sci. Food Technol., 14(1), 36-48.
 1957. El-Khamsa, M.S. (2017). Caractérisation et activités biologiques de substances

- naturelles, cas de la propolis (Doctoral dissertation, Universite Ferhat Abbas–Setif).
1958. Farooqui, T., A.A. Farooqui. Apitherapy: therapeutic effects of propolis on neurological disorders. In: Neuroprotective Effects of Phytochemicals in Neurological Disorders (2017):335-358.
 1959. Fatima, Jamali, Baserisalehi Majid, and Bahador Nima. Antimicrobial activity and chemical screening of propolis extracts." Am. J. Life Sci., 2.2 (2014):72-75.
 1960. Grajales-Conesa, J. et al. Stingless bees propolis antimicrobial activity in combination with garlic, *Allium sativum* (Amaryllidaceae). Revista Biol. Trop., 69.1 (2021):22-35.
 1961. Grenho, L., Barros, J., Ferreira, C. et al. In vitro antimicrobial activity and biocompatibility of propolis containing nanohydroxyapatite Biomed. Materials (Bristol) 2015, 10(2), 025004.
 1962. Hakkim, F.L., J. Achankunju, S.S. Hasan. In vitro DPPH radical scavenging and antibacterial activity of Oman's cymbopogon. J. Ethnopharmacol., 72 (2000):191-205.
 1963. Hamza, A.M. (2014). Chromatographical profiles, antioxidant, antimicrobial and antimycetomal activities of selected propolis samples of Sudanese origin (Doctoral dissertation, University of Science and Technology).
 1964. Hasan, A.E.Z., Ambarsari, L., Widjaja, W.K. et al. Potency of nanopropolis stinglessbee *Trigona* spp. Indonesia as antibacterial agent. IOSR J. Pharm., 2014, 4, 12, 1-9.
 1965. Herrera-López, M.G. et al. Resorcinolic lipids from Yucatecan propolis. J. Brazilian Chem. Soc., 31.1 (2020):186-192.
 1966. Hossain, R. et al. Propolis: An update on its chemistry and pharmacological applications. Chinese Med., 17.1 (2022):1-60.
 1967. Iyen, S.I., Anyam, J.V., Igoli, J.O. et al. (2022). Cycloartanes and pentacyclic triterpenes from Awka and ijebuode propolis and evaluation of their antimicrobial activity. Scientia Africana, 21(3), 121-132.
 1968. Iftikhar, F., R. Mahmood. Propolis an antibacterial agent against clinical isolates of wound infection. Int. J. Sci. Res. Sci. Eng. Technol., 2015, 1(6), 47-52.
 1969. Jumare, F.I., Sirajo, M., Hashimu, T. et al. (2021). Antibacterial activity and phytochemical analyses of propolis (bee glue) extract against *Escherichia coli* and *Staphylococcus aureus*. Nigerian J. Microbiol., 35(2), 5752-5758.
 1970. Kardar, M.N., Zhang, T., Coxon, G.D., et al. Characterisation of triterpenes and new phenolic lipids in Cameroonian propolis. Phytochemistry, 2014, 106, 156-163.
 1971. Kerek, Á., Csanády, P., Tuska-Szalay, B. et al. 2023. In vitro efficacy of Hungarian propolis against bacteria, yeast, and *Trichomonas gallinae* Isolated from pigeons - A possible antibiotic alternative?. Resources, 12(9), p.101.
 1972. Kerek, A., P. Csanady, and A. Jerzsele. Antibacterial efficiency of propolis-Part 1. Magyar Allatorvosok Lapja 144.5 (2022):285-298.
 1973. King, D.I. Kangaroo island propolis: improved characterisation and assessment of chemistry and botanical origins through metabolomics. 2017, Doctoral dissertation.
 1974. Kubina, R., Kabała-Dzik, A., Dziedzic, A. et al. The ethanol extract of polish propolis exhibits anti-proliferative and/or pro-apoptotic effect on HCT 116 colon cancer and Me45 Malignant melanoma cells in vitro conditions Adv. Clin. Exp. Med., 2015, 24, 2, 203-212.
 1975. López, M.G.H., Irabien, L.M.C. (2019). Caracterización química y actividad biológica de propóleos producidos en el estado de Yucatán (Doctoral dissertation, Centro de Investigación Científica de Yucatán).

1976. Machado, A., Zamora-Mendoza, L., Alexis, F. et al. 2023. Use of plant extracts, bee-derived products, and probiotic-related applications to fight multidrug-resistant pathogens in the post-antibiotic era. *Future Pharmacol.*, 3(3), 535-567.
1977. Maragou, N.C., Strati, I.F., Gialouris, P.L. et al. 2023. Honey and bee products. In: *Emerging Food Authentication Methodologies Using GC/MS* (pp. 137-213). Cham: Springer International Publishing.
1978. Massaro, C.F. Antibacterial natural products of propolis and honeys from Australian stingless bees (*Tetragonula carbonaria*). Doctoral dissertation, University of the Sunshine Coast, Queensland, 2014.
1979. Massaro, C.F. et al. Chemical composition and antimicrobial activity of honeybee (*Apis mellifera ligustica*) propolis from subtropical eastern Australia. *The Science of Nature* 102.11 (2015):1-11.
1980. Matson Robles, Á.A. (2013). Efecto antibacteriano in vitro de maclura tinctoria y azadirachta indica sobre streptococcus mutans y porphyromonas gingivalis. Doctoral dissertation, Universidad de Cartagena.
1981. Maureira, N. et al. Susceptibilidad de cepas de candida oral a extracto etanólico del propóleo chileno de olmué. *Int. J. Odontostomatol.*, 11.3 (2017):295-303.
1982. Merchan Ortega, K.F., Valle Quinde, V.A. (2023). Caracterización química y capacidad antioxidante de la muestra de propóleos ecuatoriano ep12.
1983. Mohiuddin, I., Kumar, T.R., Zargar, M.I. et al. (2022). GC-MS analysis, phytochemical screening, and antibacterial activity of *Cerana indica* propolis from Kashmir region. *Separations*, 9(11), 363.
1984. Moghim, H. et al. Comparative study on the antifungal activity of hydroalcoholic extract of Iranian propolis and Royal jelly against *Rhizopus oryzae*. *J. Herbmed Pharmacol.*, 4.3 (2015):89-92.
1985. Morales Muñoz, H. Perfil de HPLC, RMN y actividad biológica de extractos etanólicos de geopropóleos de abejas meliponas de la Región Centro de Veracruz. (2015).
1986. Mukaide, K. et al. Prenylflavonoids from propolis collected in Chiang Mai, Thailand. *Phytochem. Lett.*, 43 (2021):88-93.
1987. Naik, R.R., Shakya, A.K., Oriquat, G A. et al. (2021). Fatty acid analysis, chemical constituents, biological activity and pesticide residues screening in Jordanian propolis. *Molecules*, 26(16), 5076.
1988. Negri, G. et al. Cardanols detected in non-polar propolis extracts from *Scaptotrigona aff. postica* (Hymenoptera, Apidae, Meliponini). *Brazilian J. Food Technol.*, 22 (2019).
1989. Nyandwi, R., Kılıç, A.S., Çelik, M. et al. (2019). Determination and quantification of gallic acid in raw propolis by high-performance liquid chromatography–diode array detector in Burundi. *East Africa Sci.*, 1(1), 43-48.
1990. Nouredine, H. et al. Chemical characterization and cytotoxic activity evaluation of Lebanese propolis. *Biomed. & Pharmacother.*, 95 (2017):298-307.
1991. Nyandwi, Ramadhan, et al. "Determination and Quantification of Gallic Acid in Raw Propolis by High-performance Liquid Chromatography–Diode Array Detector in Burundi." *East Africa Science* 1.1 (2019): 43-48.
1992. Osés, S.M., et al. Design of a food product composed of honey and propolis. *J. Apicult. Res.*, 54.5 (2015):461-467.
1993. Oyagbemi, A.A. et al. Protective effect of Azadirachta indica and vitamin E against

- arsenic acid-induced genotoxicity and apoptosis in rats. J. Diet. Suppl., 15.3 (2018): 251-268.
1994. Oyagbemi, Ademola Adetokunbo, et al. "Protective Effect of Azadirachta indica and Vitamin E Against Arsenic Acid-Induced Genotoxicity and Apoptosis in Rats." Journal of dietary supplements (2017): 1-18.
1995. Peixoto, M. et al. Antioxidant and antimicrobial activity of blends of propolis samples collected in different years. LWT 145 (2021): 111311.
1996. Pratami, D.K. et al. Phytochemical profile and antioxidant activity of propolis ethanolic extract from tetragonula bee. Pharmacogn. J. 10.1 (2018):128-135.
1997. Przybyłek, I., T.M. Karpiński. Antibacterial properties of propolis. Molecules 24.11 (2019):2047.
1998. Rebiai, A. et al. Fatty acid composition of Algerian propolis. J. Fund. Appl. Sci., 9.3 2017:1656-1671.
1999. Renteria, I.P.D.G. (2018). Caracterización química y actividad biológica de extractos etanólicos de propóleo del estado de Sinaloa (Doctoral dissertation, Universidad Autónoma De Sinaloa).
2000. Saadawi, S.S., Amara, R.O., Mughrbi, H.N. et al. (2022). Green synthesis and characterization of Libyan propolis nanoparticles and its biological activity. South Asian Res J Pharm Sci, 4(2), 28-35.
2001. Salama, S.F., A.A. Hassan. Effect of Egyptian propolis extract as an adjuvant with irradiated cancer vaccine against Ehrlich ascites carcinoma in mice. Egypt J Radiat Sci Appl 27 (2014):1-15.
2002. Sanpa, Sirikarn, et al. "Chemical Profiles and Antimicrobial Activities of Thai Propolis Collected from Apis mellifera."
2003. Schmidt E., Stock D., Chada F.J. et al. A Comparison between characterization and biological properties of Brazilian fresh and aged propolis. BioMed Res. Int., 2014, 257617, 1-10.
2004. Schmidt, E.M. et al. A comparison between characterization and biological properties of Brazilian fresh and aged propolis. BioMed Res. Int., 2014 (2014).
2005. Selvaraj, R. et al. Phytochemical profiling and antibacterial activity of propolis. Int. J. Sci. Res., 2018, 7, 6, 4-7.
2006. Serafim, M. S. (2019). Própolis marrom e resina de Eucalyptus botryoides da região de Bambuí-MG: isolamento de compostos fenólicos e avaliação da atividade antimicrobiana (Doctoral dissertation, Universidade de São Paulo).
2007. Siddiqui, S. et al. Interaction of bioactive compounds of *Moringa oleifera* leaves with SARS-CoV-2 proteins to combat COVID-19 pathogenesis: A phytochemical and in silico analysis. Appl. Biochem. Biotechnol., 194.12 (2022):5918-5944.
2008. Siddiqui, S., Ahmad, R., Alaidarous, M. et al. (2022). Phytoconstituents from *Moringa oleifera* fruits target ACE2 and open spike glycoprotein to combat SARS-CoV-2: An integrative phytochemical and computational approach. J. Food Biochem., 46(5), e14062..
2009. Siddiqui, Sahabjada, et al. Phytoconstituents from *Moringa oleifera* fruits target ACE2 and open spike glycoprotein to combat SARS-CoV-2: An integrative phytochemical and computational approach. J. Food Biochem., (2022): e14062.
2010. Siheri, W. et al. Chemical and antimicrobial profiling of propolis from different regions within Libya. PLoS One 11.5 (2016).
2011. Silva-Carvalho, R., Baltazar, F., Almeida-Aguiar, C. Propolis: a complex natural

- product with a plethora of biological activities that can be explored for drug development. Evidence-based Compl. Altern. Med., 2015, Art. n. 206439.
2012. Silva, C.C.F.D., Salatino, A., Motta, L.B.D. et al.. (2019). Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from Ceará state. Revista Brasileira de Farmacognosia, 29, 309-318.
 2013. Soltani, E.K. (2018). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation).
 2014. Sönmez E. Investigation of chemical content and antimicrobial activities of different plant sources of Anatolian propolis samples. Uludağ Arıcılık Dergisi. 2023, 5;23(1):37-48
 2015. Sorucu, A. (2015). Marmara bölgesindeki propolislerde biyolojik etkisi olan fenolik madde ve miktarlarının mevsim ve rakım farkına bağlı olarak belirlenmesi (Doctoral dissertation, Bursa Uludag University (Turkey)).
 2016. Šturm, L, N.P. Ulrih. Advances in the propolis chemical composition between 2013 and 2018: A review. eFood 1.1 (2019):24-37.
 2017. Suleman, T, van Vuuren, S., Sandasi, M. et al. Antimicrobial activity and chemometric modelling of South African propolis. J. Appl. Microbiol., 2015, 119, 4, 981-990.
 2018. Suleman, T. The antimicrobial and chemical properties of South African propolis. Doctoral dissertation, 2016.
 2019. Syaifie, P.H. et al. Computational study of Asian propolis compounds as potential anti-type 2 diabetes mellitus agents by using inverse virtual screening with the DIA-DB web server, tanimoto similarity analysis, and molecular dynamic simulation. Molecules 27.13 (2022):3972.
 2020. Talla, E. et al. New mono-ether of glycerol and triterpenes with DPPH radical scavenging activity from Cameroonian propolis. Nat. Product Res., 31.12 (2017): 1379-1389.
 2021. Talom, Benjamin Tangué, et al. "Antibacterial Activities of some Medicinal Plants Used for Treatment of Infectious Diseases in the Vina and Mayo-Louti Divisions of Cameroon." European Journal of Medicinal Plants (2018): 1-13.
 2022. Tamfu, A. N., et al. "A new isoflavonol and other constituents from Cameroonian propolis and evaluation of their anti-inflammatory, antifungal and antioxidant potential " Journal of Natural Products and Resources. (2020)
 2023. Tatli Seven, P. et al. Nanotechnology and nano-propolis in animal production and health: an overview. Italian J. Animal Sci., 17.4 (2018):921-930.
 2024. Tran, T.D. et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020):4988., 1-35.
 2025. Wali, A.F. et al. Bee propolis (bee's glue): a phytochemistry review. J. Crit. Rev., 4.4, 2017.
 2026. Wali, A.F. et al. In vitro antioxidant and antimicrobial activities of propolis from Kashmir Himalaya region. Free Rad. Antioxid., 1 (2016):51-57.
 2027. Wang, X. et al. Determination of 14 lipophilic pesticide residues in raw propolis by selective sample preparation and gas chromatography–tandem mass spectrometry. Food Analyt. Methods 13.9 (2020):1726-1735.
 2028. Xu, X. et al. Chemical compositions of propolis from China and the United States and their antimicrobial activities against *Penicillium notatum*. Molecules 24.19 (2019):3576.
 2029. Xu, X. et al. The chemical composition of Brazilian green propolis and its protective effects on mouse aortic endothelial cells against inflammatory injury. Molecules 25.20 (2020):4612.

2030. Zabaïou, N. et al. Biological properties of propolis extracts: Something new from an ancient product. *Chem. Phys. Lipids*, 2017, 207, 214-222.
2031. Zagmutt, S. et al. Protective effect of propolis extract on pancreatic β Cell under oxidative stress in vitro. *J. Food Nutr. Res* 4 (2016): 400-407.
2032. Zeitoun, R. et al. Chemical composition, antioxidant and anti-inflammatory activity evaluation of the Lebanese propolis extract. *Curr. Pharm. Biotechnol.*, 20.1 (2019):84-96.
2033. Zuhendri, F. et al. The potential use of propolis as a primary or an adjunctive therapy in respiratory tract-related diseases and disorders: a systematic scoping review. *Biomed. & Pharmacother.*, 146 (2022):112595.
2034. Zulkiflee, N. et al. Antibacterial and antioxidant activities of ethanolic and water extracts of stingless bees *Tetrigona binghami*, *Heterotrigona itama*, and *Geniotrigona thoracica* propolis found in Brunei. *Philipp. J. Sci* 151 (2022):1455-1462.
- 87). Petrova, A., M. Popova, C. Kuzmanova, I. Tsvetkova, H. NAYDENSKI, E. Muli, V. Bankova. New biologically active compounds from Kenyan propolis, *Fitoterapia*, 2010, 81, 6, 509-514. ISSN: 0367-326X. IF 1.36**
2035. Abdou, H.S., Salah, S.H., Boolesand, H.F., et al. Effect of pomegranate pretreatment on genotoxicity and hepatotoxicity induced by carbon tetrachloride (CCl₄) in male rats. *J. Med. Plants Res.*, 6 (17), 2012, 3370-3380.
2036. Abu-Mellal, A., Koolaji, N., Duke, R.K., et al. Prenylated cinnamate and stilbenes from Kangaroo Island propolis and their antioxidant activity. *Phytochem.*, 77, 2012, 251-259.
2037. Afata T.N., Dekebo A. Chemical composition and antimicrobial effect of Western Ethiopian propolis. *Chem. Biodiv.*, 20(2), art. no. e202200922, 2023.
2038. Afata, Tariku Neme, and Aman Dekebo. "Chemical composition and antimicrobial effect of western Ethiopian propolis." *Chemistry & Biodiversity*
2039. Al Bratty, M. et al. Chemical characterization and in-vitro antimicrobial screening of ethanolic extract of propolis collected from Jazan, Saudi Arabia. *Pakistan J. Zool.*, 52.1 (2020):121-130.
2040. Alghamdi, A. S. (2020). Applications of multivariate statistics in honey bee research, analysis of metabolomics data from samples of honey bee propolis. PhD thesis, University of Strathclyde, Glasgow.
2041. Ali M.T., Blicharska N., Shilpi J.A. et al. (2018). Investigation of the anti-TB potential of selected propolis constituents using a molecular docking approach. *Sci. Rep.*, 8 (1), 12238.
2042. Aliyazicioglu, R., Kanbolat, S. (2018). Therapeutic effects of propolis. *Chem. Sci. Int. J.*, 24(2), 1-6.
2043. Almutairi, S., Eapen, B., Chundi, S.M., et al. New anti-trypanosomal active prenylated compounds from African propolis. *Phytochem. Lett.*, 2014, 10, 35-39.
2044. Almutairi, S., Edrada-Ebel, R., Fearnley, J. et al. Isolation of diterpenes and flavonoids from a new type of propolis from Saudi Arabia. *Phytochem. Lett.*, 2014, 10, 160-163.
2045. Almutairi, S. S. (2015). Anti-trypanosomal and metabolomic effects of propolis constituents. Doctoral dissertation, University of Strathclyde.
2046. Asfaram, S. et al. Promising anti-protozoan activities of propolis (bee glue) as natural product: a review. *Acta Parasitol.*, 2020, 66, 1-12.
2047. Avula, B. et al. Quantification and characterization of phenolic compounds from northern Indian propolis extracts and dietary supplements. *J. AOAC Int.* 103.5

- (2020):1378-1393.
2048. Bava, R., Castagna, F., Lupia, C. et al. (2024). Hive products: composition, pharmacological properties, and therapeutic applications. *Pharmaceuticals*, 17(5), 646.
 2049. Betoloum, S.M. et al. Phytochemical tests, assessment of antioxidant properties and isolation of two compounds of ethyl acetate extract of Chadian propolis: case of Bebotho (Southern Chad). *Am. J. Anal. Chem.*, 13.7 (2022):241-254.
 2050. Blicharska, N., Seidel, V. (2019). Chemical diversity and biological activity of African propolis. *Progress in the Chemistry of Organic Natural Products* 109, 415-450.
 2051. Bo, S. et al. Naturally occurring prenylated stilbenoids: food sources, biosynthesis, applications and health benefits. *Crit. Rev. Food Sci. Nutrit.*, 2023, 63(26), 8083-8106.
 2052. Bratty, M.A., Alhazmi, H.A., Reddy, D.N. et al. (2020). Chemical characterization and in-vitro antimicrobial screening of ethanolic extract of propolis collected from jazan, Saudi Arabia. *Pakistan J. Zool.*, 52(1).
 2053. Campos, V.A.C. et al. Antibacterial activity of propolis produced by *Frieseomelitta varia*. *Ciência e Agrotecnologia* 35.6 (2011):1043-1049.
 2054. Celik, T.A. Potential genotoxic and cytotoxic effects of plant extracts, a compendium of assays on alternative therapy. Dr. Arup Bhattacharya (Ed.), 2012, ISBN 978-953-307-863-2
 2055. Cooper, Rose. *Natural Products*. Russell, Hugo & Ayliffe's: Principles and Practice of Disinfection, Preservation and Sterilization (2013): 550-564.
 2056. Cornara, L.J. et al. Therapeutic properties of bioactive compounds from different honeybee products. *Front. Pharmacol.*, 8 (2017):412.
 2057. Cui, J. et al. Extraction, purification, structural character and biological properties of propolis flavonoids: A review. *Fitoterapia* (2021):105106.
 2058. Curifuta, M., Vidal, J., Sánchez-Venegas, J., et al. The in vitro antifungal evaluation of a commercial extract of Chilean propolis against six fungi of agricultural importance. *Cien. Inv. Agr.*, 2012, 39 (2), 347-359.
 2059. Davin, L.B. et al. Lignan chirality roadmap a scientific art in danger of being lost?. *The Lignan Handbook*. CRC Press, 2022. 191-933.
 2060. de Brito, D.Z., Cassemiro, N.S., de Souza, J.M. et al. (2021). Screening of 20 pantanal wetland plants for anti-Candida activity using HPLC-DAD-MS/MS and bioautography to characterize active compounds. *Planta Med. Int. Open*, 8(03), e96-e103.
 2061. De Groot, A.C. Propolis: A review of properties, applications, chemical composition, contact allergy, and other adverse effects, *Dermatitis*, 6, 24, 2013, 263-282.
 2062. El-Bassiony, T.A., Saad N.M., El-Zamkan M.A. Study on the antimicrobial activity of ethanol extract of propolis against enterotoxigenic methicillin-resistant staphylococcus aureus in lab prepared ice-cream. *Vet. World*, 2012, 5(3), 155-159.
 2063. El-Khamsa, M.S. (2017). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation, Université Ferhat Abbas–Setif).
 2064. El-Mossalami, H., and Y. A. Abdel-Hakeim. "Using of Propolis Extract as a Trial to Extend the Shelf-life and Improving the Quality Criteria of Fresh Egyptian Sausage." *Assiut Vet. Med. J* 59.139 (2013): 23-33.
 2065. Falcao, S.I.D.M. (2013). Chemical composition of Portuguese propolis bioactive properties (Doctoral dissertation, Universidade do Porto (Portugal)).
 2066. Fasolo, D. et al. Determination of benzophenones in lipophilic extract of Brazilian red propolis, nanotechnology-based product and porcine skin and mucosa: Analytical and

- bioanalytical assays. J. Pharm. Biomed. Anal., 124 (2016):57-66.
2067. Fraire-Reyes, I.A. et al. Use and effectiveness of propolis on chronic periodontitis: a systematic review. Odovtos-Int. J. Dental Sci., 24.1 (2022):32-43.
2068. Ha El-Mossalami, H.A.N.A.A., Abdel-Hakeim, Y.A. (2013). Using of propolis extract as a trial to extend the shelf-life and improving the quality criteria of fresh Egyptian sausage. Assiut Vet. Med. J., 59(139), 23-33.
2069. Hamilton, K.D. et al. Natural products isolated from *Tetragonula carbonaria cerumen* modulate free radical-scavenging and 5-lipoxygenase activities in vitro. BMC Compl. Altern. Med., 2017, 17, 1-8.
2070. Hamilton, K. D. Evaluation of the anti-inflammatory, anti-oxidant and wound-healing potential of cerumen from the Australian native stingless bee, *Tetragonula carbonaria* (Doctoral dissertation, University of the Sunshine Coast).
2071. Harmalkar D.S., Mali J.R., Sivaraman A. et al. Schweinfurthins A–Q: isolation, synthesis, and biochemical properties. RSC Advances, 8 (38), 21191-21209, 2018.
2072. Hossain, R. et al. Propolis: An update on its chemistry and pharmacological applications. Chinese Med., 17, 1, 2022:1-60.
2073. Huang, S. et al. Recent advances in the chemical composition of propolis. Molecules 19(12), 2014:19610-19632.
2074. Huang, Z.C., & Hu, F. Progress study the chemical composition of propolis (2008-2012). Nat. Prod. Res. Developm., 2013, 25(8), 1146-1153
2075. İlknur, U. Ç. A. K. "Influence of propolis extract on microbiological and sensory quality of rainbow trout fillets." Eurasian Journal of Food Science and Technology 2.2 (2019): 93-103.
2076. Jerz, G., Elnakady, Y.A., Braun, A., et al. Preparative mass-spectrometry profiling of bioactive metabolites in Saudi-Arabian propolis fractionated by high-speed countercurrent chromatography and off-line atmospheric pressure chemical ionization mass-spectrometry injection. Journal of Chromatography A, 2014, 1347, 17-29.
2077. Jerz, G. et al. Preparative mass-spectrometry profiling of bioactive metabolites in Saudi-Arabian propolis fractionated by high-speed countercurrent chromatography and off-line atmospheric pressure chemical ionization mass-spectrometry injection. J. Chromatogr. A 1347 (2014):17-29.
2078. Kardar, M.N., et al. Characterisation of triterpenes and new phenolic lipids in Cameroonian propolis. Phytochemistry 106 (2014):156-163.
2079. Kasiotis, K. M. Propolis non-volatile constituents: A Review. Hygeia J. Drugs Med., 2014, 6, 12, 111-121.
2080. Kenji, S., N. Takaaki. Alternative medicine safety: Agaricus blazei and Propolis. Comb. Chem. & High Throughput Screen., 2011, 14, 7, 616-621.
2081. King, D. I. (2017). Kangaroo island propolis: improved characterisation and assessment of chemistry and botanical origins through metabolomics (Doctoral dissertation).
2082. Kinghorn, A.D. et al., eds. Progress in the chemistry of organic natural products. Cham, Switzerland: Springer, 2017.
2083. Koloren, Z., Ertürk, Ö., Şekeroğlu, Z.A et al. 2023. Amoebicidal and cytotoxic activity of propolis collected from different regions in Turkey on *Acanthamoeba castellanii* trophozoites. Middle Black Sea J. Health Sci., 9(2), 312-324.

2084. Li, W. (2019). Chemical synthesis of anti-HIV compounds based on the aryl naphthalene lignans identified from justicia plants (Doctoral dissertation, Hong Kong Baptist University).
2085. Massaro, C.F., Katouli, M., Grkovic, T., et al. Anti-staphylococcal activity of C-methyl flavanones from propolis of Australian stingless bees (*Tetragonula carbonaria*) and fruit resins of *Corymbia torelliana* (Myrtaceae). *Fitoterapia*, 2014, 95, 247-257.
2086. Massaro, C. F. Antibacterial natural products of propolis and honeys from Australian stingless bees (*Tetragonula carbonaria*) (Doctoral dissertation, University of the Sunshine Coast).
2087. Matoso, L. M.L., M.B.L. Matoso. Extrato de própolis no combate ao COVID-19: um relato de experiência em nível da atenção básica em saúde. *Uniciencias* 24.1 (2020):94-103.
2088. Moosavi MR. Nematicidal effect of some herbal powders and their aqueous extracts against *meloidogyne javanica*. *Nematropica*, 2012, 42 (1), 48-56.
2089. Nedji, N., Loucif-Ayad, W. Antimicrobial activity of Algerian propolis in foodborne pathogens and its quantitative chemical composition. *Asian Pacific J. Trop. Dis.*, 2014, 4(6), 433-437.
2090. Nweze, N.E., et al. Effects of Nigerian red propolis in rats infected with *Trypanosoma brucei brucei*. *Comp. Clin. Pathol.*, 26.5 (2017):1129-1133.
2091. Nyandwi, R. et al. Determination and quantification of gallic acid in raw propolis by high-performance liquid chromatography–diode array detector in Burundi. *East Africa Sci.*, 1.1 (2019):43-48.
2092. Okhale, S.E., et al. Bee propolis: Production optimization and applications in Nigeria." *J. Pharmacogn. Phytother.*, 13.1 (2021):33-45.
2093. Oliveira, Samuel Catalino. "Relatório de Estágio realizado na Farmácia Santo António." (2017).
2094. Omar, R.M. et al. Chemical characterisation of Nigerian red propolis and its biological activity against *Trypanosoma brucei*. *Phytochem. Anal.*, 27.2 (2016):107-115.
2095. Oryan A., Alemzadeh E., Moshiri A. (2018). Potential role of propolis in wound healing: Biological properties and therapeutic activities. *Biomed. Pharmacother.*, 98, 469-483, 2018.
2096. Pant, K., Sharma, A., Chopra, H.K. et al. 2023. Impact of biodiversification on propolis composition, functionality, and application in foods as natural preservative: A review. *Food Control*, p.110097.
2097. Paul, S., Emmanuel, T., Matchawe, C., et al. Pentacyclic triterpenes and crude extracts with antimicrobial activity from Cameroonian brown propolis samples. *J. Appl. Pharm. Sci.*, 2014, 4, 7.
2098. Paula, V.M.B. Caracterização química e biológica do própolis da “serra de Bornes” por TLC. Master’s thesis, Instituto Politecnico de Braganca, 2012.
2099. Rastrelli, L. (2012). Tropical propolis: recent advances in chemical components and botanical origin: Osmany Cuesta-Rubio, Anna Lisa Piccinelli. *Medicinal Plants*, 235-266.
2100. Reyes, I.A.F., Fonseca, C.G., Argüelles, Ó.C. et al. (2022). Use and effectiveness of propolis on chronic periodontitis: a systematic review. *Odovtos: Int. J. Dental Sci.*, 160-171.
2101. Rivera-Yañez, N. et al. Biomedical properties of propolis on diverse chronic diseases and its potential applications and health benefits. *Nutrients*, 13.1 (2021):78.
2102. Robles-Zepeda, R.E. et al. Botanical origin and biological activity of propolis.

- Medicinal plants: Biodiversity and Drugs. 1st ed. New York: CRC Press, Taylor & Francis Group (2012):570-597.
2103. Rodríguez, Y., Sánchez-Catalán, F., Rojano, B., et al. Physicochemical characterization and evaluation of antioxidant activity of propolis collected in the Atlántic department, Colombia. *Revista UDCA Actualidad & Divulgación Científica*, 2012, 15(2), 303-311.
 2104. Sahoo, R., V. Sivaram, M. Anusha. Effect of solvents on phytochemical profiling by GC-MS analysis and in vitro cytotoxicity of bee propolis against leukemia and lung cancer cell lines. *J. Biol. Nature*, 2022, 14.1:16-26.
 2105. Salatino A.; C. Fernandes-Silva, A. R. Adne, et al. Propolis research and the chemistry of plant products. *Nat. Prod. Rep.*, 2011, 28, 5, 925-936.
 2106. Salatino, A. Perspectives for uses of propolis in therapy against infectious diseases. *Molecules* 27.14 (2022):4594.
 2107. Sami, B. et al. Isolation of isosativan from Nigerian red propolis. *Trop. J. Nat. Prod. Res.*, (2020): 4 (3):77-79.
 2108. Santos-Buelga, C., A.M. González-Paramás. Phenolic composition of propolis. *Bee Products-Chemical and Biological Properties*. Springer, Cham, 2017. 99-111.
 2109. Sari, L.M., et al. Acute and sub-chronic toxicity studies on the methanol leaf extract of *Leptadenia hastata* in Wistar rats *Trop. J. Nat. Prod. Res.*, (2019).
 2110. Schroeder, C.M., Dey, P.N., Beutler, J.A. et al. (2021). Synthesis of a coumarin-based analogue of Schweinfurthin F. *The J. Org. Chem.*, 86(23), 16824-16833.
 2111. Shahinozzaman, M, D.N. Obanda, S. Tawata. Chemical composition and pharmacological properties of Macaranga-type Pacific propolis: A review. *Phyther. Res.*, 2021, 35.1, 207-222.
 2112. Siheri, W., Ebiloma, G.U., Igoli, J.O. et al. (2019). Isolation of a novel flavanonol and an alkylresorcinol with highly potent anti-trypanosomal activity from Libyan propolis. *Molecules*, 24(6), 1041.
 2113. Silva, F.G.C. et al. Alimentos, nutracêuticos e plantas medicinais utilizados como prática complementar no enfrentamento dos sintomas do coronavírus (Covid-19): uma revisão *SciELO, Preprints*, 2020.
 2114. Silva, H. et al. The cardiovascular therapeutic potential of propolis—a comprehensive review. *Biology* 10.1 (2021):27.
 2115. Silva-Carvalho, R., Baltazar, F., Almeida-Aguiar, C. (2015). Propolis: a complex natural product with a plethora of biological activities that can be explored for drug development. *Evidence-Based Compl. Altern. Med.*, 2015, 206439, 1-29.
 2116. Soltani, E. K. (2018). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation).
 2117. Sorimachi, K., & Nakamoto, T. Alternative medicine safety: *Agaricus blazei* and propolis. *Combinatorial Chemistry & High Throughput Screening*, 2011, 14(7), 616-621.
 2118. Suleman, T., Vuuren, S., Sandasi, M. et al. (2015). Antimicrobial activity and chemometric modelling of South African propolis. *J. Appl. Microbiol.*, 119(4), 981-990.
 2119. Suleman, T. (2016). The antimicrobial and chemical properties of South African propolis (Doctoral dissertation).
 2120. Talla, E. et al. Phytochemical screening, antioxidant activity, total polyphenols and flavonoids content of different extracts of propolis from Tekel (Ngaoundal, Adamawa region, Cameroon). *The J. Phytopharm.*, 3.5 (2014):321-329.

2121. Tamas-Krumpe, O. et al. Evaluation of the therapeutic potential of some apicultural products with essential oils for cutaneous wounds in cats and dogs. (2019), Articles of "Scientific Papers" Iasi University of Life Sciences, Romania, 126-134 .
2122. Tamfu, A.N. et al. A new isoflavonol and other constituents from Cameroonian propolis and evaluation of their anti-inflammatory, antifungal and antioxidant potential. Saudi J. Biol. Sci., 27.6 (2020):1659-1666.
2123. Toreti V.C., Sato H.H., Pastore G.M. et al. Recent progress of propolis for its biological and chemical compositions and its botanical origin. Evidence-based Compl. Altern. Med., 2013, art. no. 697390.
2124. Tran, T.D., et al. Lessons from exploring chemical space and chemical diversity of propolis components. Int. J. Mol. Sci., 21.14 (2020):4988.
2125. Tsopmo, A., Awah, F. M., Kuete, V. (2013). Lignans and stilbenes from African medicinal plants. In Medicinal plant research in Africa (pp. 435-478). Elsevier.
2126. Tsuda, T., & Kumazawa, S. (2021). Propolis: chemical constituents, plant origin, and possible role in the prevention and treatment of obesity and diabetes. J. Agr. Food Chem., 69(51), 15484-15494.
2127. Tytkowski, B. et al. Concentration and fractionation of polyphenols by membrane operations. Curr. Pharm. Design 23.2 (2017):231-241.
2128. Uçak, İ. (2019). Influence of propolis extract on microbiological and sensory quality of rainbow trout fillets. Eurasian J. Food Sci. Technol., 2(2), 93-103.
2129. Ugariogu, S.N., Duru, I.A., Onwumere, F.C. et al. (2020). Physicochemical assessment and drug potential of some phenylpropanoid and flavonoid compounds of ethyl acetate eluate from Umudike propolis. Trop. J. Nat. Prod. Res (TJNPR), 4(12), 1208-1214.
2130. Vranješ, M., 2023. Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa (Doctoral dissertation, University of Zagreb. Faculty of Veterinary Medicine).
2131. Wali, Adil F., et al. Bee propolis (bee's glue): a phytochemistry review. J. Crit. Rev. 4.4 (2017).
2132. Wang, Q., Xue, X., Zhao, J. Application of mass spectrometry detection technology in honey traceability. J. Agric. Sci. Technol., 2013, 15(4), 42-47.
2133. Wang G., D. Jie, Z. Xiaoxiong, et al. (2011). Inhibitory effects of different volume fractions of propolis ethanol extract on maltase and sucrase in mouse intestine. Food Science, 32(19), 268-272.
2134. Yan S, Zhang H-C, Dong J. Analysis of key aroma-active components of propolis and poplar tree gum, Food Science, 2012,33 (04), 157-161.
2135. Yusop, S.A. et al. Cytotoxicity and antimicrobial activity of propolis from *Trigona itama* stingless bees against *Staphylococcus aureus* and *Escherichia coli*. Indonesian J. Pharmac. Sci. Technol., 1.1 (2018):13-20.
2136. Zabaoui, N. et al. Biological properties of propolis extracts: Something new from an ancient product. Chem. Phys. Lipids, 2017, 207, 214-222.
2137. Zainullin R.A., Kunakova R.V., Gareev V.F. et al. Flavanones and flavones from Bashkir propolis. Chem. Nat. Compd., 2018, 54(5), 975-977.
2138. Zhang , C., Hu, F. Study abroad of propolis: overview 2010. Bee Mag., 2011, 31(7), 5-8.
2139. Zhang , C., P.S. Huang, Hu, F. Study of the geographical origin of propolis, plant origin and chemical composition. Chinese Pharm. J., 2013, 48, 1889-1892.
2140. Zhang J-L., Wang K., Hu F-L. Advance in studies on antioxidant activity of propolis

- and its molecular mechanism. *Zhongguo Zhongyao Zazhi*, 2013, 16, 38, 2645-2652.
2141. Zhang, J., Chen, J., Liang, Z., & Zhao, C. New lignans and their biological activities. *Chem. & Biodiv.*, 2014, 11(1), 1-54.
2142. Zhang J., W. Kai, & H. Fuliang. (2013). Research progress on the antioxidant activity and molecular mechanism of propolis. *Chinese J. Tradit. Chinese Med.*, 38(16), 2645-2652.
2143. Zhang, J. et al. Antioxidant activities and molecular mechanisms of the ethanol extracts of *Baccharis propolis* and *Eucalyptus propolis* in RAW64. 7 cells. *Pharm. Biol.* 54.10 (2016):2220-2235.
2144. Zhang, T., Omar, R., Siheri, W. et al. Chromatographic analysis with different detectors in the chemical characterisation and dereplication of African propolis. *Talanta*, 2014, 120, 181-190.
2145. Белозерова, О.А. Синтез и биологическая активность природного лигнана севанола и его аналогов. Doctoral dissertation, Moscow, 2021.
- 88). NAJDENSKI, H., L . Gigova , I . Iliev , P . Pilarski , J . Lukavský, I . Tsvetkova , M . Ninova and V . Kusssovski . Antibacterial and antifungal activity of selected microalgae and cyanobacteria. *Int . J. Food Sci. Technol.*, 2013, 48, 7, 1533-1540.**
2146. Abd Ali, Ghaidaa H. Biofilm inhibitory potential of *westiellopsis prolifica* extract against some pathogenic microorganisms. *Eng. Technol. J.* 37.1 Part C (2019).
2147. Abd El-Hack, M.E., Abdelnour, S., Alagawany, M. et al. (2019). Microalgae in modern cancer therapy: Current knowledge. *Biomed & Pharmacotherapy*, 111, 42-50.
2148. Abdo, S. M., Youssef, M., El Nagar, I. et al. (2024). Processing and characterization of antimicrobial bioplastic films based on green microalgae *Scenedesmus obliquus* extract-loaded polyurethane. *International Journal of Biological Macromolecules*, 257, 128711.
2149. Abdella, B., Shokrak, N.M., Abozakra, N.A. et al. (2024). Aquaculture and *Aeromonas hydrophila*: a complex interplay of environmental factors and virulence. *Aquaculture International*, 1-11.
2150. Acién Fernández, F.G., C. Gómez-Serrano, J.M.Fernández-Sevilla. Recovery of nutrients from wastewaters using microalgae. *Front. Sustain. Food Systems* 2 (2018):59.
2151. Acosta, J. S., Valenzuela, C. C., & Leal, S. R. (2021). Fitodepuración mixotrófica en sistemas de recirculación acuícola (RAS) para el manejo sustentable de nutrientes contaminantes. *AquaTechnica: Revista Iberoamericana de Acuicultura*, 3(1), 37-54.
2152. Acurio L. P., Salazar D. M., Valencia A. F. et al. Antimicrobial potential of *Chlorella* algae isolated from stacked waters of the Andean Region of Ecuador. In *IOP Conf Ser Earth Environ Sci*, 151 (1), 012040, 2018
2153. Adikaram, Chamila Priyangan. "Overview of Non Tuberculosis Mycobacterial Lung Diseases." *Mycobacterium: Research and Development* 257 (2018).
2154. Afreen S., Fatma T. (2018). Extraction, purification and characterization of phycoerythrin from *Microchaete* and its biological activities. *Biocatal. Agric. Biotechnol.*, 13, 84-89, 2018.
2155. Afreen, Sumbul, and Tasneem Fatma. "Extraction, purification and characterization of phycoerythrin from *Microchaete* and its biological activities." *Biocatalysis and agricultural biotechnology* 13 (2018): 84-89.
2156. Ahmed, E.A. (2016). Antimicrobial activity of microalgal extracts isolated from

- Baharia Oasis, Egypt. Glob Adv Res J Microbiol, 5, 033-041.
2157. Akhoundian, Maryam, and Seyed Danial Mirhasannia. "Microalgal Biodiversity as a Biotechnology and Environmental Potential." Human & Environment 15.2 (2017): 39-70.
 2158. Albayati, M.A.F. (2020). Bazı makroalg türlerinin antimikrobiyal aktivitelerinin belirlenmesi.
 2159. Ali, Imane HAMOUDA, and Amel Doumandji. "Comparative phytochemical analysis and in vitro antimicrobial activities of the cyanobacterium *Spirulina platensis* and the green alga *Chlorella pyrenoidosa*: potential application of bioactive components as an alternative to infectious diseases." Bulletin de l'Institut Scientifique, Rabat, Section Sciences de la Vie 39 (2017): 41-49.
 2160. Ali, A.H., Moustafa, E.E., Abdelkader, S.A. et al. (2020). Antibacterial potential of macro and microalgae extracts against pathogens relevant to human health. Plant Archives, 20(2), 9629-9642.
 2161. Alsammarraie, O.F.A. (2020). Muscari macrocarpum sweet ve muscari racemosum mill. türlerinin genotip ve sitotiplerinin kıyaslanması.
 2162. Alsenani, F. et al. Evaluation of microalgae and cyanobacteria as potential sources of antimicrobial compounds. Saudi Pharm. J., 2020, 28, 12, 1834-1841.
 2163. Al Khawli, F. (2021). Ultrasound and supercritical fluids as useful tools to recover nutrients and bioactive compounds from aquaculture and marine side streams (Doctoral dissertation, Universitat de València).
 2164. Al-Nedawe¹, R. A. D., & Yusof², Z. N. B. Cyanobacteria As A Source Of Bioactive Compounds With Anticancer, Antibacterial, Antifungal, And Antiviral Activities: A Review.
 2165. Al-Tmimi S.L. Antibiofilm activity of intracellular extracts of *Westiellopsis prolifica* isolated from local environment in Baghdad. J. Global Pharm. Technol., 10 (3), 281-288, 2018.
 2166. Álvarez-Gómez, F. (2017). Producción de compuestos bioactivos a partir de biomasa algal basada en la biofiltración y la biorrefinería.
 2167. Amrei, H. D. (2021). The Effect of Different Light Conditions on Antimicrobial Activity of the Microalgae *Chlorella* sp. Ethanolic Extract Against *Streptococcus mutans*. Journal ISSN, 2766, 2276.
 2168. Androutsopoulou C., Makridis P. Antibacterial Activity against Four Fish Pathogenic Bacteria of Twelve Microalgae Species Isolated from Lagoons in Western Greece. Microorganisms, 11 (6), art. no. 1396, 2023. DOI: 10.3390/microorganisms11061396.
 2169. Arica, Şükran Çakır, Ayşe Ozyilmaz, and Sevil Demirci. "A study on the rich compounds and potential benefits of algae: A review." Pharm. Innov 6 (2017): 42-51.
 2170. Asan-Ozusaglam, Meltem, Yavuz Selim Cakmak, and Murat Kaya. "Bioactivity and antioxidant capacity of *Anabaenopsis* sp.(Cyanobacteria) extracts." J Algal Biomass Utln 2013.4 (2013): 50-58.
 2171. Ávila Velasco, C. I. (2021). Producción y recuperación de exopolisacáridos de *Neochloris oleoabundans* mediante fermentación extractiva con agentes termosensibles (Doctoral dissertation, Universidad Autónoma de Nuevo León).
 2172. Ayswaria R., Vijayan J., Nathan V.K. Antimicrobial peptides derived from microalgae for combating antibiotic resistance: Current status and prospects. Cell Biochemistry and Function, 41 (2), pp. 142 – 151, 2023. DOI: 10.1002/cbf.3779.
 2173. Bahrulolum, H. et al. Green synthesis of metal nanoparticles using microorganisms

- and their application in the agrifood sector. *J. Nanobiotechnol.*, 19.1 (2021):1-26.
2174. Bakku, Ranjith Kumar, and Randeep Rakwal. "Applications of cyanobacterial compounds in the energy, health, value-added product, and agricultural sectors: A perspective." *Cyanobacterial Physiology*. Academic Press, 2022. 149-164.
 2175. Bashir K.M.I., Lee J.H., Petermann M.J. et al. (2018). Estimation of antibacterial properties of Chlorophyta, Rhodophyta and Haptophyta Microalgae Spe. *Microbiol. Biotech. Lett.*, 46(3), 225-233, 2018.
 2176. Bastia, A.K. (2022). In vitro antioxidant and antibacterial activity of *Scenedesmus obliquus* collected from Similipal biosphere reserve, Odisha, India. *J. Indian Bot. Soc.*, 102(3), 218-228.
 2177. Bastidas-Oyanedel, J. R., Bonk, F., Thomsen, M. H., Schmidt, J. E. Dark fermentation biorefinery in the present and future (bio) chemical industry. *Reviews in Environmental Science and Bio/Technology*, 14, 2015, 3, 473-498.
 2178. Bastidas-Oyanedel, J. R., Bonk, F., Thomsen, M. H., & Schmidt, J. E. (2019). The future perspectives of dark fermentation: moving from only biohydrogen to biochemicals. *Biorefinery: Integrated Sustainable Processes for Biomass Conversion to Biomaterials, Biofuels, and Fertilizers*, 375-412.
 2179. Belyagoubi, L., Chaibi, R., Gouzi, H. et al. (2023). Phycochemistry study and antimicrobial activity of *Spirogyra* freshwater green microalgae from Algeria. *Journal of Natural Product Research and Applications*, 3(02), 47-60.
 2180. Bernal, I.N.V.O. (2002). TESIS.
 2181. Bhatnagar, M., & Bhatnagar, A. (2019). Diversity of polysaccharides in cyanobacteria. *Microbial Diversity in Ecosystem Sustainability and Biotechnological Applications: Volume 1. Microbial Diversity in Normal & Extreme Environments*, 447-496.
 2182. Bhowmick, Sukanya, et al. "Algal metabolites: An inevitable substitute for antibiotics." *Biotechnology Advances* (2020): 43, 107571.
 2183. Biondi N., Martina M.R., Centini M. et al. Hot springs cyanobacteria endowed with biological activities for cosmetic applications: evaluation of on-site collected communities and isolated strains. *Cosmetics*, 10 (3), art. no. 81. 2023.
 2184. Biru, Talago, and Koto Baru. "www. scholarsresearchlibrary. com t Available online."
 2185. Bishoyi, Ajit Kumar, Chita Ranjan Sahoo, and Rabindra Nath Padhy. "Recent progression of cyanobacteria and their pharmaceutical utility: an update." *Journal of Biomolecular Structure and Dynamics* (2022): 1-34
 2186. Bishoyi, A.K., Lakra, A., Mandhata, C.P. et al. (2024). Prospective Phycocompounds for Developing Therapeutics for Urinary Tract Infection. *Current Microbiology*, 81(1), 35.
 2187. Biswas D., M.H. Siddiqui, S. Mahfooz, A. Shamim, A. Farooqui. Partial purification of bioactive compounds from different cyanobacterial strains and its biological potential. *IJBPAS*, 4, 2015, 10, 6107-6115.
 2188. Bogdanovic, Sanja. Establishment of methods for bioprospecting of marine algae for antimicrobial agents. MS thesis. 2018.
 2189. Bulimaga, Valentina, et al. "Metode de separare a metaboliților secundari bioactivi din biomasa unor cianobacterii și proprietățile lor curative și toxicologice." *Studia Universitatis (Seria Științe Reale și ale Naturii)* 71.1 (2014): 57-66.
 2190. Carolina, L. T. (2016). Efecte antimicrobiene ale unor substanțe chimice din produse autohtone.

2191. Chaïb, Slimane, et al. "Allelopathy and allelochemicals from microalgae: An innovative source for bio-herbicidal compounds and biocontrol research." *Algal Research* 54 (2021): 102213.
2192. Chaidir, Z., Hillman PF Syafrizayanti, and R. Zainul. "Isolation and identification of freshwater microalgae potentially as antibacterial from Talago Biru." *Koto Baru, West Sumatera Der Pharmacia Lettre* 8 (2016): 157-165.
2193. Chen, Wei Ning, et al. "Effect of Supercritical Carbon Dioxide Extraction Parameters on the Biological Activities and Metabolites Present in Extracts from *Arthrospira platensis*." (2017).
2194. Chen, S., Xie, J., & Wen, Z. (2021). Microalgae-based wastewater treatment and utilization of microalgae biomass. *Advances in Bioenergy*, 6(1), 165-198.
2195. da Rocha Neto, Argus Cezar, et al. "Atividade antimicrobiana de extratos etanólicos de algas no controle de *Penicillium expansum* Link (Trichocomaceae, Ascomycota)." *Biotemas* 28.4 (2015): 23-33.
2196. Dahech, P., Schlömann, M., & Ortiz, C. (2021). Light intensity stimulates the production of extracellular polymeric substances (EPS) in a culture of the desert cyanobacterium *Trichormus* sp. *Journal of Applied Phycology*, 33, 2795-2804.
2197. Das, S. Genetic regulation, biosynthesis and applications of extracellular polysaccharides of the biofilm matrix of bacteria. *Carbohydrate Polymers* (2022):119536
2198. Davoodbasha M., Edachery B., Nooruddin T., Lee S. Y., Kim J. W. An evidence of C16 fatty acid methyl esters extracted from microalga for effective antimicrobial and antioxidant property. *Microb. Pathog.*, 115, 233-238, 2018.
2199. de Lima Barizão A.C., de Oliveira Gomes L.E., Brandão L.L., Sampaio I.C.F., de Moura I.V.L., Gonçalves R.F., de Oliveira J.P., Cassini S.T. Microalgae as tertiary wastewater treatment: Energy production, carbon neutrality, and high-value products. *Algal Research*, 72, art. no. 103113, 2023. DOI: 10.1016/j.algal.2023.103113.
2200. de Moraes M.G., Muniz Bezerra P.Q., Deamici K.M. et al. Algal Bioreactors for Polysaccharides Production. *Plants as Bioreactors: An Overview*, pp. 485 – 501. 2023. DOI: 10.1002/9781119875116.ch20.
2201. Dehmani, S., Djamal Zerrouki. Culture des microorganismes dans les eaux usées. Doctoral dissertation, 2018.
2202. Demirel Z., Yilmaz F.F., Ozdemir G. et al. Influence of media and temperature on the growth and the biological activities of *Desmodesmus protuberans* (FE Fritsch & MF Rich) E. Hegewald. *Turk. J. Fish. Aquat. Sci.*, 18 (10), 1195-1203, 2018.
2203. do Amaral S.C., Xavier L.P., Vasconcelos V. et al. Cyanobacteria: a promising source of antifungal metabolites. *Marine Drugs*, 21 (6), art. no. 359, 2023, 359.
2204. Dibaei, F., Fazilati, M., Moenzadeh, F. et al. (2018). Anti-angiogenesis effect of C-phycocyanin of *Spirulina platensis* on B16-F10 melanoma tumors in C57BL/6 mouse. *Pathobiology Research*, 21(3), 141-146.
2205. Dinh, N. T., Dat, N. T., Toan, N. K., & Le Anh, P. (2022). The roles of microalgae and bacteria in wastewater treatment. *Vietnam Journal of Biotechnology*, 20(3), 573-588.
2206. Dineshkumar, R., Narendran, R., Jayasingam, P. et al. (2017). Cultivation and chemical composition of microalgae *Chlorella vulgaris* and its antibacterial activity against human pathogens. *J Aquac Mar Biol*, 5(3), 00119.
2207. Effendi, S.S.W. et al. Development and fabrication of disease resistance protein in

- recombinant *Escherichia coli*. *Bioresources and Bioprocessing* 7.1 (2020):1-10.
2208. El Semary, Nermin, Haifa Al Naim, and Munirah F. Aldayel. "A Novel Application of Laser in Biocontrol of Plant Pathogenic Bacteria." *Applied Sciences* 12.10 (2022): 4933.
2209. El-Hack, Mohamed E. Abd, et al. "Microalgae in modern cancer therapy: Current knowledge." *Biomedicine & Pharmacotherapy* 111 (2019): 42-50.
2210. Elif, Ç. İ. L. (2023). A Popular Dietary Supplement: Chlorella. *Versatile Approaches to Engineering and Applied Sciences: Materials and Methods*, 37.
2211. Elkomy, R., et al. "Optimal conditions for antimicrobial activity production from two microalgae chlorella marina and Navicula F. Delicatula." *Journal of Pure and Applied Microbiology* 9.4 (2015): 2725-2733.
2212. ELWE, S.A.F. (2021). *Prospecção tecnológica de patentes sobre compostos bioativos de microalgas*.
2213. Emparan, Q., Harun, R., & Sing Jye, Y. (2021). Efficiency of pollutants removal in treated palm oil mill effluent (TPOME) using different concentrations of sodium alginate-immobilized Nannochloropsis sp. cells. *Int. J. Phytoremed.*, 23(5), 454-461.
2214. Esquivel-Hernández, Diego A., et al. "Effect of Supercritical Carbon Dioxide Extraction Parameters on the Biological Activities and Metabolites Present in Extracts from *Arthrospira platensis*." *Marine Drugs* 15.6 (2017): 174.
2215. Evans, Laurence J. *Water neutral developments: How to successfully integrate microalgae systems into wastewater management*. Diss. Heriot-Watt University, 2018.
2216. Fadjria, N., Arfiandi, A., Nofita, D., & Fadhila, M. (2023). Screening Phytochemicals and Antibacterial Activity of Microalgae Strain *Scenedesmus* sp. Auma-020. *Journal of Pharmaceutical and Sciences*, 46-51.
2217. Falaise C., C. François, M-A. Travers et al. Antimicrobial compounds from eukaryotic microalgae against human pathogens and diseases in aquaculture. *Mar. Drugs* 2016, 14, 159
2218. Feng, Yanzhang, et al. "Microalgae as a potential conditioner for continuous cropping obstacles for taro (*Colocasia esculenta* L. Schott) production." *Journal of Cleaner Production* 369 (2022): 133356
2219. Fernández, F.G.A. et al. The role of microalgae in the bioeconomy. *New Biotechnol.*, 61 (2021):99-107.
2220. Fleurence, Joël. *Les microalgues: De l'aliment du futur à l'usine cellulaire*. ISTE Group, 2021.
2221. Forján, E., Navarro, F., Cuaresma, M. et al. (2015). Microalgae: fast-growth sustainable green factories. *Critical Reviews in Environmental Science and Technology*, 45(16), 1705-1755.
2222. Fu Tingting, Ma Ning, Meng Jianzong, & Zhang Yunkai. (2015). Optimization of extraction technology of intracellular polysaccharides from a marine green algae based on response surface methodology. *Chinese Brewing*, 34(9), 115-120.
2223. Furmaniak, Magda A., et al. "Edible cyanobacterial genus *Arthrospira*: Actual state of the art in cultivation methods, genetics, and application in medicine." *Frontiers in microbiology* 8 (2017): 2541.
2224. Gaignard, Clément, et al. "Screening of marine microalgae: Investigation of new exopolysaccharide producers." *Algal Research* 44 (2019): 101711.
2225. García, J. C. A. (2021). Efecto inhibitorio de dos especies de diatomeas contra *Vibrio harveyi*, *V. alginolyticus* y *V. campbellii*.

2226. Gautam, Shristry, and M. Amin-ul Mannan. "The Role of Algae in Nutraceutical and Pharmaceutical Production." *Bioactive Natural products in Drug Discovery*. Springer, Singapore, 2020. 665-685.
2227. Gavalás-Olea, A., Siol, A., Sakka, Y. et al. (2021). Potential of the Red Alga *Dixoniella grisea* for the Production of Additives for Lubricants. *Plants*, 10(9), 1836.
2228. Ghaidaa, H. A., et al. "The Biofilm Inhibitory Potential of Compound Produced from *Chlamydomonas reinhardtii* Against Pathogenic Microorganisms." *Baghdad Science Journal* 17.1 (2020): 34-41.
2229. Gheda, Saly F., and Gehan A. Ismail. "Natural products from some soil cyanobacterial extracts with potent antimicrobial, antioxidant and cytotoxic activities." *Anais da Academia Brasileira de Ciências* 92.2 (2020), e20190934, 1-18.
2230. Gomes, L., Cotas, J., Fernandes, C. et al. (2024). Seaweed *Calliblepharis jubata* and *Fucus vesiculosus* Pigments: Anti-Dermatophytic Activity. *Applied Sciences*, 14(4), 1456.
2231. Gonçalves A.L., J.C. Pires, M. Simões. A review on the use of microalgal consortia for wastewater treatment, *Algal Res.*, 2017, 24, 403-415.
2232. Gonçalves, A.L., Santos, F.M., & Pires, J.C. (2019). Microalgal consortia: From wastewater treatment to bioenergy production. *Grand Challenges in Algae Biotechnology*, 371-398.
2233. González, A.K.V. (2023). Bioprospección de extractos microalgales con potencial aplicación biomédica.
2234. Guo, Taohong, et al. "Using OMICS technologies to analyze the mechanisms of synthetic microbial co-culture systems: a review." *Sheng wu Gong Cheng xue bao = Chinese Journal of Biotechnology* 38.2 (2022): 460-477
2235. Hamad G.M., Abd El-Baky N., Sharaf M.M., Amara A.A. Volatile Compounds, Fatty Acids Constituents, and Antimicrobial Activity of Cultured *Spirulina* (*Arthrospira fusiformis*) Isolated from Lake Mariout in Egypt. *Scientific World Journal*, 2023, art. no. 9919814, 2023. DOI: 10.1155/2023/9919814.
2236. Hashim M.S. Identification and characterization of bioactive compounds in two algae species of *Haematococcus pulvialis* and *Dunatiella* saline from the waterbodies of Basrah region, Iraq. *Iranian Journal of Ichthyology*, 10 (Special Issue 1), pp. 77 – 84. 2023.
2237. Hassan, Saqib, et al. "Identification and characterization of the novel bioactive compounds from microalgae and cyanobacteria for pharmaceutical and nutraceutical applications." *Journal of Basic Microbiology* (2022)
2238. Hassi, Mohammed, et al. "A Review of Moroccan Microalgae and Their Exploitation Fields."
2239. Heikmat, N.Z., Abd Ali, G.H., Al-Razaq, S.L. et al. (2019). Biofilm inhibitory potential of *Westiellopsis prolifica* extract against some pathogenic microorganisms. *Engineering and Technology Journal*, 37(1C), 101-108.
2240. Hidehiko, K. (2022). Effect of temperature on growth and gene expression of *Tetrademus* sp. for application in wastewater treatment (Master's thesis, Nord universitet).
2241. Hinterholz, Camila Larissa, et al. "Computational fluid dynamics applied for the improvement of a flat-plate photobioreactor towards high-density microalgae cultures." *Biochemical Engineering Journal* 151 (2019): 107257.
2242. Hisham, Nur Eliza Badrul, and Nor Hanuni Ramli. "TROPICAL AGRICULTURAL

- SCIENCE." *Pertanika J. Trop. Agric. Sci* 44.1 (2020): 221-236.
2243. Horváth, Á.N., Németh, L., Vörös, L. et al. (2022). Microalgae strains of the Mosonmagyaróvár Algal Culture Collection with activity against plant fungal pathogens.
2244. Horváth Á.N., Németh L., Vörös L., Stirk W.A., van Staden J., Ördög V. Cataloguing microalgae and Cyanobacteria strains from the Mosonmagyaróvár Algal Culture Collection with in vitro antagonistic activity against phytopathogenic fungi and oomycetes. *Phytoparasitica* 2023. DOI: 10.1007/s12600-023-01045-2.
2245. Ibarra, A.C. (2023). Efecto inhibitorio de diatomeas contra bacterias patógenas del género *Vibrio* en acuicultura.
2246. Ikram, Sana F., et al. "Prospects and constraints in studying the biodiversity of agriculturally important microalgae and cyanobacteria and useful statistical tools." *Biodiversity and Conservation* (2022): 1-30.
2247. Immanuela, J. Pengaruh Jenis Pelarut Dan Lama Waktu Maserasi Terhadap.
2248. Imran Bashir, Khawaja Muhammad, et al. "Estimation of Antibacterial Properties of Chlorophyta, Rhodophyta and Haptophyta Microalgae Species." *Microbiology and Biotechnology Letters* 46.3 (2018): 225-233.
2249. Ishaq, A. G., Matias-Peralta, H. M., Basri, H., & Muhammad, M. N. (2015). Antibacterial Activity of Freshwater Microalga *Scenedesmus* sp. on Foodborne Pathogens *Staphylococcus aureus* and *Salmonella* sp. *Journal of Science and Technology*, 7(2).
2250. Ishaq, A. G., H. M. Matias-Peralta, and H. Basri. Bioactive Compounds from Green Microalga – *Scenedesmus* and its Potential Applications: A Brief Review "Tropical Agricultural Science." *Pertanika J. Trop. Agric. Sci* 39.1 (2016): 1-16.
2251. Jain, A. "Algae-mediated synthesis of biogenic nanoparticles." *Advances in Natural Sciences: Nanoscience and Nanotechnology* 13.4 (2022): 043001.
2252. Jawaji A., Zilberg D., Khozin-Goldberg I. Using byproduct of fucoxanthin production by *Phaeodactylum tricornutum* for the development of a treatment against monogenean infection in fish. *Journal of Applied Phycology*. 2023. DOI: 10.1007/s10811-023-02990-5.
2253. Jena, Jayanti, and Enketeswara Subudhi. "Microalgae: An untapped resource for natural antimicrobials." *The role of microalgae in wastewater treatment*. Springer, Singapore, 2019. 99-114.
2254. Jha, Durga, et al. Microalgae-based Pharmaceuticals and Nutraceuticals: An Emerging Field with Immense Market Potential. *ChemBioEng Rev.*, 4.4 (2017): 257-272.
2255. Kadimpati, Kishore Kumar, et al. "Microalgae Potential Feedstock for the Production of Biohydrogen and Bioactive Compounds." *Microbial Strategies for Techno-economic Biofuel Production*. Springer, Singapore, 2020. 171-206.
2256. Kalwani, Mohneesh, et al. "Microalgae-mediated wastewater treatment and enrichment of wastewater-cultivated biomass for biofuel production." *Expanding Horizon of Cyanobacterial Biology*. Academic Press, 2022. 259-281
2257. Kanaan, Chirine Issam. Assessing methods for bioprospecting of marine algae for antimicrobial agents. MS thesis. 2019.
2258. Kang, Y. et al. Potential of algae–bacteria synergistic effects on vegetable production. *Front. Plant Sci.*,12, 2021, 656662.
2259. Kar, Joyeeta, et al. "Revisiting the role of cyanobacteria-derived metabolites as antimicrobial agent: A 21st century perspective." *Frontiers in Microbiology* 13 (2022).

2260. Karmakar, A., Ghosh, S., Islam, R., & Rajak, P. (2022). Natural Antimicrobial as an Alternative Food Preservatives in the Dietary Systems of Human and Livestock.
2261. Kaur M., Bhatia S., Gupta U., Decker E., Tak Y., Bali M., Gupta V.K., Dar R.A., Bala S. Microalgal bioactive metabolites as promising implements in nutraceuticals and pharmaceuticals: inspiring therapy for health benefits. *Phytochemistry Reviews* 2023. DOI: 10.1007/s11101-022-09848-7.
2262. Kiran, B.R., S. Venkata Mohan. Microalgal cell biofactory - therapeutic, nutraceutical and functional food applications. *Plants* 10.5 (2021):836.
2263. Koutra, Eleni, et al. "Assessing the potential of *Chlorella vulgaris* for valorization of liquid digestates from agro-industrial and municipal organic wastes in a biorefinery approach." *Journal of Cleaner Production* 280 (2021): 124352.
2264. Koutra, Eleni, et al. "Microalgal Biorefinery." *Microalgae Cultivation for Biofuels Production*. Academic Press, 2020. 163-185.
2265. Kovač, Dajana. "Biotehnoški potencijal filamentoznih sojeva cijanobakterija sa područja Vojvodine." (2017).
2266. Krohn, Ines, et al. "Health benefits of microalgae and their microbiomes." *Microbial Biotechnology* (2022).
2267. Lai, Yu-Cheng, et al. "Towards protein production and application by using *Chlorella* species as circular economy." *Bioresource technology* (2019): 121625.
2268. Lage, V.M.G.B., Deegan, K.R., Santos, G.F. et al. (2022). Atividade biológica das microalgas em dermatófitos: Revisão. *Research, Society and Development*, 11(11), e126111133404-e126111133404.
2269. Lakshmidevi, Rajendran, Nagarajan Nagendra Gandhi, and Karuppan Muthukumar. "Bioelectricity and bioactive compound production in an algal-assisted microbial fuel cell with immobilized bioanode." *Biomass Conversion and Biorefinery* (2020): 1-17.
2270. Lakshmidevi, Rajendran, Nagarajan Nagendra Gandhi, and Karuppan Muthukumar. "Bioelectricity and bioactive compound production in an algal-assisted microbial fuel cell with immobilized bioanode." *Biomass Conversion and Biorefinery* (2020): 1-17.
2271. Lamprinou, V., Tryfinopoulou, K., Velonakis, E.N. et al. Cave Cyanobacteria showing antibacterial activity. *International Journal of Speleology*, 44, 2015, 231-238.
2272. Laroche, C. Exopolysaccharides from microalgae and cyanobacteria: diversity of strains, production strategies, and applications. *Marine Drugs* 20.5 (2022):336.
2273. Lekshmi, S., et al. "Antibacterial activity of *Chroococcus minutus* (Kützinger) Nägeli isolated from Cochin estuary against selected pathogens." *Int. J. Fish. Aquat. Stud* 4.3 (2016): 700-703.
2274. Li A., L. Zhang, Z. Zhao, S. Ma, M. Wang, P. Liu. Prescreening, identification and harvesting of microalgae with antibacterial activity. *Biologia* 71/10: 1—, 2016.
2275. Li, X. (2018). Development of robust cyanobacterial strains for biotechnological applications: Stress tolerance and cell-specific expression in heterocyst-forming cyanobacteria (Doctoral dissertation, Acta Universitatis Upsaliensis).
2276. Little, Shannon M., et al. "Antibacterial compounds in green microalgae from extreme environments: a review." *Algae* 36.1 (2021): 61-72.
2277. Liu, Wei, and Roger Ruan. "Microalgae-based biomaterials for environmental remediation and functional use." *Algae-Based Biomaterials for Sustainable Development*. Elsevier, 2022. 277-290.

2278. Lortou, U., & Gkelis, S. (2023). Antibacterial activity, pigments, and biomass content of microalgae isolated from Greece. *Journal of Biological Research-Thessaloniki*, 30.
2279. Lu, Y.-Z., et al. "Research progress of pharmacological active substances from diazotrophic cyanobacteria", *Chinese Traditional and Herbal Drugs* Volume 49.18, (2018): 4453-4460.
2280. Mahesh, R., Panda, S.K., Das, M., Yashavanth, P.R. et al. (2021). Advances in biotechnological tools for bioremediation of wastewater using bacterial–algal symbiotic system. In *Wastewater Treatment* (pp. 385-411). Elsevier.
2281. Mehmood, S., Huo, S., Kubar, A.A., Kumar, S. et al. (2023). *Bioscience Research*.
2282. Mamun M.A., Hossain M.A., Saha J., Khan S., Akter T., Banu M.R. Effects of spirulina *Spirulina platensis* meal as a feed additive on growth performance and immunological response of Gangetic mystus *Mystus cavasius*. *Aquaculture Reports*, 30, art. no. 101553, 2023. DOI: 10.1016/j.aqrep.2023.101553.
2283. Man, Yu Bon, et al. "Growth and intestinal microbiota of Sabah giant grouper reared on food waste-based pellets supplemented with spirulina as a growth promoter and alternative protein source." *Aquaculture Reports* 18 (2020): 100553.
2284. Manoharan, D., Natesan, S., Billamboz, M., & Jawhara, S. (2024). Role of Bacteria-Derived Exopolysaccharides in Inflammatory Bowel Disease with a Special Focus on Cyanobacterial Exopolysaccharides. *Applied Microbiology*, 4(1), 250-274.
2285. Manzaneda Choque, F.P. (2022). Aprovechamiento de nutrientes en aguas residuales de curtiembres como medio de crecimiento para la obtención de biomasa de microalgas.
2286. Martínez-Francés, E., & Escudero-Oñate, C. (2018). Cyanobacteria and microalgae in the production of valuable bioactive compounds. *Microalgal Biotechnol*, 6, 104-128.
2287. Marrez, Diaa A., et al. "Antimicrobial and anticancer activities of *Scenedesmus obliquus* metabolites." *Heliyon* 5.3 (2019): e01404.
2288. Maruthanayagam, V., et al. "Effects of surface material on growth pattern and bioactive exopolymers production of intertidal cyanobacteria *Phormidium* sp." (2020), *Indian Journal of Geo-Marine Sciences* 49(10), pp. 1669-1677.
2289. Mashhadinejad, Ahmad, Hojjatolah Zamani, and Jannat Sarmad. "Effect of growth conditions and extraction solvents on enhancement of antimicrobial activity of the microalgae *Chlorella vulgaris*." *Pharmaceutical and Biomedical Research* 2.4 (2016):65-73.
2290. Matos, Â. P., Saldanha-Corrêa, F.M.P., da Silva Gomes, R., & Hurtado, G. R. (2023). Exploring microalgal and cyanobacterial metabolites with antiprotozoal activity against *Leishmania* and *Trypanosoma* parasites. *Acta Tropica*, 107116.
2291. Matulich, Patrick. "Screening of Crude Microalgal Extracts for Antimicrobial Activity." (2019).
2292. Mohsenpour, S.F. et al. Integrating micro-algae into wastewater treatment: A review. *Science of The Total Environment* (2020):142168.
2293. Montalvão, S., Z. Demirel, P. Devi et al. Large-scale bioprospecting of cyanobacteria, micro-and macroalgae from the Aegean Sea. *New biotechnology*, 33, (2016), (3), 399-406.
2294. Morales-Jiménez, Mónica, et al. "Production, preparation and characterization of microalgae-based biopolymer as a potential bioactive film." *Coatings* 10.2 (2020): 120.
2295. Moreira, J. B., Kuntzler, S. G., Bezerra, P. Q. M. et al. (2022). Recent advances of microalgae exopolysaccharides for application as biofloculants. *Polysaccharides*, 3(1),

2296. Morsi, H.H., El-Sabbagh, S.M., Mehesen, A.A., & Mohamed, A. D. (2022). Bioactivity of natural compounds extracted from *Scenedesmus obliquus* toward some pathogenic bacteria. *International Journal of Alternative Fuels and Energy*, 6(1), 1-11.
2297. Mukherjee, C., Suryawanshi, P.G., Kalita, M.C. et al. (2024). Polarity-wise successive solvent extraction of *Scenedesmus obliquus* biomass and characterization of the crude extracts for broad-spectrum antibacterial activity. *Biomass Conversion and Biorefinery*, 14(2), 2467-2483.
2298. Narayanan, Sharmila, and Santhi Raju Pilli. "10 Algal-derived pharmaceuticals." *Algal Biorefinery: Developments, Challenges and Opportunities* (2021): 231.
2299. Navarro, Francisco, et al. "Antimicrobial activity of the acidophilic eukaryotic microalga *Coccomyxa onubensis*." *Phycological Research* 65.1 (2017): 38-43.
2300. Navvabi, A., Homaei, A., Navvabi, N. et al. (2022). Exopolysaccharides from Marine Microalgae. In *Marine Biochemistry* (pp. 325-337). CRC Press.
2301. Neto, N. O., Mota, R., Carvalho, J., Porto, A., Marques, D. V., & Bezerra, R. (2015). Extraction of antibacterial substances from *arthrospiraplatensis* biomass against antibiotic-resistant *staphylococcus* sp. Isolated from bovine mastitis. *Blucher chemical engineering proceedings*, 1(2), 881-887.
2302. Nowruzi, Bahareh, Hossein Fahimi, and Adriana Sturion Lorenzi. "Recovery of pure C-phycoerythrin from a limestone drought tolerant cyanobacterium *Nostoc* sp. and evaluation of its biological activity." (2020).
2303. Nuhu, Abdulmumin A. "Spirulina (*Arthrospira*): An important source of nutritional and medicinal compounds." *Journal of Marine biology* 2013 (2013).
2304. Ognistaia, A. V., Zh V. Markina, and T. Yu Orlova. "Antimicrobial Activity of Marine Microalgae." *Russian Journal of Marine Biology* 48.4 (2022): 217-230.
2305. Ördög, V. (2015). *Mikroalgák biotechnológiai alkalmazása a növénytermesztésben és növényvédelemben* (Doctoral dissertation, Nyugat-magyarországi Egyetem).
2306. Orejuela-Escobar, L., Gualle, A., Ochoa-Herrera, V. et al. (2021). Prospects of microalgae for biomaterial production and environmental applications at biorefineries. *Sustainability*, 13(6), 3063.
2307. Oviedo, Jineth Arango, et al. "A half-century of research on microalgae-bacteria for wastewater treatment." *Algal Research* (2022): 102828.
2308. Oyeronke A.O., Mojisola A.-A.A., Funmilola A.S., Bunmi A.J. Natural Compounds of Algae Origin with Potential Anticarcinogenic Benefits. *Next-Generation Algae: Volume II: Applications in Medicine and the Pharmaceutical Industry*, pp. 153 – 175, 2023. DOI: 10.1002/9781119857860.ch7.
2309. ÖZOĞUL, İlyas, et al. "Inhibitory impacts of *Spirulina platensis* and *Chlorella vulgaris* extracts on biogenic amine accumulation in sardine fillets." *Food Bioscience* (2021): 101087.
2310. Parwani, Laxmi, Medha Bhatt, and Jaspreet Singh. "Potential Biotechnological Applications of Cyanobacterial Exopolysaccharides." *Brazilian Archives of Biology and Technology* 64 (2021).
2311. Pašukonytė, N. (2022). Melsvabakterių (cyanobacteria) biomasės baltymų kiekinė-kokybinė analizė ir priešmikrobinio poveikio tyrimas in vitro.
2312. Peng, J., Cao, K. L., Lv, S. B. et al. (2023). Algal strains, treatment systems and

- removal mechanisms for treating antibiotic wastewater by microalgae. *Journal of Water Process Engineering*, 56, 104266.
2313. Pereira, Sara B., et al. "Strategies to Obtain Designer Polymers Based on Cyanobacterial Extracellular Polymeric Substances (EPS)." *International journal of molecular sciences* 20.22 (2019): 5693.
 2314. Petrova, D., Yocheva, L., Petrova, M. et al. (2020). Antimicrobial and Antioxidant Activities of Microalgal Extracts. *Oxid Commun*, 43(1), 103.
 2315. Petrova, M.N. Phd Thesis Abstract.
 2316. Pina-Pérez, M.C., et al. Antimicrobial potential of macro and microalgae against pathogenic and spoilage microorganisms in food. *Food Chem.*, 2017, 235, 34-44.
 2317. Prihanto, Asep A., et al. "Freshwater Microalgae as Promising Food Sources: Nutritional and Functional Properties." *The Open Microbiology Journal* 16.1 (2022).
 2318. Potocki, L., Oklejewicz, B., Kuna, E. et al. (2021). Application of green algal planktochlorella nurekis biomasses to modulate growth of selected microbial species. *Molecules*, 26(13), 4038.
 2319. Quintero-Pacheco, N., Vásquez-García, A., Polanía, A. M., & Londoño-Hernandez, L. (2024). Potential Applications of Bioactive Compounds from Arid Zone Microorganisms. In *Exploration and Valorization of Natural Resources from Arid Zones* (pp. 65-100). Apple Academic Press.
 2320. Rajvanshi, Meghna, et al. "Biomolecules from microalgae for commercial applications." *Sustainable Downstream Processing of Microalgae for Industrial Application*. CRC Press, 2019. 3-38.
 2321. Rendón, L., Obando, E., Martínez, T., & Zapata, P. (2022). Photoautotrophic microorganisms with biotechnological potential unexplored. *Revista de la Facultad de Agronomía*, 121.
 2322. Reveillon D., Tunin-Ley A., Grondin I., Othmani A., Zubia M., Bunet R., Turquet J., Culioli G., Briand J. F. Exploring the chemodiversity of tropical microalgae for the discovery of natural antifouling compounds. *J. Appl. Phycol.*, 1-15, 2018.
 2323. Réveillon, Damien, et al. "Exploring the chemodiversity of tropical microalgae for the discovery of natural antifouling compounds." *Journal of applied phycology* 31.1 (2019): 319-333.
 2324. Righini, Hillary, et al. "Preliminary Study on the Activity of Phycobiliproteins against *Botrytis cinerea*." *Marine drugs* 18.12 (2020): 600.
 2325. Righini, H., Francioso, O., Di Foggia, M. et al. (2021). Assessing the potential of the terrestrial cyanobacterium *Anabaena minutissima* for controlling *Botrytis cinerea* on tomato fruits. *Horticulturae*, 7(8), 210.
 2326. Rojas, V, et al. Cyanobacteria and eukaryotic microalgae as emerging sources of antibacterial peptides. *Molecules*, 25.24 (2020):5804.
 2327. Rompin, T.W.O.E. (2016). antibacterial activities of lipid and pigment (Doctoral Dissertation, Universiti Tun Hussein Onn Malaysia).
 2328. Rudic, V., Pisov, M., Zosim, L., & Bulimaga, V. (2014). Metode de separare a metaboliților secundari bioactivi din biomasa unor cianobacterii și proprietățile lor curative și toxicologice.
 2329. Saad, M. H., Sidkey, N. M., & El-Fakharany, E. M. (2023). Identification and statistical optimization of a novel alginate polymer extracted from newly isolated

- Synechocystis algin* MNE ON864447 with antibacterial activity. *Microbial Cell Factories*, 22(1), 229.
2330. Safari, Moein, Salman Ahmady-Asbchin, and Pantea Zamanifar. "In vitro evaluation of antimicrobial activities from aqueous and methanolic extracts of cyanobacteria." *European Journal of Biological Research* 9.3 (2019): 184-192.
 2331. Sahoo, C.R., Swain, S., Luke, A.M. et al. (2021). Biogenic synthesis of silver-nanoparticles with the brackish water cyanobacterium *Nostoc sphaeroides* and assessment of antibacterial activity against urinary tract infecting bacteria. *Journal of Taibah University for Science*, 15(1), 805-813.
 2332. Saka, Cafer, Mustafa Kaya, and Mesut Bekiroğullari. "Chlorella vulgaris microalgae strain modified with zinc chloride as a new support material for hydrogen production from NaBH₄ methanolysis using CuB, NiB, and FeB metal catalysts." *International Journal of Hydrogen Energy* 45.3 (2020): 1959-1968.
 2333. Salem, O. M., Hoballah, E. M., Ghazi, S. M., & Hanna, S. N. Antimicrobial activity of microalgal extracts with special emphasize on *Nostoc* sp. *Life Science Journal*, 11(12), 2014, 752-758.
 2334. Salimi F., Farrokh P. Recent advances in the biological activities of microbial exopolysaccharides. *World Journal of Microbiology and Biotechnology*, 39 (8), art. no. 213. 2023. DOI: 10.1007/s11274-023-03660-x.
 2335. Saliu, Toyin Dunsin, et al. "Biocoagulant with Frother Properties for Harvesting Invasive Microalgae Colonies from the Eutrophicated System." *ACS Sustainable Chemistry & Engineering* 10.15 (2022): 5024-5034.
 2336. Sanmukh, S., Bruno, B., Ramakrishnan, U. et al. (2014). Bioactive compounds derived from microalgae showing antimicrobial activities. *J. Aquacult. Res. Developm.*, 2014, 5(3), 1-5.
 2337. Saulia, E.A. (2019). *Cyanobactéries diazotrophes du Pacifique Sud: variabilité saisonnière, caractérisation morpho-génétique/chimique et potentiel de valorisation* (Doctoral dissertation, Nouvelle Calédonie).
 2338. Sawant S. S., Mane V. K. Nutritional profile, antioxidant, antimicrobial potential, and bioactives profile of *Chlorella emersonii* KJ725233. *Asian J. Pharm. Clin. Res.*, 11 (3), 220-225, 2018.
 2339. Sawant, Sneha Sunil, and Varsha Kelkar-Mane. "Study of the changes in the growth, protein, and bioactive profile of *Chlorella emersonii* KJ725233 in response to sodium and ammonium nitrate." *Journal of Applied Biology & Biotechnology* Vol 7.04 (2019): 19-25.
 2340. Scare, M.S. Blog-Latest News.
 2341. Schmid, B. (2021). Assessment of microalgal biomass as potential feedstock for sustainable, eco-friendly biostimulants and biopesticides in plant production (Doctoral dissertation, Universidade do Algarve (Portugal)).
 2342. Schuelter, Adilson Ricken, et al. "Isolation and identification of new microalgae strains with antibacterial activity on food-borne pathogens. Engineering approach to optimize synthesis of desired metabolites." *Biochemical engineering journal* 144 (2019): 28-39.
 2343. Segers C., Mysara M., Coolkens A. et al. *Limnospira indica* PCC 8005 Supplementation Prevents Pelvic Irradiation-Induced Dysbiosis but Not Acute Inflammation in Mice. *Antioxidants*, 12 (3), art. no. 572, 2023. DOI:

- 10.3390/antiox12030572.
2344. Segers, C., Mysara, M., Coolkens, A. et al. (2022). *Limnospira indica* PCC 8005 or *Lactacaseibacillus rhamnosus* GG Dietary Supplementation Modulate the Gut Microbiome in Mice. *Applied Microbiology*, 2(3), 636-650.
2345. Senhorinho, G.N.A., Ross, G.M., Scott, J.A. Cyanobacteria and eukaryotic microalgae as potential sources of antibiotics *Phycologia* 2015, 54, 3, 271-282.
2346. Senhorinho, Gerusa NA, Carita Lannér, and John A. Scott. "The Importance of Harvesting Time on the Screening of *Chlamydomonas* spp. Extracts for Antibacterial Activity." *International Journal on Algae* 22.3 (2020), 269-278.
2347. Senhorinho, Gerusa Neyla Andrade. Antibacterial activity of freshwater green microalgae isolated from water bodies near abandoned mine sites in Ontario, Canada. Diss. Laurentian University of Sudbury, 2018.
2348. Shah, Sayed Asmat Ali, et al. "Chemically Diverse and Biologically Active Secondary Metabolites from Marine Phylum chlorophyta." *Marine Drugs* 18.10 (2020): 493.
2349. Schuelter, A. R., Kroumov, A. D., Hinterholz, C. L. et al. (2019). Isolation and identification of new microalgae strains with antibacterial activity on food-borne pathogens. Engineering approach to optimize synthesis of desired metabolites. *Biochemical engineering journal*, 144, 28-39.
2350. Shuttleworth, Beryl. "Can spirulina cause cancer?."
2351. Shuttleworth, B. Cutting edge nature.
2352. Silva Acosta, J.A. (2021). Fitodepuración mixotrófica en sistemas de recirculación acuícola (RAS): desempeño de un cocultivo de microalgas y bacterias nitrificantes inmovilizadas en matriz polimérica.
2353. Singh M., Gupta N., Gupta P. et al. Discovery of Novel and Biologically Active Compounds from Algae. *Next-Generation Algae: Volume II: Applications in Medicine and the Pharmaceutical Industry*, pp. 1 – 40, 2023. DOI: 10.1002/9781119857860.ch1.
2354. Singh U., Laxmi, Singh P., Singh A.K., Singh S., Kumar D., Shrivastava S.K., Asthana R.K. In silico and in vitro evaluation of extract derived from *Dunaliella salina*, a halotolerant microalga for its antifungal and antibacterial activity. *Journal of Biomolecular Structure and Dynamics*, 41 (15), pp. 7069 – 7083, 2023. DOI: 10.1080/07391102.2022.2115556.
2355. Singh, P.K., Fillat, M.F., Sither, V., & Kumar, A. (Eds.). (2022). *Expanding Horizon of Cyanobacterial Biology*. Academic Press..
2356. Sinha, Surbhi, et al. "Applications and Efficacy of Exceptional Bioactive Compounds from Microalgae."., *Handbook of Environmental Chemistry*, 104, (2020), 161-176.
2357. Sivakumar, N., & Selvakumar, G. (2015). Marine cyanobacteria: A prolific source of antimicrobial natural products. In *Antimicrobials* (pp. 220-249). CRC Press.
2358. Sreenikethanam, A. et al. Genetic engineering of microalgae for secondary metabolite production: Recent developments, challenges, and future prospects. *Front. Bioeng. Biotechnol.*, 10 2022, 836056.
2359. Stejskal, Nadia, et al. "Quality Enhancement of Refrigerated Hake Muscle by Active Packaging with a Protein Concentrate from *Spirulina platensis*", *Food and Bioprocess Technology*, 2020, 13, 1110–1118.
2360. Stirk, W.A., J. van Staden. Bioprospecting for bioactive compounds in microalgae: Antimicrobial compounds. *Biotechnol. Adv.*, 2022, 59:107977.

2361. Sun X.-L., Wang Y., Xiong H.-Q. et al. Removal of environmental estrogens from wastewater by microalgae under the influence of bacteria. *Journal of Cleaner Production*, 414, art. no. 137635. 2023. DOI: 10.1016/j.jclepro.2023.137635.
2362. Swapnil, Sanmukh, et al. "Bioactive compounds derived from microalgae showing antimicrobial activities." *Journal of Aquaculture research and Development* 5.3 (2014).
2363. Tang, D.Y.Y. et al. Potential utilization of bioproducts from microalgae for the quality enhancement of natural products. *Bioresource Technol.*, 304 (2020):122997.
2364. Tang, D.Y.Y. (2023). Evaluation of mixed microalgae species biorefinery of *Desmodesmus* sp. And *Scenedesmus* sp. For bioproducts synthesis (Doctoral dissertation, University of Nottingham).
2365. Terra, Ana Luiza Machado, et al. "Microalgae biosynthesis of silver nanoparticles for application in the control of agricultural pathogens." *Journal of Environmental Science and Health - Part B Pesticides, Food Contaminants and Agricultural Wastes*, 54.8 (2019): 709-716.
2366. Tiwari D., Mishra P., Gupta N. Bioactive Compounds Derived from Microalgae Showing Diverse Medicinal Activities. *Next-Generation Algae: Volume II: Applications in Medicine and the Pharmaceutical Industry*, pp. 77 – 94, 2023. DOI: 10.1002/9781119857860.ch3.
2367. Tokgöz, M., Yarkent, Ç., Köse, A., & Oncel, S. S. (2023). The potential of microalgal sources as coating materials: A case study for the development of biocompatible surgical sutures. *Letters in Applied Microbiology*, 76(8), ovad086.
2368. Toshkova-Yotova, Tanya, et al. Antitumor and antimicrobial activity of fatty acids from green microalga *Coelastrella* sp. BGV. *South African J. Botany* (2022).
2369. Tounsi, L et al. Microalgae as feedstock for bioactive polysaccharides. *Int. J. Biol. Macromol.*, 2022, 221, 1238-1250.
2370. Tounsi, L., Hlima, H. B., Elhadeif, K. et al. (2024). B-phycoerythrin of *Porphyridium cruentum* UTEX 161: A multifunctional active molecule for the development of biodegradable films. *European Polymer Journal*, 112851.
2371. Trentin, Riccardo, et al. Total phenolic levels, in vitro antioxidant properties, and fatty acid profile of two microalgae, *Tetraselmis marina* strain IMA043 and *Naviculoid diatom* strain IMA053, isolated from the North Adriatic Sea. *Marine Drugs* 20.3 (2022):207.
2372. Tsianta, Angeliki. "Pharmaceutical Applications of Eukaryotic Microalgae." (2020).
2373. Ugarković, T. Đ. (2021). Партиципативна права радника као вид ограничавања послодавчеве власти и претпоставка индустријске демократије. Универзитет у Београду.
2374. Uğuz, S., & Şimşek, E. (2023). Biological treatment of livestock wastewater by photobioreactor systems. *Advances in Agriculture, Forestry and Aquaculture Sciences*, 318.
2375. Uhliaríková, I., Šutovská, M., Barboríková, J. et al. (2020). Structural characteristics and biological effects of exopolysaccharide produced by cyanobacterium *Nostoc* sp. *International Journal of Biological Macromolecules*, 160, 364-371.
2376. Vanlalsangi, Rebecca, and Rosie Lalmuanpuui. "Cyanobacteria-derived bioactive compounds: A beneficial aspects." *Expanding Horizon of Cyanobacterial Biology*. Academic Press, 2022. 195-208.
2377. Vehapi, M. Makro ve mikroalglerin antimikrobiyal ve antioksidan etkilerinin incelenmesi (Master's thesis, Fen Bilimleri Enstitüsü).

2378. Wan, Dan, Qinghua Wu, and Kamil Kuča. "Spirulina." Nutraceuticals. Academic Press, 2016. 569-583.
2379. Widowati, I., Zainuri, M., Kusumaningrum, H. P., & Hardivillier, Y. (2021). Antibacterial Activity of Microalgae *Dunaliella salina*, *Tetraselmis chuii* and *Isochrysis galbana* Against Aquatic Pathogens.
2380. Witthohn M., Strieth D., Ulber R., Muffler K. Continuous production of an antimicrobial metabolite with the terrestrial cyanobacterium *Chroococcidiopsis cubana*. *Algal Research*, 74, art. no. 103193, 2023. DOI: 10.1016/j.algal.2023.103193.
2381. Yadavalli R., Valluru P., Raj R., Reddy C.N., Mishra B. Biological detoxification of mycotoxins: Emphasizing the role of algae. *Algal Research*, 71, art. no. 103039. DOI: 10.1016/j.algal.2023.103039, 2023.
2382. Yarkent, Ç., Aslanbay Güler, B., Imamoglu, E., & Oncel, S. S. (2024). Microalgae-factories as potential antimicrobial agents: a comprehensive review. *Biologia*, 1-14.
2383. Yi, Z., Su, Y., Brynjolfsson, S. et al. (2021). Bioactive polysaccharides and their derivatives from microalgae: biosynthesis, applications, and challenges. *Studies in Natural Products Chemistry*, 71, 67-85.
2384. Yun, Mira, et al. "Contaminated bacterial effects and qPCR application to monitor a specific bacterium in *Chlorella* sp. KR-1 culture." *Biotechnology and Bioprocess Engineering* 22.2 (2017): 150-160.
2385. Zahan, N., Hossain, M.A., Islam, M.R. et al. (2024). Effects of dietary *Spirulina platensis* on growth performance, body composition, haematology, immune response, and gut microflora of stinging catfish *Heteropneustes fossilis*. *Aquaculture Reports*, 35, 101997.
2386. Zayadan, B. K. (2021). Huma Balouch (Doctoral Dissertation, Al-Farabi Kazakh National University).
2387. Zainul, R. (2016). Isolation and identification of freshwater microalgae potentially as antibacterial from Talago Biru, Koto Baru, West Sumatera. *Der Pharmacia Lettre*, 8(20), 157-165.
2388. Zhang Liuting, Lü Jinyue, Zhang Gang et al. (2021). Optimization of ultrasound-assisted extraction of polyphenols from *Euryale ferox* and its inhibitory effect on *Staphylococcus aureus*. *Journal of Southern Agriculture*, 52(1), 189-197.
2389. Zhang Z., Chen Y., Klausen L.H. et al. The Rational Design and Development of Microalgae-Based Biohybrid Materials for Biomedical Applications. *Engineering* 2023. DOI: 10.1016/j.eng.2022.09.016.
2390. Zhou, J. et al. Extraction of lipids from microalgae using classical and innovative approaches. *Food Chem.*, 384 (2022):132236.
2391. Zhou, Yin, et al. A review of the antibacterial activity and mechanisms of plant polysaccharides. *Trends Food Sci. & Technol.*, (2022), 123, 264-280.
2392. Ziaziabari, F., & Fadaei, V. (2022). Characterization of a traditional egg-free crème caramel dessert containing *Chlorella protothecoides*. *Journal of Food Biosciences and Technology*, 12(2), 1-14.
2393. Жемчужин, С. Г., Ю. Я. Спиридонов, and Г. С. Босак. "Биопестициды: Современное состояние проблемы (дайджест публикаций за 2012–2017 гг.)." *Агрохимия* 11 (2019): 77-85.
2394. Огнистая, Альбина Васильевна. "Воздействие экстрактов микроводорослей

японского моря на патогенные микроорганизмы, вызывающие заболевания у человека и животных." Актуальные проблемы освоения биологических ресурсов Мирового океана. 2020.

89). NAJDENSKI, H., Iteman, I., Carniel, E. Efficient subtyping of pathogenic *Yersinia enterocolitica* strains by pulsed-field gel electrophoresis (1994) *Journal of Clinical Microbiology*, 32 (12), pp. 2913-2920. IF 3.474

2395. Aroma compounds of mountain tea (*Sideritis scardica* and *S. raeseri*) from Western Balkan. *Natural Product Communications*, 9 (9), 2014, pp. 1369-1372.
2396. Asplund K, Johansson T, Siitonen A. Evaluation of pulsed-field gel electrophoresis of genomic restriction fragments in the discrimination of *Yersinia enterocolitica* O:3. *Epidemiol. Infect.*, 1998, 121, 3, 579-586.
2397. Baraúna, Rafael A., et al. "Gel-Based Approaches in Genomic and Proteomic Sciences." *Polymer Gels*. Springer, Singapore, 2018. 185-195.
2398. Bhagat, N., & Virdi, J. S. (2010). The Enigma of *Yersinia enterocolitica* biovar 1A. *Critical Reviews in Microbiology*, 1, 15.
2399. Bing Qi, Wang Zhonghui, & Zhao Bin. (2015). Research progress on identification of plague bacteria. *Chinese Journal of Endemic Disease Control*, (5), 345-346.
2400. Blixt Y, Knutsson R, Borch E, et al. Interlaboratory random amplified polymorphic DNA typing of *Yersinia enterocolitica* and *Y. enterocolitica*-like bacteria. *Int. J. Food Microbiol.*, 2003, 83, 1, 15-26.
2401. Boghenbor, K.K., Stephen L., W. On, B. Kokotovic, A. Baumgartner, T.M. Wassenaar, M. Wittwer, B. Bissig-Choisat, and J. Frey. Genotyping of human and porcine *Yersinia enterocolitica*, *Yersinia intermedia*, and *Yersinia bercovieri* strains from Switzerland by amplified fragment length polymorphism analysis. *Appl. Environm. Microbiol.*, 2006, 72, 6, 4061-4066.
2402. Bosák, J., Micenková, L., Vrba, M., et al. Unique activity spectrum of colicin FY: All 110 characterized *Yersinia enterocolitica* isolates were colicin FY susceptible. *PloS one*, 2013, 8(12), e81829.
2403. Bottone EJ. *Yersinia enterocolitica*: overview and epidemiologic correlates. *Microbes Infect.*, 1999, 1, 4, 323-333.
2404. Boxall, Naomi. "pulsed-field gel electrophoresis: a tool for molecular epidemiology." New Zealand: Massey University (1999).
2405. Brack, Manfred, et al. "Fact sheet compiled by Last update." (2008).
2406. Campioni, F. Tipagem molecular e caracterização do potencial patogênico de linhagens de *Yersinia enterocolitica* biotipo 1A de origens diversas. Teses, Universidade De São Paulo, Faculdade De Ciências Farmacêuticas De Ribeirão Preto, 2009.
2407. Campioni, F., & Falcao, J. P.. Genotyping of *Yersinia enterocolitica* biotype 1A strains from clinical and non-clinical origins by Pulsed-field gel electrophoresis. *Can. J. Microbiol.*, 2014, 60, 419-424.
2408. Campioni, F., & Falcão, J.P. Genotypic diversity and virulence markers of *Yersinia enterocolitica* biotype 1A strains isolated from clinical and non-clinical origins. *Apmis*, 2014, 122(3), 215-222.
2409. Capilla S, Goni P, Rubio MC, et al. Epidemiological study of resistance to nalidixic acid and other antibiotics in clinical *Yersinia enterocolitica* O:3 isolates. *J. Clin.*

- Microbiol., 2003, 41, 10, 4876-4878.
2410. Cerdá, J. Sahagún, et al. "Epidemiological Study of Resistance to Nalidixic Acid and Other Antibiotics in Clinical *Yersinia enterocolitica* O:3 Isolates."
 2411. Ceylan, E. (2015). *Yersinia*.
 2412. Chae, H.-S., J.-Y. Kim, J.-E. Kim, Y.-M. Yang, K.-S. Jin, B.-W. Shin, S.-H. Kim, J.-H. Lee. Characteristics of *Yersinia enterocolitica* isolates from beef and pork carcass. Korean J. Vet. Serv., 2008, 31(2), 195-205.
 2413. Chaiyaroj SC, Kotrnon K, Koonpaew S, et al. Differences in genomic macrorestriction patterns of arabinose-positive (*Burkholderia thailandensis*) and arabinose-negative *Burkholderia pseudomallei*. Microb. Immunol., 1999, 43, 7, 625-30.
 2414. De Souza, R.A. Identificação de linhagens atípicas de *Yersinia* spp. por métodos moleculares, Dissertação, Universidade de São Paulo, Faculdade de Ciências Farmacêuticas de Ribeirão Preto, 2009.
 2415. Dhar, M. S.. Interaction of strains of two clonal groups of *Yersinia enterocolitica* with cultured cells in vitro. Indian ETD Repository, 2014
 2416. Donovan WH, Hull R, Rossi CD. Analysis of Gram negative recolonization of the neuropathic bladder among patients with spinal cord injuries. Spinal Cord, 1996, 34, 10, 587-591.
 2417. Estrada CSML, Velazquez LD, Escudero ME, et al. Pulsed field, PCR ribotyping and multiplex PCR analysis of *Yersinia enterocolitica* strains isolated from meat food in San Luis Argentina. Food Microbiol., 2011, 28, 1, 21-28.
 2418. Fakruddin, M., Bin Mannan, K.S., Mohammad M.RM., et al. Identification and characterization of microorganisms: DNA-fingerprinting methods. Songklanakarin J. Sci. Technol., 2013, 4, 35.
 2419. Falcão, J.P., Falcão, D.P., Pitondo-Silva, A., Malaspina, A.C., Brocchi, M. Molecular typing and virulence markers of *Yersinia enterocolitica* strains from human, animal and food origins isolated between 1968 and 2000 in Brazil (2006) Journal of Medical Microbiology, 55 (11), pp. 1539-1548.
 2420. Favier GI, Escudero ME, Velazquez L, et al. Reduction of *Yersinia enterocolitica* and mesophilic aerobic bacteria in eggshell by washing with surfactants and their effect on the shell microstructure. Food Microbiol., 2000, 17, 1, 73-81.
 2421. Fearnley, C., On, S.L.W., Kokotovic, B., Manning, G., Cheasty, T., Newell, D.G. Application of fluorescent amplified fragment length polymorphism for comparison of human and animal isolates of *Yersinia enterocolitica* (2005) Applied and Environmental Microbiology, 71 (9), pp. 4960-4965.
 2422. Franzin L, Cabodi D. Molecular typing of *Yersinia* strains by pulsed-field gel electrophoresis and RAPD-PCR. 8th International Symposium on *Yersinia*, Sept. 04-08, 2002 Turku, Finland. Genus *Yersinia*: entering the functional genomic era, Book Series: Adv. Exper. Med. Biology, 2003, 529, 349-352.
 2423. Franzin, Laura, and Daniela Cabodi. "Molecular typing of *Yersinia* strains by pulsed-field gel electrophoresis and RAPD-PCR." The Genus *Yersinia*. Springer, Boston, MA, 2004. 349-352.
 2424. Frazão, M. R. Tipagem molecular e caracterização do potencial patogênico de linhagens de *Yersinia enterocolitica* biotipo 2 de origens diversas, PhD Thesis, Universidade de São Paulo, 2013

2425. Fredriksson-Ahomaa M, Autio T, Korkeala H. Efficient subtyping of *Yersinia enterocolitica* bioserotype 4/O:3 with pulsed-field gel electrophoresis. *Lett. Appl. Microbiol.*, 1999, 29, 5, 308-312.
2426. Fredriksson-Ahomaa M, Bjorkroth J, Hielm S, et al. Prevalence and characterization of pathogenic *Yersinia enterocolitica* in pig tonsils from different slaughterhouses. *Food Microbiol.*, 2000, 17, 1, 93-101.
2427. Fredriksson-Ahomaa M, Hallanvuori S, Korte T, et al. Correspondence of genotypes of sporadic *Yersinia enterocolitica* bioserotype 4/O:3 strains from human and porcine sources. *Epidemiol. Infect.*, 2001, 127, 1, 37-47.
2428. Fredriksson-Ahomaa, M. Tracing of enteropathogenic *Yersinia*. In: *Yersinia: Systems Biology and Control*, Elisabeth Carniel & Bernard Joseph Hinnebusch eds. 2012, 201.
2429. Fredriksson-Ahomaa, M., S. Wacheck, R. Bonke, R. et al. Different enteropathogenic *Yersinia* strains found in wild boars and domestic pigs. *Foodborne Pathog Dis.* 2011; 8(6):733-737.
2430. Fredriksson-Ahomaa, M., Stolle, A., Korkeala, H. Molecular epidemiology of *Yersinia enterocolitica* infections (2006) *FEMS Immunology and Medical Microbiology*, 47 (3), pp. 315-329.
2431. Fredriksson-Ahomaa, M., Stolle, A., Stephan, R. Prevalence of pathogenic *Yersinia enterocolitica* in pigs slaughtered at a Swiss abattoir (2007) *International Journal of Food Microbiology*, 119 (3), 207-212.
2432. Fredriksson-Ahomaa, Maria. "Molecular epidemiology of *yadA*-positive *Yersinia enterocolitica*." (2001).
2433. Fukushima H, Hoshina K, Itogawa H, et al. Introduction into Japan of pathogenic *Yersinia* through imported pork, beef and fowl. *Int. J. Food Microbiol.*, 1997, 35, 3, 205-212.
2434. Fukushima, H., M. Gomyoda, S. Aleksic. Genetic variation of *Yersinia enterocolitica* serotype O:9 strains detected in samples from Western and Eastern countries. *Zbl. Bacteriol.*, 1998, 288, 2, 167-174.
2435. Galindo, C., J. A. Rosenzweig, M. L. Kirtley, et al. Pathogenesis of *Y. enterocolitica* and *Y. pseudotuberculosis* in human yersiniosis. *J. Pathogens* 2011, Article ID 182051.
2436. Garzetti, Debora. Genome-based characterization of *Yersinia enterocolitica*. *Diss. Imu*, 2015.
2437. Gauthier, Françoise Maurin, and Yves Richard. "Pulsed-Field Gel Electrophoresis Is More." *J. Clin. Microbiol* 37.2 (1999): 380.
2438. Gierczyński, R., Golubov, A., Neubauer, H., Pham, J.N., Rakin, A. Development of multiple-locus variable-number tandem-repeat analysis for *Yersinia enterocolitica* subsp. *paleartica* and its application to bioserogroup 4/O3 subtyping (2007) *Journal of Clinical Microbiology*, 45 (8), pp. 2508-2515.
2439. Gierczyński, R. Filogeneza, chorobotwórczość i zróżnicowanie pałeczek *Yersinia enterocolitica* Phylogeny, pathogenicity and genetic diversity of *Yersinia enterocolitica*.
2440. Goering, R.V. Molecular epidemiology of nosocomial infection. In: *Rapid detection of infectious agents* (S. Specter, M. Bendinelli, and H. Friedman, eds.), Plenum Press, New York and London, 1998, 131-157.
2441. Goering, Richard V. "The molecular epidemiology of nosocomial infection." *Rapid detection of infectious agents*. Springer, Boston, MA, 2002. 131-157.

2442. Gulati, P., Varshney, R.K., Viridi, J.S. Multilocus variable number tandem repeat analysis as a tool to discern genetic relationships among strains of *Yersinia enterocolitica* biovar 1A (2009) *Journal of Applied Microbiology*, 107 (3), pp. 875-884.
2443. Gulati, P.S., Viridi, J.S. The *rrn* locus and *gyrB* genotyping confirm the existence of two clonal groups in strains of *Yersinia enterocolitica* subspecies *polarctica* biovar 1A (2007) *Research in Microbiology*, 158 (3), pp. 236-243.
2444. Hallanvuo, S. Foodborne *Yersinia*: Identification and Molecular Epidemiology of Isolates from Human Infections. Academic Dissertation. University of Helsinki, Faculty of Agriculture and Forestry, 2009.
2445. Hosaka, S., M.Uchiyama, M.Ishikawa, T.Akahoshi, H.Kondo, C.Shimauchi, T.Sasahara, M.Inoue. *Yersinia enterocolitica* serotype 0:8 septicemia in an otherwise healthy adult: analysis of chromosome of DNA pattern by pulsed-field gel electrophoresis. *J. Clin. Microbiol.*, 1997, 35, 12, 3346-3347.
2446. Hunter, S.B., P. Vauterin, M.A. Lambert-Fair, M.S. Van Duyne, K. Kubota, L. Graves, D. Wrigley, T. Barrett, and E. Ribot. Establishment of a universal size standard strain for use with the PulseNet standardized pulsed-field gel electrophoresis protocols: converting the National Databases to the new size standard. *J. Clin. Microbiol.*, 2005, 43, 3, 1045-1050.
2447. Iteman, I., Guiyoule, A., & Carniel, E. (1996). Comparison of three molecular methods for typing and subtyping pathogenic *Yersinia enterocolitica* strains. *Journal of Medical Microbiology*, 45(1), 48-56.
2448. Jalkanen, Kaisa. "*Yersinia enterocolitica* biotyypin 1A ja *Y. enterocolitica* kaltaisten bakteerikantojen geneettisen monimuotoisuuden ja lämpökestoisten enterotoksiinigeenien kartoitus." (2007).
2449. Kardos, G., Turcsányi, I., Bistyák, A., Nagy, J., Kiss, I. DNA fingerprinting analysis of breakthrough outbreaks in vaccine-protected poultry stocks (2007) *Clinical and Vaccine Immunology*, 14 (12), pp. 1649-1651.
2450. Kardos, G., Turcsányi, I., Bistyák, A., Nagy, J., & Kiss, I. (2007). Application of molecular epidemiology methods to study breakthrough outbreaks occurring in vaccine-protected poultry stocks. *Clinical and Vaccine Immunology*.
2451. Kasimir, S. Verlaufsuntersuchungen zum Vorkommen potentiell humanpathogener *Yersinia enterocolitica* und *Campylobacter* spp. in Schweinebeständen von der Geburt bis zur Schlachtung sowie genotypisierung ausgewählter isolate. Dissertation, Veterinärmedizinischen Fakultät, Universität Leipzig, 2005.
2452. Kodjo, Angeli, et al. "Pulsed-field gel electrophoresis is more efficient than ribotyping and random amplified polymorphic DNA analysis in discrimination of *Pasteurella haemolytica* strains." *Journal of clinical microbiology* 37.2 (1999): 380-385.
2453. Koonpaew S, Ubol MN, Sirisinha S, et al. Genome fingerprinting by pulsed-field gel electrophoresis of isolates of *Burkholderia pseudomallei* from patients with melioidosis in Thailand. *Int. Congress on Melioidosis*, Nov. 1998, Bangkok, Thailand, *Acta Tropica*, 2000, 2-3, 187-191.
2454. Kotetishvili, M., Kreger, A., Wauters, G., Morris Jr., J.G., Sulakvelidze, A., Stine, O.C. Multilocus sequence typing for studying genetic relationships among *Yersinia* species (2005) *Journal of Clinical Microbiology*, 43 (6), pp. 2674-2684.
2455. Kozan, B. Kromozomal DNA ayırımında kullanılan düşük maliyetli atımlı alan

- elektroforez cihazının tasarımı ve optimizasyonu (Master's thesis, Fen Bilimleri Enstitüsü).
2456. Kuehni-Boghenbor, K., On, S.L.W., Kokotovic, B., Baumgartner, A., Wassenaar, T.M., Wittwer, M., Bissig-Choisat, B., Frey, J. Genotyping of human and porcine *Yersinia enterocolitica*, *Yersinia intermedia*, and *Yersinia bercovieri* strains from Switzerland by amplified fragment length polymorphism analysis (2006) *Applied and Environmental Microbiology*, 72 (6), pp. 4061-4066.
 2457. Κυρατσά, Α. (2013). Φαινοτυπική και γονοτυπική ανίχνευση λοιμογόνων παραγόντων σε στελέχη *Yersinia enterocolitica* που απομονώθηκαν από κλινικά περιστατικά (Doctoral dissertation, Εθνικό και Καποδιστριακό Πανεπιστήμιο Αθηνών (ΕΚΠΑ). Σχολή Επιστημών Υγείας. Τμήμα Ιατρικής. Τομέας Κλινικοεργαστηριακός. Εργαστήριο Μικροβιολογίας).
 2458. Kwak, Hyo-Shun, and Chong-Sam Lee. "Characterization of the Serotyping and the Plasmid Profile of *E. coli* Isolated from Foods and Clinical Specimens." *Korean Journal of Biological Sciences* 3.4 (1999): 399-405.
 2459. Lambertz ST, Danielsson-Tham ML. Identification and characterization of pathogenic *Yersinia enterocolitica* isolates by PCR and pulsed-field gel electrophoresis. *Appl. Environm. Microbiol.*, 2005, 71, 7, 3674-3681.
 2460. Laukkanen-Ninios, R., & Fredriksson-Ahomaa, M. Epidemiology, virulence genes, and reservoirs of enteropathogenic *Yersinia* species. *Foodborne and Waterborne Bacterial Pathogens: Epidemiology, Evolution and Molecular Biology*, 2012, 269-287.
 2461. Leal TCA, Leal NC, de Almeida AMP. RAPD-PCR typing of *Yersinia enterocolitica* (Enterobacteriaceae) O : 3 serotype strains isolated from pigs and humans. *Gen. Mlecul. Biol.*, 1999, 22, 3, 315-319.
 2462. Leon-Velarde, Carlos G. The Application of Bacteriophage Host Recognition Binding Proteins for the Isolation of *Yersinia enterocolitica* in Foods. Diss. 2017.
 2463. Lucero Estrada, C.S.M., L.C. Velázquez, M.E. Escudero, G.I. Favier, V. Lazarte and A.M.S. de Guzmán. Pulsed field, PCR ribotyping and multiplex PCR analysis of *Yersinia enterocolitica* strains isolated from meat food in San Luis Argentina, *Food Microbiology*, 2010 28(1), 21-28.
 2464. Lukinmaa S, Nakari UM, Eklund M, et al. Application of molecular genetic methods in diagnostics and epidemiology of food-borne bacterial pathogens. *APMIS*, 2004, 112, 11-12, 908-929.
 2465. Lyte, M., K.T. Nguyen. Alteration of *Escherichia coli* O157-H7 growth and molecular fingerprint by the neuroendocrine hormone noradrenaline. *Microbios*, 1997, 89, 360, 197-213.
 2466. Mallik, Sarita, and Jugsharan S. Virdi. "Genetic relationships between clinical and non-clinical strains of *Yersinia enterocolitica* biovar 1A as revealed by multilocus enzyme electrophoresis and multilocus restriction typing." *BMC microbiology* 10.1 (2010): 1-13.
 2467. Matic, I., M. Radman, F. Taddei, B. Picard, C. Doit, E. Bingen, E. Denamur, J. Elion. Highly variable mutation rates in commensal and pathogenic *Escherichia coli*. *Science*, 1997, 277, 5333, 1833-1834.
 2468. Mohandas, S., T.Rang. Relapsing infection due to *Burkholderia pseudomallei* detected by pulsed-field gel electrophoresis of sequential clinical isolates. *Asia Pacific Journal of Molecular Biology and Biotechnology*, 3, 1995, 3, 224-229.

2469. Muramatsu, K. (1999). Study of epidemiological markers for *Vibrio parahaemolyticus*. *Journal of Infectious Diseases*, 73(2), 179-186.
2470. Murru, N., Kodjo, A., Villard, L., Fratino, L., Perrelli, G., Tozzi, M., Cortesi, M.L. Identification of coagulase negative Staphylococci isolated from dairy products using molecular methods (2005) *Revue de Medecine Veterinaire*, 156 (8-9), pp. 455-459.
2471. Nagano, T. & Tsubokura, M. (1996). Serogroups of *Yersinia pseudotuberculosis* and some related problems. *Journal of the Japan Veterinary Medical Association*, 49(8), 509-515.
2472. Nesbakken, T. *Yersinia enterocolitica*. In: *Foodborne pathogens: microbiology and molecular biology*, P.M. Framaticco, A.K. Bhunia, and J.L. Smith, eds.), 2005, 227-250.
2473. Nesbakken, T. *Yersinia* infections. In: *Foodborne infections and intoxications*, H.P. Riemann, and D.O. Cliver, eds.), 3rd Edition, 2006, 187-199.
2474. Nesbakken, Truls. "Yersinia infections." *Foodborne Infections and Intoxications*, 3rd edn. Elsevier, Amsterdam (2006): 289-312.
2475. Nikolova S, Wesselinova D, Vesselinova A. Pretreatment of guinea-pigs with iron or Desferal influences the course of *Yersinia enterocolitica* infections. *J. Vet. Med. B-Infect. Dis. Vet. Public Health*, 2001, 48, 3, 167-178.
2476. Novoslavskij, A., Kudirkienė, E., Marcinkutė, A., Bajoriūnienė, A., Korkeala, H., & Malakauskas, M. (2013). Genetic diversity and antimicrobial resistance of *Yersinia enterocolitica* isolated from pigs and humans in Lithuania. *Journal of the Science of Food and Agriculture*, 93(8), 1858-1862.
2477. Odaert, M., P.Berche, M.Simonet. Molecular typing of *Yersinia pseudotuberculosis* by using an IS 200-like element. *J. Clin. Microbiol.*, 34, 1996, 9, 2231-2235.
2478. Okatani AT, Uto T, Taniguchi T, et al. Pulsed-field gel electrophoresis in differentiation of *Erysipelothrix* species strains. *J. Clin. Microbiol.*, 2001, 11, 4032-36.
2479. Okwori, J.A.E. Studies on yersiniosis in human and selected animal populations in Jos and its environs. A Thesis in the Department of Botany, Faculty of Natural Sciences, University of Jos, 2010.
2480. Okwori, Joseph Ameh Eleyi. Studies on Yersiniosis in Human and Selected Animal Populations in Jos and its Environs. Diss. 2008.
2481. Paixão, R., Moreno, L.Z., Sena De Gobbi, D.D., et al. Characterization of *Yersinia enterocolitica* biotype 1A strains isolated from swine slaughterhouses and markets. *Sci. World J.*, 2013, art. no. 769097.
2482. Petsios, Stefanos, et al. "Conventional and molecular methods used in the detection and subtyping of *Yersinia enterocolitica* in food." *International journal of food microbiology* 237 (2016): 55-72.
2483. Pilon J, Higgins R, Quessy S. Epidemiological study of *Yersinia enterocolitica* in swine herds in Quebec. *Can. Vet. J. – Rev. Vet. Can.*, 2000, 41, 5, 383-387.
2484. Qazimi, B., Stefkov, G., Karapandzova, M., Cvetkovikj, I., Kulevanova, S.
2485. Rahman, A., T. S. Bonny, S. Stonsaovapak, et al. *Yersinia enterocolitica*: epidemiological studies and outbreaks. *J. Pathogens*, 2011, Article ID 239391
2486. Raymond, Pierre, et al. "Diversity of *Yersinia enterocolitica* isolated from pigs in a French slaughterhouse over 2 years." *MicrobiologyOpen* 8.6 (2019): e00751.
2487. Renzi, F. Detection of *Yersinia enterocolitica* in Food by Biomolecular Techniques, PhD Thesis, University of Camerino, 2012.

2488. Renzi, Francesco. Detection of *Yersinia enterocolitica* in Food by Biomolecular Techniques. Diss. SCHOOL OF ADVANCED STUDIES-Doctoral course in Veterinary Sciences (XXIV cycle), 2012.
2489. Ringwood, Tamara. Epidemiology and comparative analysis of *Yersinia* in Ireland. Diss. University College Cork, 2013.
2490. Sacchini, Lorena, et al. "The prevalence, characterization, and antimicrobial resistance of *Yersinia enterocolitica* in pigs from Central Italy." *Veterinaria italiana* 54.29 (2018): 115-123.
2491. Sachdeva P, Viridi JS. Repetitive elements sequence (REP/ERIC)-PCR based genotyping of clinical and environmental strains of *Yersinia enterocolitica* biotype 1A reveal existence of limited number of clonal groups. *FEMS Microbiol. Lett.*, 2004, 240, 2, 193-201.
2492. Sanchez-Céspedes J, Navia MM, Martínez R, et al. Clonal dissemination of *Yersinia enterocolitica* strains with various susceptibilities to nalidixic acid. *J. Clin. Microbiol.*, 2003, 41, 4, 1769-1771.
2493. Sharma, G., Chauhan, U. K., & Rathore, R. S. (2012). Pulsed-Field Gel Electrophoresis: A Tool for Molecular Epidemiology of Bacteria. *Journal of Immunology and Immunopathology*, 14(2), 63-71.
2494. Serra, T., González De Cárdenas, M., Plovins, J., Ballesteros, Á., Vindel, A., Sáez-Nieto, J.A. Three cases of *Yersinia pseudotuberculosis* gastrointestinal infection having no apparent epidemiological relationship, caused by identical strains (2005) *Enfermedades Infecciosas y Microbiología Clínica*, 23 (1), pp. 19-21.
2495. Sihvonen LM, Toivonen S, Haukka K, et al. Multilocus variable-number tandem-repeat analysis, pulsed-field gel electrophoresis, and antimicrobial susceptibility patterns in discrimination of sporadic and outbreak-related strains of *Y. enterocolitica*. *BMC Microbiol.*, 2011, 11, 42.
2496. Sihvonen, L.M., Jalkanen, K., Huovinen, E., et al. Clinical isolates of *Yersinia enterocolitica* biotype 1A represent two phylogenetic lineages with differing pathogenicity-related properties. *BMC Microbiology*, 12, 2012, art. no. 208.
2497. Sihvonen, Leila M., et al. "Multilocus variable-number tandem-repeat analysis, pulsed-field gel electrophoresis, and antimicrobial susceptibility patterns in discrimination of sporadic and outbreak-related strains of *Yersinia enterocolitica*." *BMC microbiology* 11.1 (2011): 1-10.
2498. Singh, A., Goering, R.V., Simjee, S., Foley, S.L., Zervos, M.J. Application of molecular techniques to the study of hospital infection (2006) *Clinical Microbiology Reviews*, 19 (3), pp. 512-530.
2499. Smirnova, N. I. "*Vibrio cholerae* adhesins: a phenotypical analysis and genetic control of synthesis." *MOLECULAR GENETICS MICROBIOLOGY AND VIROLOGY C/C OF MOLEKULIARNAIA GENETIKA MIKROBIOLOGIIA I VIRUSOLOGIIA* (1995): 1-18.
2500. Souza, R.A., Pitondo-Silva, A., Falcão, D.P., Falcão, J.P. Evaluation of four molecular typing methodologies as tools for determining taxonomy relations and for identifying species among *Yersinia* isolates (2010) *Journal of Microbiological Methods*, 82 (2), pp. 141-150.
2501. Steinhart,C.E., Doyle,M.E., and Cochrane,B.A., eds., *Food Safety*, 1996, Marcel

- Dekker, Inc., New York, Basel, Hong Kong
2502. Tadesse, D.A. Molecular epidemiology of *Campylobacter* and *Yersinia enterocolitica* isolates from pigs reared in conventional and antibiotic free farms from different geographic regions, PhD dissertation, The Ohio State University, 2009.
 2503. Tennant SM, Grant TH, Robins-Browne RM. Pathogenicity of *Yersinia enterocolitica* biotype 1A. *FEMS Immunol. Med. Microbiol.*, 2003, 38, 2, 127-137.
 2504. Thang, K.L., S.D.Puthuchery, T.Rang. In vivo genetic stability of *Salmonella typhi* detected by ribotyping and pulsed-field gel electrophoresis. *Asia Pacific J. Mol. Biol. Biotechnol.*, 3, 1995, 4, 356-361.
 2505. Thang,K.L., M.Passey, A.Clegg, B.G.Combs, R.M.Yassin, T.Rang. Molecular analysis of isolates of *Salmonella typhi* obtained from patients with fatal and non-fatal typhoid fever. *J. Clin. Microb.*, 34, 1996, 4, 1029-1033.
 2506. Thibodeau V, Frost EH, Chenier S, et al. Presence of *Yersinia enterocolitica* in tissues of orally-inoculated pigs and the tonsils and feces of pigs at slaughter. *Can. J. Vet. Res.- Rev. Can. Rech. Vet.*, 1999, 63, 2, 96-100.
 2507. Thisted Lambertz, S., Danielsson-Tham, M.-L. Identification and characterization of pathogenic *Yersinia enterocolitica* isolates by PCR and pulsed-field gel electrophoresis (2005) *Applied and Environmental Microbiology*, 71 (7), pp. 3674-3681.
 2508. Thong, K. L., Passey, M., Clegg, A., Combs, B. G., Yassin, R. M., & Pang, T. (1996). Molecular analysis of isolates of *Salmonella typhi* obtained from patients with fatal and nonfatal typhoid fever. *Journal of Clinical Microbiology*, 34(4), 1029-1033.
 2509. Toutounian-Mashhad, Kaveh. Molekularbiologische Feintypisierung von *Yersinia* spp.-Stämmen aus Mastgeflügel. Diss. 2007.
 2510. Uysal, Ahmet. Çeşitli *Escherichia coli* suşlarının pulsed field jel elektroforez yöntemi ile tiplendirilmesi, plazmit profilleri ve antibiyotik duyarlılıklarının araştırılması. Diss. Selçuk Üniversitesi Fen Bilimleri Enstitüsü, 2012.
 2511. Velazquez, L.D.C., M.E. Escudero, A.M.S. Deguzman. Prevalence of *Yersinia enterocolitica* in hake (*Merluccius-Hubbsi*) fillets. *J. Food Protection*, 1996, 59, 7, 781-783.
 2512. Viridi, J.S., Sachdeva, P. Molecular heterogeneity in *Yersinia enterocolitica* and 'Y. enterocolitica-like' species - Implications for epidemiology, typing and taxonomy (2005) *FEMS Immunology and Medical Microbiology*, 45 (1), pp. 1-10.
 2513. Virtanen, S., Laukkanen-Ninios, R., Martínez, PO., et al. Multiple-locus variable-number tandem-repeat analysis in genotyping *Yersinia enterocolitica* strains from human and porcine origins. *J. Clin. Microbiol.*, 7, 51, 2013, 2154-2159.
 2514. Wang, Zengguo, Shao, Shi-he, Jing, Huai-qi. Molecular epidemiologic features of the *ystB* gene-bearing *Yersinia enterocolitica*. *Chinese Journal of Zoonoses*, 2008, 24(3).
 2515. Wang Zengguo, Shao Shihe, & Jing Huaiqi. (2008). Molecular typing of *Yersinia*. *Disease Surveillance*, 23(3), 183-187.
 2516. Wilfert, Franziska Gabriele. Evaluierung eines beadbasierten Immunoassays zum Nachweis enteropathogener *Yersinia* spp. bei Schlachtschweinen. Diss. lmu, 2014.
 2517. Wojciech L, Staroniewicz Z, Jakubczak A, et al. Typing of *Yersinia enterocolitica* isolates by ITS profiling, REP- and ERIC-PCR. *J. Vet. Med. B-Infect. Diseases Vet. Public Health*, 2004, 51, 5, 238-244.
 2518. Won-yong Kim, In-hwan Yoo, Mi-ok Song et al. (1999). Molecular evolution and

- identification of *Yersinia* bacteria by 16S rDNA analysis. Korean Journal of Microbiology, 34(4), 337-345.
2519. Woods Jr, C. R. (2009). Other *Yersinia* species. In Feigin and Cherry's Textbook of Pediatric Infectious Diseases (pp. 1586-1603). WB Saunders.
2520. Wrigley, Timothy Barrett, et al. "Susan B. Hunter, Paul Vauterin, Mary Ann Lambert-Fair, M."
2521. Zhu, J., X. Ting, W. Peng, & S. Zhizhong. Research progress on *Yersinia pestis* molecular typing methods. Chinese J. Contr. Endem. Dis., 2013, 3, 177-180.
2522. Воскресенская, Екатерина Александровна. Новые подходы к микробиологическому мониторингу популяций *Yersinia pseudotuberculosis* и лабораторной диагностике псевдотуберкулеза. Diss. Российский научно-исследовательский противочумный институт "Микроб", 2014.
2523. Каримова, Татьяна Викторовна. "Энтеропатогенные иерсинии: микробиологический мониторинг, молекулярно-биологические особенности, алгоритм лабораторной диагностики." (2017).
2524. Митов, И. Дисертация "Доктор на медицинските науки", София, 2005
2525. Николова, С. Роля на желязото върху патогенността на бактериите от вид *Yersinia enterocolitica*. Канд. дисертация, 1997, София
2526. Орозова, П. Физиологични особености на щамове *Yersinia enterocolitica* и *Yersinia pseudotuberculosis* с различна вирулентност. Дисертация за присъждане на образователната и научна степен "доктор". София, 2001.
2527. Смирнова, Елена Юрьевна. Совершенствование лабораторного обеспечения системы эпидемиологического надзора за иерсиниозами. Diss. Санкт-Петербургская государственная медицинская академия им. ИИ Мечникова, 2003.
2528. Смирнова, Н. И. "Адгезины *Vibrio cholerae*: фенотипический анализ и генетический контроль синтеза." Молекуляр. ген. 6 (1995): 18.
2529. Чеснокова, М. В. "Энтеропатогенные иерсинии: микробиологический мониторинг, молекулярно-биологические особенности, алгоритм лабораторной диагностики." (2017).
2530. Чеснокова, Маргарита Валентиновна. Псевдотуберкулез в Сибири: Теоретические и прикладные аспекты эпидемиологии, лабораторной диагностики и профилактики. Diss. Науч. центр мед. экологии ВШЦ СО РАМН, 2004.
2531. Чужебаева, Г. Д., et al. "Диагностика и идентификация штаммов *Yersinia enterocolitica* и *Yersinia pseudotuberculosis* методом полимеразной цепной реакции (ПЦР)." (2017): 110-110.
- 90). Kamenarska, Z., Serkedjieva, J., NAJDENSKI, H., Stefanov, K., Tsvetkova, I., Dimitrova-Konaklieva, S., Popov, S. Antibacterial, antiviral, and cytotoxic activities of some red and brown seaweeds from the Black Sea (2009) Botanica Marina, 52 (1), pp. 80-86.**
2532. Ahmed, H.H., M.M. Hegazi, H.I. Abd-Alla, E.F. Eskander and M.S. Ellithey. Antitumour and antioxidant activity of some red sea seaweeds in Ehrlich ascites carcinoma in vivo. Zeitschr. Naturforsch.- Sect. C J. Biosci., 2011, 66 C(7-8), 367-376.
2533. Akremi, N., et al. "Phytochemical and in vitro antimicrobial and genotoxic activity in the brown algae Dictyopteris membranacea." South African Journal of Botany 108 (2017):

- 308-314.
2534. AliAboutabl, Elsayed, et al. "Secondary metabolites and certain bioactivities of *Pterocladia capillacea* (S. Gmelin) Bornet and *Dictyopteris membranacea* (Stackhouse) Batters." *Medicinal and Aromatic Plant Science and Biotechnology* 4.1 (2010): 41-48.
 2535. Aoun, Z.B., Said, R.B., Farhat, F. Anti-inflammatory, antioxidant and antimicrobial activities of aqueous and organic extracts from *Dictyopteris membranacea* (2010) *Botanica Marina*, 53 (3), pp. 259-264.
 2536. Arunkumar, K., Sivakumar, S. R., & Shanthi, N. *Int Ernatiol Journal Of Advances In Pharmacy, Biology And Chemistry*.
 2537. Blanco, Andreu, et al. "Mapping invasive macroalgae in the Western Iberian Peninsula: a methodological guide." (2020).
 2538. Bhardwaj, M., & Vasanthi, H.R. (2021). Biomedical Potential of Marine Macroalgae in Modulating Chronic Disease Pathologies. *Algae for Food*, 239-260.
 2539. Bouameur, S., Menad, A., Hamadou, T. et al. (2022). *Gelidium spinosum* red algae ameliorates oxidative/nitrosative stress and inflammation in DSS-induced ulcerative colitis in mice. *Egyptian Journal of Chemistry*, 65(3), 341-352.
 2540. Celenk, Fatma, and Atakan Sukatar. "Macroalgae of Izmir Gulf: *Cystoseira barbata*, *Cystoseira compressa* and *Cystoseira crinita* species have high α -glucosidase and moderate pancreatic lipase inhibition activities." *Iranian Journal of Pharmaceutical Research* (2020).19(2), 391-402
 2541. Cornish, M. L., & Garbary, D. J. Antioxidants from macroalgae: Potential applications in human health and nutrition. *Algae*, 2010, 25(4), 155-171.
 2542. Dang, V. T., Speck, P., Doroudi, M., et al. Variation in the antiviral and antibacterial activity of abalone *Haliotis laevis*, *H. rubra* and their hybrid in South Australia. *Aquaculture*, 2011, 315(3), 242-249.
 2543. Dang, V.T., Y. Li, P. Speck and K. Benkendorff. Effects of micro and macroalgal diet supplementations on growth and immunity of greenlip abalone, *Haliotis laevis*. *Aquaculture*. 2011, 320(1-2), 91-98.
 2544. Dang, V. (2012). Antiviral immune responses in abalone and influence of potential abiotic and biotic factors. Flinders University of South Australia, School of Biological Sciences.
 2545. De Felício, R., de Albuquerque, S., Young, M.C.M., Yokoya, N.S., Deboni, H.M. Trypanocidal, leishmanicidal and antifungal potential from marine red alga *Bostrychia tenella* J. Agardh (Rhodomelaceae, Ceramiales) (2010) *Journal of Pharmaceutical and Biomedical Analysis*, 52 (5), pp. 763-769.
 2546. Devi, G. K., Manivannan, K., & Anantharaman, P. Evaluation of antibacterial potential of seaweeds occurring along the coast of Mandapam, India against human pathogenic bacteria. *Journal of Coastal Life Medicine*, 2014, 2(3), 196-202.
 2547. Dhaouafi, J., Romdhani, M., Deracinois, B. et al. (2024). Fractionation and identification of bioactive peptides from red macroalgae protein hydrolysates: in silico analysis and in vitro bioactivities. *Biocatalysis and Agricultural Biotechnology*, 103211.
 2548. El-Saharty, A., Farghaly, O.A., Hamed, A.R. et al. (2018). Anticancer activity of some marine macroalgae in hepatocellular carcinoma cell lines (HepG2). *Int. J. Ecotoxicol. Ecobiol*, 3, 22-30.
 2549. Gerasimenko, N. I., Martyyas, E. A., & Busarova, N. G. Composition of lipids and

- biological activity of lipids and photosynthetic pigments from algae of the families Laminariaceae and Alariaceae. *Chem. Nat. Comp.*, 2012, 48(5), 737-741.
2550. Gerasimenko, N. I., Martyyas, E. A., Logvinov, S. V., & Busarova, N. G. Biological activity of lipids and photosynthetic pigments of *Sargassum pallidum* C. Agardh. *Applied Biochemistry and Microbiology*, 2014, 50(1), 73-81.
2551. Hernández, Oscar E., et al. "Species diversity and biogeographical patterns of *Laurencia sensu stricto* (Rhodophyta) in the Atlantic Ocean." *Hidrobiológica* 27.3 (2017): 301-314.
2552. Jha, Durga, et al. "Microalgae-based Pharmaceuticals and Nutraceuticals: An Emerging Field with Immense Market Potential." *ChemBioEng Reviews*.
2553. Koyande, A.K., Chew, K.W., Manickam, S. et al. (2021). Emerging algal nanotechnology for high-value compounds: a direction to future food production. *Trends in Food Science & Technology*, 116, 290-302.
2554. Lomartire, S., Pacheco, D., Araújo, G.S. et al. (2021). Wastewater utilization as growth medium for seaweed, microalgae and cyanobacteria, defined as potential source of human and animal services. In *Phycology-based approaches for wastewater treatment and resource recovery* (pp. 25-70). CRC Press.
2555. Latorre, Nicolás, et al. "First approach of characterization of bioactive compound in *Pyropia orbicularis* during the daily tidal cycle." *Latin american journal of aquatic research* 47.5 (2019): 826-840.
2556. Macroalga, O. B. (2014). Antimicrobial, Antioxidant And Haemolytic Potential.
2557. Maghin, Federica, Sabrina Ratti, and Carlo Corino. "Biological functions and health promoting effects of brown seaweeds in swine nutrition." *J. Dairy Vet. Anim. Res* 1 (2014): 14-16.
2558. Marino, Fabio, et al. "Preliminary study on the in vitro and in vivo effects of *Asparagopsis taxiformis* bioactive phycoderivates on teleosts." *Frontiers in physiology* 7 (2016): 459.
2559. Mariya, V., & Ravindran, V. S. Biomedical and Pharmacological significance of marine macro algae-review. *Indian J. Geo-Marine Sciences*, 2013, 42(5), 527-537.
2560. Martyyas, E. A., Gerasimenko, N. I., Busarova, N. G., et al. Seasonal changes in biological activity of lipids and photosynthetic pigments of *Saccharina cichorioides* (Miyabe) (Laminariaceae Family). *Russian J. Bioorg. Chem.*, 2013, 39(7), 720-727.
2561. McReynolds, Colin. *Invasive marine macroalgae and their current and potential use in cosmetics*. Diss. 2017.
2562. Meinita M.D.N., Harwanto D., Amron, Hannan M.A., Jeong G.-T., Moon I.S., Choi J.-S. A concise review of the potential utilization based on bioactivity and pharmacological properties of the genus *Gelidium* (Gelidiales, Rhodophyta). *Journal of Applied Phycology*, 35 (4), pp. 1499 – 1523, 2023. DOI: 10.1007/s10811-023-02956-7.
2563. Milchakova, N. A. *Marine plants of the Black Sea. An illustrated field guide*. Digit Print, Sevastopol. 2011. 144.
2564. Mohamed S, Hashim SN, Rahman HA. Seaweeds: a sustainable functional food for complementary and alternative therapy. *Trends Food Sci. Technol.*, 2012, 23(2), 83-96.
2565. Namjoyan, Foroogh, et al. "The Anti-melanogenesis Activities of Some Selected Red Macroalgae from Northern Coasts of the Persian Gulf." *Iranian Journal of Pharmaceutical Research* 18.1 (2019): 383-390.

2566. Nawi, M.N.B.M. (2015). Nutritional composition and heavy metal content of farmed and wild seaweed (*Gracilaria changii*) (Doctoral dissertation, MSc thesis, University Putra Malaysia).
2567. Neamat-Allah, Ahmed NF, Abd elhakeem I. El-Murr, and Yasser Abd El-Hakim. "Dietary supplementation with low molecular weight sodium alginate improves growth, haematology, immune reactions and resistance against *Aeromonas hydrophila* in *Clarias gariepinus*." *Aquaculture Research* 50.5 (2019): 1547-1556.
2568. Nuñez, M.L.S., Rabelo, V.W.H., de Palmer Paixao, I. C. N. et al. (2022). Brown macroalgae: Promising sources of bioactive products against human herpesviruses. *Journal of Medicinal Plants Research*, 16(3), 82-96.
2569. Paradas, WLADIMIR COSTA. Mecanismos de armazenamento, biossíntese e liberação de metabólitos secundários em macroalgas vermelhas (Rhodophyta) in Brazil. Diss. Ph. D. dissertation, Universidade Federal Fluminense, Niterói, Brazil, 2013.
2570. Patel M., Bazaid A.S., Azhar E.I. et al. Novel phytochemical inhibitors targeting monkeypox virus thymidine and serine/threonine kinase: integrating computational modeling and molecular dynamics simulation. *Journal of Biomolecular Structure and Dynamics* 2023. DOI: 10.1080/07391102.2023.2179547.
2571. Pereira, Leonel. "Seaweed flora of the European north Atlantic and Mediterranean." *Springer handbook of marine biotechnology*. Springer, Berlin, Heidelberg, 2015. 65-178.
2572. Pereira, Leonel. *Edible seaweeds of the world*. CRC Press, 2016.
2573. Pereira, Leonel. *Therapeutic and nutritional uses of algae*. CRC Press, 2018.
2574. Pinho, P. G., et al. "Head space-solid phase micro extraction and gas chromatography mass spectrometry applied to determination of volatiles in natural metrices." *Funct Plant Sc Biotech* 3.1 (2009): 1-15.
2575. Pirian, Kiana, et al. "Antidiabetic and antioxidant activities of brown and red macroalgae from the Persian Gulf." *Journal of Applied Phycology* (2017): 1-9.
2576. Prabha, V.S., Lekshmi, J.L., Jerin, S. (2024). GC-MS and antimicrobial activity analysis of the ethanolic extract of *Caulerpa taxifolia* (M. Vahl) C. Agardh. *Journal of Pharmacognosy and Phytochemistry*, 13(2), 620-626.
2577. Pushparaj, A., Ronald, J., & Tomas, A. (2014). An antibacterial activity of selected brown, green and red seaweeds from Manapad, Thoothukudi, Tamil Nadu, India. *World Journal of Pharmaceutical Sciences*, 854-856.
2578. Quitério, P.L.R. (2016). Caracterização de polissacarídeos presentes no sal e Salinas marinhas (Doctoral dissertation, Universidade de Aveiro (Portugal)).
2579. Raj, Sujitra Raj Genga, et al. "Antibacterial Potential of Aqueous Extracts and Compounds from Selected Brown Seaweeds." *INNOSC Theranostics and Pharmacological Sciences* 2.1 (2019).
2580. Rangaiah, G. Subba, P. Lakshmi, and K. Sruthikeerthi. "Antimicrobial activity of the crude extracts of Chlorophycean seaweeds *Ulva*, *Caulerpa* and *Spongomorpha* sps. against clinical and phytopathogens." *Drug Invention Today* 2.6 (2010).
2581. Reddy, P.B., Das, A., & Verma, A.K. (2024). Seaweed as a functional feed supplement in animal diet—A review. *The Indian Journal of Animal Sciences*, 94(4), 291-300.
2582. Santoro, Ilaria, et al. "Sustainable and Selective Extraction of Lipids and Bioactive Compounds from Microalgae." *Molecules* 24.23 (2019): 4347.
2583. Saranya, C., Parthiban, C., & Anantharaman, P. Evaluation of antibacterial and

- antioxidant activities of seaweeds from Pondicherry coast. *Adv. Appl. Sci. Res.*, 2014, 5, 4, 82-90.
2584. Sayın, S., Aydın, B., Çimen, B.A., Açıık, L. (2021). Evaluation of Antimicrobial Effects of Four Selected Marine Macroalgae from Iskenderun Bay.
2585. Sirakov, I., Velichkova, K., Rusenova, N., & Dinev, T. (2019). In vitro test of inhibition effect of extracts from three seaweed species distributed at Black sea on different pathogens potentially dangerous for aquaponics. *Romanian Biotechnological Letters*, 24(1), 176-183.
2586. Suryanarayanan, T.S. Fungal endosymbionts of seaweeds. *Biol. Mar. Fungi*, 2012, 53, 53-69.
2587. Tan, S. P., O'Sullivan, L., Prieto, M.L., et al. 13 Seaweed antimicrobials: isolation, characterization, and potential use in functional foods. *Bioactive Compounds from Marine Foods: Plant Anim. Sourc.*, 2013, 269-312.
2588. Todorov, D., Hinkov, A., Shishkova, K., & Shishkov, S. Antiviral potential of Bulgarian medicinal plants. *Phytochemistry Reviews*, 2014, 13(2), 525-538.
2589. Vakarelova, Martina. Microencapsulation of bioactive molecules from *Spirulina platensis* and *Haematococcus pluvialis*. Diss. University of Verona, 2017.
2590. Van Doan, H., Doolgindachbaporn, S., Suksri, A. Effects of low molecular weight agar and *Lactobacillus plantarum* on growth performance, immunity, and disease resistance of basa fish (*Pangasius bocourti*, Sauvage 1880). *Fish & Shellfish Immunology*, 2014, 41, 2, 340–34
2591. Van Doan, Hien, et al. "Combined administration of low molecular weight sodium alginate boosted immunomodulatory, disease resistance and growth enhancing effects of *Lactobacillus plantarum* in Nile tilapia (*Oreochromis niloticus*)."*Fish & shellfish immunology* 58 (2016): 678-685.
2592. Van Doan, Hien, et al. "Effects of low molecular weight sodium alginate on growth performance, immunity, and disease resistance of tilapia, *Oreochromis niloticus*." *Fish & shellfish immunology* 55 (2016): 186-194.
2593. Van Doan, Hien, et al. "The effects of dietary kefir and low molecular weight sodium alginate on serum immune parameters, resistance against *Streptococcus agalactiae* and growth performance in Nile tilapia (*Oreochromis niloticus*)."*Fish & shellfish immunology* 62 (2017): 139-146.
2594. Venugopal, Vazhiyil. *Marine polysaccharides: Food applications*. CRC Press, 2016.
2595. Vicente, T.F., Lemos, M.F., Félix, R. et al. (2021). Marine macroalgae, a source of natural inhibitors of fungal phytopathogens. *Journal of Fungi*, 7(12), 1006.
2596. Vicente T.F.L., Félix C., Félix R., Valentão P., Lemos M.F.L. Seaweed as a Natural Source against Phytopathogenic Bacteria. *Marine Drugs*, 21 (1), art. no. 23, 2023. DOI: 10.3390/md21010023.
2597. Vitale, Fabrizio, et al. "Effectiveness of red alga *Asparagopsis taxiformis* extracts against *Leishmania infantum*." *Open Life Sciences* 10.1 (2015).
2598. Yamamoto, R., Toriumi, S., Kawagoe, C. et al. (2024). Extraction and antioxidant capacity of mycosporine-like amino acids from red algae in Japan. *Bioscience, Biotechnology, and Biochemistry*, zbae051.
2599. Zahmatkesh, Fatemeh. Sampling, mapping and adding value to marine invasive seaweeds of the Iberian Peninsula. MS thesis. 2017.

2600. Анисимов, М. М., Мартыяс, Е. А., Чайкина, Е. Л., & Герасименко, Н. И. Противомикробная, гемолитическая и фиторегулирующая активность липидных экстрактов из морских водорослей. *Химия растительного сырья*, 2010, 4, 125-130.
2601. Герасименко, Н.И., Мартыяс, Е.А., Логвинов, С.В., Бусарова, Н.Г. (2014). Биологическая активность липидов и фотосинтетических пигментов бурой водоросли *Sargassum pallidum* (Turner) C. Agardh. *Прикладная биохимия и микробиология*, 50(1), 85-85.
2602. Мартыяс, Е. А., Герасименко, Н. И., Бусарова, Н. Г., и сътр. Биологическая активность липидов и фотосинтетических пигментов *Saccharina cichorioides* (Miyabe)(сем. *Laminariaceae*). Сезонные изменения активности. *Химия растительного сырья*, 2012, 1, 123-131.
- 91). Ivanova, A., Mikhova, B., NAJDENSKI, H., Tsvetkova, I., Kostova, I. Antimicrobial and cytotoxic activity of *Ruta graveolens*. *Fitoterapia*, 2005, 76(3-4), 344-347. IF 0.845**
2603. Abdallat, A. R. A. T. Evaluation Of Antimicrobial Activity Of Callus, In Vitro And Ex Vitro Extracts Of *Ruta Graveolens* L.
2604. Abdelrahim, Dana N., Hamed R. Takruri, and Khalid M. Al-Ismail. "Effect of Introducing the Jordanian Common Rue (*Ruta chalepensis* L.) on Liver Enzymes and Lipid Peroxidation in Adult Male Sprague Dawley Rats Toxicated With Paracetamol." *Journal of Agricultural Science* 12.4 (2020).
2605. Abou Elkhair, Emad, et al. "Antibacterial Effect of Some Palestinian Plant Extracts against Clinical Multidrug-Resistant Gram-Negative Bacteria: A possible synergism with antibiotics." *Bangladesh Journal of Medical Science* 19.3 (2020): 509-519.
2606. Al Qaisi, Yaseen T., et al. "*Ruta graveolens*, *Peganum harmala*, and *Citrullus colocynthis* methanolic extracts have in vitro protoscolocidal effects and act against bacteria isolated from echinococcal hydatid cyst fluid." *Archives of Microbiology* 204.4 (2022): 1-13.
2607. Al-Alouni, Z.I., Abbas, S., Shatnawi, M.A., Al-Makhadmeh, I. (2016). Effect of plant growth regulators on in vitro micropropagation and evaluation of antimicrobial activity extracts from ex vitro, in vitro and callus of rue (*Ruta graveolens* L.).
2608. Al-Ajlouni, Z.I., Abbas, S., Al-Ghzawi, A.L., Al-Makhadmeh, I.B.R.A.H.I.M., Shatnawi, M., Al-Abdallat, A.M. And Al-Tawaha, A.R., 2023. Evaluation of antimicrobial activity of callus, in vitro and ex vitro extracts of *ruta graveolens* L. *Farmacia*, 71(4).
2609. Alonso, Jessica, et al. "A arruda na hipercolesterolemia: Informações e relevância etnobotânica na investigação pré-clínica." (2015).
2610. Al Qaisi, Y.T., Khleifat, K.M., Oran, S.A. et al. (2022). *Ruta graveolens*, *Peganum harmala*, and *Citrullus colocynthis* methanolic extracts have in vitro protoscolocidal effects and act against bacteria isolated from echinococcal hydatid cyst fluid. *Archives of Microbiology*, 204(4), 228.
2611. AL-Qurainy, F. S. Khan, M. Ali, F. Al-Hemaid, et al. Authentication of *Ruta graveolens* and its adulterant using internal transcribed spacer (its) sequences of nuclear ribosomal DNA. *Pak. J. Bot*, 2011, 43, 3, 1613-1620.
2612. Alviz, E. R. L. Guerrero, E. Valencia. Estudio etnobotánico de especies medicinales utilizadas por la comunidad de la vereda campo alegre del corregimiento de Siberia –

- Cauca (Colombia). *Revista de Ciencias*, 2013, 17, 2, 35-49.
2613. Amaral, A.C.F., J.R.A. de Silva, D.Q. Falcão, L.G. Ferreirinha, A.R. dos Santos, R.B. Araújo and J.L.P. Ferreira. Ferreira. Chemical analysis of toxic principles in preparations of *ruta graveolens* and *petiveria alliacea*. In: *Poisoning by Plants, Mycotoxins, and Related Toxins*. CABI Publishing, 2011, pp. 698-704. ISBN:9781845938338 (ISBN).
 2614. Almeida, A., & Silveira, J. *Perspectivas Da Utilização De Arruda (Ruta Graveolens) No Armazenamento De Grãos*.
 2615. Amit Baran, Sharangi. "Medicinal Plants: The Magic of Wound Healing Activity." *Current Traditional Medicine* 2.3 (2016): 186-206.
 2616. An, Z.Y., Y.Y. Yan, D. Peng et al. Synthesis and evaluation of graveoline and graveoline derivatives with potent anti-angiogenesis activities. *Eur. J. Med. Chem.*, 2010, 45(9), 3895-3903.
 2617. Antil, A., Mukhopadhyay, S., Kumar, M. (2022). Aloe vera's effect on wound healing. *Biochemical & Cellular Archives*, 22(2).
 2618. Ardeshtirlarijani, E., Namazi, N., B Jalili, et al. (2019). Potential anti-obesity effects of some medicinal herb: In vitro α -amylase, α -glucosidase and lipase inhibitory activity. *International Biological and Biomedical Journal*, 5(2), 0-0.
 2619. Attou, A. Contribution à l'étude phytochimique et activités biologiques des extraits de la plante *Ruta chalepensis* (Fidjel) de la région d'Ain Témouchent. MS Thesis, Université Abou Bekr Belkaid Tlemcen (Algeria), 2011.
 2620. Ayil-Gutiérrez, Benjamín A., et al. "Ruta graveolens extracts and metabolites against *Spodoptera frugiperda*." *Natural product communications* 10.11 (2015): 1934578X1501001137.
 2621. Azalework, Henok Gulilat, et al. "Phytochemical investigation, GC-MS profile and antimicrobial activity of a medicinal plant *Ruta graveolens* L. from Ethiopia." (2017).
 2622. Bandatmakuru S. R., Arava V. R. Novel synthesis of graveoline and graveoline. *Synth. Commun.*, 48 (20), 2635-2641, 2018.
 2623. Bejaoui, I., T. Karmous. Tunisian *Ruta graveolens* essential oil: influence of factors on its yield and composition. *J. Essent. Oil Bear. Plants*, 2012, 15, 2, 276-282.
 2624. Bielikova, Olena, et al. "Antimicrobial Activity of Extracts of" Hairy" Root Culture and Regenerated Plants of *Ruta Graveolens* L. Against Some Soil and Pathogenic Bacteria." *Agrobiodiversity for improving nutrition, health and life quality* 1 (2017).
 2625. Bocardi, J.M.B. (2007). *Etnofarmacologia Das Plantas Medicinais De Céu Azul E Composição Química Do Óleo Essencial De Plectranthus neochilus* Schltr.
 2626. Bohidar, S., M. Thirunavoukkarasu, T.V. Rao. Propagation of *Ruta graveolens* L. by in vitro culture of nodal explants. *Indian Journal of Plant Physiology*, 2008, 13, 2, 125-129.
 2627. Cardona, E.R.C., García, P. P., & Álvarez, M.C.P. (2011). *La Informática al servicio de la Traducción*. Lenguaje, 33.
 2628. Cartaxo, S.L., M.M. de Almeida Souza, U.P. de Albuquerque. Medicinal plants with bioprospecting potential used in semi-arid northeastern Brazil. *J. Ethnopharmacol.*, 2010, 131, 2, 326-342.
 2629. Coimbra, Alexandra T., Susana Ferreira, and Ana Paula Duarte. "Genus *Ruta*: A natural source of high value products with biological and pharmacological properties." *Journal of Ethnopharmacology* (2020): 113076.

2630. Costa, V.P. and M.A.S. Mayworm. Medicinal plants used by the community of Tenentes District - extrema municipality, Minas Gerais State, Brazil. *Rev. Brasileira Plant. Med.*, 2011, 13(3), 282-292.
2631. Cunha, Micael R., et al. Analgesic activity of *Ruta graveolens* L.(Rue) extracts. *African Journal of Pharmacy and Pharmacology* 9.1 (2015): 1-5.
2632. da Costa, A.M.J. (2020). Pesquisa e Caracterização de inseticidas naturais em células de insectos.
2633. da Silva, Diego Romário, et al. "Atividade antimicrobiana do extrato de *Chenopodium ambrosioides* e *Ruta graveolens* sobre *Streptococcus mutans*." *Archives of Health Investigation* 7.4 (2018).
2634. Da Silva, F.G.E., F.R.S. Mendes, J. C. da C. Assunção, et al. Seasonal variation, larvicidal and nematicidal activities of the leaf essential oil of *Ruta graveolens* L. *J. Essent. Oil Res.*, 2014, 26, 3, 204-209.
2635. de Luna Antonio, Raquel, et al. "Smoke of Ethnobotanical Plants used in Healing Ceremonies in Brazilian Culture." *Ethnomedicinal Plants: Revitalizing of Traditional Knowledge of Herbs* (2011): 166.
2636. De Medeiros PM., Ladio AH., Albuquerque UP. Patterns of medicinal plant use by inhabitants of Brazilian urban and rural areas: A macroscale investigation based on available literature. *J. Ethnopharmacol.*, 2013, 2, 150, 729-746.
2637. De Obtención, E.D.U.P. (2010). Facultad De Ingeniería Química Y Agroindustria (Doctoral dissertation, ESCUELA POLITÉCNICA NACIONAL).
2638. Dob, T., Dahmane, D., Gauriat-Desrudy, B., Daligault, V. Volatile constituents of the essential oil of *Ruta chalepensis* L. subsp. *angustifolia* (Pers.) P. Cout. (2008) *Journal of Essential Oil Research*, 20 (4), pp. 306-309.
2639. Dongyue, J. I. A. N. G., L. I. Yonghong, and S. H. E. N. Xin. "Components and variations of volatile organic compounds released from leaves and flowers of *Ruta graveolens*." *Journal of Zhejiang A&F University* 35.3 (2018): 572-580.
2640. Ebadimanas, G., & Najafi, G. (2020). The in vitro effects of *Ruta graveolens* L. extract on the sperm DNA, fertilization and early embryonic development in mice. *Iranian Journal of Medicinal and Aromatic Plants*, 36(3).
2641. El-Bashiti, Tarek A., Emad Abou Elkhair, and Wesam S. Abu Draz. "The antibacterial and synergistic potential of some Palestinian plant extracts against multidrug resistant *Staphylococcus aureus*." *Journal of Medicinal Plants* 5.2 (2017): 54-65.
2642. El-Sayed MA., Kamel MM., El-Raei MA., Osman SM., Gamil L., Abbas HA. Study of antibacterial activity of some plant extracts against Enterohemorrhagic *Escherichia coli* O157:H7. *Res. J. Pharm. Technol.*, 8, 6, 2013, 916-919.
2643. Espadero, Mónica, et al. "Toxicological Evaluation of Essential Oil and Ethanolic Extracts of *Ruta graveolens* in *Artemia salina*." *Communication, Smart Technologies and Innovation for Society*. Springer, Singapore, 2022. 165-174.
2644. Flačar, Dora. Antibakterijska i antioksidativna aktivnost ekstrakata rutvice (*Ruta graveolens* L.) Dobivenih optimiranom ekstrakcijom u eutektičkom otapalu kolin-klorid/limunska kiselina. Diss. Josip Juraj Strossmayer University of Osijek. Department of biology, 2017.
2645. Foronda, Cervando Christian Gutierrez. Facultad de ciencias farmacéuticas y bioquímicas carrera de bioquímica. Diss. Universidad Mayor De San Andres, 2017.

2646. Forsatkar M. N., HedayatiRad M., Luchiari A. C. "Not tonight zebrafish": the effects of *Ruta graveolens* on reproduction. *Pharm. Biol.* , 56 (1), 60-66, 2018.
2647. Freire, R.B., H.R. Borba, C.D. Coelho. *Ruta graveolens* L. toxicity in *Vampirolepis nana* infected mice. *Indian J. Pharmacol.*, 2010, 42, 6, 345.
2648. Garcia, Daniel, and Lin Chau Ming. The Influence of Displacement by Human Groups Among Regions in the Medicinal Use of Natural Resource: A Case Study in Diadema, São Paulo-Brazil. *Pharmacology* (2012): 479.
2649. Garcia, Daniel, Marcus Vinicius Domingues, and Eliana Rodrigues. Ethnopharmacological survey among migrants living in the southeast Atlantic forest of Diadema, São Paulo, Brazil. *Journal of Ethnobiology and Ethnomedicine* 6.1 (2010): 1-19.
2650. Ghramh, Hamed A., et al. Silver nanoparticle production by *Ruta graveolens* and testing its safety, bioactivity, immune modulation, anticancer, and insecticidal potentials. *Bioinorganic Chemistry and Applications* 2020:5626382
2651. Gibka, J. Acykliczne ketony C7-C13 i wybrane ich pochodne: synteza i aktywność biologiczna. *Zeszyty Naukowe. Rozprawy Naukowe/Politechnika*. 2011, Z408, 3-143.
2652. Gibka, J., Kunicka-Styczyńska, L., Gliński, M. Antimicrobial activity of undecan-2-one, undecan-2-ol and their derivatives (2009) *Journal of Essential Oil-Bearing Plants*, 12 (5), pp. 605-614.
2653. Giresha, A.S., Anitha, M.G., Dharmappa, K.K. Phytochemical composition, antioxidant and in-vitro anti-inflammatory activity of ethanol extract of *Ruta Graveolens* L. leaves. *International Journal of Pharmacy and Pharmaceutical Sciences* 2015, 7, 272-276.
2654. Ghandour, A., Abdel-Rahim, M., Bayoumi, S.A.L. et al. (2021). Evaluation of antimicrobial and synergistic effects of some medicinal plant extracts on antimicrobial resistant organisms. *Microbes and Infectious Diseases*, 2(4), 807-818.
2655. Gonzales, G.F.N. Establecimiento de un proceso de obtención de extracto de ruda (*Ruta graveolens*), con alto contenido de polifenoles. Escuela Politécnica Nacional, Facultad de Ingeniería Química y Agroindustria, 2010.
2656. Gomides, N.A.M.T.P., Neto, G.G., Martins, M.P. et al. (2022). Estudio etnobotánico y etnofarmacológico de especies medicinales utilizadas en la Comunidad de Coqueiros, Brasil. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, 21(6), 671-715.
2657. Gu Guanyun, & Jiang Yu. (2008). Overview of research on plant anti-tumor drugs VI. *Foreign Medicine: Plant Medicine*, 23(6), 240-251.
2658. Guidi, L., M. Landi. Aromatic plants: use and nutraceutical properties. In: *Novel Plant Bioresources: Applications in Food, Medicine and Cosmetics*, Editor: Ameenah Gurib-Fakim, 2014, 303-345
2659. Gutiérrez Foronda, C.C., & Quisberth Barrera, S.R.T. (2017). Evaluación de la actividad antifúngica de extractos etanólicos de paico (*Chenopodium ambrosioides*), khoa (*Clinopodium bolivianum*) y ruda (*Ruta graveolens*) frente a *Moniliophthora* spp aislada a partir de muestras de cacao con moniliasis, La Paz-Bolivia, 2015 (Doctoral dissertation).
2660. Hashemi Karouei, S.M. (2015). Antifungal and antibacterial effects of *Ruta-graveolens* extracts. *International Journal of Molecular and Clinical Microbiology*, 5(1), 475-480.
2661. Hammiche V., Azzouz M. The rues: ethnobotany, phytopharmacology and toxicity.

- Phytotherapie, 1, 11, 2013, 22-30.
2662. Hammiche, V., R. Merad, and M. Azzouz. "Rues: Plantes toxiques à usage médicinal du pourtour méditerranéen. Springer Paris, 2013, 197-226.
 2663. Harat, Z.N., Sadeghi, M.R., Sadeghipour, H.R., Kamalinejad, M., Eshraghian, M.R. Immobilization effect of *Ruta graveolens* L. on human sperm: A new hope for male contraception (2008) *Journal of Ethnopharmacology*, 115 (1), pp. 36-41.
 2664. Honarmand, H.R., Falah-Delavar, S., Saeidinia, A. (2014). Antimicrobial effect of *Ruta graveolens* extract on pathogenic bacteria. *Journal of Gorgan University of Medical Sciences*, 16(3), 133-137.
 2665. Hoshiari, Aref, Gholamreza Najafi, and Leila Zareai. "Evaluation of oxidant effect of aqueous extract of *Ruta graveolens* on mice ovary." *Scientific Journal of Kurdistan University of Medical Sciences* 22.3 (2017): 40-48.
 2666. Huertas, J.M., García, J.P., Benavides, N.N., Yépez, M. C. (2008). Evaluación de la variación de los compuestos presentes en aceite. *Universidad y Salud*, 1(10).
 2667. Ivancheva, S., M. Nikolova and R. Tsvetkova. Pharmacological activities and biologically active compounds of Bulgarian medicinal plants. In: *Phytochemistry: Advances in Research*, (F. Imperato, ed.), 2006: 87-103.
 2668. Jabir, N. A. *Ruta Graveolens* (Hurb Of Grace)—Anti Bacterial Effects, A.
 2669. Jafar, S.M.J. (2008). In vitro thrombolytic/fibrinolytic effects of rue aqueous distilled extract. *Al-Nahrain Journal of Science*, 11(1), 28-33.
 2670. Jalali Moghadam, M. Ahmadi, et al. "Antimicrobial effect of *Ruta graveolens* extract on pathogenic bacteria." *Journal of Gorgan University of Medical Sciences* 16.3 (2014).
 2671. Jalali Moghadam, M. Ahmadi, et al. "Study on antibacterial effect of *Ruta graveolens* extracts on pathogenic bacteria." *Annals of Biological Research* 3.9 (2012): 4542-4545.
 2672. Jaradat, Nidal. "Quantitative estimations for the volatile Oil by using hydrodistillation and microwave accelerated distillation methods from *Ruta graveolens* L. and *Ruta chalepensis* L. leaves from Jerusalem area/Palestine." *Moroccan Journal of Chemistry* 4.1 (2016): 4-1.
 2673. Jeon, J.H., J.H. Park, H.S. Lee. 2-Isopropyl-5-methylphenol isolated from *Ruta graveolens* and its structural analogs show antibacterial activity against food-borne bacteria. *J. Korean Soc. Appl. Biol. Chem.*, 57.4 (2014): 485-490
 2674. Jiménez Posada, E.V. (2013). Efectos de los extractos polares de *ruta graveolens*, *Nicotiana tabacum* y *Chrysanthemum morifolium* sobre el hongo *Botrytis cinerea* de la mora de castilla (*Rubus glaucus* Benth).
 2675. Jorge, T.C.M., Lenartovicz, V., Andrade, M.W., Bonafin, T., Giordani, M.A., Bueno, N.B.C., Schneider, D.S.L.G. Pediculicidal activity of hydroethanolic extracts of *Ruta graveolens*, *Melia azedarach* and *Sambucus australis* (2009) *Latin American Journal of Pharmacy*, 28 (3), pp. 457-459.
 2676. Kacem, M, Kacem, I. Simon, G. Ben Mansour, A Chaabouni, S Elfeki, A. Bouaziz, M. Phytochemicals and biological activities of *Ruta chalepensis* L. growing in Tunisia *Food Bioscience* 2015, 12, 73-83.
 2677. Kane, A.M., Al-Darwesh, A.A., Al-Shammar, S.M. et al. (2024). Inhibitor Properties of Rue (*Ruta graveolens* L.) on Spermatogenesis in Guppy *Poecilia reticulata* Peters, 1859. *Basrah Journal of Agricultural Sciences*, 37(1), 224-235.
 2678. KAPLAN, A. (2021). Radical Scavenging and Antibacterial Activities of *Ruta*

- buxbaumii Poir.(Rutaceae) Growing in Raman Mountain-Batman. Sakarya University Journal of Science, 25(5), 1148-1158.
2679. Khan, S., K.J. Mirza, and M.Z. Abdin. DNA fingerprinting for the authentication of *Ruta graveolens*. African J. Biotechnol., 2011, 10 (44), 8709-8715.
2680. Kiani, I.E. Anti-viral compositions and method. US Patent 7700137 B1, 2010.
2681. Kiani, I.E. Composition and method for treating viral conditions. US Patent 8637094 B2, 2014
2682. Kiani, I.E. Method of treating viral conditions. US Patent 7850998,B2 2010
2683. Krause, M.S. A de Fatima Bonetti, J. de Melo Turnes et al. Phytochemistry and biological activities of *Zanthoxylum rhoifolium* Lam., Rutaceae - mini review. Visão Acadêmica Curitiba, 2013, 14, 4, 118-127.
2684. Krause, M.S. Estudos morfoanatômico, fitoquímico e de atividades biológicas de folha e caule de *Zanthoxylum rhoifolium* LAM., Rutaceae. MS Thesis, Universidade Federal do Parana, 2013.
2685. Krause, Mariana Saragioto. "Estudos morfoanatômico, fitoquímico e de atividades biológicas de folha e caule de *Zanthoxylum rhoifolium* Lam., Rutaceae." (2013).
2686. Kunicka-Styczyńska, A. and J. Gibka. Antimicrobial activity of undecan-x-ones (x = 2-4). Polish Journal of Microbiology. 2010, 59(4), 301-306.
2687. Law S., Sanyal S., Chatterjee R., Law A., Law A., Chattopadhyay S. Therapeutic management of peritoneal ascitic sarcomatosis by *Ruta graveolens*: A study in experimental mice. Pathol. Res. Pract., 214 (9), 1282-1290, 2018.
2688. Lopes, L.T.A., J.R. de Paula, L.M.F. Tresvenzol, M.T.F. Bara, S. de Sá, P.H. Ferri and T.S. Fiuza. Chemical composition and antimicrobial activity of essential oil and anatomy of citrus limettoides tanaka (rutaceae) leaves and stem. Revista de Ciencias Farmaceuticas Basica e Aplicada. 2013, 34(4), 503-511. ISSN:1808-4532.
2689. Luo, Wen, et al. "Synthesis, in vitro and in vivo biological evaluation of novel graveoline derivatives as potential anti-Alzheimer agents." Bioorganic & Medicinal Chemistry 28.1 (2020): 115190.
2690. Magno-Silva, E.R., Rocha, T.T. et al. (2020). Etnobotánica y etnofarmacología de plantas medicinales utilizadas en las comunidades de la Reserva Extractiva Marina de Soure-Pará, Brasil. Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas, 19(1), 29-64.
2691. Mahboubi, M., & Feizabadi, M.M. (2009). Antimicrobial activity of *Ducrosia anethifolia* essential oil and main component, decanal against methicillin-resistant and methicillin-susceptible *Staphylococcus aureus*. Journal of Essential Oil Bearing Plants, 12(5), 574-579.
2692. Malik, A.A., Ahmad, J., Suryapani, S., et al. Effect of inorganic and biological fertilizer treatments on essential oil composition of *Ruta graveolens* L. J. Herbs, Spices Med. Plants, 18 (2), 2012, 191-202.
2693. Malik, Sonia, et al. "Ruta graveolens: Phytochemistry, Pharmacology, and Biotechnology." Transgenesis and Secondary Metabolism (2017): 177-204.
2694. Mariotti, K.C., R.S. Schuh, J.M. Nunes, S.P. Salamoni, G. Meirelles, F. Barreto, G.L. Von Poser, R.B. Singer, E. Dallegrave, S.T. Van Der Sand and R.P. Limberger. Chemical constituents and pharmacological profile of *Gunnera manicata* L. extracts. Brazilian Journal of Pharmaceutical Sciences. 2014, 50(1), 147-154.

2695. Matvieieva, Nadiia A. "Regeneration of *Ruta graveolens* Transgenic Plants." *Ekin Journal of Crop Breeding and Genetics* 3.1 (2017): 66-71.
2696. Meccia, C., G. Rojas, and L.B. Usabillaga. Study on the essential oil of *Ruta graveolens* L. that grows in Mérida state, Venezuela. *Revista de la Facultad de Farmacia, Universidad de los Andes, Merida, Venezuela*, 2009, 50, 7-9.
2697. Mguis, K., A. Albouchi, N.B. Brahim. Effect of temperature and salinity on germination of *Ruta graveolens* L. *Acta Bot. Gallica: Bot. Let.*, 2011, 158, 4, 645-652.
2698. Mirrezaee, Noshin, and SHahin Mehrpour. "Effects of antifungal Rue on the candida albicans isolated from patients with vaginitis on in vitro during spring and winter seasons and comparison with two antibiotics." *Yafte* 19.2 (2017).
2699. Moghadam MAJ, Honarmand H, Falah-Delavar S, Saeidinia A. Study on antibacterial effect of *Ruta graveolens* extracts on pathogenic bacteria. *Ann. Biol. Res.* 2012, 3(9), 4542-4545.
2700. Mulat, M. Chali, K Tariku, Y Bacha, K. Evaluation for in-vitro antibacterial activity of selected medicinal plants against food-borne pathogens. *International Journal of Pharmaceutical Sciences Review and Research* 2015, 32, 2, 45-50.
2701. Muñiz, P.D.M., Ladio, A.H., Albuquerque, U.P. (2013). Patterns of medicinal plant use by inhabitants of Brazilian urban and rural areas: A macroscale investigation based on available literature.
2702. Naghibi Harat, Z., Kamalinejad, M., Sadeghi, M.R., Sadeghipour, H.R., Eshraghian, M.R. A review on *Ruta graveolens* L.; its usage in traditional medicine and modern research data (2009) *Journal of Medicinal Plants*, 8 (30), pp. 1-19+173.
2703. Nasir, K.F., Abduljabbar, I.A., Al-Mamoori, A.H. et al. (2021, June). Effects of phase change material on the performance of solar dryer used for *Eruca Sativa*. In *IOP Conference Series: Materials Science and Engineering* (Vol. 1105, No. 1, p. 012048). IOP Publishing.
2704. Naveda González, G.F. (2010). Establecimiento de un proceso de obtención de extracto de ruda (*Ruda Graveolens*), con alto contenido de polifenoles (Bachelor's thesis, QUITO/EPN/2010).
2705. Negri, G. and E. Rodrigues. Essential oils found in the smoke of "tira-capeta", a cigarette used by some quilombolas living in pantanal wetlands of Brazil. *Brazilian J. Pharmacognosy*. 2010, 20(3), 310-316.
2706. Pandey, P., A. Mehta, S. Hajra, et al. Anthelmintic activity of *Ruta graveolens* L. leaves extract. *Med. Plants*, 2010, 2, 3, 241-243.
2707. Pandey, P., A. Mehta, S. Hajra. Evaluation of antimicrobial activity of *Ruta graveolens* stem extracts by disc diffusion method. *J. Phytol.*, 2011, 3, 3, 92-95.
2708. Parray, S.A., Bhat, J.U., Ahmad, G., et al. *Ruta graveolens*: from traditional system of medicine to modern pharmacology: an overview. *Am. J. Pharm. Tech. Res.*, 2012, 2 (2), 239-252.
2709. Pathak, R., Panayides, J.-L., Jeftic, T.D., De Koning, C.B., Van Otterlo, W.A.L. The synthesis of 5-, 6-, 7- and 8-membered oxygen-containing benzo-fused rings using alkene isomerization and ring-closing metathesis reactions (2007) *South African Journal of Chemistry*, 60, pp. 1-7.
2710. Patil, A.G., Jobanputra, A.H. Rutin-Chitosan Nanoparticles: Fabrication, Characterization and Application in Dental Disorders. *Polymer - Plastics Technology and*

2711. Pavić, Valentina, et al. "Assessment of Total Phenolic Content, In Vitro Antioxidant and Antibacterial Activity of *Ruta graveolens* L. Extracts Obtained by Choline Chloride Based Natural Deep Eutectic Solvents." *Plants* 8.3 (2019): 69.
2712. Pavlović, D.R., M. Vukelić, S. Najman, M. Kostić, et al. Assessment of polyphenol content, in vitro antioxidant, antimicrobial and toxic potentials of wild growing and cultured rue. *J. Appl. Botany Food Qual.*, 2014, 87, 175-181.
2713. Pomagualli Quinchuela, Fanny Yolanda. Actividad antimicrobiana del extracto alcohólico y aceite esencial de *Rosmarinus officinalis* "Romero" frente a la cepa *Pseudomona aeruginosa*. MS thesis. 2018.
2714. Poonkodi, Kathirvel, et al. "Gas chromatography-mass spectrometry analysis and in vitro antioxidant activities of *Ruta graveolens* L. From western ghats region-south India." *GAS* 10.5 (2017).
2715. Posada, E.V.J. Efectos de los extractos polares de *ruta graveolens*, *Nicotiana tabacum* y *Chrysanthemum morifolium* sobre el hongo *Botrytis cinerea* de la mora de castilla (*Rubus glaucus* Benth). MS Thesis, Universidad Tecnológica De Pereira, 2013.
2716. Preethi, K.C., Nair, C.K.K., Kuttan, R. Clastogenic potential of *Ruta graveolens* extract and a homeopathic preparation in mouse bone marrow cells (2008) *Asian Pacific Journal of Cancer Prevention*, 9 (4), pp. 763-769.
2717. Prudente, R.C.C. and R.B. de Moura. Evidências científicas para a indicação popular de algumas espécies da família Rutaceae no tratamento de doenças respiratórias na região Sudeste do Brasil. *Infarma-Ciências Farmacêuticas*, 2013, 21, 1, 24-31.
2718. Rahim, F., G. Saki and M. Bazrafkan. Effect of alcohol extracts of the *Ruta graveolens* L. on the count, motility and in vitro fertilization capacity of rat's sperm. *Asian Journal of Plant Sciences*. 2010, 9(1), 63-66.
2719. Rai, M., Acharya, D., Rios, J.L. (Eds.). (2011). *Ethnomedicinal plants: Revitalizing of traditional knowledge of herbs*. CRC press.
2720. Ramulu, D.R., & Murthy, K.S.R. (2024). In vitro Propagation of *Ruta graveolens* L. *Micropropagation of Medicinal Plants: Volume 1*, 349.
2721. Rezk, S., Alqabbasi, O., Ramadan, A., Turkey, M. (2022). Effect of *ruta graveolens* extract on the major virulence factors in methicillin resistant *Staphylococcus aureus*. *Infection and Drug Resistance*, 7147-7156.
2722. Ribeiro Magno-Silva, Elis, Tainá Teixeira Rocha, and Ana Cláudia Caldeira Tavares-Martins. "Ethnobotany and ethnopharmacology of medicinal plants used in communities of the Soure Marine Extractive Reserve, Pará State, Brazil." *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas* 19.1 (2020).
2723. Sabale, P., Bhimani, B., Prajapati, C., et al. An overview of medicinal plants as wound healers, *J. Appl. Pharm. Sci.*, 2 (11), 2012, 143-150.
2724. Saeidinia, Amin, et al. "Lack of antibacterial activity of *Ruta graveolens* extracts against *Enterococcus faecalis*." *Pak J Pharm Sci* 29.4 (2016): 1371-1374.
2725. SANCHEZ, GILDARDO RIVERA. "Ruta graveolens extracts and metabolites against *Spodoptera frugiperda*." (2015).
2726. Santos, Rodrigo Souza, et al. "Registros de ocorrência e novos hospedeiros de *Gargaphia lunulata* (Mayr)(Hemiptera: Tingidae) nos estados do Acre e Paraná." *Embrapa Acre-Nota Técnica/Nota Científica (ALICE)* (2017).

2727. Semerdjieva, Ivanka B., et al. "Essential Oil Composition of *Ruta graveolens* L. Fruits and *Hyssopus officinalis* Subsp. *aristatus* (Godr.) Nyman Biomass as a Function of Hydrodistillation Time." *Molecules* 24.22 (2019): 4047.
2728. Silva, D.R., Ferreira, S.A.M., Silva, T.S. et al. (2018). Atividade antimicrobiana do extrato de *Chenopodium ambrosioides* e *Ruta graveolens* sobre *Streptococcus mutans*. *Arch Health Invest*, 7(4).
2729. Stachurska X., Mizielińska M., Ordon M., Nawrotek P. Combinations of *Echinacea* (*Echinacea purpurea*) and Rue (*Ruta graveolens*) Plant Extracts with Lytic Phages: A Study on Interactions. *Applied Sciences* (Switzerland), 13 (7), art. no. 4575, 2023. DOI: 10.3390/app13074575.
2730. Stafford, G.I., Pedersen, P.D., Jäger, A.K., Van Staden, J. Monoamine oxidase inhibition by southern African traditional medicinal plants (2007) *South African Journal of Botany*, 73 (3), pp. 384-390.
2731. STAFFORD, Gary Ivan. Southern African plants used to treat central nervous system related disorders. 2009. PhD Thesis.
2732. Shahrajabian, M.H. (2024). A Candidate for Health Promotion, Disease Prevention and Treatment: Common Rue (*Ruta graveolens* L.), an Important Medicinal Plant in Traditional Medicine. *Current Reviews in Clinical and Experimental Pharmacology Formerly Current Clinical Pharmacology*, 19(1), 2-11.
2733. Shilpa, V. (2021). Evaluation Of Hepatoprotective Activity Of *Ruta Graveolens* L. Against Paracetamol Induced Liver Damage.
2734. Sundar, G., Joseph, J., Sundar, R., John et al. (2022). Phyto-Nano Bioengineered Scaffolds: A Promise to Tissue Engineering Research. In *Handbook of Research on Nano-Drug Delivery and Tissue Engineering* (pp. 489-509). Apple Academic Press.
2735. Taheriazam, Afshin, Amin Saeidinia, and Faeze Keihanian. "Antibacterial activity of hexanic extract of *Ruta graveolens* on *Klebsiella pneumoniae* common postorthopedic surgery infection." *International Journal Of Pharmaceutical Sciences And Research* 9.4 (2018): 1650-1653.
2736. Teixeira Pires Gomides, Núbia Alves Mariano, et al. "Ethnobotanical and ethnopharmacological survey of medicinal species utilized in the Coqueiros Community, Brazil." *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas* 21.6 (2022).
2737. Torres Delgado, N.R. (2021). Revisión del uso tradicional como antimicrobianos agrícolas de la flora canaria versus la investigación fitoquímica.
2738. Tosun, G., Arslan, T., Iskefiyeli, Z., Küçük, M., Karaoğlu, Ş.A., Yayli, N. Synthesis and biological evaluation of a new series of 4-alkoxy-2-arylquinoline derivatives as potential antituberculosis agents. *Turkish Journal of Chemistry* 2015, 39, 850-866.
2739. Twej-Thu-Alfeqar, R., and Hameed-Sarmad Adil. "Comparative study between pre and post bacterial growth of periodontal infections by treatment with extracts Rue. An in vitro study." *Journal of Pharmaceutical Sciences and Research* 11.1 (2019): 104-109.
2740. Wubuli A., Abdulla R., Zang D., Jiang L., Chen L., Aisa H.A. Spectrum-effect relationship between UPLC fingerprints and melanogenic effect of *Ruta graveolens* L. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 1221, art. no. 123683, 2023. DOI: 10.1016/j.jchromb.2023.123683.
2741. Zanella, C.Â., Lunardi, P.S., Weber, T. et al. (2020). Administração aguda do óleo

- essencial de *Ruta graveolens* L. sobre a memória e sobre a amnésia causada pela escopolamina em camundongos machos. *Revista Perspectiva*, 44(165), 135-152.
2742. Вайновская, И. Ф., and Т. И. Фоменко. "Культура клеток и тканей in vitro фармакопейного растения *Ruta graveolens* L. Для биотехнологического использования." Биологически активные вещества растений-изучение и использование.
2743. Кароматов, Ином Джураевич. "Простые лекарственные средства." (2012).
2744. Кароматов, Иномжон Джураевич, and Суръат Сирожович Мавлонов. "Рута душистая перспективное лекарственное растение." Биология и интегративная медицина 6 (2018).
2745. Ястремська, Лариса Сергіївна. "Antimicrobial activity of extracts of "hairy" root culture and regenerated plants of *Ruta graveolens* L. against some soil and pathogenic bacteria." (2017).
- 92). Ivanova A., Mikhova B., NAJDENSKI H., Tsvetkova I., Kostova I. Chemical composition and antimicrobial activity of wild garlic *Allium ursinum* of Bulgarian origin. *Natural Product Communications*, 8, 4, 2009, 1059-1062, ISSN: 1934-578X IF 0.810**
2746. Ali, Akbar, et al. "Exploration of structural, electronic and third order nonlinear optical properties of crystalline chalcone systems: Monoarylidene and unsymmetrical diarylidene cycloalkanones." *Journal of Molecular Structure* 1241 (2021): 130685.
2747. Amany, S.Y., Suzan, A.E.F., Samy, A.E.A., & Rasha, A.A.A. Microorganisms isolated from surgical wounds infection and treatment with different natural products and antibiotics. *African J. Microbiol. Res.*, 2013, 7(30), 3895-3902.
2748. Bagiu RV, Vlaicu B, Butnariu M. Chemical composition and in vitro antifungal activity screening of the *Allium ursinum* L. (Liliaceae). *Int. J. Mol. Sci.*, 13 (2), 2012, 1426-1436.
2749. Barringer, S.A. (2020). Variety Differences in Garlic Volatile Sulfur Compounds, by Application of Selected Ion Flow Tube Mass Spectrometry (Sift–ms) with Chemometrics.
2750. Bârlă G.F., Poroch-Serițan M., Sănduleac (Tudosî) E., Ciornei (Ștefăroi) S.E. Antioxidant activity and total phenolic content in *Allium ursinum* and *Ranunculus ficari*. Volume XIII, Issue 4 – 2014, pag. 349 – 353
2751. BÂRLĂ, Gheorghe-Florin, et al. "Antioxidant activity and total phenolic content in *Allium ursinum* and *Ranunculus Ficaria*." *Food and Environment Safety Journal* 13.4 (2016).
2752. Benkerroum N. Traditional Fermented Foods of North African countries: technology and food safety challenges with regard to microbiological risks. *Comprehens. Rev. Food Sci.. Food Safety*, 2013, 1, 12, 54-89.
2753. Biswal, Priyabrata, et al. "Cobalt (II) porphyrin-Mediated Selective Synthesis of 1, 5-Diketones via an Interrupted-Borrowing Hydrogen Strategy Using Methanol as a C1 Source." *The Journal of Organic Chemistry* 86.9 (2021): 6744-6754.
2754. Bombicz, Mariann, et al. "A novel therapeutic approach in the treatment of pulmonary arterial hypertension: *Allium ursinum* liophylisate alleviates symptoms comparably to sildenafil." *International journal of molecular sciences* 18.7 (2017): 1436.
2755. Christiana, Jesumirhewe, Utomi Ozioma Lilian, and Ogunlowo Oladejo Peter.

- "Comparison of the phytochemicals and antimicrobial activity of leaf extracts of *Calotropis procera*, *Momordica charantia* and *Allium ascalonicum*." *African Journal of Biological Sciences* 2.2 (2020): 112-117.
2756. Da Silva, F.G.E., F.R. Da S. Mendes, J.C. Da C. Assunção, et al. Seasonal variation, larvicidal and nematocidal activities of the leaf essential oil of *Ruta graveolens* L. *J. Essent. Oil Res.*, 2014, 26(3), 204-209.
2757. Danquah, Cynthia Amaning, et al. "The Phytochemistry and Pharmacology of *Tulbaghia*, *Allium*, *Crinum* and *Cyrtanthus*: 'Talented' Taxa from the Amaryllidaceae." *Molecules* 27.14 (2022): 4475.
2758. Đurđević-Milošević, D. Antibacterial Activity Of *Allium Sativum* And *Allium Ursinum* On Selected Foodborne Pathogens.
2759. Džaferović, Aida, et al. "Antimikrobno Djelovanje Biljnih Ekstrakata Iz Roda *Allium* Antimicrobial Activity Of Plant Extracts From Genus *Allium*." *Proceedings/Zbornik Radova*: 65.
2760. El-Newary S.A., Abd Elkarim A.S., Abdelwahed N.A.M., Omer E.A., Elgamal A.M., ELsayed W.M. *Chenopodium murale* Juice Shows Anti-Fungal Efficacy in Experimental Oral Candidiasis in Immunosuppressed Rats in Relation to Its Chemical Profile. *Molecules*, 28 (11), art. no. 4304, 2023. DOI: 10.3390/molecules28114304.
2761. Farshbaf-Khalili, Azizeh, et al. "Comparing the effect of garlic, *zataria multiflora* and clotrimazole vaginal cream 2% on improvement of fungal vaginitis: a randomized controlled trial." *Iranian Red Crescent Medical Journal* 18.12 (2016).
2762. Finder, G.T., & Mapper, G.S. References Associated with Literature Topic: *Candida albicans*.
2763. Furdak, P., Pieńkowska, N., Kapusta, I., Bartosz, G. and Sadowska-Bartos, I., 2023. Comparison of Antioxidant and Antiproliferative Effects of Various Forms of Garlic and Ramsons. *Molecules*, 28(18), p.6512.
2764. Gargi, B., Singh, P., Painuli, S. et al. (2024). Literature-based screening and bibliometric analysis of the chemical composition, antioxidant and antimicrobial potential of essential oils isolated from *Allium* genus: 23 years of investigation. *Pharmacological Research-Modern Chinese Medicine*, 100354.
2765. Greplová, M., Polzerová, H., Ptáček, J., Domkářová, J. (2015). Preliminary experience with protoplast culture of *Allium ursinum*. *Acta Horticulturae* (Conference Paper), 1083, 461-467.
2766. Grzesiak B., Kołodziej B., Głowacka A., Krukowski H. The Effect of Some Natural Essential Oils Against Bovine Mastitis Caused by *Prototheca zopfii* Isolates In Vitro. *Mycopathologia*, 183(3), 541-550, 2018.
2767. Hamidović, S., Čakić, E., Bašić, F., Stojanova, M. and Lalević, B., 2023. Potencijal biljnih ekstrakata vlasca, medvjede luka. *Glasnik Zaštite Bilja*, 46(3.), pp.110-115.
2768. Harouak, Hazim, Jamal Ibjibjen, and Laila Nassiri. "Chemical profile of *Tetralin* *articulata* (Vahl) Masters, and *Juglans regia* L. and *Olea europaea* L. var. *Sylvestris* used against oral diseases: in vitro analysis between polyphenolic content and aqueous extraction optimization." *Heliyon* 7.5 (2021): e07118.
2769. Helbig, C., & Heber, T. (2020). Chemical sabotage in forest health management: Effects of *Allium ursinum* L. based semiochemicals on the catches of bark beetles and their antagonists in stands of European ash (*Fraxinus excelsior* L.). *Mitt. Dtsch. Ges. Allg.*

- Angew. Entomol, 22, 269-272.
2770. Hyun-woo Kim, Ha-neul Jo, Byeong-wan Yoo et al. (2018). Physiological activity and cosmetic preservative effect of ethanol extract of wild rose. *Journal of Korean Medicinal Crops Society*, 26(4), 308-316.
 2771. Jgerenaia, G., 2023. Chemistry and biological activity of secondary metabolites of plants *Allium saxatile* and *Allium ponticum* widespread in Georgia.
 2772. Jiang, Z., L. Liu, X. Zheng, et al. Effects of three relay intercropping patterns on watermelon growth and soil microflora in continuous cropping. *Acta Agricult. Changhai*, 2012, 1, 60-64.
 2773. Joardder, Mohammad UH, and Mahadi Hasan Masud. "Food preservation techniques in developing countries." *Food Preservation in Developing Countries: Challenges and Solutions*. Springer, Cham, 2019. 67-125.
 2774. Jung, Hyo Young, et al. "Essential oils from two *Allium* species exert effects on cell proliferation and neuroblast differentiation in the mouse dentate gyrus by modulating brain-derived neurotrophic factor and acetylcholinesterase." *BMC complementary and alternative medicine* 16.1 (2016): 431.
 2775. Khumaidi, Akhmad, Kumalahayati Maulina, and Arsa Wahyu Nugrahani. "Antibacterial Activity of *Allium ascalonicum* Linn Fractions A562275sal from the Palu Valley against *Shigella dysenteriae*." *JURNAL ILMU KEFARMASIAN INDONESIA* 17.2 (2019): 199-209.
 2776. Kılınç G., Yalçın S., Yalçın S. Effects of supplemental dried wild leek (*Allium scorodoprasum* L. subsp. *rotundum*) leaves on laying performance, egg quality characteristics, and oxidative stability in laying hens. *Tropical Animal Health and Production*, 55 (3), art. no. 169, 2023. DOI: 10.1007/s11250-023-03583-7.
 2777. Kim, Hyun Woo, et al. "Biological activity and cosmetic preservative effects of *Rosa multiflora* ethanol extracts." *Korean Journal of Medicinal Crop Science* 26.4 (2018): 308-316.
 2778. Kim, Yangseon, et al. "Identification and Comparison of Bioactive Components of Two *Dryopteris* sp. Extract Using LC-QTOF-MS." *Plants* 11.23 (2022): 3233.
 2779. Kısa, Dursun, et al. "Assessment of antimicrobial and enzymes inhibition effects of *Allium kastambulense* with in silico studies: Analysis of its phenolic compounds and flavonoid contents." *Arabian Journal of Chemistry* 15.6 (2022): 103810.
 2780. Kovačević T.K., Major N., Sivec M., Horvat D., Krpan M., Hruškar M., Ban D., Išić N., Goreta Ban S. Phenolic Content, Amino Acids, Volatile Compounds, Antioxidant Capacity, and Their Relationship in Wild Garlic (*A. ursinum* L.). *Foods*, 12 (11), art. no. 2110, 2023. DOI: 10.3390/foods12112110.
 2781. Krivokapić, Miloš Z. Испитивање антимикробних, антиинфламацијских, антиоксидационих и кардиопротективних ефеката екстракта сремуша, *Allium Ursinum* L "Univeristy Of Kragujevac Faculty Of Medical Sciences."
 2782. Krivokapic, Milos, et al. "Phytochemical and Pharmacological Properties of *Allium ursinum*." *Serbian Journal of Experimental and Clinical Research* 1.ahead-of-print (2018).
 2783. Kunicka-Styczyńska, A. and J. Gibka. Antimicrobial activity of undecan-x-ones (x = 2-4). *Polish J. Microbiol.*, 2010, 59(4), 301-306.
 2784. Kurćubić V.S., Stajić S.B., Miletić N.M., Petković M.M., Dmitrić M.P., Đurović V.M., Heinz V., Tomasevic I.B. Techno-Functional Properties of Burgers Fortified by

- Wild Garlic Extract: A Reconsideration. *Foods*, 12 (11), art. no. 2100, 2023. DOI: 10.3390/foods12112100.
2785. Kurnia, D., Ajiati, D., Heliawati, L. et al. (2021). Antioxidant properties and structure-antioxidant activity relationship of *Allium* species leaves. *Molecules*, 26(23), 7175.
2786. Kyung, KH. Antimicrobial properties of allium species. *Curr. Opin. Biotechnol.*, 23 (2), 2012, 142–147.
2787. Lachowicz, Sabina, et al. "Comparison of phenolic content and antioxidant capacity of bear garlic (*Allium ursinum* L.) in different maturity stages." *Journal of Food Processing and Preservation* 41.1 (2017).
2788. Lachowicz, Sabina, et al. "Influence of Maturity on the Content of Phenolic Compounds of *Alium ursinum* L." *Journal of Food Processing and Preservation* 41.1 (2017).
2789. Laurent, Emilien. "L'aromathérapie, la phytothérapie et les médicaments homéopathiques pour la prise en charge thérapeutique du sportif amateur: conseils à l'officine." (2017).
2790. Lazarević, J.S., A.S. Đorđević, B.K. Zlatković, et al. Chemical composition and antioxidant and antimicrobial activities of essential oil of *Allium sphaerocephalon* L. subsp. *sphaerocephalon* (Liliaceae) inflorescences. *J. Sci. Food Agric.*, 2011, 91, 2, 322-329.
2791. Li, Meiping, Xiying Zhao, and Manjun Xu. "Chemical Composition, Antimicrobial and Antioxidant Activity of Essential Oil from *Allium tenuissimum* L. Flowers." *Foods* 11.23 (2022): 3876.
2792. Li, R., Wang, Y.-F., Sun, Q., Hu, H.-B. Chemical composition and antimicrobial activity of the essential oil from *Allium hookeri* consumed in Xishuangbanna, Southwest China, *Nat. Prod. Commun.*, 2014, 9, 6, 863-864.
2793. Lopes, L.T.A., J.R. de Paula, L.M.F. Tresvenzol, et al. Chemical composition and antimicrobial activity of essential oil and anatomy of citrus *limettioides tanaka* (rutaceae) leaves and stem. *Revista de Ciencias Farmaceuticas Basica e Aplicada*. 2013, 34(4), 503-511.
2794. Mariotti, K.C., R.S. Schuh, J.M. Nunes, et al. Chemical constituents and pharmacological profile of *Gunnera manicata* L. extracts. *Brazilian J. Pharm. Sci.*, 2014, 50(1), 147-154.
2795. Masato Fukuyama, & Masato Fukuyama. (2012). Informatics study of plant taxonomy for the application of the Indonesian traditional medicine Jamu.
2796. Mekonnen, H., A. Lemma. Plant species used in traditional smallholder dairy processing in East Shoa, Ethiopia. *Trop. Anim. Health Prod.*, 2011, 43, 833-841.
2797. Mihaylova, D.S., Lante, A., Tinello, F., & Krastanov, A.I. Study on the antioxidant and antimicrobial activities of *Allium ursinum* L. pressurised-liquid extract. *Natural Product Research*, (ahead-of-print), 2014, 1-6. DOI:10.1080/14786419.2014.923422
2798. Mihaylova, Dasha Sp, et al. "Study on the antioxidant and antimicrobial activities of *Allium ursinum* L. pressurised-liquid extract." *Natural product research* 28.22 (2014): 2000-2005
2799. Mitra, S., N. Maryam, H. Zeinab. Antibacterial effect of *Allium akaka* herbal extract on planktonic and biofilm cells of pathogen bacteria in laboratory conditions. *Ann. Rev. & Res. Biol.*, 2014, 4, 20.

2800. Mitra, Salehi, Navidi Maryam, and Hatami Zeinab. "Antibacterial Effect of Allium akaka Herbal Extract on Planktonic and Biofilm Cells of Pathogen Bacteria in Laboratory Conditions." *Annual Research & Review in Biology* (2014): 3087-3095.
2801. Mladenović, K.G., Grujović, M.Ž., Čomić, L.R. et al. Microbiological Safety Evaluation And Biological Properties Of The Allium Ursinum Sauce (Original Product From Kučaj Mountain).
2802. Mok, S.Y., Kim, H.M., Lee, S. Isolation of astragalin from flowers of *Rhododendron mucronulatum* for albiflorum. *Horticulture, Environm. Biotechnol.*, 2013, 54.5: 450-455.
2803. Mousavi, S. M., L. Najafian, and M. Farsi. "Effect of carboxymethyl cellulose and sodium alginate-based edible coating containing wild garlic (*Allium ursinum* L.) extract on the shelf-life of lactic cheese." *Food Hygiene* 10.1 (37) (2020): 73-89.
2804. Najgebauer-Lejko, Dorota, et al. "Effect of Bear Garlic Addition on the Chemical Composition, Microbiological Quality, Antioxidant Capacity, and Degree of Proteolysis in Soft Rennet Cheeses Produced from Milk of Polish Red and Polish Holstein-Friesian Cows." *Molecules* 27.24 (2022): 8930.
2805. Negri, G. and E. Rodrigues. Essential oils found in the smoke of "tira-capeta", a cigarette used by some quilombolas living in pantanal wetlands of Brazil. *Brazilian J. Pharmacognosy*, 2010, 20(3), 310-316.
2806. Nićetin, M., Filipović, V., Filipović, J. et al. (2022). Osmotic dehydration of wild garlic in sucrose–salt solution. *Acta Universitatis Sapientiae, Alimentaria*, 15(1), 27-39.
2807. Oszmiański J., Kolniak-Ostek J., Wojdyło A. Characterization and content of flavonol derivatives of *Allium ursinum* L. plant. *J. Agr. Food Chem.*, 2013, 1, 61, 176-184.
2808. Ozcan, Mehmet Musa. "Bioactive properties of garlic (*Allium sativum* L.)-A review." *Zeitschrift Fur Arznei- & Gewurzpflanzen* 21.4 (2016): 174-182.
2809. Özcan, F. Bazı liliaceae familyası üyelerinin antimikrobiyal, antibiyofilm, antioksidan, antimutajenik ve sitotoksik aktiviteleri (Master's thesis, Fen Bilimleri Enstitüsü).
2810. Pacirc, M., A.E. Pacirc, L. Vlase, et al. Antifungal properties of *Allium ursinum* L. ethanol extract. *J. Med. Plants Res.*, 2011, 5, 10, 2041-2046.
2811. Packia Lekshmi, N. C. J., et al. "Antimicrobial spectrum of *Allium* species—a review." *History* 15.44 (2015): 1-5.
2812. Packia Lekshmi, N.C.J., Viveka, S., Jeeva, S. et al. (2015). Efficacy of crude extracts of *Allium sativum* and *Allium cepa* against human pathogens. *Adv. Appl. Sci. Res*, 6(1), 72-78.
2813. Pant, H.M., Pant, N., & Negi, J.S. Study on ethno-medicinal practices and system of cure among the people of rath region of Garhwal Himalaya, Uttarakhand. *Nature & Science*, 2011, 9(6). <http://www.sciencepub.net/nature>
2814. Parvu, A.E., Catoi, F., Deelawar, S., et al. Anti-inflammatory effect of *Allium ursinum*. *Notulae Scientia Biologicae*, 2014, 6(1), 20-26.
2815. Pârvu, M., A.E. Pârvu. Antifungal plant extracts. In *Science against microbial pathogens: communicating current research and technological advances*. A. Mendez-Vilaz (ed.), Formatex Research Center, Badajoz (Spain), 2011, 1055-1062.
2816. Pavlić B., Aćimović M., Sknepnek A., Miletić D., Mrkonjić Ž., Kljakić A.C., Jerković J., Mišan A., Pojić M., Stupar A., Zeković Z., Teslić N. Sustainable raw materials for efficient valorization and recovery of bioactive compounds. *Industrial Crops and Products*, 193, art. no. 116167, 2023. DOI: 10.1016/j.indcrop.2022.116167.

2817. Periferakis, Argyrios, et al. "Kaempferol: Antimicrobial Properties, Sources, Clinical, and Traditional Applications." *International Journal of Molecular Sciences* 23.23 (2022): 15054.
2818. Petkova, D. T., et al. "Green approach to obtain extracts of seven edible flowers." *IOP Conference Series: Materials Science and Engineering*. Vol. 1031. No. 1. IOP Publishing, 2021, 012101.
2819. Petkova, N. Tr, et al. "Fructans and antioxidants in leaves of culinary herbs from Asteraceae and Amaryllidaceae families." *Food Research* 3.5 (2019): 407-415.
2820. Petropoulos, Spyridon A., et al. "Natural antioxidants, health effects and bioactive properties of wild *Allium* species." *Current Pharmaceutical Design* 26.16 (2020): 1816-1837
2821. Piatkowska, E., A. Kopec, and T. Leszczynska. "Basic chemical composition, content of micro-and macroelements and antioxidant activity of different varieties of garlic's leaves Polish origin." *Żywność Nauka Technologia Jakość* 22.1 (2015).
2822. Pop, Raluca Maria, et al. "Evaluation of the Antioxidant Activity of *Nigella sativa* L. and *Allium ursinum* Extracts in a Cellular Model of Doxorubicin-Induced Cardiotoxicity." *Molecules* 25.22 (2020): 5259.
2823. Popova A., Mihaylova D., Alexieva I. GC-MS chemical composition of volatile oil and mineral element content of *Allium ursinum* and *Nectaroscordum siculum*. *Pak. J. Bot*, 50 (6), 2351-2354, 2018.
2824. Rabah, Samia, et al. "Unveiling the bioactivity of *Allium triquetrum* L. lipophilic fractions: chemical characterization and in vitro antibacterial activity against methicillin-resistant *Staphylococcus aureus*." *Food & Function* 11(6), (2020) 1-10.
2825. Radusin, Tanja, et al. "Hybrid Pla/wild garlic antimicrobial composite films for food packaging application." *Polymer Composites* 40.3 (2019): 893-900.
2826. Radusin, Tanja, et al. "Preparation, characterization and antimicrobial properties of electrospun polylactide films containing *Allium ursinum* L. extract." *Food Packaging and Shelf Life* 21 (2019): 100357.
2827. Rahim, F., G. Saki and M. Bazrafkan. Effect of alcohol extracts of the *Ruta graveolens* L. on the count, motility and in vitro fertilization capacity of rat's sperm. *Asian J. Plant Sci.*, 2010, 9(1), 63-66.
2828. Rashid, Khalid H., Anees A. Khadom, and Salman H. Abbas. "Optimization, kinetics, and electrochemical investigations for green corrosion inhibition of low-carbon steel in 1 M HCl by a blend of onion-garlic leaves wastes." *Bioresource Technology Reports* 19 (2022): 101194.
2829. Riaz A., Rasul A., Hussain G., Zahoor M. K., Jabeen F., Subhani Z., Selamoglu Z. Astragalin: A Bioactive Phytochemical with Potential Therapeutic Activities. *Adv. Pharmacol. Sci.* 2018, 9794625, 2018.
2830. Sadeghi Dinani, Masoud, and Narges Zakeri Tehrani. "Bioassay Guided Fractionation of *Allium austroiranicum* by Cytotoxic Effects against Ovary and Cervical Cancer Cell Lines." *Research Journal of Pharmacognosy* 7.1 (2020): 1-6.
2831. Saptarini, N.M., Mustarichie, R., Hasanuddin, S. et al. (2024). *Cassia alata* L.: A Study of Antifungal Activity against *Malassezia furfur*, Identification of Major Compounds, and Molecular Docking to Lanosterol 14-Alpha Demethylase. *Pharmaceuticals*, 17(3), 380.
2832. Sayed, Shima MA, et al. "Identification of a hydroxygallic acid derivative,

- zingibroside R1 and a sterol lipid as potential active ingredients of *Cuscuta Chinensis* extract that has neuroprotective and antioxidant effects in aged *Caenorhabditis Elegans*." *Nutrients* 14.19 (2022): 4199.
2833. Sharifi-Rad, J., et al. "Plants of the genus *Allium* as antibacterial agents: From tradition to pharmacy." *Cellular and Molecular Biology* 62.9 (2016): 57-68.
2834. SİNİR, GÜLŞAH ÖZCAN, and Sheryl Ann Barringer. "Variety differences in garlic volatile sulfur compounds, by application of selected ion flow tube mass spectrometry (SIFT-MS) with chemometrics." *Turkish Journal of Agriculture and Forestry* 44.4 (2020): 408-416.
2835. Škrovánková S., Mlček J., Snopek L., Planetová T. Polyphenols and antioxidant capacity in different types of garlic. *Potravinárstvo Slovak J. Food Sci.*, 12 (1), 267-272, 2018.
2836. Sobolewska D., Podolak I., Makowska-Was J. *Allium ursinum*: botanical, phytochemical and pharmacological overview. *Phytochem Rev* (2015) 14:81–97
2837. Sobolewska, Danuta, Irma Podolak, and Justyna Makowska-Was. "Allium ursinum: botanical, phytochemical and pharmacological overview." *Phytochemistry reviews* 14.1 (2015): 81-97.
2838. Šobot, Kosana, et al. "Contribution of Osmotically Dehydrated Wild Garlic on Biscuits' Quality Parameters." *Periodica Polytechnica Chemical Engineering* 63.3 (2019): 499-507.
2839. Stanisavljević, Nemanja, et al. "Antioxidant and antiproliferative activity of *Allium ursinum* and their associated microbiota during simulated in vitro digestion in the presence of food matrix." *Frontiers in microbiology* 11 (2020).601616
2840. Stupar, Alena, et al. "Antibacterial Potential of *Allium ursinum* Extract Prepared by the Green Extraction Method." *Microorganisms* 10.7 (2022): 1358.
2841. Taslimi, P., & Kisa, D. Assessment of Antimicrobial and Enzymes Inhibition Effects of *Allium kastambulense* With In Silico Studies: Analysis of Its Phenolic Compounds and Flavonoid Contents.
2842. Ting, J., Liu, L., Z. Qing, et al. Three intercropping patterns of watermelon growth and soil microbial flora at continuous cropping. *Shanghai Agr. Sci.*, 2012, 28 (1), 60-64.
2843. Tomovic, Marina T., et al. "BIOLOGICAL ACTIVITIES OF DIFFERENT EXTRACTS FROM *ALLIUM URSINUM* LEAVES." *Acta Poloniae Pharmaceutica-Drug Research* 77.1 (2020): 121-129.
2844. Tomšik A., Šarić L., Bertoni S., Protti M., Albertini B., Mercolini L., Passerini N. (2018). Encapsulations of wild garlic (*Allium ursinum* L.) extract using spray congealing technology. *Food Res. Int.*, 2019, 119, 941-950.
2845. Uddin, Shaikh Jamal. Cytotoxicity Screening of Bangladeshi Medicinal Plants and Isolation and Structural Elucidation of Novel Anti-Cancer Compounds from *Acrostichum aureum*. Griffith University, 2011.
2846. Vidović, Senka, et al. "Supercritical Carbon Dioxide Extraction of *Allium ursinum*: Impact of Temperature and Pressure on the Extracts Chemical Profile." *Chemistry & Biodiversity* 18.4 (2021): e2100058.
2847. Vasile, D.I.A.N.A., Dincă, L.U.C.I.A.N., Voiculescu, I. (2015). Collecting medicinal plants from spontaneous flora of forest fund managed by National Forest Administration Romsilva.

2848. Vlase, L., M. Parvu, E.A. Parvu et al. Phytochemical analysis of *Allium fistulosum* L. and *A. ursinum* L. Digest J. Nanomat.Biostruct., 2012, 8(1), 457-467.
2849. Xiao, Yingcong. "The crystal structure of (3Z, 3' Z)-4, 4'-((1, 4-phenylenebis (methylene)) bis (azanediyl)) bis (pent-3-en-2-one), C₁₈H₂₄N₂O₂." Zeitschrift für Kristallographie-New Crystal Structures 236.4 (2021): 757-758.
2850. Xu X-Y., Song G-Q., Yu Y-Q., et al. Apoptosis and G2/M arrest induced by *Allium ursinum* (ramson) watery extract in an AGS gastric cancer cell line. OncoTargets and Therapy, 6, 2013, 779-783.
2851. Yang FL, Zhu F, Lei CL. Insecticidal activities of garlic substances against adults of grain moth, *Sitotroga cerealella* (Lepidoptera: Gelechiidae). Insect Science, 19, 2012, 205-212.
2852. Youssef, Amany S., et al. "Microorganisms Isolated from Surgical Wounds Infection and Treatment with Different Natural Products and Medications." International Journal of Medical and Health Sciences 7.6 (2013): 236-239.
2853. Zadeh Hosseingholi, Elahesh, et al. "In silico identification and characterization of antineoplastic asparaginase enzyme from endophytic bacteria." IUBMB life 72.5 (2020): 991-1000.
2854. Zhang Wanping, & Zhao Li. (2012). Antibacterial effects of garlic extract and root exudates on three soil-borne pathogens. Chinese Vegetables, (01X), 66-71.
2855. Zardzewiały, M., Matłok, N., Piechowiak, T., Antos, P. and Balawejder, M., 2023. The Impact of Convection Drying Temperature on Content of Selected Phytochemicals in Dried Wild Garlic Leaves (*Allium ursinum* L.). Acta Universitatis Cinbinesis, Series E: Food Technology, 27(1).
2856. Zhu, X., Zhang, F., Zhou, L., et al. Diallyl trisulfide attenuates carbon tetrachloride-caused liver injury and fibrogenesis and reduces hepatic oxidative stress in rats. Naunyn-Schmiedeberg's Archives of Pharmacology, 2014, 387(5), 445-455.
- 93). Bengoechea, J.A., NAJDENSKI, H., Skurnik, M. Lipopolysaccharide O antigen status of *Yersinia enterocolitica* O:8 is essential for virulence and absence of O antigen affects the expression of other *Yersinia* virulence factors (2004) Mol. Microbiol. 52 (2), 451-469.**
2857. Abeyrathne, P.D., Daniels, C., Poon, K.K.H. et al. Functional characterization of WaaL, a ligase associated with linking O-antigen polysaccharide to the core of *Pseudomonas aeruginosa* lipopolysaccharide (2005). J. Bacteriol., 187 (9), 3002-3012.
2858. Ahrens, P. et al. Characterization and biological role of the O-polysaccharide gene cluster of *Yersinia enterocolitica* serotype O:9. J. Bacteriol 189.20 (2007):7244.
2859. Aich, Udayanath, K.J. Yarema. Glycobiology and immunology. Carbohydrate-Based Vaccines and Immunotherapies. Hoboken, New Jersey: John Wiley & Sons (2009):1-54.
2860. Alenezi, D. Comparative pathogenesis of *Yersinia enterocolitica* biotypes. Doctoral dissertation. Nottingham Trent University, 2015.
2861. Asad, Z. Role of iNOS in septic pulmonary microvascular endothelial cell activation. PhD thesis, University of Ontario (2013).
2862. Asadishad, B., S. Ghoshal, N. Tufenkji. Role of cold climate and freeze-thaw on the survival, transport, and virulence of *Yersinia enterocolitica*. Environ. Sci. Technol., 2013, 47 (24), 14169-14177.

2863. Augustin, D. K., et al. Presence or absence of lipopolysaccharide O antigens affects type III secretion by *Pseudomonas aeruginosa*. J. Bacteriol., 189.6 (2007): 2203-2209.
2864. Augustin, D.K. Identifying mechanism of traversal of corneal epithelial cells by *Pseudomonas aeruginosa*. PhD Thesis, University of California, Berkeley, 2011.
2865. Bartra, S.S. et al. Resistance of *Yersinia pestis* to complement-dependent killing is mediated by the Ail outer membrane protein. Infect. Immun., 76.2 (2008):612-622.
2866. Beims, Hannes, et al. "Discovery of Paenibacillus larvae ERIC V: Phenotypic and genomic comparison to genotypes ERIC I-IV reveal different inventories of virulence factors which correlate with epidemiological prevalences of American Foulbrood." International Journal of Medical Microbiology 310.2 (2020): 151394.
2867. Biedzka-Sarek, M., Molecular Details of Serum Resistance of *Yersinia enterocolitica*. Doctoral dissertation. University of Helsinki, Faculty of Medicine, Haartman Institute, Bacteriology and Immunology, 2008.
2868. Böer, T., Bengelsdorf, F.R., Bömeke, M., Daniel, R. and Poehlein, A., 2023. Genome-based metabolic and phylogenomic analysis of three *Terrisporobacter* species. Plos one, 18(10), p.e0290128.
2869. Bozcal, E. et al. LuxCDE-luxAB-based promoter reporter system to monitor the *Yersinia enterocolitica* O: 3 gene expression in vivo. PloS one 12.2 (2017):e0172877.
2870. Bozcal, E. A general view on virulence determinants and infection strategies of *Yersinia enterocolitica*. Minerva Biotechnol., 32.1 (2020):29-37.
2871. Bravo, D., Silva, C., Carter, J.A. et al. Growth-phase regulation of lipopolysaccharide O-antigen chain length influences serum resistance in serovars of *Salmonella* (2008) J. Med. Microbiol., 57 (8), 938-946.
2872. Byvalov, A. A., V. L. Kononenko, I. V. Konyshchev. Effect of lipopolysaccharide O-side chains on the adhesiveness of *Yersinia pseudotuberculosis* to J774 macrophages as revealed by optical tweezers. Appl. Biochem. Microbiol., 53.2 (2017):258-266.
2873. Byvalov, A.A., V.L. Kononenko, I.V. Konyshchev. Single-cell force spectroscopy of interaction of lipopolysaccharides from *Yersinia pseudotuberculosis* and *Yersinia pestis* with J774 macrophage membrane using optical tweezers. Biochemistry (Moscow), Supplement Series A: Membrane and Cell Biology 12.2 (2018): 93-106.
2874. Byvalov, A.A. et al. Single-cell force spectroscopy of interaction of lipopolysaccharide from *Yersinia pseudotuberculosis* and *Yersinia pestis* with j774 macrophage using optical tweezers", Biologicheskie Membrany 35.2(2018):115-130.
2875. Canals, R. et al. The UDP N-acetylgalactosamine 4-epimerase gene is essential for mesophilic *Aeromonas hydrophila* serotype O34 virulence. Inf. Immun., 74.1 (2006):537-548.
2876. Carlsson, K.E., Liu, J., Edqvist, P.J., Francis, M.S. Influence of the Cpx extracytoplasmic-stress-responsive pathway on *Yersinia* sp.-eukaryotic cell contact (2007) Infection and Immunity, 75 (9), pp. 4386-4399.
2877. Changchang, X.C., B. Liu, B. Hu, et al. Biochemical characterization of UDP-gal:glcnac-pyrophosphate-lipid β -1,4-galactosyltransferase WfeD, a new enzyme from *Shigella boydii* type 14 that catalyzes the second step in O-antigen repeating-unit synthesis. J. Bacteriol., 2011, 193, 2 449-459
2878. Crimmins, Gregory T., et al. "Identification of MrtAB, an ABC transporter specifically required for *Yersinia pseudotuberculosis* to colonize the mesenteric lymph nodes." PLoS Pathog 8.8 (2012): doi:10.1371/journal.ppat.1002828.

2879. Czuchry, D., W.A. Szarek, I. Brockhausen. Identification and biochemical characterization of WbwB, a novel UDP-Gal: Neu5Ac-R α 1, 4-galactosyltransferase from the intestinal pathogen *Escherichia coli* serotype O104. *Glycoconjugate J.*, 35.1 (2018): 65-76.
2880. Czuchry, Diana, Walter A. Szarek, and Inka Brockhausen. "Identification and biochemical characterization of WbwB, a novel UDP-Gal: Neu5Ac-R α 1, 4-galactosyltransferase from the intestinal pathogen *Escherichia coli* serotype O104." *Glycoconjugate journal* (2017): 1-12.
2881. de Melo, Adma Nadja Ferreira, et al. "Genomic investigation of antimicrobial resistance determinants and virulence factors in *Salmonella enterica* serovars isolated from contaminated food and human stool samples in Brazil." *International Journal of Food Microbiology* 343 (2021): 109091.
2882. DebRoy, C., E. Roberts, and P.M. Fratamico. Detection of O antigens in *Escherichia coli*. *Anim. Health Res. Rev.*, 2011, 12 :169-185.
2883. Dhar, Mahesh Shanker, and Jugsharan Singh Viridi. "Strategies used by *Yersinia enterocolitica* to evade killing by the host: thinking beyond Yops." *Microbes and infection* 16.2 (2014): 87-95.
2884. Di Lorenzo, F., De Castro, C., Lanzetta, R. et al. Lipopolysaccharides as microbe-associated molecular patterns: A structural perspective RSC Drug Discovery Series, 2015-43, 38-63
2885. Domfeh, S., Amoah, M., Sefa, L., 2023. Comparative study of Widal test to stool culture for the diagnosis of suspected typhoid fever: a study in a primary health centre, Ghana. *Microbes Inf. Dis.*, 4(4), 1409-1415.
2886. Dow, G.T. et al. Structural investigation on WlaRG from *Campylobacter jejuni*: A sugar aminotransferase. *Protein Science* 26.3 (2017):586-599.
2887. Dudek, B. et al. Proteomic analysis of outer membrane proteins from *Salmonella Enteritidis* strains with different sensitivity to human serum. *PloS one* 11.10 (2016):e0164069.
2888. Duncombe, Lucy, et al. The tip of *Brucella* O-polysaccharide is a potent epitope in response to Brucellosis infection and enables short synthetic antigens to be superior diagnostic reagents. *Microorganisms* 10.4 (2022):708.
2889. Fàbrega, A., Ballesté-Delpierre, C., Vila, J. (2015). Antimicrobial Resistance in *Yersinia enterocolitica*. *Antimicrobial Resistance and Food Safety: Methods and Techniques*, 77-104.
2890. Fàbrega, A., J. Vila. *Yersinia enterocolitica*: pathogenesis, virulence and antimicrobial resistance. *Enfermedades infecciosas y microbiología clínica* 30.1 (2012):24-32.
2891. Fälker, Stefan, et al. "Overproduction of DNA adenine methyltransferase alters motility, invasion, and the lipopolysaccharide O-antigen composition of *Yersinia enterocolitica*." *Infection and immunity* 75.10 (2007): 4990-4997.
2892. Feodorova, V.A., Golova, A.B. Antigenic and phenotypic modifications of *Yersinia pestis* under calcium and glucose concentrations simulating the mammalian bloodstream environment. (2005). *J. Med. Microbiol.*, 54 (5), 435-441.
2893. Focà A, Liberto MC, Quirino A, et al. Lipopolysaccharides: from erinyes to charites. *Mediators Inflamm.*, 2012, doi:10.1155/2012/684274.
2894. Francis, M.S. Secretion systems and metabolism in the pathogenic yersiniae. In. *Adv. Mol.Cell. Microbiol.: Stress Response in Pathogenic Bacteria*. CABI Publishing, 2011,

182-220.

2895. Futoma-Kołoch, B., G. Bugla-Płoskońska, J. Sarowska. Searching for outer membrane proteins typical of serum-sensitive and serum-resistant phenotypes of *Salmonella*." *Salmonella-Distribution, Adaptation, Control Measures, and Molecular Technologies*; Annous, BA, Ed (2012):265-290.
2896. Fux A.C., Casonato Melo C., Michelini S. et al. Heterogeneity of lipopolysaccharide as source of variability in bioassays and LPS-binding proteins as remedy. *Int. J. Mol. Sci.*, 24 (9), art. no. 8395, 2023.
2897. Goebel, Elizabeth M., et al. "O antigen protects *Bordetella parapertussis* from complement." *Infection and immunity* 76.4 (2008): 1774-1780.
2898. Goldstone, R.J. Investigating the relationship between quorum sensing, motility, and the type 3 secretion system of *Yersinia pseudotuberculosis*. PhD Thesis, University of Nottingham, 2012.
2899. Grabenstein, J.P. (2005). Identification and characterization of genes involved in survival and replication of *Yersinia pestis* and *Yersinia pseudotuberculosis* in macrophages. Doctoral dissertation, State University of New York at Stony Brook.
2900. Gu, W, Wang, X, Qiu, H, et al. Comparative antigenic proteins and proteomics of pathogenic *Yersinia enterocolitica* bio-serotypes 1B/O: 8 and 2/O: 9 cultured at 25°C and 37°C. *Microbiol. Immunol.*, 2012, 56 (9), 583-594.
2901. Gu, W.P. et al. Comparison of lipopolysaccharide and protein immunogens from pathogenic *Yersinia enterocolitica* bio-serotype 1B/O: 8 and 2/O: 9 using SDS-PAGE." *Biomed. Env.Sci.*, 25.3 (2012):282-290.
2902. Gutierrez, J. G., et al. Interleukin-12p40 contributes to protection against lung injury after oral *Yersinia enterocolitica* infection. *Inflamm. Res.*, 57.11 (2008):504-511.
2903. Haiko, J. et al. Invited review: breaking barriers—attack on innate immune defences by omptin surface proteases of enterobacterial pathogens. *Innate Immun.*, 15.2 (2009):67-80.
2904. Hallanvuori, S. (2009). Foodborne *Yersinia*: Identification and molecular epidemiology of isolates from human infections. Doctoral dissertation, University of Helsinki.
2905. Hallanvuori, S. Foodborne *Yersinia*: identification and molecular epidemiology of isolates from human infections. Doctoral Dissertation, University of Helsinki, 2009.
2906. Halvorsen, T.M. (2020). Complexities in crossing membrane barriers: new members of the CdiA and CDI ionophore protein families reveal novel mechanisms for receptor-binding and intoxication of target cells. Doctoral Dissertation, University of California, Santa Barbara.
2907. Heisdorf, C. J., Griffiths, W. A., Thoden, J. B. et al. (2021). Investigation of the enzymes required for the biosynthesis of an unusual formylated sugar in the emerging human pathogen *Helicobacter canadensis*. *Protein Science*, 30(10), 2144-2160.
2908. Hoare, A. et al. The outer core lipopolysaccharide of *Salmonella enterica* serovar Typhi is required for bacterial entry into epithelial cells. *Inf. Immun.*, 74.3 (2006):1555-1564.
2909. Holzer, S. Funktionelle analyse von typ 3 sekretionssystemen in *Salmonella enterica*. Der Naturewissenschaftlichen Fakultät der Friedrich-Alexander Universität Erlangen-Nürnberg, Doctoral dissertation, 2010.
2910. Hong, L.. Biochemical studies of the enzymes involved in deoxysugar D-forosamine biosynthesis. Doctoral dissertation, University at Texas at Austin, 2004.
2911. Ieva, R. Interfering with outer membrane biogenesis to fight Gram-negative bacterial

- pathogens. *Virulence* (2017):1-4.
2912. Ilg, K.C. Glycoengineering and Glycomimicry: *Campylobacter jejuni* carbohydrate structures on *Salmonella enterica* serovar Typhimurium. A dissertation submitted to ETH Zurich for the degree of Doctor of Sciences, 2009.
 2913. Ilg, Karin, et al. "O-antigen-negative *Salmonella enterica* serovar Typhimurium is attenuated in intestinal colonization but elicits colitis in streptomycin-treated mice." *Infection and immunity* 77.6 (2009): 2568-2575.
 2914. Jun, J.W., Park, S C., Wicklund, A. et al. (2018). Bacteriophages reduce *Yersinia enterocolitica* contamination of food and kitchenware. *Int. J. Food Microbiol.*, 271, 33-47.
 2915. Kakoschke, T.K. (2021). Role of the RNA chaperone Hfq in the virulence of *Yersinia enterocolitica* (Doctoral dissertation, lmu).
 2916. Kasperkiewicz, K Swierzko, A.S. Bartlomiejczyk, M.A. Cedzynski, M., Noszczyńska, M., Duda, K.A., Michalski, M., Skurnik, M. Interaction of human mannose-binding lectin (MBL) with *Yersinia enterocolitica* lipopolysaccharide *International Journal of Medical Microbiology* 2015, 305, 544-552.
 2917. Kaszowska M., Jachymek W., Lukasiewicz J., et al. The unique structure of complete lipopolysaccharide isolated from semi-rough *Plesiomonas shigelloides* O37 (strain CNCTC 39/89) containing (2S)-O-(4-oxopentanoic acid)- α -D-Glcp (α -D-Lenose). *Carbohydr. Res.*, 2013, 378, 98-107.
 2918. Kenyon, J.J., M.M. Cunneen, P.R. Reeves. Genetics and evolution of *Yersinia pseudotuberculosis* O-specific polysaccharides: a novel pattern of O-antigen diversity. *FEMS Microbiol. Rev.*, 41.2 (2017):200-217.
 2919. Leskinen, K. (2016). The roles of YbeY, RfaH, and Hfq in gene regulation and virulence of *Yersinia enterocolitica*O: 3. Doctoral dissertation, University of Helsinki.
 2920. Lewis, V.G., M.P. Ween, C.A. McDevitt. The role of ATP-binding cassette transporters in bacterial pathogenicity. *Protoplasma* 249.4 (2012):919-942.
 2921. Li, Y. et al. Structural and genetic relationships of two pairs of closely related O-antigens of *Escherichia coli* and *Salmonella enterica*: *E. coli* O11/S. enterica O16 and *E. coli* O21/S. enterica O38. *FEMS Immunol. & Med. Microbiol.*, 61.3 (2011):258-268.
 2922. Li, D.G., Liu, B., Zhou, D.W. (2013). Structural characterization of enzymatic products in the dTDP-d-Qui4Nfo biosynthetic pathway using electrospray ionization tandem mass spectrometry. *Rapid Communicat. Mass Spectrometry*, 27(6), 681-690.
 2923. Liles, M.R., Terhune, J.S., Newton, J.C. et al (2016). U.S. Patent No. 9,492,521. Washington, DC: U.S. Patent and Trademark Office.
 2924. Liu Y., Koudelka G. O-Polysaccharides of LPS Modulate *E. coli* Uptake by *Acanthamoeba castellanii*. *Microorganisms*, 11 (6), art. no. 1377. 2023.
 2925. Liu, B. et al. Structural and genetic relationships between the O-antigens of *Escherichia coli* O118 and O151. *FEMS Immunol. & Med. Microbiol.*, 60.3 (2010):199-207.
 2926. Liu, D. (Ed.). (2011). Molecular detection of human bacterial pathogens. CRC press.
 2927. Liu, B. et al. Structural diversity in *Salmonella* O antigens and its genetic basis. *FEMS Microbiol. Rev.*, 38.1 (2014):56-89.
 2928. Liu, B. et al. Structure and genetics of *Escherichia coli* O antigens." *FEMS Microbiol. Rev.*, 44.6 (2020):655-683.
 2929. Liu, B. et al. Structure and genetics of *Shigella* O antigens. *FEMS Microbiol. Rev.*, 32.4 (2008):627-653.

2930. Liu, Y., 2023. Carbohydrates of lipopolysaccharide: *Acanthamoeba castellanii* predation determinant and bacterial defense strategy (Doctoral dissertation, State University of New York at Buffalo).
2931. Llobet, E. et al. *Klebsiella pneumoniae* OmpA confers resistance to antimicrobial peptides. *Antimicrob. Agents Chemother.*, 53.1 (2009):298-302.
2932. Marradi, M., Chiodo, F., García, I. (2015). Glyconanotechnology and disease: Gold nanoparticles coated with glycosides as multivalent systems for potential applications in diagnostics and therapy, 5, 89-131.
2933. McGiven, J. Immunoselection and structural evaluation of *Brucella* O-polysaccharide epitopes and their application to the serodiagnosis of bovine brucellosis. (2013).
2934. McNally, A. et al. An aflagellate mutant *Yersinia enterocolitica* biotype 1A strain displays altered invasion of epithelial cells, persistence in macrophages, and cytokine secretion profiles in vitro. *Microbiology* 153.5 (2007):1339-1349.
2935. Meng, J., Xu, J., Huang, C. et al. (2020). Rcs phosphorelay responses to truncated lipopolysaccharide-induced cell envelope stress in *Yersinia enterocolitica*. *Molecules*, 25(23), 5718.
2936. Miguel AV, Hanuszkiewicz A. Proteins involved in the membrane translocation of lipopolysaccharide O antigen. *Mini-Rev. Org. Chem.*, 2012, 9 (3), 261-269.
2937. Monahan, A. M., J. J. Callanan, and J. E. Nally. "Host-pathogen interactions in the kidney during chronic leptospirosis." *Veterinary pathology* 46.5 (2009): 792-799.
2938. Murray, Gerald L., et al. "Mutations affecting *Leptospira interrogans* lipopolysaccharide attenuate virulence." *Molecular microbiology* 78.3 (2010): 701-709.
2939. Nally, Jarlath E., et al. "Changes in lipopolysaccharide O antigen distinguish acute versus chronic *Leptospira interrogans* infections." *Infection and immunity* 73.6 (2005): 3251-3260.
2940. Nieckarz, M. et al. The role of OmpR in the expression of genes of the KdgR regulon involved in the uptake and depolymerization of oligogalacturonides in *Yersinia enterocolitica*. *Front. Cell. Inf. Microbiol.*, 7 (2017):366.
2941. Pal, M., Lema, A.G. (2022). Current perceptive on the virulence factors of *Yersinia enterocolitica*: a critical review. *Am. J. Inf. Dis.*, 10(1), 7-10.
2942. Paunova-Krasteva, T., Stoitsova, S.R., Topouzova-Hristova, T. et al. *Escherichia coli* O157: Effects of growth temperature on concanavalin a binding and the adherence to cultured cells, *Compt. Rend. Acad. Bulg. Sci.*, 2014, 67, 3, 349-354
2943. Paulson, A.R. (2020). Temperature-and host-dependent transcriptional responses in the entomopathogenic bacterium, *Yersinia entomophaga* MH96: a thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Genetics at Massey University, Albany Campus, New Zealand (Doctoral dissertation, Massey University).
2944. Perez, M.F. et al. First report on the plasmidome from a high-altitude lake of the Andean Puna. *Front. Microbiol.*, 11 (2020):1343.
2945. Pérez-Gutiérrez, C. et al. Role of lipid A acylation in *Yersinia enterocolitica* virulence. *Infect. Immun.*, 78.6 (2010):2768-2781.
2946. Pieretti, G. et al. O-chain structure from the lipopolysaccharide of the human pathogen *Halomonas stevensii* strain S18214. *Carbohyd. Res.*, 346.2 (2011):362-365.
2947. Pieretti, G. et al. Structural determination of the O-specific polysaccharide from

- Aeromonas hydrophila* strain A19 (serogroup O: 14) with S-layer. Carbohydr. Res., 346.15 (2011):2519-2522.
2948. Pinta, E. Biosynthesis of *Yersinia enterocolitica* serotype O: 3 lipopolysaccharide outer core. PhD Thesis, University of Turku (Finland), 2010.
 2949. Pinta, Elise, et al. "Characterization of the six glycosyltransferases involved in the biosynthesis of *Yersinia enterocolitica* serotype O: 3 lipopolysaccharide outer core." Journal of Biological Chemistry 285.36 (2010): 28333-28342.
 2950. Pinta, Elise, et al. "Identification of three oligo-/polysaccharide-specific ligases in *Yersinia enterocolitica*." Molecular microbiology 83.1 (2012): 125-136.
 2951. Rapisavoli, Jeannette N., et al. "O antigen modulates insect vector acquisition of the bacterial plant pathogen *Xylella fastidiosa*." Applied and environmental microbiology 81.23 (2015): 8145-8154.
 2952. Regué, Susana Merino, and Juan M. Tomás. "The UDP."
 2953. Reines, M., E. Llobet, K.M. Dahlström, C. Perez-Gutierrez, C.M. Llompарт, N. Torrecabota, T.A. Salminen and J.A. Bengoechea. Deciphering the Acylation Pattern of *Yersinia enterocolitica* Lipid A. PLoS Pathogens. 2012, 8(10).
 2954. Reuter, Sandra, et al. "Directional gene flow and ecological separation in *Yersinia enterocolitica*." Microbial genomics 1.3 (2015).
 2955. Robert, G.J. Investigating the relationship between quorum sensing, motility, and the type 3 secretion system of *Yersinia pseudotuberculosis*. PhD thesis, University of Nottingham, 2012.
 2956. Robins-Browne, R.M. *Yersinia enterocolitica*. Food Microbiology: fundamentals and frontiers (2012): 339-376.
 2957. Robins-Browne, R.M. (2001). *Yersinia enterocolitica*. In Microbiología de los alimentos: Fundamentos y fronteras (pp. 199-223). Acribia.
 2958. Ruan, X. M.A. Valvano. In vitro O-antigen ligase assay. Glycosyltransferases. Humana Press, Totowa, NJ, 2013. 185-197.
 2959. Ruan, Xiang, et al. "The WaaL O-antigen lipopolysaccharide ligase has features in common with metal ion-independent inverting glycosyltransferases." Glycobiology 22.2 (2012): 288-299.
 2960. Salinger, A.J., et al. Biochemical studies on WbcA, a sugar epimerase from *Yersinia enterocolitica*. Protein Sci., 24.10 (2015):1633-1639.
 2961. Sánchez-Gómez, S., R. Conde-Alvarez, J. Antonio, et al. Modifications in lipopolysaccharide that reduce interaction of bacterial pathogens with the innate immune system and cause resistance to antimicrobial peptides. In. Antimicrobial Peptides: Properties, Functions and Role in Immune Response. Nova Science Publishers, Inc., 2013, 1-30.
 2962. Saticioglu, Izzet Burcin, Muhammed Duman, and Soner Altun. "Genome analysis and antimicrobial resistance characteristics of *Chryseobacterium aquaticum* isolated from farmed salmonids." Aquaculture 535 (2021): 736364.
 2963. Schütz, M., et al. "Trimer stability of YadA is critical for virulence of *Yersinia enterocolitica*." Infection and immunity 78.6 (2010): 2677-2690.
 2964. Seabaugh, J.A., & Anderson, D.M. (2024). Pathogenicity and virulence of *Yersinia*. Virulence, 15(1), 2316439.
 2965. Senchenkova, S.N., Feng, L., Wang, Q., Perepelov, A.V., Qin, D., Shevelev, S.D., Ren, Y., Shashkov, A.S., Knirel, Y.A., Wang, L. Structural and genetic characterization of

- Shigella boydii* type 17 O antigen and confirmation of two new genes involved in the synthesis of glucolactilic acid (2006) Biochemical and Biophysical Research Communications, 349 (1), pp. 289-295.
2966. Skorek, K., A. Raczowska, B. Dudek, et al. Regulatory protein OmpR influences the serum resistance of *Yersinia enterocolitica* O: 9 by modifying the structure of the outer membrane. PloS one, 2013, DOI: 10.1371/journal.pone.0079525
2967. Sofya, N.S. et al. Structural and genetic characterization of *Shigella boydii* type 17 O antigen and confirmation of two new genes involved in the synthesis of glucolactilic acid. Biochem. Biophys. Res. Commun., 349.1 (2006):289-295.
2968. Spahich, N.A. A tale of two proteins: insights into the *Haemophilus influenzae* Hap and Hia. Doctoral dissertation, Duke University, 2011
2969. Spahich, N.A. et al. Inactivation of *Haemophilus influenzae* lipopolysaccharide biosynthesis genes interferes with outer membrane localization of the hap autotransporter. J. Bacteriol., 194.7 (2012):1815-1822.
2970. Stanton C.R., Batinovic S., Petrovski S. Burkholderia contaminans bacteriophage CSP3 requires O-Antigen polysaccharides for infection. Microbiol. Spectrum, 11(3) 2023.
2971. Sterba, J., Dupejova, J., Fiser, M. et al. (2011). Fibrinogen-related proteins in ixodid ticks. Parasites & vectors, 4, 1-10.
2972. Stevenson, R. M. W. Investigating survival mechanisms of *Yersinia ruckeri* in rainbow trout, *Oncorhynchus mykiss*. Doctoral dissertation, 2008.
2973. Storz, E. Über die Antibiotikaresistenz bei *Yersinia enterocolitica*: in vitro und in vivo Untersuchungen (Doctoral dissertation, München, Ludwig-Maximilians-Universität, 2018).
2974. Suomalainen, M. Molecular factors affecting the activity and substrate selectivity of the Pla protease of *Yersinia pestis*. Doctoral Dissertation, University of Helsinki, 2014.
2975. Suomalainen, Marjo. "Molecular Factors Affecting the Activity and Substrate Selectivity of the Pla Protease of *Yersinia pestis*." (2014).
2976. Tang, Gaoyan, Keith P. Mintz. Glycosylation of the collagen adhesin EmaA of *Aggregatibacter actinomycetemcomitans* is dependent upon the lipopolysaccharide biosynthetic pathway. J. Bacteriol., 192.5 (2010):1395-1404.
2977. Tang, G. et al. Lipopolysaccharides mediate leukotoxin secretion in *Aggregatibacter actinomycetemcomitans*. Mol. Oral Microbiol., 27.2 (2012):70-82.
2978. Taylor, P.D. Computational prediction of bacterial protein subcellular location and characterisation of transmembrane protein topology. PhD thesis, The University of Manchester (United Kingdom), 2004.
2979. Teng, Lin, et al. "Phage controlling method against novel freshwater-derived *Vibrio parahaemolyticus* in ready-to-eat crayfish (*Procambarus clarkii*)." Food Research International 162 (2022): 111986.
2980. Thoden, J.B. et al. Bacterial sugar 3, 4-ketoisomerases: structural insight into product stereochemistry. Biochemistry 54.29 (2015):4495-4506.
2981. Thomas, Rebecca M., et al. "The immunologically distinct O antigens from *Francisella tularensis* subspecies *tularensis* and *Francisella novicida* are both virulence determinants and protective antigens." Infection and immunity 75.1 (2007): 371-378.
2982. Thomson, N.R., et al. Directional gene flow and ecological separation in *Yersinia enterocolitica*. Microb. Genomics (2015), ISSN:2057-5858.
2983. Tils, D.V. (2015). Untersuchung und Charakterisierung der Typ-II-Sekretionssysteme

- von *Yersinia enterocolitica* (Doctoral dissertation, Universität Münster, 2015).
2984. Tran, E. N. H., Papadopoulos, M., & Morona, R. (2014). Relationship between O-antigen chain length and resistance to colicin E2 in *Shigella flexneri*. *Microbiology*, 2014, 160(Pt 3), 589-601.
 2985. Troughs, Water. "Escherichia coli Involvement of the."
 2986. Uliczka F, Dersch P. Unique virulence properties of *Yersinia enterocolitica* O:3. *Adv. Yersinia Res./Adv. Exper. Med. Biol.*, 2012, 954, 281-287.
 2987. Uliczka, Frank, et al. "Unique cell adhesion and invasion properties of *Yersinia enterocolitica* O: 3, the most frequent cause of human Yersiniosis." *PLoS Pathog* 7.7 (2011): e1002117.
 2988. Valentin-Weigand, P., Heesemann, J., & Dersch, P. (2014). Unique virulence properties of *Yersinia enterocolitica* O: 3—An emerging zoonotic pathogen using pigs as preferred reservoir host. *International Journal of Medical Microbiology*, 304(7), 824-834.
 2989. Valvano MA, Hanuszkiewicz A. Proteins involved in the membrane translocation of lipopolysaccharide O antigen. *Mini-Rev. Org. Chem.*, 2012, 9 (3), 261-269.
 2990. Vilches, S. et al. *Aeromonas hydrophila* AH-3 type III secretion system expression and regulatory network. *Appl. Env. Microbiol.*, 75.19 (2009):6382-6392.
 2991. Vinogradov EV., Bogdanove AJ. Requirement of the lipopolysaccharide O-chain biosynthesis gene *wxocB* for type III secretion and virulence of *Xanthomonas oryzae* pv. *Oryzicola* Wang, L. J. *Bacteriol.*, 9, 195, 2013, 1959-1969.
 2992. Vogel, Ulrike, Koen Beerens, and Tom Desmet. "Nucleotide sugar dehydratases: structure, mechanism, substrate specificity and application potential." *Journal of Biological Chemistry* (2022): 101809.
 2993. von Tils, Dominik. Untersuchung und Charakterisierung der Typ-II-Sekretionssysteme von *Yersinia enterocolitica*. Diss. Universitäts-und Landesbibliothek Münster, 2015.
 2994. Wang, Lei, Quan Wang, and Peter R. Reeves. "The variation of O antigens in gram-negative bacteria." *Endotoxins: Structure, Function and Recognition*. Springer, Dordrecht, 2010. 123-152.
 2995. Wang, Li, et al. Novel candidate virulence factors in rice pathogen *Xanthomonas oryzae* pv. *oryzicola* as revealed by mutational analysis. *Appl. Env. Microbiol.*, 73.24 (2007):8023-8027.
 2996. Wang, Li, Evgeny V. Vinogradov, and Adam J. Bogdanove. "Requirement of the lipopolysaccharide O-chain biosynthesis gene *wxocB* for type III secretion and virulence of *Xanthomonas oryzae* pv. *Oryzicola*." *Journal of bacteriology* 195.9 (2013): 1959-1969.
 2997. Wang, Q. et al. Identification of the two glycosyltransferase genes responsible for the difference between *Escherichia coli* O107 and O117 O-antigens. *Glycobiol.*, 22.2 (2012): 281-287.
 2998. Wang, H., Shi, C., Yang, B. et al. (2024). Characterization of the genome and cell invasive phenotype of *Vibrio diabolus* Cg5 isolated from mass mortality of Pacific oyster, *Crassostrea gigas*. *Microbial Pathogenesis*, 186, 106466.
 2999. Weirich, Johanna, et al. "Identifying components required for OMP biogenesis as novel targets for anti-infective drugs." *Virulence* (2017): 1-20.
 3000. Xu CC, Liu B, Hu B, et al. Biochemical characterization of UDP-Gal:GlcNAc-pyrophosphate-lipid beta-1,4-galactosyltransferase WfeD, a new enzyme from *Shigella boydii* type 14 that catalyzes the second step in O-antigen repeating-unit synthesis. *J.*

- Bacteriol., 2011, 193, 2, 449-459.
3001. Xu, Yy, Cy Dong, and Dw Zhou. Characterization of biosynthetic pathway of the dtdp-l-rha by electrospray ionization tandem mass spectrometry. J. Chinese Mass Spectrometry Soc., 37.1 (2016):17-22.
 3002. Xue, Ying, et al. "Proteus mirabilis targets atherosclerosis plaques in human coronary arteries via DC-SIGN (CD209)." Frontiers in Immunology 11 (2020).
 3003. Yang, K., He, Y., Park, C. G. Et al. (2019). Yersinia pestis interacts with SIGNR1 (CD209b) for promoting host dissemination and infection. Frontiers in immunology, 10, 96.
 3004. Yi, Xuan. Genetic analysis of c-di-GMP signaling and virulence in Dickeya dadantii 3937. Doctoral dissertation, The University of Wisconsin-Milwaukee, 2009.
 3005. Yoon, J.W. et al. Involvement of the Escherichia coli O157: H7 (pO157) ecf operon and lipid A myristoyl transferase activity in bacterial survival in the bovine gastrointestinal tract and bacterial persistence in farm water troughs. Inf. Immun., 73.4 (2005):2367-2378.
 3006. Yoon, J.W., Kwon, K.H., Park, Y. H. (2013). Whole genome-scale transcriptome analysis of the amyristolyated enterohemorrhagic *Escherichia coli* O157: H7. J. Prevent. Vet. Med., 37(4), 153-162.
 3007. Zhang, Pei, et al. "Human dendritic cell-specific intercellular adhesion molecule-grabbing nonintegrin (CD209) is a receptor for Yersinia pestis that promotes phagocytosis by dendritic cells." Infection and immunity 76.5 (2008): 2070-2079.
 3008. Zhang, Y., Li, X., Qi, X., Jiang, R., Guo, L., Zhang, R., & Li, Y. Identification and functional analysis of the gene ste9 involving in Ebosin biosynthesis from Streptomyces sp. 139. FEMS Microbiol. Lett., 2014, 350(2), 257-264.
 3009. Бывалов, А.А., Кононенко, В.Л., Коньшев, И.В. (2017). Влияние О-боковых цепей липополисахарида на адгезивность *Yersinia pseudotuberculosis* к макрофагам J774, установленное методом оптической ловушки. Прикладная биохимия и микробиология, 53(2), 234-243.
 3010. Бывалов, А.А., Кононенко, В.Л., Коньшев, И.В. (2018). Исследование взаимодействия липополисахаридов *Yersinia pseudotuberculosis* и *Yersinia pestis* с мембраной макрофага J774 методом силовой спектроскопии с использованием оптического пинцета. Биологические мембраны, 35(2), 115-130.
 3011. Дентовская, С.В. (2012). Молекулярно-генетические механизмы образования и функциональная значимость липополисахарида *Yersinia pestis* (Doctoral dissertation, диссертация на соискание ученой степени доктора медицинских наук: 03.02. 03-микробиология 03.01. 04—биохимия. М).
 3012. Каримова, Т.В. Энтеропатогенные иерсинии: микробиологический мониторинг, молекулярно-биологические особенности, алгоритм лабораторной диагностики. Диссертация, Иркутск, 2017.
 - 94). Batovska, D., Parushev, St., Slavova, A., Bankova, V., Tsvetkova, I., Ninova, M., NAJDENSKI, H. Study on the substituents' effects of a series of synthetic chalcones against the yeast Candida albicans (2007) European Journal of Medicinal Chemistry, 42 (1), pp. 87-92.**
 3013. Agent, P A.I. (2015). Siti Munirah Mohd Faudzi.
 3014. Ahmad, K. et al. Synthesis and spectroscopic characterization of medicinal azo

- derivatives and metal complexes of Indandion. *J. Mol. Struct.*, 1198 (2019):126885.
3015. Aksöz, B.E., & Ertan, R. (2012). Spectral properties of chalcones II. *Fabad J. Pharm. Sci.*, 37(4), 205-216.
 3016. Alberton, E. Influence of chalcone analogues, xanthenes and monosaccharides on glycemia in an experimental animal model, Ph.D. thesis, Federal University of Santa Catarina, Brazil, 2007.
 3017. Andrade J.T., Santos F.R.S., Lima W.G. et al. Design, synthesis, biological activity and structure-activity relationship studies of chalcone derivatives as potential anti-Candida agents. *J. Antibiot.*, 71 (8), 702-712, 2018.
 3018. Arshad, L. et al. Immunosuppressive effects of natural α , β -unsaturated carbonyl-based compounds, and their analogs and derivatives, on immune cells: A review." *Front. Pharmacol.*, 8 (2017).
 3019. Ashok, D., K. Sudershan, & M. Khalilullah. Solvent-free microwave-assisted synthesis of E-(1)-(6-benzoyl-3,5-dimethylfuro[3',2':4,5]benzo[b]furan-2-yl)-3-(aryl)-2-propen-1-ones and their antibacterial activity. *Green Chem. Lett. Rev.*, 2011. DOI: 10.1080/17518253.2011.584912
 3020. Assolini, J.P. et al. 4-nitrochalcone exerts leishmanicidal effect on *L. amazonensis* promastigotes and intracellular amastigotes, and the 4-nitrochalcone encapsulation in beeswax copaiba oil nanoparticles reduces macrophages cytotoxicity. *Eur. J. Pharmacol.*, 884 (2020):173392.
 3021. Bai X, Shi WO, Chen HF, Zhang P, Li Y, Yin SF. Synthesis and antitumor activity of 1-acetyl-3-(4-phenyl)-4,5-dihydro-2-pyrazoline-5-phenylursolate and 4-chalcone ursolate derivatives. *Chem Nat Compounds*, 48 (1), 2012, 60-65, ISSN 0009-3130
 3022. Banedar, P.N. Derivatives of 1-chloromethyl naphthalene: synthesis and microbiological evaluation as potential antifungal agents. *Der Pharma Chemica*, 2011, 3, 1, 105-111.
 3023. Basic, J., Kalinic, M., Ivkovic, B., Eric, S., Milenkovic, M., Vladimirov, S., Vujic, Z. Synthesis, QSAR analysis and mechanism of antibacterial activity of simple 2'-hydroxy chalcones. *Digest Journal of Nanomaterials and Biostructures* 9, 2014, 1537-1546.
 3024. Bašić, J.V. (2016). Ispitivanje korelacije između hemijske strukture, fizičko-hemijskih i retencionih parametara i antimikrobne aktivnosti novosintetisanih derivata propiofenona. Универзитет у Београду.
 3025. Begum, S., Bharathi, K., & KVSRRG, P. *Journal of Global Trends in Pharmaceutical Sciences*.
 3026. Bist, Ganesh, et al. "Dihydroxylated 2, 6-diphenyl-4-chlorophenylpyridines: Topoisomerase I and II α dual inhibitors with DNA non-intercalative catalytic activity." *European Journal of Medicinal Chemistry* 133 (2017): 69-84.
 3027. Bist, Ganesh, et al. "Inhibition of LPS-stimulated ROS production by fluorinated and hydroxylated chalcones in RAW 264.7 macrophages with structure-activity relationship study." *Bioorganic & Medicinal Chemistry Letters* 27.5 (2017): 1205-1209.
 3028. Bukhari, S.N.A., M. Jasamai and I. Jantan. Synthesis and biological evaluation of chalcone derivatives (mini review). *Mini-Rev. Med. Chem.*, 2012, 12(13), 1394-1403.
 3029. Caboni P., Aissani N., Demurtas M., Ntalli N., Onnis V. Nematicidal activity of acetophenones and chalcones against *Meloidogyne incognita* and structure-activity considerations. *Pest Manag Sci*, 2015 doi: 10.1002/ps.3978.

3030. Campos-Buzzi, F., Correa, R., & Cechinel Filho, V. Antinociceptive Activity of a New Benzofuranone Derived from a Chalcone.
3031. Chen, D., Z. Chen, X. Xiao, Z. Yang, L. Lin, X. Liu, X. Feng. Highly enantioselective michael addition of malonate derivatives to enones catalyzed by an n,n'-dioxide–scandium(iii) complex. *Chemistry – A European Journal*, 2009, 15, 28, 6807-6810.
3032. Chimenti, F., Fioravanti, R., Bolasco, A., Chimenti, P., Secci, D., Rossi, F., Yáñez, M., Orallo, F., Ortuso, F., Alcaro, S. Chalcones: A valid scaffold for monoamine oxidases inhibitors (2009) *Journal of Medicinal Chemistry*, 52 (9), pp. 2818-2824.
3033. CO2CH, E. (2021). 2.2. 1 Unexpected Side Products in the Base-Mediated Castagnoli-Cushman Reaction. *Diastereoselective Reactions of Imines with Anhydrides and their Derivatives and Mechanistic Investigation of the Multicomponent Castagnoli-Cushman Reaction*, 25.
3034. Cordeiro, M.N.S. Síntese e caracterização de chalconas derivadas da 3, 4, 5-trimetoxiacetofenona com potencial atividade antileucêmica. MS Thesis, Universidade Federal De Santa Catarina, Centro De Ciências Físicas E Matemáticas, Departamento De Química. Florianópolis 2010.
3035. Corrêa, R., Fenner, B.P., Buzzi, F.D.C., Cechinel Filho, V., Nunes, R.J. Antinociceptive activity and preliminary structure-activity relationship of chalcone-like compounds (2008) *Zeitschrift fur Naturforschung - Section C Journal of Biosciences*, 63 (11-12), pp. 830-836.
3036. Cushniea, T.P.T., and A.J. Lambb. Recent advances in understanding the antibacterial properties of flavonoids. *Int.. J. Antimicrob. Agents* 38 (2011) 99– 107.
3037. Da Silva, V. Estudo da syntese da chalcona 1(4'-n-fenil-sulfonilamidafenil)-3-(4-metilfenil)-2propen-1-ona, Universidade Estadual de Goiás (ueg), Unidade Universitária de Ciências Exatas e Tecnológicas (UnUCET), ANaPOLIS-GO, 2008.
3038. Daglia, M. Polyphenols as antimicrobial agents. *Curr. Opin. Biotechnol.*, 2012, 23, 2, 174-181.
3039. Damazio, R.G., Zanatta, A.P., Cazarolli, L.H., Chiaradia, L.D., Mascarello, A., Nunes, R.J., Yunes, R.A., Barreto Silva, F.R.M. Antihyperglycemic activity of naphthylchalcones (2010) *European Journal of Medicinal Chemistry*, 45 (4), pp. 1332-1337.
3040. Dangi, L.L., M.S. Dulawat, P. Tiwari, et al. New substituted m-phenoxy chalcones; their synthesis by microwave irradiation and antifungal activity. *Asian J. Res. Chem.*, 2013, 6, 5, 461-463.
3041. Demin, Konstantin A., et al. "Mechanisms of Candida Resistance to Antimycotics and Promising Ways to Overcome It: The Role of Probiotics." *Probiotics and antimicrobial proteins* (2021): 1-23.
3042. Din, Z.Ud, T.P. Fill, F.F. de Assis, D Lazarin-Bidoia, et al. Unsymmetrical 1, 5-diaryl-3-oxo-1, 4-pentadienyls and their evaluation as antiparasitic agents. *Bioorg. Med. Chem.*, 2014, 22, 3, 1121-1127.
3043. Dinakaran VS, Jacob D, Mathew JE.. Synthesis and biological evaluation of novel pyrimidine-2(1H)-ones/thiones as potent anti-inflammatory and anticancer agents. *Med. Chem. Res.*, 12 (11), 2012, 3598-3606.
3044. Domínguez Rivera, R.A. (2021). Síntesis, caracterización y determinación de la actividad biológica de Chalconas y Flavonas (Doctoral dissertation, Universidad de El Salvador).

3045. Eddarir, S, Kajjouta M, Rolandoa C. An efficient synthesis of (Z)- α -fluorochalcones via the palladium-catalyzed cross-coupling reaction of (Z)- α -fluorocinnamoyl chloride with boronic acids. *Tetrahedron*, 69 (6), 2013, 1735–1738.
3046. Emami, S., Foroumadi, A., Falahati, M., Lotfali, E., Rajabalian, S., Ebrahimi, S.-A., Farahyar, S., Shafiee, A. 2-Hydroxyphenacyl azoles and related azolium derivatives as antifungal agents (2008) *Bioorganic and Medicinal Chemistry Letters*, 18 (1), pp. 141-146.
3047. Evranos, B. (2010). Yeni bazı flavonoid türevlerinin sentezi, kimyasal yapılarının aydınlatılması ve monoamin oksidaz enzimleri üzerine etkilerinin araştırılması.
3048. Fadhil, G.F. (2023). Myasar Kh. Ibrahim. *Journal of Survey in Fisheries Sciences*, 10(3S), 5376-5386.
3049. Fang, Wan-Yin, et al. "Synthetic approaches and pharmaceutical applications of chloro-containing molecules for drug discovery: A critical review." *European journal of medicinal chemistry* 173 (2019): 117-153.
3050. Farias, I.F.D. (2017). Síntese, caracterização e avaliação do potencial biológico de Chalconas e análogos.
3051. Faudzi, S. M., Leong, S. W., Abas, F., Aluwi, M. M., Rullah, K., Lam, K. W., ... & Lajis, N. H. (2015). Synthesis, biological evaluation and QSAR studies of diarylpentanoid analogues as potential nitric oxide inhibitors. *MedChemComm*.
3052. Fedorova, Galina F., et al. "Exogenous and endogenous mediators of oxygen metabolism: alternatives for chemical and biological activity." *Studies in Natural Products Chemistry*. Vol. 47. Elsevier, 2016. 357-385.
3053. Feng J., Qi H., Sun X., Feng S., Liu Z., Song Y., Qiao X. (2018). Synthesis of novel pyrazole derivatives as promising DNA-binding agents and evaluation of antitumor and antitopoisomerase I/II activities. *Chem. Pharm. Bull.*, 66 (11), 1065-1071, 2018
3054. Feng, X., D. Yan, K.-J. Zhao, et al. Applications of microcalorimetry in the antibacterial activity evaluation of various *Rhizoma coptidis*. *Pharm Biol.* 2011, 49 (4):348-53.
3055. Fernandes, W.B., L.A. Malaspina, F.T. Martins, et al. Conformational variability in a new terpenoid-like bischalcone: Structure and theoretical studies. *J. Struct. Chem.*, 2013, 54, 6, 1112-1121.
3056. Fun, H.-K., T. Kobkeathhawin, P. Ruanwas, S. Chantrapromma, (E)-1-(4-Aminophenyl)-3-(2,4,5-trimethoxyphenyl)prop-2-en-1-one, *Acta Cryst.*, 2010, 66 (8), 1973-1974.
3057. Fun, H.-K., T. Suwunwong, S. Chantrapromma, and C. Karalai. (E)-1-(2-Furyl)-3-(3,4,5-trimethoxyphenyl)prop-2-en-1-one. *Acta Cryst.*, 2010, E66, 12, 3070-3071
3058. Garg S., Raghav N. Spectrophotometric analysis of bovine serum albumin in presence of some 1-(naphthalen-3-yl)-3-phenylprop-2-en-1-ones. *Int. J. Chem.l Sci.*, 2, 11, 2013, 1137-1145.
3059. Garg S., Raghav N. Synthesis of novel chalcones of schiff' s bases and to study their effect on bovine serum albumin. *Asian J. Pharm. Clin. Res.*, 6 (suppl.4), 2013, 181-184.
3060. Gatabazi, A. (2019). Irrigation and nitrogen management of African (*Siphonochilus aethiopicus* (Schweinf.) BL Burt) and commercial ginger (*Zingiber officinale* Roscoe). University of Pretoria (South Africa).
3061. Geweely N.S., Hassaneen H.M., Ali R.A., Soliman M.M., Abdelhamid I.A. New

- Inhibitory Effect by Green Synthesized Chalcone Derivatives on the Fungal Deterioration of Archaeological Egyptian Mummy, Egypt. Polycyclic Aromatic Compounds 2023. DOI: 10.1080/10406638.2023.2231595.
3062. Ghoneim, Amira A., Rehab M. Elbargisy, and Afaf Manoer. "Design and synthesis of heterocyclic Compounds from 1, 4-diacetylbenzene with Expected Antimicrobial Activity." *Egyptian Journal of Chemistry* 63.8 (2020): 9-10.
3063. Gonçalves CJ, Lenoir AS, Padaratz P, et al. Benzofuranones as potential antinociceptive agents: Structure–activity relationships. *Eur. J. Med. Chem.* 56, 2012, 120–126.
3064. Guan L-P., Zhao D-H., Chang Y., et al. Design, synthesis and antidepressant activity evaluation 2'-hydroxy-4',6'-diisoprenyloxychalcone derivatives. *Med. Chem. Res.*, 11, 22, 2013, 5218-5226.
3065. Guan L-P., Zhao D-H., Chang Y., et al. Synthesis of 2,4-dihydroxychalcone derivatives as potential antidepressant effect. *Drug Res.*, 1, 63, 2013, 46-51.
3066. Guerrero Villalobos, L.R. (2017). Estudio de la reactividad de chalconas como precursores en la síntesis de nuevos compuestos pirazolínicos, betalactámicos y tiazolidínicos fusionados (Doctoral dissertation).
3067. Guo, T., Jiang, Q., Yu, L., Yu, Z. Synthesis of chalcones via domino dehydrochlorination/Pd(OAc)₂-catalyzed Heck reaction *Cuihua Xuebao/Chinese Journal of Catalysis* 36, 2015, 78-85
3068. Hu, J., Liu, D., Xu, W., hang, F., Zheng, H. One-pot reaction for the concise synthesis of spiro[benzofuran-2,2'-naphthalen]-1'-one derivatives *Tetrahedron*, 2014, 70, 7511-7517.
3069. Hui, Y., J. Jiang, W.Wang, W.Chen, Y.Cai, L.Lin, X.Liu, X.Feng, Highly enantioselective conjugate addition of thioglycolate to chalcones catalyzed by lanthanum: low catalyst loading and remarkable chiral amplification, *Angewandte Chemie* 122 (25), 4386 – 4389, 2010.
3070. Ibrahim, M.K. and Fadhil, G.F., 2023. Synthesis and Photochromism of 3-(2, 6-dichlorophenyl)-1-(Naphthalen-1-yl) Prop-2-en-1-one (Part II). *Journal of Survey in Fisheries Sciences*, 10(2S), pp.4082-4094.
3071. Ibrahim, M.K. and Fadhil, G.F., 2023. Synthesis and Photochromism of (E)-3-(2, 3-dichlorophenyl)-1-(naphthalen-1-yl) prop-2-en-1-one. *Journal of Survey in Fisheries Sciences*, 10(3S), pp.5376-5386
3072. INSIGHTS, M. (2017). University of Malaya (Doctoral dissertation, UNIVERSITY OF MALAYA KUALA LUMPUR).
3073. Jia Yongbing, Zhang Rui, Xie Bin et al. (2014). Study on the asymmetric Michael addition reaction of chalcone derivatives with malonic acid diesters. *Journal of Guangzhou University: Natural Science Edition*, 13(3), 47-51.
3074. Jiang. W.-Q. An improved experiment for bis-benzalacetones synthesis. *CJTD*, 2010, 25, 2, 42 – 45.
3075. Jin, H., Xiang, L., Wen, F., Tao, K., Liu, Q., Hou, T. Improved synthesis of chalconoid-like compounds under ultrasound irradiation (2008) *Ultrasonics Sonochemistry*, 15 (5), pp. 681-683.
3076. Jin, Yong-Sheng. "Recent advances in natural antifungal flavonoids and their derivatives." *Bioorganic & medicinal chemistry letters* 29.19 (2019):126589.

3077. Jun, K.Y., H. Kwon, S.E. Park, et al. Discovery of dihydroxylated 2, 4-diphenyl-6-thiophen-2-yl-pyridine as a non-intercalative DNA-binding topoisomerase II-specific catalytic inhibitor. *Eur. J. Med. Chem.*, 2014, 80, 428-438.
3078. Kadayat, T.M., M.J. Kim, T. Nam, P.H. Park, et al. Thiény/furanyl-hydroxyphenylpropenones as inhibitors of LPS-induced ROS and NO production in RAW 264.7 macrophages, and their structure-activity relationship study. *Bull. Korean Chem. Soc.*, 2014, 35, 8, 2482-2486.
3079. Kadayat, T.M., Song, C., Kwon, Y., Lee, E.-S Modified 2,4-diaryl-5H-indeno[1,2-b]pyridines with hydroxyl and chlorine moiety: Synthesis, anticancer activity, and structure-activity relationship study. *Bioorganic Chemistry* 2015, 62, 30-40
3080. Kadayat, Tara Man, et al. "Effect of chlorine substituent on cytotoxic activities: Design and synthesis of systematically modified 2, 4-diphenyl-5H-indeno [1, 2-b] pyridines." *Bioorganic & medicinal chemistry letters* 26.7 (2016): 1726-1731.
3081. Kalogiros, C., and L.P. Hadjirapoglou. Facile preparation of bicyclo [2.2. 2] octenone derivatives via Diels–Alder cycloadditions of in situ-generated masked o-benzoquinones. *Tetrahedron*, 2011, 67, 18, 3216-3225.
3082. Kamal, A., A. Mallareddy, P. Suresh, V. et al. Synthesis and anticancer activity of 4b-alkylamidochalcone and 4b-cinnamido linked podophyllotoxins as apoptotic inducing agents. *Eur. J. Med. Chem.*, 2011, 1-16.
3083. Kancheva, V.D., O.T. Kasaikina. Bio-antioxidants—a chemical base of their antioxidant activity and beneficial effect on human health. *Curr. Med. Chem.*, 2013, 20, 37, 4784-4805.
3084. Karki R, Kang Y, Kim CH, et al. Hydroxychalcones as potential anti-angiogenic agent. *Bull. Korean Chem. Soc.*, 33 (9), 2012, 2925, ISSN 0253-2964.
3085. Karki, R., P. Thapa, H.Y. Yoo, TM Kadayat, et al. Dihydroxylated 2, 4, 6-triphenyl pyridines: Synthesis, topoisomerase I and II inhibitory activity, cytotoxicity, and structure–activity relationship study. *Eur. J. Med. Chem.*, 2012, 49, 219-228.
3086. Karki, R., Park, C., Jun, K. Y., Kadayatt, T. M., Lee, E. S., & Kwon, Y. Synthesis and Biological Activity of 2, 4-Di-I p/i-Phenolyl-6- i 2/i-Furanyl-Pyridine As a Potent Topoisomerase II Poison. *Eur. J. Mmed.Chem.*, 2015, 90, 360–378.
3087. Karki, Radha, et al. "A new series of 2-phenol-4-aryl-6-chlorophenyl pyridine derivatives as dual topoisomerase I/II inhibitors: Synthesis, biological evaluation and 3D-QSAR study." *European Journal of Medicinal Chemistry* 113 (2016): 228-245.
3088. Kim, M. J., Kadayat, T., Da Eun Kim, E. S. L., & Park, P. H.. TI-I-174, a Synthetic Chalcone Derivative, Suppresses Nitric Oxide Production in Murine Macrophages via Heme Oxygenase-1 Induction and Inhibition of AP-1. *Biomolecules & therapeutics*, 22(5), 2014, 390.
3089. Kim, M.J., Kadayat, T., Um, Y.J., Jeong, T.C., Lee, E.-S., Park, P.-H. Inhibitory effect of 3-(4-hydroxyphenyl)-1-(thiophen-2-yl) prop-2-en-1-one, a chalcone derivative on MCP-1 expression in macrophages via inhibition of ROS and Akt signaling. *Biomolecules and Therapeutics*, 2015, 23, 2, 119-127
3090. Karki, R., Thapa, P., Yoo, H.Y. et al. (2012). Dihydroxylated 2, 4, 6-triphenyl pyridines: Synthesis, topoisomerase I and II inhibitory activity, cytotoxicity, and structure–activity relationship study. *European journal of medicinal chemistry*, 49, 219-228.
3091. Kong W-J, Xing X-Y, Xiao X-H, et al. Effect of berberine on *Escherichia coli*,

- Bacillus subtilis*, and their mixtures as determined by isothermal microcalorimetry. *Appl. Microbiol. Biotechnol*, 96 (2), 2012, 503-510.
3092. Kong, W.J., Wang, J.B., Jin, C., Zhao, Y.L., Dai, C.M., Xiao, X.H., Li, Z.L. Effect of emodin on *Candida albicans* growth investigated by microcalorimetry combined with chemometric analysis (2009). *Appl. Microbiol. Biotechnol.*, 83 (6), pp. 1183-1190.
 3093. Kong, W.-J., Zhao, Y.-L., Xiao, X.-H., Li, Z.-L., Jin, C., Li, H.-B. Investigation of the anti-fungal activity of coptisine on *Candida albicans* growth by microcalorimetry combined with principal component analysis (2009) *Journal of Applied Microbiology*, 107 (4), pp. 1072-1080.
 3094. Kotan, G., & Medetalibeyoğlu, H. Antifungal Properties Of 1, 2, 4-Triazoles.
 3095. Kumar, R., P. Sharma, A. Shard, et al. Chalcones as promising pesticidal agents against diamondback moth (*Plutella xylostella*): microwave-assisted synthesis and structure–activity relationship. *Med. Chem. Res.*, 2011, DOI 10.1007/s00044-011-9602-8
 3096. Li X, Jin C, Liu W, et al. A microcalorimetric method to determine antimicrobial effects of two bile acid derivatives on *Staphylococcus aureus*. *J. Thermal Anal. Calorim.*, 108 (3), 2012, 1293-1301.
 3097. Lu Xinhua, Zou Jianping, Bian Guoqing, & Jiang Wenqing. (2009). Improvement of the preparation experiment of dibenzylideneacetone. *Laboratory Science*, (6), 68-70.
 3098. Lobo, C.I.V., Lopes, A.C.U.D.A., Klein, M.I. (2021). Compounds with distinct targets present diverse antimicrobial and antibiofilm efficacy against *Candida albicans* and *Streptococcus mutans*, and combinations of compounds potentiate their effect. *Journal of Fungi*, 7(5), 340.
 3099. Ma, Z., Xie, F., Yu, H. et al. (2013). Copper-catalyzed asymmetric 1, 4-conjugate addition of Grignard reagents to linear α , β , γ , δ -unsaturated ketones. *Chemical communications*, 49(46), 5292-5294.
 3100. Manoer, A., & Elbargisy, R. M. (2021). Synthesis of bis chalcones and transformation into bis heterocyclic compounds with expected antimicrobial activity. *Indian Journal of Chemistry-Section B (IJC-B)*, 60(9), 1272-1278.
 3101. Meghwal, K., Gupta, S., & Gomber, C. (2018). *Inorganic, Coordination and Organometallic Compounds. Sonochemistry*, 115.
 3102. Mellado M., Madrid A., Reyna M., Weinstein-Opppenheimer C., Mella J., Salas C. O., Sánchez E., Cuellar M. Synthesis of chalcones with antiproliferative activity on the SH-SY5Y neuroblastoma cell line: Quantitative Structure–Activity Relationship Models. *Med. Chem. Res.*, 27 (11-12), 2414-2425, 2018.
 3103. Mellado-Garcia, M., Reyna, M., Weinstein-Opppenheimer, C. et al. (2018). Preliminary Evaluation of Cytotoxicity for Small Chalcones on Breast and Colorectal Cancer Cell Lines: Synthesis and Structure-Activity Relationship. *J Pharmacol Ther Forecast*. 2018; 1 (1), 1003.
 3104. Mellado, Marco, et al. "Design, synthesis, antifungal activity, and structure–activity relationship studies of chalcones and hybrid dihydrochromane–chalcones." *Molecular diversity* (2019): 1-13.
 3105. Messier, C., F. Epifano, S. Genovese, D. Grenier. Inhibition of *Candida albicans* biofilm formation and yeast-hyphal transition by 4-hydroxycordoin. *Phytomedicine*, 2011, 18 (5):380-383.
 3106. Midoricava, L.R.O. (2015). Prospecção de substâncias anti histoplasma capsulatum

nas formas planctônica e biofilme e análise proteômica.

3107. Mirzaei, Hassan, Mahdi Abastabar, and Saeed Emami. "Indole-derived chalcones as anti-dermatophyte agents: In vitro evaluation and in silico study." *Computational Biology and Chemistry* 84 (2020): 107189.
3108. Mohd Faudzi, S.M., Leong, S.W., Abas, F., Mohd Aluwi, M.F.F., Rullah, K., Lam, K.W., Ahmad, S., Tham, C.L., Shaari, K., Lajis, N.H. Synthesis, biological evaluation and QSAR studies of diarylpentanoid analogues as potential nitric oxide inhibitors *MedChemComm* 2015, 6, 1069-1080
3109. Motta, L.F., W.P. Almeida. Ketone derivatives as anti-Candida albicans. *Int. J. Drug Discov.*, 2011, 3, 2, 100-117.
3110. Muñoz, V.A., Kretek, C.G., Montaña, M.P., Pappano, N.B., Debattista, N.B., Ferrari, G.V. Chalcones as analytical reagents of aluminum: Stability, thermodynamic and kinetic study *Zeitschrift fur Physikalische Chemie*, 2015, 229, 417-426
3111. Nasir Abbas B.S., M. Jasamai, J. Ibrahim. Synthesis and biological evaluation of chalcone derivatives (mini review). *Mini Rev. Med. Chem.*, 2012, 12, 13, 1394-1403.
3112. Navarini, A. Avaliação do efeito de chalcones sobre a linhagem celular B16-F10 de melanoma, Ph.D. Thesis, Federal University of Santa Catarina, Brasil, 2007.
3113. Nithya, R., N. Santhanamoorthi, P. Kolandaivel, and K. Senthilkumar. Structural and spectral properties of 4-bromo-1-naphthyl chalcones: a quantum chemical study. *J. Phys. Chem.*, 2011, 115, 24, 6594-6602.
3114. Nishimura, T., Noishiki, A., & Hayashi, T. (2012). Electronic tuning of chiral diene ligands in iridium-catalyzed asymmetric 1, 6-addition of arylboroxines to δ -aryl- α , β , γ , δ -unsaturated ketones. *Chemical communications*, 48(7), 973-975.
3115. Osmaniye D., Avuso Glu, B. L. K., Sa glik B. M. N., Levent S., Acar evik U., Atli Z., Zkay Y., Kaplancikli Z. A. Synthesis and Anticandidal Activity of New Imidazole-Chalcones. *Molecules*, 23 (4), 831, 2018
3116. Padaratz, P., Fracasso, M., De Campos-Buzzi, F., Corrêa, R., Niero, R., Monache, F.D., Cechinel-Filho, V. Antinociceptive activity of a new benzofuranone derived from a chalcone (2009) *Basic and Clinical Pharmacology and Toxicology*, 105 (4), pp. 257-261.
3117. Park, Sejeong, et al. "Novel 2-aryl-4-(4'-hydroxyphenyl)-5H-indeno [1, 2-b] pyridines as potent DNA non-intercalative topoisomerase catalytic inhibitors." *European journal of medicinal chemistry* 125 (2017): 14-28.
3118. Patel, D.V. (2020). Synthesis and Evaluation of Novel Antifungal Agents Targeting the Fungal Plasma Membrane H⁺-ATPase.
3119. Phang, C.W. (2017). The bioefficacy of flavokawain C against colon cancer and the underlying mechanistic insights/Phang Chung Weng (Doctoral dissertation, University of Malaya).
3120. Philip Parker (Ed.), *Candida: Webster's Quotations, Facts and Phrases*, ICON Group International, Inc., 2008.
3121. Pitt, D.W. (2015). Design of polymer systems and surface-active agents for the improvement of cell attachment for treatment of ocular diseases (Doctoral dissertation, University of Southampton).
3122. Prescott, Thomas AK, et al. "A simplified and easy-to-use HIP HOP assay provides insights into chalcone antifungal mechanisms of action." *FEBS letters* (2022).
3123. Raghav, N., S. Garg. Synthesis of novel chalcones of Schiff's bases and to study their

- effect on bovine serum albumin. *Asian J. Pharm. Clin. Res.*, 2013, 6, 4, 181-184.
3124. Rao, Y.K., Fang, S.-H., Tzeng, Y.-M. Synthesis and biological evaluation of 3',4',5'-trimethoxychalcone analogues as inhibitors of nitric oxide production and tumor cell proliferation (2009) *Bioorganic and Medicinal Chemistry*, 17 (23), pp. 7909-7914.
 3125. Richterová, L. (2016). Chalkony a jejich analogy jako potenciální léčiva X.
 3126. Ruanwas, P., Chantrapromma, S., Fun, H.-K. Synthesis, Characterization, Antioxidant, and Antibacterial Activities of 2-Aminochalcones and Crystal Structure of (2E)-1-(2-aminophenyl)-3-(4-ethoxyphenyl)-2-propen-1-one. *Molecular Crystals and Liquid Crystals* 2015, 609, 126-139.
 3127. Russell, M., & Hossain Soiket, M.I. (2014). Some new azole type heterocyclic compounds as antifungal agents. *Organic Communications*, 7(4).
 3128. Ruzie, C., M.Krayer, J.Lindsey, Fast and robust route to hydroporphyrin-chalcones with extended red or near-infrared absorption, *Organic Letters*, 11 (8), 1761-1764, 2009.
 3129. Saengsuwan, Nikorn, et al. "Photophysical Properties of Various Substituted Thiophene-based Heterocyclic Chalcone: Experimental and DFT Studies." *CHIANG MAI JOURNAL OF SCIENCE* 46.6 (2019): 1176-1190.
 3130. Saha, P., Biswas, A., Molleti, N. et al. (2015). Enantioselective synthesis of highly substituted chromans via the oxa-Michael–Michael cascade reaction with a bifunctional organocatalyst. *The Journal of Organic Chemistry*, 80(21), 11115-11122.
 3131. Salehi, Bahare, et al. "Pharmacological Properties of Chalcones: A Review of Preclinical Including Molecular Mechanisms and Clinical Evidence." *Frontiers in Pharmacology* 11 (2021): 2068.
 3132. Sardi, J.D.C.O. (2020). Antifungal activity and pharmacokinetic prediction of chalcone on phospholipase-producing *C. Albicans* isolates. *EC Pharmacology and Toxicology*, 8, 29-37.
 3133. Saroj, M. K., N. Sharma, and R. C. Rastogi. Solvent effect profiles of absorbance and fluorescence spectra of some indole based chalcones. *J. Fluoresc*, 2011, 21(6), 2213-2227.
 3134. Sathiyamoorthi, K., V. Mala, R. Suresh, et al. Synthesis, spectral correlations and antimicrobial activities of some 2-hydroxyphenyl-styrylketone. *Int. Let. Chem. Phys. Astronomy*, 2013, 7, 2, 102-119.
 3135. Shahnaz, M., S. Kaura, S. Lata, et al. Synthesis, characterization and antifungal activity of 1-naphthylmethylamine derivatives. *Int. J. Bioassays*, 2013, 2, 10, 1317-1321.
 3136. Shakhathreh, M.A.K., et al. "Study of the antibacterial and antifungal activities of synthetic benzyl bromides, ketones, and corresponding chalcone derivatives" *Drug Design, Development and Therapy* 10 (2016): 3653-3660.
 3137. Shan, Y., Lei, J., Zhang, L., Fan, T., Wang, M., Ma, Y Design, Synthesis, and Biological Evaluation of Chalcone Derivatives as Novel Anticandidal Agents *Chemistry of Natural Compounds* 2015, 51, 4, 620-625
 3138. Sharma, A., N. Sharma, A. Shard, et al. Tandem allylic oxidation–condensation/esterification catalyzed by silica gel: an expeditious approach towards antimalarial diaryldienones and enones from natural methoxylated phenylpropenes. *Org. Biomol. Chem.*, 2011, 9, 5211-5219.
 3139. Shrestha A., Park S., Shin S., Kadayat T. M., Bist G., Katila P., Kwon Y., Lee E. S. Design, synthesis, biological evaluation, structure-activity relationship study, and mode of action of 2-phenol-4, 6-dichlorophenyl-pyridines. *Bioorg. Chem.*, 79, 1-18, 2018.

3140. Silva, S.J.D. (2008). Síntese de análogos de chalconas do tipo retinóide com potencial atividade biológica.
3141. Silva, P. T., et al. "Cytotoxic and Antifungal Activity of Chalcones Synthesized from Natural Acetophenone Isolated from *Croton anisodontus*." *Revista Virtual de Química* 12.3 (2020).
3142. Singh, K., Manisha, M., Likhawat, M., & Singh, L. (2011). Synthesis of Novel Functionalized Chalcones and their In-vitro Antimalarial Evaluation. *Journal of Chemical, Biological and Physical Sciences (JCBPS)*, 1(2), 335.
3143. Singh K, Sahu A, Manisha J, Singh L. In-vitro antimalarial evaluation of novel functionalized chalcones. *J. Chem. Biol. Phys. Sci.*, 2, 2012, 782-791.
3144. Singh, S., Fatima, Z., & Hameed, S. (2020). Plant phenolics for overcoming multidrug resistance in human fungal pathogen. *Plant Phenolics in Sustainable Agriculture: Volume 1*, 407-430.
3145. Sivakumar, P.M., Muthu Kumar, T., Doble, M. Antifungal activity, mechanism and QSAR studies on chalcones (2009) *Chem. Biol. Drug Design*, 74 (1), pp. 68-79.
3146. Srinivasan, B., Johnson, T.E., Lad, R. et al. (2009). Structure– activity relationship studies of chalcone leading to 3-hydroxy-4, 3', 4', 5'-tetramethoxychalcone and its analogues as potent nuclear factor κ B inhibitors and their anticancer activities. *Journal of medicinal chemistry*, 52(22), 7228-7235.
3147. Sulis P.M., Bittencourt Mendes A.K., Fernandes T.A., Frederico M.J.S., Rey D.P., Aragón M., Ruparelia K.C., Silva F.R.M.B. Signal transduction of the insulin secretion induced by the chalcone analogue, (E)-3-(phenyl)-1-(3,4,5-trimethoxyphenyl)prop-2-en-1-one, and its role in glucose and lipid metabolism. *Biochimie*, 212, pp. 85 – 94, 2023. DOI: 10.1016/j.biochi.2023.04.006.
3148. Sultana F., Bonam S. R., Reddy V. G., Nayak V. L., Akunuri R., Routhu S. R., Alarifi A., Halmuthur M. S. K., Kamal A. (2018). Synthesis of benzo [d] imidazo [2, 1-b] thiazole-chalcone conjugates as microtubule targeting and apoptosis inducing agents. *Bioorg. Chem.*, 76, 1-12, 2018.
3149. Suwunwong, T., S. Chantrapromma, H.K. Fun. Influence of trimethoxy-substituted positions on fluorescence of heteroaryl chalcone derivatives. *Chem. Papers*, 2011, 65, 6, 890-897.
3150. Tailor, N.K. One pot synthesis and antibacterial activity of tetrahydrochalcones.
3151. Tailor, N.K. Synthesis and antifungal activity of certain chalcones and their reduction. *Indo Global J. Pharm. Sci.*, 2014; 4, 1 25-28.
3152. Tang C, Zhu L, Li J, et al. Synthesis and structure elucidation of five new conjugates of oleanolic acid derivatives and chalcones using 1D and 2D NMR spectroscopy. *Magn. Reson. Chem*, 50 (3), 2012, 236–241.
3153. Tristão, T.C. (2008). TRIAGEM PARA A ATIVIDADE ANTIMICROBIANA E POTENCIAL TÓXICO DE UMA NOVA SÉRIE DE CHALCONAS SINTÉTICAS E SEUS DERIVADOS ANÁLOGOS.
3154. Unoh Y., Hirano K., Satoh T., Miura M. Palladium-catalyzed decarboxylative arylation of benzoylacrylic acids toward the synthesis of chalcones. *J. Org.Chem.*, 10, 78, 2013, 5096-5102.
3155. Vásquez-Martínez, Yesseny A., et al. "Antimicrobial, Anti-Inflammatory and Antioxidant Activities of Polyoxygenated Chalcones." *Journal of the Brazilian Chemical*

Society 30.2 (2019): 286-304.

3156. Venkatesan, P., & T. Maruthavanan. Piperidine-mediated synthesis of thiazolyl chalcones and their derivatives as potent antimicrobial agents. *Nat. Prod. Res.*, 2011, 26, 3, 223-234.
3157. Wang, C., Wu, H., Tan, Y. et al. Synthesis of chalcone and azachalcone through mechanical vibration. *J. Nantong University (Nat. Sci. Edition)*, 2013, 12, 1, 40-45.
3158. Wang, J., D. Cheng, N. Zeng, H. Xia, et al. Application of microcalorimetry and principal component analysis. *J. Therm. Anal. Calorim.*, 2010, 102, 137-142.
3159. Wang, S., W. Hao, T. Yajun, & Z. Ze. Mechanical method for chalcone synthesis. *Nantong University, Nat. Sci.*, 2013, 1, 007
3160. Wang, W., X. Liu, W. Cao, J. Wang, L. Lin, X. Feng. Highly enantioselective synthesis of β -heteroaryl-substituted dihydrochalcones through friedel–crafts alkylation of indoles and pyrrole. *Chemistry – A European Journal*, 2010, 16, 5, 1664-1669.
3161. Weng, P.C. (2017). The Bioefficacy of Flavokawain C Against Colon Cancer and the Underlying Mechanistic Insights (Doctoral dissertation, University of Malaya (Malaysia)).
3162. Wu, J., C. Wang, Y. Cai, et al. Liang. Synthesis and crystal structure of chalcones as well as on cytotoxicity and antibacterial properties. *Med. Chem. Res.*, 2011, DOI 10.1007/s00044-011-9549-9
3163. Xin, Y., Zang, Z.-H., Chen, F.-L. Ultrasound-promoted synthesis of 1,5-diarylpenta-2,4-dien-1-ones catalyzed by activated barium hydroxide (2009). *Synthetic Communications*, 39 (22), pp. 4062-4068.
3164. Xue, Y. Xu, K., Liu, Y. DFT study on the structures and electronic spectra of furan chalcones. *J. Mol. Sci.*, 2010, 26, 3, 12-17.
3165. Xue, Y., Mou, J., Liu, Y., Gong, X., Yang, Y., An, L. An ab initio simulation of the UV/Visible spectra of substituted chalcones (2010). *Centr. Eur. J. Chemistry*, 8 (4), pp. 928-936.
3166. Xie, C., Peng, Z., Zhao, S.L. et al. (2014). Synthesis of 2-hydroxy-4-isoprenyloxychalcone derivatives with potential antidepressant-like activity. *Medicinal Chemistry*, 10(8), 789-799.
3167. Zhang, Z., Deng, J.A., L. Zhihong, et al. Pharmacological activity of Coptisine. *Progr. Chinese Mat. Med.*, 2013, 38 (017), 2750-2754.
3168. Zhao Y., Jia L., Wang J., et al. Microcalorimetry with correspondence analysis for studying the antibacterial effect of ephedrine on *Escherichia coli*. *Thermochim. Acta*, 557, 2013, 50-54, ISSN: 0040-6031.
3169. Zhao, Y., D. Yan, J. Wang, P. Zhang and X. Xiao. Anti-fungal effect of berberine on *Candida albicans* by microcalorimetry with correspondence analysis. *Journal of Therm. Anal. Calorim.*, 2010, 102(1), 49-55.
3170. Валова, Марина Сергеевна. Халконо-поданды в реакциях с ацетоуксусным эфиром и аминокетонами: диссертация на соискание ученой степени кандидата химических наук: 02.00. 03. Diss. б. и., 2017.
3171. Курьянов, В.О., Токарев, М.К., & Чупахина, Т.А. (2013). Синтез глюкозаминидов халконов. *Ученые записки Крымского федерального университета имени В.И. Вернадского. Биология. Химия*, 26(1 (65)), 312-322.

95). Batovska, D.I., Todorova, I.T., Tsvetkova, I.V., NAJDENSKI, H.M. Antibacterial study of the medium chain fatty acids and their 1-monoglycerides: Individual effects and

synergistic relationships (2009) Polish Journal of Microbiology, 58 (1), pp. 43-47.

3172. Abbasi, Zaeem Arif, et al. "Efficacy of oil-pulling versus chlorhexidine mouthwash in reducing oral Streptococcus mutans count: A systemic review and meta-analysis." *Journal of Advances in Medicine and Medical Research* (2019): 1-9.
3173. Adewuyi, Adewale, Omolola H. Fasusi, and Rotimi A. Oderinde. "Antibacterial activities of acetanilides prepared from the seed oils of *Calophyllum inophyllum* and *Pterocarpus osun*." *Journal of Acute Medicine* 4.2 (2014): 75-80.
3174. Agarwal, Priyanka, et al. "Topical semifluorinated alkane based azithromycin suspension for the management of ocular infections." *European Journal of Pharmaceutics and Biopharmaceutics* 142 (2019) 83-91.
3175. Agarwal, Priyanka. *Application of Semifluorinated Alkanes in Dry Eye Therapy*. Diss. ResearchSpace@ Auckland, 2018.
3176. Akula, Satya Tejaswi, et al. "Antifungal efficacy of lauric acid and caprylic acid—Derivatives of virgin coconut oil against *Candida albicans*." *Biomedical and Biotechnology Research Journal (BBRJ)* 5.2 (2021): 229.
3177. Alcock, Joe, Melissa L. Franklin, and Christopher W. Kuzawa. "Nutrient signaling: evolutionary origins of the immune-modulating effects of dietary fat." *The Quarterly review of biology* 87.3 (2012): 187-223.
3178. Ali, M.M., Elashry, M.A., Mohammady, E.Y. et al. 2023. Dietary Alpha-Monolaurin for Nile Tilapia (*Oreochromis niloticus*): Stimulatory Effects on Growth, Immunohematological Indices, and Immune-Related Gene Expressions. *Aquaculture Research*, 2023.
3179. Al-Ismael, Maria Ibrahim. *The Antimicrobial Effect of Coconut Oil and its Fatty Acids on Oral Microorganisms Compared to Chlorhexidine Mouth Rinse: An in vitro Study*. Diss. Tufts University School of Dental Medicine, 2013.
3180. Alkhalaf, M.I., Churchill, G.C. and Mirghani, M.E., 2023. Chemical composition and antioxidant/antibacterial depictions of Zahidi date palm (*Phoenix dactylifera*) kernel Oil. *Journal of King Saud University-Science*, 35(7), p.102817.
3181. Almeida, A.B., Araujo, D.N., Strapazzon, J.V. et al. (2021). Use of blend based on an emulsifier, monolaurin, and glycerides of butyric acid in the diet of broilers: Impacts on intestinal health, performance, and meat. *Anais da Academia Brasileira de Ciências*, 93(suppl 4), e20210687.
3182. Almutawif, Yahya Ahmad. "Staphylococcus aureus and its Enterotoxin in Donor Human Milk." (2019).
3183. Amorn Suwan Phiwat, A.S. Effects of a self-care oral care program combined with coconut oil gargle on gingivitis in cancer patients receiving chemotherapy. *Thai Journal of Nursing Council*, 32.1 (2017): 18-31.
3184. Anacarso I., Quartieri A., De Leo R., Pulvirenti A. Evaluation of the antimicrobial activity of a blend of monoglycerides against *Escherichia coli* and *Enterococci* with multiple drug resistance. *Arch. Microbiol.*, 200 (1), 85-89, 2018.
3185. Anderson, R.C., Poole, T.L., Beier, R.C. Inhibition Of Multi-Drug Resistant Staphylococci By Sodium Chlorate And Select Nitro-And Medium Chain Fatty Acid Compounds.
3186. Ara, Ismet, et al. "Evaluation of antimicrobial properties of two different extracts of *Juglans regia* tree bark and search for their compounds using gas chromatography-mass

- spectrum." *International Journal of Biology* 5.2 (2013): 92.
3187. Arellano, H., Nardello-Rataj, V., Szunerits, S., Boukherroub, R. and Fameau, A.L., 2023. Saturated long chain fatty acids as possible natural alternative antibacterial agents: Opportunities and challenges. *Advances in Colloid and Interface Science*, p.102952.
 3188. Arif Abbasi, Z., Saher, F., Ahmed Qureshi, J. et al. (2020). Efficacy of oil-pulling versus chlorhexidine mouthwash in reducing oral *Streptococcus mutans* count: A systemic review and meta-analysis. *Journal of Advances in Medicine and Medical Research*, 31(12), 1-9.
 3189. Armita, Devi. Uji Daya Hambat VCO yang disuplementasi Metabolit BAL terhadap Bakteri Patogen. Diss. Universitas Islam Negeri Alauddin Makassar, 2014.
 3190. Arroyo-Urea, E.M., Lázaro-Díez, M., Garmendia, J. et al. (2023). Lipid-based nanomedicines for the treatment of bacterial respiratory infections: current state and new perspectives. *Nanomedicine*, (0).
 3191. Appleton, S.R., Ballou, A., Watkins, K.L. (2024). Use of Monoglycerides and Diglycerides to Mitigate Poultry Production Losses: A Review. *Veterinary Sciences*, 11(3), 101.
 3192. Bain, Jerald, Et Al. Compositions For Treating Sufficient Or Illnesses Provided By Neurokinin-2-Receptor Activity. 2013.
 3193. Baltić, B., et al. "Importance of medium chain fatty acids in animal nutrition." IOP Conference Series: Earth and Environmental Science. Vol. 85. No. 1. IOP Publishing, 2017.
 3194. Baltić, Branislav. "Ispitivanje uticaja dodavanja srednjelančanih masnih kiselina na zdravstveno stanje, proizvodne rezultate i kvalitet mesa brojlera." Универзитет у Београду (2019).
 3195. Bhattacharyya, Anamika, et al. "Mechanistic Insight Into the Antifungal Effects of a Fatty Acid Derivative Against Drug-Resistant Fungal Infections." *Frontiers in microbiology* 11 (2020): 2116.
 3196. Bhattacharyya, J., Chatterjee, J.K., Chatterjee, P.N., Pal, A. (2024). Influence of Incorporating Full-Fat Black Soldier Fly Larvae Meal (BSFLM) in Meat Type Chicken Diets on the Lower Gut Short Chain Fatty Acids profile, Gut Morphology and Intestinal Lesion Score. *bioRxiv*, 2024-04.
 3197. Blackshaw, K., Wu, J., Valtchev, P. et al. (2021). The effects of thermal pasteurisation, freeze-drying, and gamma-irradiation on the antibacterial properties of donor human milk. *Foods*, 10(9), 2077.
 3198. Blackshaw, K. (2021). A Non-Thermal Process for Increasing the Shelf-Life and Preserving Functionality of Donor Human Milk (Doctoral dissertation).
 3199. Boondireke, Sirirat, et al. "Encapsulation of monomyristin into polymeric nanoparticles improved its in vitro antiproliferative activity against cervical cancer cells." *Colloids and Surfaces B: Biointerfaces* 176 (2019): 9-17.
 3200. Božić, Aleksandar, et al. "Inhibition of multidrug-resistant *Staphylococci* by sodium chlorate and select nitro-and medium chain fatty acid compounds." *Journal of applied microbiology* 126.5 (2019): 1508-1518.
 3201. Brasil, J.S.M.P. Grasas saturadas: una perspectiva de la lactancia y de la composición de la leche-Parte 3/3.
 3202. Bunyovimonnat, R. (2018). Preparation and antimicrobial activity of caprylic acid

emulsion for food coating.

3203. Bulut, Nurefşan. Türkiye'de yaşayan Nogay Türkleri'nin nogay çay tüketimi, beslenme ve sağlık üzerine etkisi. MS thesis. Biruni Üniversitesi Sağlık Bilimleri Enstitüsü, 2019.
3204. Cais-Sokolińska, D., Bielska, P., Rudzińska, M. et al. (2024). Water thermodynamics and lipid oxidation in stored whey butter. *Journal of Dairy Science*, 107(4), 1903-1915.
3205. Canli Kerem, et al. "In vitro antimicrobial screening of *Lycoperdon lividum* and determination of the ethanol extract composition by gas chromatography/mass spectrometry." *Bangladesh Journal of Pharmacology* 11.2 (2016): 389-394.
3206. Canli, K., Cetin, B., Altuner, E.M., Türkmen, Y., Uzek, U., Dursun, H. In vitro antimicrobial screening of *Hedwigia ciliata* Var. *leucophaea* and determination of the ethanol extract composition by gas chromatography/mass spectrometry (GC/MS). *Journal of Pure and Applied Microbiology* 2014, 8, 2987-2998.
3207. Cayona, R. and Yu, G., 2023. Selective Synthesis of Monolaurin: a Preliminary Investigation. *Philippine Journal of Science*, 152(3).
3208. Chantadee, Takron, et al. "Vancomycin hydrochloride-loaded stearic acid/lauric acid in situ forming matrix for antimicrobial inhibition in patients with joint infection after total knee arthroplasty." *Materials Science and Engineering: C* 115 (2020): 110761.
3209. Chaouat, Clara, et al. "Antimicrobial catanionic vesicular self-assembly with improved spectrum of action." *Journal of Surfactants and Detergents* 16.5 (2013): 717-722.
3210. Chen, Jianwei, et al. "Potential applications of biosurfactant rhamnolipids in agriculture and biomedicine." *Applied microbiology and biotechnology* 101.23-24 (2017): 8309-8319.
3211. Chen, Kai Yu, et al. "Monoacylglycerol of 7, 10-Dihydroxy-8 (E)-octadecenoic Acid Enhances Antibacterial Activities against Food-Borne Bacteria." *Journal of agricultural and food chemistry* 67.29 (2019): 8191-8196.
3212. Christen, Lukas. "Pasteurization of donor human milk for the use in the neonatal intensive care unit." Crawley: The University of Western Australia (2014).
3213. Chung D., Cho T. J., Rhee M. S. Citrus fruit extracts with carvacrol and thymol eliminated 7-log acid-adapted *Escherichia coli* O157: H7, *Salmonella typhimurium*, and *Listeria monocytogenes*: A potential of effective natural antibacterial agents. *Food Res. Int.*, 107, 578-588, 2018.
3214. Çınar, Mehmet Ulaş, et al. "Milk and fatty acid composition of Anatolian water buffalo (*Bubalus bubalis*) from different provinces." *Buffalo Bulletin* 38.1 (2019): 107-118.
3215. Cochrane, Roger A., et al. "Determining the Minimum Inhibitory Concentration of Medium Chain Fatty Acids for generic *Escherichia coli*, Enterotoxigenic *Escherichia coli*, *Salmonella Typhimurium*, *Campylobacter coli*, and *Clostridium perfringens*." (2020).
3216. Coelho, Olívia Gonçalves Leão, Flávia Galvão Cândido, and Rita de Cássia Gonçalves Alfenas. "Dietary fat and gut microbiota: mechanisms involved in obesity control." *Critical reviews in food science and nutrition* 59.19 (2019): 3045-3053.
3217. Córdova Albores, L.C. (2014). Composición química del aceite de *Jatropha curcas* y su actividad sobre el crecimiento, morfología y viabilidad de *Fusarium oxysporum* f. sp. *gladioli*.
3218. Dabbou, Sihem, et al. "Modified Black Soldier Fly Larva Fat in Broiler Diet: Effects on Performance, Carcass Traits, Blood Parameters, Histomorphological Features and Gut

- Microbiota." *Animals* 11.6 (2021): 1837.
3219. Dahlke, S. Threat posed by multi-drug resistant *Staphylococcus aureus* and possible new treatment options. MS Thesis, University of Wisconsin-La Crosse, 2013.
 3220. Dayrit, F. M. The Properties of Lauric Acid and Their Significance in Coconut Oil. *Journal of the American Oil Chemists' Society*, 2015, 92, 1-15.
 3221. De Leo, Riccardo, and Andrea Pulvirenti. "Immacolata Anacarso, Andrea Quartieri."
 3222. de Oliveira, Filipe Silva Nunes, and Ana Rita Cruz Duarte. "A look on target-specificity of eutectic systems based on natural bioactive compounds." 2021, 97, 271-307.
 3223. Demirci, Mehmet. "Broyler rasyonlarına orta zincirli yağ asitleri ilavesinin performans ve bazı kan parametreleri üzerine etkileri." (2018).
 3224. Deschepper, Katrien, et al. "A balanced mixture of medium chain fatty acids improves zootechnical performances and slaughter results of broilers." *Proceedings of the 19th European Symposium of Nutrition*. 2013.
 3225. Devan, Krishnapriya, et al. "Antimicrobial efficacy of medium-chain fatty acids, 2% Chlorhexidine, and 5% sodium hypochlorite against *Enterococcus faecalis*: An in vitro study." *Indian Journal of Oral Health and Research* 4.2 (2018): 47.
 3226. Dhakal, Janak, and Charles G. Aldrich. "Use of Medium Chain Fatty Acids To Mitigate *Salmonella Typhimurium* (ATCC 14028) on Dry Pet Food Kibbles." *Journal of Food Protection* 83.9 (2020): 1505-1511.
 3227. Doležalová, Magda, et al. "Antimicrobial properties of 1-monoacylglycerols prepared from undecanoic (C11: 0) and undecenoic (C11: 1) acid." *European journal of lipid science and technology* 112.10 (2010): 1106-1114.
 3228. Dong, Liyou, et al. "Clostridium perfringens suppressing activity in black soldier fly protein preparations." *LWT* (2021): 111806, 147.
 3229. Duranova, H., Kuzelova, L., Fialkova, V. Et al. (2024). Coconut-sourced MCT oil: its potential health benefits beyond traditional coconut oil. *Phytochemistry Reviews*, 1-42.
 3230. Dutoit, A., Decourcelle, N., Mathot, A.G. et al. (2024). Relationships between the inhibitory efficacy and physicochemical properties of six organic acids and monolaurin against *Bacillus weihenstephanensis* KBAB4 growth in liquid medium. *Food Microbiology*, 104498.
 3231. El-Kholy, K.H., Ghonim, A.I.A., Ahmed, M.A. et al. (2020). Physiological and reproductive responses of Domyati ducks to different dietary levels of coconut oil as a source of medium-chain fatty acids during laying period. *Journal of World's Poultry Research*, 10(3), 485-492.
 3232. El-Kholy, K.H., Ghonim, A.I.A., Ahmed, M.A. et al. (2021). Determining the Physiological and Reproductive Responses of Domyati Ducks to Different Dietary Levels of Coconut Oil as a Source of Medium-Chain Fatty Acids during Laying Period.
 3233. Elshikh, M., et al. "Rhamnolipids and lactonic sophorolipids: natural antimicrobial surfactants for oral hygiene." *Journal of applied microbiology* 123.5 (2017): 1111-1123.
 3234. Elshikh, Mohamed, et al. "Rhamnolipids from non-pathogenic *Burkholderia thailandensis* E264: Physicochemical characterization, antimicrobial and antibiofilm efficacy against oral hygiene related pathogens." *New biotechnology* 36 (2017): 26-36.
 3235. EM-Sub Pin Valley et al. The effect of the coconut oil for oral care to thick Valle mouth and Bossier oral disease in patients with cancer receiving chemotherapy: a pilot study. *Thai Cancer J.*, 2013, 33 (2), 41-52.

3236. Fife, B. Health properties of coconut oil. *Agro Food Ind. Hi-Tech*, 3, 24, 2013, 7-10.
3237. Frutis-Murillo, Minerva, et al. "Immunomodulatory molecules regulate adhesin gene expression in *Staphylococcus aureus*: Effect on bacterial internalization into bovine mammary epithelial cells." *Microbial pathogenesis* 131 (2019): 15-21.
3238. Gantois, I., E. Van Meenen, L. Maertens. L'effet d'un mélange spécifique d'acides gras a chaîne moyenne sur les performances zootechniques des poulets de chair. *Dixièmes Journées de la Recherche Avicole et Palmipèdes à Foie Gras (JRA-JRFG)*, La Rochelle, du 26 au 28 mars 2013, 805-809.
3239. Gayatri, A. D. I. T. A., E. V. A. Fauziah, and M. A. R. G. A. R. E. T. H. A. Suharsini. "Antibacterial effect of virgin coconut oil on the viability of chromogenic bacteria that causes dental black stain in children." *Int J Appl Pharm* 9.2 (2017).
3240. German, J.B. and C.J. Dillard. Saturated fats: A perspective from lactation and milk composition. *Lipids*. 2010, 45(10), 915-923. ISSN:0024-4201
3241. Ghany, S.S.H.A.E., Ibrahem, R.A., El-Gendy, A.O. et al. (2024). Novel synergistic interactions between monolaurin, a mono-acyl glycerol and β lactam antibiotics against *Staphylococcus aureus*: an in vitro study. *BMC Infectious Diseases*, 24(1), 379.
3242. Gharib-Naseri, Kosar, et al. "Buffered formic acid and a monoglyceride blend coordinately alleviate subclinical necrotic enteritis impact in broiler chickens." *Poultry Science* (2021): 101214.
3243. Goc, A., A. Niedzwiecki, and M. Rath. "In vitro evaluation of antibacterial activity of phytochemicals and micronutrients against *Borrelia burgdorferi* and *Borrelia garinii*." *Journal of applied microbiology* 119.6 (2015): 1561-1572.
3244. Gómez González, J.A. (2019). Compuestos con actividad antimicrobiana en plantas del género *Kalanchoe* como tratamiento promisorio para infecciones producidas por bacterias.
3245. Gomez-Osorio, L.M., Yepes-Medina, V., Ballou, A. et al. (2021). Short and medium chain fatty acids and their derivatives as a natural strategy in the control of necrotic enteritis and microbial homeostasis in broiler chickens. *Frontiers in veterinary science*, 8, 773372.
3246. Ham, Youngseok, and Tae-Jong Kim. "Inhibitory activity of monoacylglycerols on biofilm formation in *Aeromonas hydrophila*, *Streptococcus mutans*, *Xanthomonas oryzae*, and *Yersinia enterocolitica*." *Springerplus* 5.1 (2016): 1-8.
3247. Hamedi, Hassan, Seyyed Mehdi Razavi-Rohani, and Hassan Gandomi. "Combination effect of essential oils of some herbs with monolaurin on growth and survival of *Listeria monocytogenes* in culture media and cheese." *Journal of Food Processing and Preservation* 38.1 (2014): 304-310.
3248. Han, Ching Enn, et al. "Growth characteristic of probiotic in fermented coconut milk and the antibacterial properties against *Streptococcus pyogenes*." *Journal of Food Science and Technology* 59.9 (2022): 3379-3386.
3249. Hanczakowska, Ewa. "The Use of Medium-Chain Fatty Acids in Piglet Feeding—A Review." *Annals of Animal Science* 17.4 (2017): 967-977.
3250. Herwita, D. (2017). Karakteristik Fisik, Kimia, Mikrobiologis Dan Organoleptik Dodol Terlapisi Minyak Nabati (Doctoral dissertation, FAKULTAS TEKNOLOGI PERTANIAN).
3251. Horincar, Georgiana, and Gabriela Bahrin. "The antimicrobial properties of enzymatic

- hydrolysates of goat milk fat." *The Annals of the University Dunarea de Jos of Galati. Fascicle VI-Food Technology* 41.1 (2017): 30-40.
3252. Horincar, Georgiana, and Gabriela Bahrin. "The antimicrobial properties of enzymatic hydrolysates of goat milk fat." *The Annals of the University Dunarea de Jos of Galati. Fascicle VI-Food Technology* 41.1 (2017): 30-40.
3253. Hovenjürgen, M. Einfluss mittelkettiger Fettsäuren auf die Entwicklung schweinespezifischer pathogener Keime in vitro.
3254. Iggman, David, and Ulf Risérus. "Role of different dietary saturated fatty acids for cardiometabolic risk." *Clinical lipidology* 6.2 (2011): 209-223.
3255. IQBAL, M., Shah, M. D., Gnanaraj, C., & Khan, M. S. (2015). *Dillenia Suffruticosa L., Impedes Carbon Tetrachloride-Induced Hepatic Damage by Modulating Oxidative Stress and Inflammatory Markers In Rats. Journal of Environmental Pathology, Toxicology and Oncology*, 24, 133-152.
3256. Jaafar S. H. S., Hashim R., Hassan Z., Arifin N. A Comparative Study on Physicochemical Characteristics of Raw Goat Milk Collected from Different Farms in Malaysia. *Tropical Life Sci. Res.*, 29 (1), 195, 2018.
3257. Jadhav, P., Manwar, S., Khose, K. et al. (2021). Effect of medium chain fatty acids as replacement to antibiotics in diets on growth performance and gut health in broiler chicken. *Indian Journal of Animal Research*, 55(8), 894-899.
3258. Kent, Jacqueline C., et al. "Role of breast milk." *Nutrition for the Preterm Neonate*. Springer, Dordrecht, 2013. 311-335.
3259. Khosravinia, H. Effect of dietary supplementation of medium-chain fatty acids on growth performance and prevalence of carcass defects in broiler chickens raised in different stocking densities. *Journal of Applied Poultry Research* 2015, 24, 1-9
3260. Kim, H.R., & Kim, I.H. (2024). 7, 10-Dihydroxy-8 (E)-octadecenoic Acid Displays a Fungicidal Activity against *Malassezia furfur*. *Journal of Oleo Science*, 73(2), 215-218.
3261. Klementavičiūtė, Jolita, et al. "EFFECT OF MEDIUM CHAIN FATTY ACIDS AND EMULSIFIER ON QUALITY PARAMETERS OF LAYING HEN 'S EGGS." *The Use of local raw material in the animal nutrition: benefits and limitations regarding digestive physiology, products quality and animal health: 6 October, 2016, Kaunas/Lietuvos sveikatos mokslų universitetas. Veterinarijos akademija. Gyvulininkystės Technologijos fakultetas. Gyvūnų auginimo technologijų institutas. Hohenheimo universiteto Gyvulių mitybos institutas (Vokietija) ir [kt.]. Kaunas: Lietuvos sveikatos mokslų universiteto Leidybos namai*, 2016.
3262. Klementavičiūtė, J. (2019). *Helianthus tuberosus L. stiebagumbių miltų, Macleaya cordata veikliųjų medžiagų ir skirtingų riebalų rūgščių įtaka viščiukų broilerių ir vištų dedeklių produktyvumui bei produkcijos kokybei: daktaro disertacija: žemės ūkio mokslai, gyvūnų mokslai (A003)*.
3263. Koçer, Bülent, and Burak Birgören. "Approaches for problem diagnosis via statistical process control charts." *Gazi University Journal of Science* 17.4 (2004): 59-69.
3264. Kovanda, Lauren, et al. "In Vitro Antimicrobial Activities of Organic Acids and Their Derivatives on Several Species of Gram-Negative and Gram-Positive Bacteria." *Molecules* 24.20 (2019): 3770.
3265. Kovanda, Lauren. *Antimicrobial Effects of Organic Acids and Their Derivatives*. Diss. University of California, Davis, 2020.

3266. Kumar, Alip, et al. "Monoglyceride Blend Reduces Mortality, Improves Nutrient Digestibility, and Intestinal Health in Broilers Subjected to Clinical Necrotic Enteritis Challenge." *Animals* 11.5 (2021): 1432.
3267. Kumar, Sandeep. "Dietary Saturated Fat: Facts and Fallacies." *Journal of Medical Academics* 1.2 (2018): 102-108.
3268. Laboratoriumgeneeskunde, U.M.C., & Amsterdam, C.V. (2012). Het faillissement van de verzaadigd vethypothese van cardiovasculaire ziektes. *Ned Tijdschr Klin Chem Labgeneesk*, 37(3), 192-211.
3269. Lalouckova, Klara, et al. "In vitro antagonistic inhibitory effects of palm seed crude oils and their main constituent, lauric acid, with oxacillin in *Staphylococcus aureus*." *Scientific Reports* 11.1 (2021): 1-12.
3270. Laloučková, Klára, et al. "In vitro antimicrobial effect of palm oils rich in medium-chain fatty acids against mastitis-causing Gram-positive bacteria." *Czech Journal of Animal Science* 64.8 (2019): 325-331.
3271. Lester, Katherine Louise. *Zoocin A and lauricidin in combination selectively inhibit Streptococcus mutans in a biofilm model*. Diss. University of Otago, 2011.
3272. Letlole, Bakang R., Ellen PCW Damen, and Christine Jansen van Rensburg. "The Effect of α -Monolaurin and Butyrate Supplementation on Broiler Performance and Gut Health in the Absence and Presence of the Antibiotic Growth Promoter Zinc Bacitracin." *Antibiotics* 10.6 (2021): 651.
3273. Li, S., Ren, R., Lyu, L., Song, J., Wang, Y., Lin, T.-W., Brun, A. L., et al. "Solid and Liquid Surface-Supported Bacterial Membrane Mimetics as a Platform for the Functional and Structural Studies of Antimicrobials." *Membranes* 12.10 (2022): 906.
3274. Li, M., Yuan, J., Yang, Q. et al. (2023). Therapeutic deep eutectic solvents based on natural product matrine and caprylic acid: Physical properties, cytotoxicity and formation of surfactant free microemulsion. *Journal of Drug Delivery Science and Technology*, 90, 105177.
3275. Ling, Zinyin, Ruth Anisah Fromming Gabriele, and Amit Zunika. "Fatty acids composition and antimicrobial activities of *Litsea garciae* pulp and seed extracts." *Medicinal Plants-International Journal of Phytomedicines and Related Industries* 14.2 (2022): 301-311.
3276. Liu, T., Ruan, S., Mo, Q., Zhao, M., Wang, J., Ye, Z., Chen, L. and Feng, F., 2023. Integrated serum metabolome and gut microbiome to decipher chicken amino acid improvements induced by medium-chain monoglycerides. *Metabolites*, 13(2), p.208.
3277. Liu, T., Ruan, S., Mo, Q., Zhao, M., Wang, J., Ye, Z., Chen, L. and Feng, F., 2023. Evaluation of dynamic effects of dietary medium-chain monoglycerides on performance, intestinal development and gut microbiota of broilers in large-scale production. *Animal Nutrition*.
3278. Liu, Tao, et al. "Dietary medium-chain 1-monoglycerides modulates the community and function of cecal microbiota of broilers." *Journal of the Science of Food and Agriculture* 102.6 (2022): 2242-2252.
3279. Liu, Tao, et al. "Dietary medium-chain α -monoglycerides increase BW, feed intake, and carcass yield in broilers with muscle composition alteration." *Poultry Science* (2020). in press
3280. Liu, Tao, et al. "Glycerol monolaurate enhances reproductive performance, egg quality

- and albumen amino acids composition in aged hens with gut microbiota alternation." *Agriculture* 10.7 (2020): 250.
3281. Liu, Tao, Jun Tang, and Fengqin Feng. "Medium-chain α -monoglycerides improves productive performance and egg quality in aged hens associated with gut microbiota modulation." *Poultry Science* 99(12), 7122-7132.
 3282. Liu MengYun, L. M., Chen XiaoQian, C. X., Zhao HaoBin, Z. H., & Feng FengQin, F. F. (2018). Effect of dietary supplementation with glycerol monolaurate on growth performance, digestive ability and chicken nutritional components of broilers.
 3283. Longmore B. Want the good oil?: It's coconuts to you! *Austr. Pharmacist*, 2013, 32, 4, 73-74.
 3284. Lopes, Leonardo Quintana Soares, et al. "Glycerol monolaurate nanocapsules for biomedical applications: in vitro toxicological studies." *Naunyn-Schmiedeberg's archives of pharmacology* 392.9 (2019): 1131-1140.
 3285. López-Colom, Paola, et al. "Efficacy of medium-chain fatty acid salts distilled from coconut oil against two enteric pathogen challenges in weanling piglets." *Journal of animal science and biotechnology* 10.1 (2019): 89.
 3286. Madrid, C., & Del, A. Efecto de ácidos grasos presentes en leche materna humana en la inducción de diferenciación de células epiteliales intestinales de origen humano. Papel de receptor nuclear PPAR γ (Doctoral dissertation).
 3287. Malá, L., et al. "Susceptibility of *Aeromonas hydrophila* to Medium-Chain Fatty Acids and their Monoesters." (2020).
 3288. Marcq, C., Ginste, J. V., & Naeyaert, W. (2017). Effect of free medium chain fatty acids on zootechnical performance and health of piglets after weaning: a meta-analysis.
 3289. Mazzone, G., Malaj, N., Galano, A., Russo, N., Toscano, M. Antioxidant properties of several coumarin-chalcone hybrids from theoretical insights. *RSC Advances* 2015, 5, 565-575.
 3290. Mbabazi, Immaculate, Phanice Wangila, and Isaac O. K'Owino. "Antimicrobial Activity of *Euclea divinorum* Hern (Ebenaceae) Leaves, Tender Stems, Root Bark and an Herbal Toothpaste Formulated from Its Ethanolic Root Bark Extract." *International Journal of Research and Reports in Dentistry* (2020): 8-16.
 3291. Mbabazi, I. (2021). Phytochemical analysis, antimicrobial activity of *Euclea divinorum* (Magic guarri) leaves, tender stems, root bark extracts and Formulation of a Herbal-based Toothpaste for Dental caries Control (Doctoral dissertation, Moi University).
 3292. Mladenoska, Irina, and Darko Dimitrovski. "Preliminary evaluation of the antimicrobial activity of some spices used as additives in tomato sauce products." *Advanced Technologies* 6.1 (2017): 14-18.
 3293. Mladenoska, I., Temkov, M., Dimitrovski, D. (2017). The effect of monolaurin on the colour and microbiological safety of nitrite reduced sausages. *Advanced Technologies*, 6(2), 11-17.
 3294. Mladenoska, I., Tolev, A., & Kadifkova-Panovska, T. Application of monolaurin as an antimicrobial agent and as an active packaging agent of bakery products.
 3295. Mladenoska, Irina, Vesna Nikolovska, and Lenče Puzderliska. "Model meat pasteurized sausages enriched with monolaurin as nutraceuticals with pronounced antimicrobial properties." *Food and Feed research* 39.2 (2012): 69-71.

3296. Moran Ramirez, Kory Nathaly. "Super-dosing Phytase and Phytogenic Feed Additives to Improve the Performance and Health Status of Young Pigs." (2018).
3297. Mudgil, Poonam. "Antimicrobial Tear Lipids in the Ocular Surface Defense." *Frontiers in Cellular and Infection Microbiology* 12 (2022): 866900.
3298. Musa, Akpemi A., et al. "Phytochemical screening and antimicrobial activity of solvent fractions of *Securidaca longepedunculata* (Fresen) root bark methanol extract." *Journal of Chemical and Pharmaceutical Research* 5.10 (2013): 28-33.
3299. Muskiet, F.A.J, Muskiet, M.H.A, Kuipers, R.S. Failure of the saturated fat hypothesis of cardiovascular diseases. *Nederlands Tijdschrift voor Klinische Chemie en Laboratoriumgeneeskunde*, 37 (3), 2012, 192-211 .
3300. Nehdi, Imededdine Arbi, et al. "Chamaerops humilis L. var. argentea André date palm seed oil: a potential dietetic plant product." *Journal of food science* 79.4 (2014): C534-C539.
3301. Nenti, S. (2023). *Kualitas Kimia Susu Kambing Peternakan Rakyat (Studi Kasus Di Kabupaten Pesawaran Dan Lampung Timur)*.
3302. Nitbani, Febri Odel, et al. "Antimicrobial properties of lauric acid and monolaurin in virgin coconut oil: A review." *ChemBioEng Reviews* (2022).
3303. Omores, R.A. (2021). Extraction, fractionation, nanoparticles formulation, and antimicrobial activity of lipids from black soldier fly larvae.
3304. Pangprasit, Noppason, et al. "Antibacterial properties of lauric acid in combination with organic acids against major pathogens causing dairy mastitis." *Veterinary Integrative Sciences* 19.1 (2020).1-8.
3305. Papadopoulos, Georgios A., et al. "Effects of a tributyrin and monolaurin blend compared to high ZnO levels on growth performance, faecal microbial counts, intestinal histomorphometry and immunohistochemistry in weaned piglets: A field study in two pig herds." *Research in Veterinary Science* 144 (2022): 54-65.
3306. Pluske J. R., Kim J. C., Black J. L. Manipulating the immune system for pigs to optimise performance. *Anim. Prod. Sci.*, 58 (4), 666-680, 2018.
3307. Portocarero, N. "Comparison of caproic acid and monolaurin with zinc oxide and benzoic acid; effect on growth performance of weaned pigs." *Journal of Applied Animal Nutrition* 10.1 (2022): 39-44.
3308. Portocarero, N. "Comparison of medium chain organic acids with zinc oxide and benzoic acid; effect on growth performance of pigs." *Journal of Applied Animal Nutrition* 10.1 (2022): 31-38.
3309. Press, S. (2015). *The Coconut Oil Cure: Essential Recipes and Remedies to Heal Your Body Inside and Out*. Callisto Media, Inc..
3310. Pupala, Sameer Shivaji, et al. "Topical application of coconut oil to the skin of preterm infants: a systematic review." *European journal of pediatrics* 178.9 (2019): 1317-1324.
3311. Qi N., Liu S., Yan F., Chen B., Wu S., Lin X., Yan Z., Zhou Q., Liao S., Li J., Lv M., Cai H., Hu J., Zhang J., Gu Y., Sun M. Study of microencapsulated fatty acid antimicrobial activity in vitro and its prevention ability of *Clostridium perfringens* induced necrotic enteritis in broiler chicken. *Gut Pathogens*, 15 (1), art. no. 1, 2023. DOI: 10.1186/s13099-022-00526-9.
3312. Ragionieri, L., et al. "Effect of the supplementation with a blend containing short and medium chain fatty acid monoglycerides in milk replacer on rumen papillae development

- in weaning calves." *Annals of Anatomy-Anatomischer Anzeiger* 207 (2016): 97-108.
3313. Ramaswamy, Lalitha. "Antimicrobial Properties of *Cocos nucifera*: A Review." *CORD* 31.1 (2015): 6-6.
3314. Ramirez, Kory Nathaly Moran. *Super-dosing Phytase and Phytogenic Feed Additives to Improve the Performance and Health Status of Young Pigs*. North Carolina State University, 2018.
3315. Rani, Sapna, et al. "Antibacterial activity and mechanism of essential oils in combination with medium-chain fatty acids against predominant bovine mastitis pathogens." *Letters in Applied Microbiology* 74.6 (2022): 959-969.
3316. Rengachari, S., Aschauer, P., Sturm, C., & Oberer, M. (2015). Purification, crystallization and preliminary X-ray diffraction analysis of a soluble variant of the monoglyceride lipase Yju3p from the yeast *Saccharomyces cerevisiae*. *Acta Crystallographica Section F: Structural Biology Communications*, 71(2), 242-245.
3317. Righi, Federico, et al. "Adding monoglycerides containing short and medium chain fatty acids to milk replacer: effects on health and performance of preweaned calves." *Italian Journal of Animal Science* 19.1 (2020): 1417-1427.
3318. Rogge, Tina, and José Vanheule. "Animal feed comprising a combination of monoglycerides." U.S. Patent No. 10,398,155. 3 Sep. 2019.
3319. Rongpan, Sudarat, et al. "Anti-Proliferative Effect of Long-Chain Monoglyceride Derivatives on Human Cervical Carcinoma Cell Line." *Journal Of The Medical Association Of Thailand* 100.10 (2017): 165.
3320. Roy, R. N. "Commentary: "Physicochemical Characterization and Antibacterial Activity of the Leaf oil of *Crotalaria pallida* Aiton." *Journal of Meningitis* 1 (2016): 110.
3321. Ruiz-Núñez, Begoña, DA Janneke Dijck-Brouwer, and Frits AJ Muskiet. "The relation of saturated fatty acids with low-grade inflammation and cardiovascular disease." *The Journal of Nutritional Biochemistry* 36 (2016): 1-20.
3322. Saarani, Nur Najiha Binti. "Antibacterial and cytocompatibility analyses on triple layered poly (lactic-co-glycolic acid)/nanoapatite/lauric acid composite membrane." (2016).
3323. Santin, J. *Gorduras saturadas: uma perspectiva da lactação e da composição do leite – Parte 2/3*. Ciencia do Leite, 2011
3324. Santin, J. *Gorduras saturadas: uma perspectiva da lactação e da composição do leite – Parte 3/3*. Ciencia do Leite, 2011.
3325. Saputra, Leo, F. A. R. I. S. Z. A. Gita, and RATNA SARI Dewi. "Effect of 12.5% virgin coconut oil (*cocos nucifera*) mouthwash on plaque index of fixed prosthetic denture users." *Int J App Pharm* 9.2 (2017).
3326. Seca, Ana ML, et al. "Comparative study by GC-MS and chemometrics on the chemical and nutritional profile of *Fucus spiralis* L. juvenile and mature life-cycle phases." *Journal of Applied Phycology* 30.4 (2018): 2539-2548.
3327. Setianto, W. B., et al. "Synthesis of glycerol mono-laurate from lauric acid and glycerol for food antibacterial additive." *IOP Conference Series: Earth and Environmental Science*. Vol. 65. No. 1. IOP Publishing, 2017.
3328. Shah, M.D., Gnanaraj, C., Khan, M.S., Iqbal, M. *Dillenia Suffruticosa* L. Impedes carbon tetrachloride-induced hepatic damage by modulating oxidative stress and

- inflammatory markers in rats. *Journal of Environmental Pathology, Toxicology and Oncology* 2015, 34, 133-152
3329. Shah, Muhammad Dawood, Urban JA D'Souza, and Mohammad Iqbal. "The potential protective effect of *Commelina nudiflora* L. against carbon tetrachloride (CCl₄)-induced hepatotoxicity in rats, mediated by suppression of oxidative stress and inflammation." *Environmental health and preventive medicine* 22.1 (2017): 66.
 3330. Shah, M.D., & Iqbal, M.O.H.A.M.M.A.D. (2018). Antioxidant activity, phytochemical analysis and total polyphenolics content of essential oil, methanol extract and methanol fractions from *Commelinanudiflora*. *Int. J. Pharm. Pharm. Sci*, 10(8), 36-43.
 3331. Shor, S.M. and Schweig, S.K., 2023. The Use of Natural Bioactive Nutraceuticals in the Management of Tick-Borne Illnesses. *Microorganisms*, 11(7), p.1759.
 3332. Siddiqui, R., Khodja, A., Ibrahim, T., Khamis, M., Anwar, A. and Khan, N.A., 2023. The increasing importance of novel deep eutectic solvents as potential effective antimicrobials and other medicinal properties. *World Journal of Microbiology and Biotechnology*, 39(12), p.330.
 3333. Silva, Joana M., et al. "A closer look in the antimicrobial properties of deep eutectic solvents based on fatty acids." *Sustainable Chemistry and Pharmacy* 14 (2019): 100192.
 3334. Sinanoglou, V.J., Koutsouli, P., Fotakis, C., Sotiropoulou, G., Cavouras, D., Bizelis, I. Assessment of lactation stage and breed effect on sheep milk fatty acid profile and lipid quality indices. *Dairy Science and Technology* 2015, 95,509-531
 3335. Sinanoglou, Vassilia J., et al. "Factors affecting human colostrum fatty acid profile: A case study." *PloS one* 12.4 (2017): e0175817.
 3336. Slozhenkina, M. I., et al. "Possible replacing antibiotics with natural feed supplements in poultry farming." *IOP Conference Series: Earth and Environmental Science*. Vol. 677. No. 2. IOP Publishing, 2021.
 3337. Smith, H. E., et al. Literatuurstudie en in vitro onderzoek naar antibacteriële werking van voeradditieven ter vermindering van de *Streptococcus suis* problematiek= Desk study and in vitro analysis of antibacterial effects of feed additives to reduce *Streptococcus suis* in the field. No. 760. Wageningen UR Livestock Research, 2014.
 3338. Strunk T., Pupala S., Hibbert J., Doherty D., & Patole, S. (2018). Topical Coconut Oil in Very Preterm Infants: An Open-Label Randomised Controlled Trial. *Neonatology*, 113(2), 146-151.
 3339. Subramaniam, S., Elz, A., Wignall, A. et al. (2023). Self-emulsifying drug delivery systems (SEDDS) disrupt the gut microbiota and trigger an intestinal inflammatory response in rats. *International Journal of Pharmaceutics*, 648, 123614.
 3340. Sumarheni, Sudir, et al. "Effect of refined coconut oil intake on blood glucose, cholesterol, and leukocyte count of rats (*Rattus norvegicus*)." *Drug Invention Today* 12.4 (2019): 721-726.
 3341. Suwanpiwat, A., W. Petpichetchian, K. Maneewat. The potential effect of the oral self-care program combining coconut oil pulling on mucositis of patients with cancer undergoing chemotherapy: a pilot study. *Thai Cancer J.*, 2013, 33, 41-52.
 3342. Szabó, R.T., Kovács-Weber, M., Zimborán, Á., Kovács, L. and Erdélyi, M., 2023. Effects of Short-and Medium-Chain Fatty Acids on Production, Meat Quality, and Microbial Attributes—A Review. *Molecules*, 28(13), p.4956.
 3343. Szabó, R.T., Kovács-Weber, M., Zimborán, Á., Kovács, L. and Erdélyi, M., 2023.

Effects of Short and Medium-Chain Fatty Acids on Production, Meat Quality and Gut Hygiene of Broiler Chickens—A Review.

3344. Tsuda, Kentaro, et al. "Effects of alkyl gallates, fatty acids, and acylglycerols on the growth of the psychrotolerant bacterium *Sporosarcina* sp. S92h." *Biocatalysis and agricultural biotechnology* 17 (2019): 294-298.
3345. Umerska, Anita, et al. "Antibacterial action of lipid nanocapsules containing fatty acids or monoglycerides as co-surfactants." *European Journal of Pharmaceutics and Biopharmaceutics* 108 (2016): 100-110.
3346. Umerska, Anita, et al. "Synergistic interactions between antimicrobial peptides derived from plectasin and lipid nanocapsules containing monolaurin as a cosurfactant against *Staphylococcus aureus*." *International journal of nanomedicine* 12 (2017): 5687-5699.
3347. van der Gaag, Ellen José, Romy Wieffer, and Judith van der Kraats. "Advising Consumption of Green Vegetables, Beef, and Full-Fat Dairy Products Has No Adverse Effects on the Lipid Profiles in Children." *Nutrients* 9.5 (2017): 518.
3348. van der Kraats, Judith. Een retrospectieve studie naar het effect van groene groenten, rundvlees, volle melk en roomboter op het lipidenprofiel bij kinderen. Diss. 2014.
3349. Vltavská, Pavlína, et al. "Antifungal and antibacterial effects of 1-monocaprylin on textile materials." *European Journal of Lipid Science and Technology* 114.7 (2012): 849-856.
3350. Wadhvani, K.N., Bhavsar, M., Islam, M.M., Patel, J.H. and Lunagariya, P.M., 2023. Modern health concept of non-bovine milk: A review. *Indian Journal of Animal Production and Management*, 37(1), pp.12-22.
3351. WANG, WENYUE, et al. "In Vitro Antibacterial Activities and Mechanisms of Action of Fatty Acid Monoglycerides against Four Foodborne Bacteria." *Journal of Food Protection* 83.2 (2020): 331-337.
3352. Wang, Yajuan, et al. "Preparation of porous polyaminobenzenesulfonic acid and synthesis of glycerol monolaurate." *Chemical Papers* 76.4 (2022): 2431-2445.
3353. Wang, Yuchao, et al. "Dietary glycerol monolaurate improved the growth, activity of digestive enzymes and gut microbiota in zebrafish (*Danio rerio*)." *Aquaculture Reports* 20 (2021): 100670.
3354. Yoon, B.K. (2018). Investigating membrane morphological responses induced by antimicrobial lipids and correlations with biological activity (Doctoral dissertation).
3355. Yoon, B.K., Tan, S.W., Tan, J.Y.B., Jackman, J.A., Cho, N.-J. "Nanoarchitectonics-based model membrane platforms for probing membrane-disruptive interactions of odd-chain antimicrobial lipids." *Nano Convergence* 9.1 (2022): 1-13.
3356. Yoon, Bo Kyeong, et al. "Competing Interactions of Fatty Acids and Monoglycerides Trigger Synergistic Phospholipid Membrane Remodeling." *The Journal of Physical Chemistry Letters* (2020) 11(13), 4951-4957.
3357. Yosief, H.O., & Sarker, M.I. (2021). Naturally derived fatty acid based antibacterial agents. In *Conversion of Renewable Biomass into Bioproducts* (pp. 91-117). American Chemical Society.
3358. Yuliana, Renita, et al. "Daya Antimikrobia Sarang Lebah Madu *Trigona* spp terhadap Mikrobia Patogen." *Bioedukasi: Jurnal Pendidikan Biologi* 8.1 (2015): 67-72.
3359. Zhang, J., Liu, T., Tang, J., Zhang, X., Feng, F., Cai, H. and Zhao, M., 2023. Food

- additive glycerol monocaprylate modulated systemic inflammation and gut microbiota without stimulating metabolic dysfunction in high-fat diet fed mice. *Food Research International*, 167, p.112734.
3360. Zhang, Song, et al. "The identification of critical lethal action in antimicrobial mechanism of glycerol monomyristate against foodborne pathogens." *bioRxiv* (2018): 336354.
3361. Zhao, Min-jie, et al. "Effects of dietary glycerol monolaurate on productive performance, egg quality, serum biochemical indices, and intestinal morphology of laying hens." *Journal of Zhejiang University-SCIENCE B* 20.11 (2019): 877-890.
- 96). Batovska, D., Parushev, S., Stamboliyska, B., Tsvetkova, I., Ninova, M., NAJDENSKI, H. Examination of growth inhibitory properties of synthetic chalcones for which antibacterial activity was predicted (2009) European Journal of Medicinal Chemistry, 44 (5), pp. 2211-2218.**
3362. Abdullah, Maryam A., Siti Munirah Mohd Faudzi, and Nadiah Mad Nasir. "A Review on Biological Properties and Synthetic Methodologies of Diarylpentadienones." *Mini reviews in medicinal chemistry* 21.9 (2021): 1058-1070.
3363. Abdullah, Maryam Aisyah, et al. "Development of diarylpentadienone analogues as alpha-glucosidase inhibitor: Synthesis, in vitro biological and in vivo toxicity evaluations, and molecular docking analysis." *Bioorganic Chemistry* 104 (2020): 104277.
3364. Ahamed, A. Asrar, and M. Mohamed Sihabudeen. "Synthesis and biological evaluation of some novel heterocyclic Chalcone derivatives." *Asian Journal of Pharmacy and Pharmacology* 3.6 (2017): 247-250.
3365. Aksöz, B. Evranos, and Rahmiye Ertan. "Chemical and structural properties of chalcones I." *FABAD J Pharm Sci* 36 (2011): 223-242.
3366. Aksöz, Begüm Evranos, and Rahmiye Ertan. "Spectral properties of chalcones II." *Fabard J. Pharm. Sci* 37.4 (2012): 205-216.
3367. Ali A., Din Z.U., Ibrahim M., Ashfaq M., Muhammad S., Gull D., Tahir M.N., Rodrigues-Filho E., Al-Sehemi A.G., Suleman M. Acid catalyzed one-pot approach towards the synthesis of curcuminoid systems: unsymmetrical diarylidene cycloalkanones, exploration of their single crystals, optical and nonlinear optical properties. *RSC Advances*, 13 (7), pp. 4476 – 4494, 2023. DOI: 10.1039/d2ra07681k.
3368. Ali, Akbar, et al. "Crystal and Quantum Chemical Exploration of the Potent Monocarbonyl Curcuminoids to Unveil Their Structural and Intriguing Electronic Properties." *ChemistrySelect* 5.12 (2020): 3735-3745.
3369. Ali, Akbar, et al. "Exploration of structural, electronic and third order nonlinear optical properties of crystalline chalcone systems: Monoarylidene and unsymmetrical diarylidene cycloalkanones." *Journal of Molecular Structure* 1241 (2021): 130685.
3370. Almutaleb, Arabya AA, et al. "Synthesis, Characterization and Biological Activity of (E)-9-Benzylidene-5-phenyl-5, 6, 7, 8, 9, 10-hexahydro-[1, 2, 4] triazolo [3, 4-a] quinazoline." *Jordan Journal of Chemistry (JJC)* 15.3 (2020): 111-118.
3371. Alrubaie, Leaqaa A., Raheem J. Muhasin, and Mazin N. Mousa. "Synthesis, characterization and evaluation of antiinflammatory properties of novel α , β -unsaturated ketones." *Tropical Journal of Pharmaceutical Research* 19.1 (2020): 147-154.
3372. Ashburn, Bradley O. "Computational Analysis of a Series of Chlorinated Chalcone

- Derivatives." Computational Chemistry 7.4 (2019): 106-120.
3373. Ashok, D., K. Sudershan, and M. Khalilullah. "Solvent-free microwave-assisted synthesis of E-(1)-(6-benzoyl-3, 5-dimethylfuro [3', 2': 4, 5] benzo [b] furan-2-yl)-3-(aryl)-2-propen-1-ones and their antibacterial activity." Green Chemistry Letters and Reviews 5.2 (2012): 121-125.
3374. Asiri, A.M. and S.A. Khan. (2E)-3-(3,5-dimethyl-1-phenyl-1H-pyrazol-4-yl)-1-(2,5-dimethyl-3-furanyl)prop-2-en-1-one. MolBank. 2010, 2010 (2), 1-3. ISSN:1422-8599
3375. Avupati, V. R., & Yejella, R. P.. Chalcones: a mini review. World J. Pharm. Pharmaceut. Sci. 2014, 3, 1713-1742
3376. Bala, Daniela, et al. "Design, Synthesis, and Biological Evaluation of New Azulene-Containing Chalcones." Materials 15.5 (2022): 1629.
3377. Bale A. T., Khan K. M., Salar U., Chigurupati S., Fasina T., Ali F., Kanwal, Wadood A., Taha M., Nanda S. S., Ghufraan M., Perveen, S. Chalcones and bis-chalcones: As potential α -amylase inhibitors; synthesis, in vitro screening, and molecular modelling studies. Bioorg. Chem., 79, 179-189, 2018.
3378. Baydere, Cemile, et al. "Crystal structure and Hirshfeld surface analysis of (E)-2-(2, 4, 6-trimethylbenzylidene)-3, 4-dihydronaphthalen-1 (2H)-one." Acta Crystallographica Section E: Crystallographic Communications 75.6 (2019): 746-750.
3379. Biológica, L. E. Q. F. Uso De Herramientas Quimioinformáticas Para La Predicción De Potenciales Actividades Biológicas De Moléculas Con Núcleo Chalcona.
3380. Biswal, Priyabrata, et al. "Cobalt (II) porphyrin-Mediated Selective Synthesis of 1, 5-Diketones via an Interrupted-Borrowing Hydrogen Strategy Using Methanol as a C1 Source." The Journal of Organic Chemistry (2021).
3381. Bogi, Ana. Priprave halkona i njihovih oksima. Diss. University of Zagreb. Faculty of Science. Department of Chemistry, 2018.
3382. Božić, Dragana D. "Antimikrobna aktivnost halkona i in vitro uticaj na fiziološko-biohemijske karakteristike i ekspresiju faktora virulencije meticilin-rezistentnih sojeva Staphylococcus aureus." Универзитет у Београду (2014).
3383. Burmaoglu, S., Kazancioglu, E.A., Kazancioglu, M.Z., et al."Synthesis, In Vitro Biological Evaluation, and Molecular Docking Studies of Novel Biphenyl Chalcone Derivatives as Antimicrobial Agents". Polycyclic Aromatic Compounds, 42(9), 2022, 5948-5961.
3384. Caboni, Pierluigi, et al. "Nematicidal activity of acetophenones and chalcones against Meloidogyne incognita and structure–activity considerations." Pest management science 72.1 (2016): 125-130.
3385. Carvalho, D. S. (2022). Estudo de chalconas e sua utilização em protetores solares.
3386. Cushnie, TP Tim, and Andrew J. Lamb. "Recent advances in understanding the antibacterial properties of flavonoids." International journal of antimicrobial agents 38.2 (2011): 99-107.
3387. da Cunha Xavier, J., dos Santos, H.S., Machado Marinho, M. et al. (2024). Chalcones As Potent Agents Against Staphylococcus aureus: A Computational Approach. Letters in Drug Design & Discovery, 21(4), 684-700.
3388. Daglia, Maria. "Polyphenols as antimicrobial agents." Current opinion in biotechnology 23.2 (2012): 174-181.
3389. Dan, Wenjia, and Jiangkun Dai. "Recent developments of chalcones as potential

- antibacterial agents in medicinal chemistry." *European Journal of Medicinal Chemistry* 187 (2020): 111980.
3390. Danova A., Nguyen D.V., Toyoda R., Mahalapbutr P., Rungrotmongkol T., Wonganan P., Chavasiri W. 3',4',5'-trimethoxy- and 3,4-dimethoxychalcones targeting A549 cells: Synthesis, cytotoxic activity, and molecular docking. *Journal of Molecular Structure*, 1275, art. no. 134572, 2023. DOI: 10.1016/j.molstruc.2022.134572.
3391. Dar, Bilal Ahmad, et al. "Synthesis and screening of ursolic acid-benzylidene derivatives as potential anti-cancer agents." *European Journal of Medicinal Chemistry* 111 (2016): 26-32.
3392. de Oliveira Santos, Giselle C., et al. "Candida infections and therapeutic strategies: mechanisms of action for traditional and alternative agents." *Frontiers in microbiology* 9 (2018): 1351.
3393. Debnath, S. (2017). Design, Synthesis, Characterization of Some New Pyrazoline Derivatives and Evaluation for Their Antimicrobial Profiles (Doctoral dissertation, Rajiv Gandhi University of Health Sciences (India)).
3394. Din, Z.Ud, T.P. Fill, F.F. de Assis, et al. Unsymmetrical 1, 5-diaryl-3-oxo-1, 4-pentadienyls and their evaluation as antiparasitic agents. *Bioorg. Med. Chem.*, 2014, 22, 3, 1121-1127.
3395. Din, Zia Ud, and Edson Rodrigues-Filho. "Optimized one-pot synthesis of monoarylidene and unsymmetrical diarylidene cycloalkanones." *Arabian Journal of Chemistry* 12.8 (2019): 4756-4763.
3396. Din, Zia Ud, et al. "Phytotoxicity, structural and computational analysis of 2-methyl-1, 5-diarylpentadienones." *Journal of Molecular Structure* 1142 (2017): 239-247.
3397. Eddarir, Said, Mohammed Kajjout, and Christian Rolando. "An efficient synthesis of (Z)- α -fluorochalcones via the palladium-catalyzed cross-coupling reaction of (Z)- α -fluorocinnamoyl chloride with boronic acids." *Tetrahedron* 69.6 (2013): 1735-1738.
3398. Evranos Aksöz, B. and R. Ertan. Chemical and structural properties of chalcones I. *Fabad J. Pharm. Sci.*, 2011, 36(4), 223-242.
3399. Evranos-Aksöz, Begüm, Fatma Kaynak Onurdağ, and Selda Özgen Özgacar. "Antibacterial, antifungal and antimycobacterial activities of some pyrazoline, hydrazone and chalcone derivatives." *Zeitschrift für Naturforschung C* 70.7-8 (2015): 183-189.
3400. Fernandes, Thaís Alves. "Estudo do efeito e do mecanismo de ação de chalconas derivativas na secreção de insulina: ilhotas pancreáticas." (2018).
3401. Fernandes, W.B., L.A. Malaspina, F.T. Martins. Conformational variability in a new terpenoid-like bischalcone: structure and theoretical studies. *J. Struct. Chem.*, 2013, 54, 6, 1112-1121.
3402. Ferraz, Carlos AN, et al. "Potentiation of antibiotic activity by chalcone (E)-1-(4'-aminophenyl)-3-(furan-2-yl)-prop-2-en-1-one against gram-positive and gram-negative MDR strains." *Microbial Pathogenesis* 148 (2020): 104453.
3403. Fourman, Cody. "PharmaFlights: Fragment based drug discovery based on chalcones with a 3, 4, 5-trimethoxy substitution on ring B." (2018).
3404. Garg, S., I. Ravish, N. Raghav. Spectrophotometric analysis of bovine serum albumin in presence of 3-phenyl-1-(pyridin-2-yl) prop-2-en-1-ones. *Int. J. ChemTech. Res.*, 2013, 5, 5, 2338-2343.
3405. Ghoneim, Amira A., Rehab M. Elbargisy, and Afaf Manoer. "Design and synthesis of

- heterocyclic Compounds from 1, 4-diacetylbenzene with Expected Antimicrobial Activity." *Egyptian Journal of Chemistry* 63.8 (2020): 9-10.
3406. Göç, F., Sarı, A., Özer, B. et al. (2024). Antimicrobial activities of *Scorzonera ketzkhovellii* Sosn. ex Grossh.(Asteraceae) and determination of natural compounds by LC-HRMS analysis. *Istanbul Journal of Pharmacy*, 54(1).
3407. Gomes, Marcelo N., et al. "QSAR-driven design, synthesis and discovery of potent chalcone derivatives with antitubercular activity." *European Journal of Medicinal Chemistry* 137 (2017): 126-138.
3408. Gonsalves, I. S., and A. R. Shaikh. "2D QSAR Analysis of 3', 4', 5'-trimethoxychalcone analogues as inhibitors of nitric oxide production and tumor cell proliferation." *J Comput Methods Mol Des* 2.1 (2012): 24-38.
3409. Gonzaga, G.R.R. (2018). Nitro-E Aminoflavonas Como Agentes Antibacterianos (Master's thesis, Universidade de Aveiro (Portugal)).
3410. Guil-Guerrero, J. L., et al. "Antimicrobial activity of plant-food by-products: A review focusing on the tropics." *Livestock Science* 189 (2016): 32-49.
3411. Guil-Guerrero, J. L., et al. "Plant-food by-products to improve farm-animal health." *Animal Feed Science and Technology* 220 (2016): 121-135.
3412. Guimarães Neto, José Jamil Aniz. "Estudo cristalográfico da chalcona fluorada C18H17FO como potencial pesticida." (2018).
3413. Harouak, Hazim, Jamal Ibijbijen, and Laila Nassiri. "Chemical profile of *Tetraclinis articulata* (Vahl) Masters, and *Juglans regia* L. and *Olea europaea* L. var. *Sylvestris* used against oral diseases: in vitro analysis between polyphenolic content and aqueous extraction optimization." *Heliyon* 7.5 (2021): e07118.
3414. Hoepfner, A., Petzer, A., Petzer, J.P. et al. (2023). In vitro and in silico antibacterial evaluation of nitrocatechol chalcone and pyrazoline derivatives. *Results in Chemistry*, 6, 101194.
3415. Jaime, V.B.J. Solvent-free synthesis of ferrocenylchalcones. *Int.J. Chem.Tech. Res.*2014,6(1), 138-146.
3416. Jain, K., & Jain, N.K. (2020). Synthesis and Anti-Microbial Activity of Novel Substituted Chalcone Derivatives. *Annals of the Romanian Society for Cell Biology*, 1426-1439.
3417. Jurić, Tatjana. Ispitivanje biljnih vrsta *Alchemilla vulgaris* L. i *Satureja hortensis* L.: fitohemijski profil i biološka aktivnost u in vitro i in vivo uslovima. Diss. Универзитет у Крагујевцу, Природно-математички факултет, 2020.
3418. Kamal, Ahmed, et al. "Synthesis and anticancer activity of 4 β -alkylamidochalcone and 4 β -cinnamido linked podophyllotoxins as apoptotic inducing agents." *European journal of medicinal chemistry* 47 (2012): 530-545.
3419. Kammari K., Devaraya K., Swain S., Kondapi A.K. The topoisomerase II β -kinase associated with HIV-1 is a potential target for pyridine-bis-chalcones' anti-HIV-1 activity. *European Journal of Medicinal Chemistry*, 259, art. no. 115623, 2023. DOI: 10.1016/j.ejmech.2023.115623.
3420. Kanchana, V., Mayavan, L., Kistan, A., & Mohan, S. (2024). SPECTRAL ELUCIDATION, ANTIMICROBIAL AND ANTIOXIDANT STUDY OF NEWLY SYNTHESIZED PYRAZOLINE DERIVATIVES. *Rasayan Journal Of Chemistry*, 17(1).
3421. Kauffmann A.C., Castro V.S. Phenolic Compounds in Bacterial Inactivation: A

- Perspective from Brazil. *Antibiotics*, 12 (4), art. no. 645, 2023. DOI: 10.3390/antibiotics12040645.
3422. Kazmi, Madiha, et al. "Developing new hybrid scaffold for urease inhibition based on carbazole-chalcone conjugates: Synthesis, assessment of therapeutic potential and computational docking analysis." *Bioorganic & Medicinal Chemistry* 27.22 (2019): 115123.
 3423. Ke, C., W. Di, X. Wei, et al. Latest progress antibacterial activity of polyphenol compounds *Food Ind. Technol.*, 2012, 33 (17), 405-408.
 3424. Khalid, Muhammad, et al. "Facile preparation, characterization, SC-XRD and DFT/DTDFT study of diversely functionalized unsymmetrical bis-aryl- α , β -unsaturated ketone derivatives." *Journal of Molecular Structure* 1206 (2020): 127755.
 3425. Kim, Daon, et al. "Synthesis of higher carbon sugars thio-functionalized with heterocycles." *Abstracts Of Papers Of The American Chemical Society*. 257. 1155 16TH ST, NW, Washington, DC 20036 USA: Amer Chemical Soc, 2019.
 3426. Kim, Yuri A., et al. "Flavonoids determine the rate of fibrillogenesis and structure of collagen type I fibrils in vitro." *International Journal of Biological Macromolecules* (2017).
 3427. Kopel, J., Julianna McD., and Abdul H. "An Assessment of the In Vitro Models and Clinical Trials Related to the Antimicrobial Activities of Phytochemicals." *Antibiotics* 11.12 (2022): 1838.
 3428. Kottapalle, Gajanan D., Nagesh J. Deshmukh, and Avinash T. Shinde. "Growth Inhibitory Properties of Synthetic Chalcones." *Current Bioactive Compounds* 16.6 (2020): 892-899.
 3429. Kumar R, Sharma P, Shard A, et al. Chalcones as promising pesticidal agents against diamondback moth (*Plutella xylostella*): microwave-assisted synthesis and structure–activity relationship. *Med. Chem. Res.*, 21 (6), 2012, 922-931.
 3430. Kumar, B., Kumari, B., Singh, N., Ram, B., Balram, B. Synthesis, characterization and antibacterial activity of some new chalcone derivatives. *Der Pharma Chemica* 2015, 7, 286-291.
 3431. Kumar, Rakesh, et al. "Reinvestigation of structure–activity relationship of methoxylated chalcones as antimalarials: Synthesis and evaluation of 2, 4, 5-trimethoxy substituted patterns as lead candidates derived from abundantly available natural β -asarone." *European journal of medicinal chemistry* 45.11 (2010): 5292-5301.
 3432. Lai, C.-H., Y.K.Rao, S.-H.Fang, Y.-T.Sing, Y.-M.Tzeng, Identification of 3',4',5'-trimethoxychalcone analogues as potent inhibitors of *Helicobacter pylori*-induced inflammation in human gastric epithelial cells, *Bioorg. Med. Chem. Lett.*, DOI: 10.1016/j.bmcl.2010.07.094.
 3433. Liu, L., Feng, S., Li, C. A green synthesis of highly substituted 1,5-diketones. *RSC Advances* 2015, 5, 56949-56953
 3434. Mahalakshmia, K., Tharmaraj, P., Sheela, C.D. Binuclear Schiff base ligand and its metal (II) complexes derived from 1, 6-diaminohexane and 2, 2'-dihydroxysalicylalacetone.
 3435. Manoer, A., & Elbargisy, R.M. (2021). Synthesis of bis chalcones and transformation into bis heterocyclic compounds with expected antimicrobial activity. *Indian Journal of Chemistry-Section B (IJC-B)*, 60(9), 1272-1278.

3436. Mansoorian, Bahareh. Effect of food matrix interaction between dietary fibre and polyphenols on their metabolism by colonic bacteria. Diss. University of Glasgow, 2014.
3437. Maravalho, Izabella Vitoria Souza. "Síntese de chalconas e seus derivados heterocíclicos com potencial fotofísico." (2019).
3438. Marcoux, Eve. Propriétés antibactériennes envers *Enterococcus faecalis* et innocuité de quatre composés naturels et de la nisine: une étude in vitro. Diss. Université Laval, 2019.
3439. Marques, B.C., Santos, M.B., Ayusso, G.M. et al. Avaliação da Inibição do Crescimento de *Paracoccidioides brasiliensis* por Chalconas Derivadas de Apocinina.
3440. Mazzone G., Malaj N., Galano A., Russo N., Toscano M. Antioxidant properties of several coumarin–chalcone hybrids from theoretical insights RSC Adv., 2015, 5, 565-575
3441. Meier, Dieter. Screening for novel antimicrobial agents and elucidation of the corresponding mechanisms. Diss. 2020.
3442. Mikłasińska-Majdanik M., Kępa M., Wojtyczka R., Idzik D., Wąsik T. (2018). Phenolic Compounds Diminish Antibiotic Resistance of *Staphylococcus Aureus* Clinical Strains. Int J Environ Res Public Health, 15 (10), 2321, 2018.
3443. Ming, Liew Suk, et al. "Synthesis, characterization, antifungal activities and crystal structure of thiophene-based heterocyclic chalcones." Chemical Data Collections 9 (2017): 104-113.
3444. Mitrev, Yavor N., et al. "Original enzyme-catalyzed synthesis of chalcones: Utilization of hydrolase promiscuity." Journal of the Serbian Chemical Society 81.11 (2016): 1231-1237.
3445. Mojarrad, J.S., G. Zarrini, H. Nazemiye, M.R. Zavareh, Z. Khoshkam and D. Asgari. Synthesis, antimicrobial and antioxidant evaluations of allyloxy chalcone derivatives. Pharm. Sci., 2011, 17(1), 65-73.
3446. Mora, C.L., J. Castaño, M.C. Jaramillo. Actividad inhibitoria de dihidroxifenilpropenona sobre β -lactamasa de *Enterobacter cloacae*: estudio preliminar en el desarrollo de fármacos para enfrentar la resistencia bacteriana. Biomédica, 2013, 34 (Supl. 1), 114-123.
3447. Mora, C.L., Castaño, J., & Jaramillo, M.C. (2014). Inhibitory activity of dihydroxy-phenyl-propenone on betalactamase of *Enterobacter cloacae*: Preliminary study for drug development to overcome bacterial resistance. Biomédica, 34, 114-23.
3448. Moreira, Caua A., et al. "Structural insights and antioxidant analysis of a tri-methoxy chalcone with potential as a diesel-biodiesel blend additive." Fuel Processing Technology 227 (2022): 107122.
3449. Mostafa, Sara M., et al. "An efficient approach for the synthesis of novel series of 1, 3-dihydrospiro [indene-2, 6-[1, 3] thiazine] derivatives." Monatshefte für Chemie-Chemical Monthly 153.1 (2022): 87-94
3450. Mowlana, M. Yaseen, and A. J. A. Nasser. "Synthesis and Molecular Docking studies of Heterocyclic Chalcone Derivatives as BRCA1 inhibitors." International Journal of Pharmaceutical Chemistry 3 (2015): 196-200.
3451. Murtaza, Shahzad, et al. "Synthesis and Evaluation of Chalcone and its Derivatives as Potential Anticholinergic Agents." Letters in Drug Design & Discovery 16.3 (2019): 322-332.
3452. Muškinja, Jovana M., et al. "Synthesis and anticancer activity of chalcone analogues

- with sulfonyl groups." *Medicinal Chemistry Research* 28.3 (2019): 279-291.
3453. Nawaz, T., Tajammal, A., Qurashi, A.W. et al. (2024). Synthesis, Antibacterial, Antibiofilm, and Docking Studies of Chalcones Against Multidrug Resistance Pathogens. *Heliyon*.
 3454. Ngameni, B., et al. "Synthesis and evaluation of anticancer activity of O-allylchalcone derivatives." *medicinal chemistry* 3.3 (2013): 233-237.
 3455. Niu, C., Li, G., Tuerxuntayi, A., Aisa, H.A. Synthesis and bioactivity of new chalcone derivatives as potential tyrosinase activator based on the click chemistry *Chinese Journal of Chemistry* 33, 2015, 486-494
 3456. Niu, Chao, and Haji A. Aisa. "Upregulation of Melanogenesis and Tyrosinase activity: potential agents for Vitiligo." *Molecules* 22.8 (2017): 1303.
 3457. Nixha, A.R., Arslan, M., Atalay, Y., et al. Synthesis and theoretical calculations of carbazole substituted chalcone urea derivatives and studies their polyphenol oxidase enzyme activity. *J. Enz. Inhib. Med. Chem.* 4, 28, 2013, 808-815.
 3458. Ogle, Wesley Guy Cheslan. Synthesis, characterization and antioxidant activity of prenylated and fluorine based flavonoids. Diss. 2013.
 3459. Olender, D., Sowa-Kasprzak, K., Pawełczyk, A., Skóra, B., Zaprutko, L. and Szychowski, K.A., 2024. Curcuminoid Chalcones: Synthesis and Biological Activity against the Human Colon Carcinoma (Caco-2) Cell Line. *Current Medicinal Chemistry*.
 3460. Pang, D., Liu, Fa, Shi, Y., Liu, J., et al. Antibacterial activity of 10 phenolic compounds from mulberry. *J. China Pharm. University*, 2014, 45, 2, 221-226.
 3461. Paramesh, M. Synthesis and biological evaluation of novel pyrazole derivatives. *Pharm. Chem.*, Rajiv Gandhi University of Health Sciences, Karnataka, Bangalore, 2010 <http://hdl.handle.net/123456789/546>
 3462. Paramesh, M., Niranjana, M.S., Niazi, S., et al. Synthesis and antimicrobial study of some chlorine containing chalcones. *Int. J. Pharm. Pharmac. Sci.*, 2010, 2(2), 113-117.
 3463. Rafiee, E., F. Rahimi. Synthesis of biologically active chalcon analogues via claisen-schmidt condensation in solvent-free conditions: supported mixed addenda heteropoly acid as a heterogeneous catalyst. *J. Chil. Chem. Soc.*, 2013, 58, 3, 1926-1929. <http://dx.doi.org/10.4067/S0717-97072013000300029>
 3464. Rai, S., P. N. Patel, and A. Chadha. "Preparation, characterisation, and crystal structure analysis of (2E, 2' E)-3, 3'-(1, 4-phenylene) bis (1-(2-aminophenyl) prop-2-en-1-one)." *Crystallography Reports* 61.7 (2016): 1086-1089.
 3465. Ramakrishna, A., V. Sirisha, M.A. Jose, B. Narasimharao, B. Deepthi, P. Ramankumar, M. Soumya and T. Ramanamma. Synthesis and evaluation of anti-bacterial activity of some new benzofuran chalcone derivatives. *Indian Drugs*. 2011, 48(4), 25-29.
 3466. Rao, Yerra Koteswara, Shih-Hua Fang, and Yew-Min Tzeng. "Synthesis and biological evaluation of 3', 4', 5'-trimethoxychalcone analogues as inhibitors of nitric oxide production and tumor cell proliferation." *Bioorganic & medicinal chemistry* 17.23 (2009): 7909-7914.
 3467. Rashid, Faisal, et al. "New Hybrid Scaffolds Based on Carbazole-Chalcones as Potent Anticancer Agents." *Anti-Cancer Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Anti-Cancer Agents)* 21.9 (2021): 1082-1091.
 3468. Raia, S., Patela, P. N., Chadha, A. (2016). Structure Of Organic Compounds. *Crystallography Reports*, 61(7).

3469. Reid, Rachael, et al. "Controlling blown pack spoilage using anti-microbial packaging." *Foods* 6.8 (2017): 67.
3470. rights are reserved by Bradley, A., & Ashburn, O. Synthesis and Antimicrobial Evaluation of a Series of Chlorinated Chalcone Derivatives.
3471. Russell, M., Soiket, M.I.H. Some new azole type heterocyclic compounds as antifungal agents. *Organic Communications* 2015, 7, 114-122
3472. Saidugari, Swamy, et al. "Synthesis, Characterization and Antimicrobial Evaluation of Novel (E)-N'-(4-(1-((3, 4-dimethoxypyridin-2-yl) methyl)-1H-1, 2, 3-triazol-4-yl) benzylidene) benzohydrazide Derivatives." *Oriental Journal of Chemistry* 32.4 (2016): 2155-2161.
3473. Salum, B., Altei, W.F., Chiaradia, L.D., et al. A. Cytotoxic 3,4,5-trimethoxychalcones as mitotic arresters and cell migration inhibitors. *Eur. J. Med. Chem.*, 63, 2013, 501-510.
3474. Sankaran S., Balasubramanian R. Insight into the lipophilicity of selected monosubstituted chalcones. *Pak. J. Pharm. Sci.*, 31 (3), 941-946, 2018.
3475. Santos G. C. D. O., Vasconcelos C. C., Lopes A. J. O., Cartagenaes M. D. S. D. S., Barros Filho A. K. D., Nascimento F. R., Ramos R. M., Pires E. R. R. B, de Andrade M. S., Rocha F. M. G., Monteiro, C. D. A. Candida infections and therapeutic strategies: mechanisms of action for traditional and alternative agents. *Front. Microbiol.*, 9, 1351, 2018.
3476. Sarbu, L. G., et al. "Synthetic flavonoids with antimicrobial activity: a review." *Journal of applied microbiology* 127.5 (2019): 1282-1290.
3477. Sarkı, Gülpınar, et al. "Synthesis, characterization and electrochemical studies of metal-free and metallophthalocyanines containing two different chalcone units substituted on peripherally positions." *Journal of Molecular Structure* 1196 (2019): 592-603.
3478. Saroj, Manju K., Neera Sharma, and Ramesh C. Rastogi. "Photophysical study of some 3-benzoylmethyleneindol-2-ones and estimation of ground and excited states dipole moments from solvatochromic methods using solvent polarity parameters." *Journal of Molecular Structure* 1012 (2012): 73-86.
3479. Saroj, Manju Kumari, Neera Sharma, and Ramesh C. Rastogi. "Solvent effect profiles of absorbance and fluorescence spectra of some indole based chalcones." *Journal of fluorescence* 21.6 (2011): 2213-2227.
3480. Sedighi V, Azerang P, Soroush Sardari S. Synthesis of dibenzalacetone derivatives and evaluation of their antimycobacterial property. *Int. Conf. Adv. Biol. Pharmac. Sci. (ICABPS'2012)* March 24-25, 2012, Dubai.
3481. Senthilkumar, G., K. Neelakandan and H. Manikandan. A convenient, green, solvent free synthesis and characterization of novel fluoro chalcones under grind-stone chemistry. *Pelagia Research Library, Der Chemica Sinica*, 2014, 5, 2, 106-113
3482. Senthilkumar, N., et al. "Synthesis And Biological Evaluation of New S-Mannich Bases of 3-Methyl-4-Phenyl-3, 4, 5, 6, 7, 8, Hexahydroquinazoline-2 (1h)-Thione." *Asian Journal of Research in Chemistry* 4.10 (2011): 1573-1577.
3483. Shah S., N.N., H. Md. Ziauddin, M. Zameer, J.A. Dhole, T. Khan and M.A. Baseer. Synthesis and antimicrobial studies of some novel pyrrolidine chalcones. *Der Pharmacia Lettre*. 2011, 3(1), 180-184.
3484. Shahbazi, .M.J., Zarrini, G., Nazemiye H. Et al. Synthesis, antimicrobial and antioxidant evaluations of allyloxy chalcone derivatives. *Pharm. Sci.*, 2011, 17, 1, 65-73.

3485. Song M. X., Deng X. Q. Synthesis and Biological Evaluation of Uncharged Chalcone Derivatives as Novel PTP1B Inhibitors. *Lat. Am. J. Pharm.*, 37 (1), 68-72, 2018.
3486. Sonmez, Fatih, et al. "Evaluation of new chalcone derivatives as polyphenol oxidase inhibitors." *Bioorganic & medicinal chemistry letters* 21.24 (2011): 7479-7482.
3487. Sood, A. and Kesavan, V., 2023. Synthesis and antibacterial activity of 2-benzylidene-3-oxobutanamide derivatives against resistant pathogens. *RSC Medicinal Chemistry*, 14(9), pp.1817-1826.
3488. Souza, P.B.D.N. (2015). Estudos visando a síntese de novos derivados azólicos substituídos análogos da chalcona com potencial atividade biológica. Trabalho de Conclusão de Curso (Bacharelado em Farmácia)-Colegiado de Ensino de Graduação-Macaé, Universidade Federal do Rio de Janeiro, Macaé.
3489. Srinivasan, B., Johnson, T.E., Lad, R., Xing, C. (2009). Structure– activity relationship studies of chalcone leading to 3-hydroxy-4, 3', 4', 5'-tetramethoxychalcone and its analogues as potent nuclear factor κ B inhibitors and their anticancer activities. *Journal of medicinal chemistry*, 52(22), 7228-7235.
3490. Steffi, I.G., & Shaikh, A.R. (2012). 2D QSAR Analysis of 3', 4', 5'-Trimethoxychalcone analogues as inhibitors of nitric oxide production and tumor cell proliferation. *Journal of Computational Methods in Molecular Design*, 2(1), 24-38.
3491. Stompor M., Potaniec B., Szumny A., Zielinski P., Zonierczyk AK., Anioł M. Microbial synthesis of dihydrochalcones using *Rhodococcus* and *Gordonia* species. *J. Mol. Catal. B: Enzymatic*, 97, 2013, 283-288.
3492. Stompor, Monika, and Barbara Żarowska. "Antimicrobial activity of xanthohumol and its selected structural analogues." *Molecules* 21.5 (2016): 608.
3493. Tailor, N.K. One pot synthesis and antibacterial activity of tetrahydrochalcones. *Int. J. Pharm. Erudition*, 2013, 3, 3, 41-47.
3494. Tailor, N.K. Synthesis and antibacterial profile of certain chalcones and their reduction. *Int. J. Pharm. Erudition*, 2014, 3, 4, 17-23.
3495. Thapa, H.B., Bajracharya, N., Thapa, S., Bajracharya, G.B. "Synthesis, Structure-Activity Relationship and Antibacterial Activity of Some Simple (E)-Chalcones", *Asian Journal of Chemistry*, 34(11), 2022, 2935-2941.
3496. Ugurlu, G. (2021). Conformational And Vibrational Analysis Of Chalcone (E)-3-(Furan-2-Yl)-1-Phenylprop-2-En-1-One By Density Functional Theory And Ab Initio Hartree-Fock. *Caucasian Journal Of Science*, 8(2), 148-164.
3497. Vaishnav, A. and Ogre, P., 2023. Chalcones as Next-generation Antimicrobial Agents: Deep Insights into Anti-infective Perspectives and Novel Synthesis Routes. *Int J Med Phar Sci| Vol*, 13(7), p.1.
3498. Vásquez-Martínez, Yesseny A., et al. "Antimicrobial, Anti-Inflammatory and Antioxidant Activities of Polyoxygenated Chalcones." *Journal of the Brazilian Chemical Society* 30.2 (2019): 286-304.
3499. Vazquez-Rodriguez, S., Figueroa-Guñez, R., Matos, M.J., et al. Synthesis of coumarin-chalcone hybrids and evaluation of their antioxidant and trypanocidal properties. *Med. Chem. Comm.*, 2013, 6, 4, 993-1000.
3500. Vazquez-Rodriguez, S., López, R. L., Matos, M. J., Armesto-Quintas, G., Serra, S., Uriarte, E., ... & Santos, Y. (2015). Design, synthesis and antibacterial study of new potent and selective coumarin–chalcone derivatives for the treatment of tenacibaculosis.

- Bioorganic & medicinal chemistry, 23(21), 7045-7052.
3501. Vazquez-Rodriguez, Saleta, et al. "Coumarin-chalcone derivatives as potential antitrypanosomal and antioxidant compounds." ECSOC 16 (2012): 1-14.
http://www.usc.es/congresos/ecsoc/ & http://www.sciforum.net/conf/ecsoc-16/
 3502. Venkatesan P, Maruthavanan T. Piperidine-mediated synthesis of thiazolyl chalcones and their derivatives as potent antimicrobial agents. Nat. Prod. Res., 2012, 26(3), 223-234.
 3503. Venkatesan, P., and T. Maruthavanan. "Piperidine-mediated synthesis of thiazolyl chalcones and their derivatives as potent antimicrobial agents." Journal of Heterocyclic Chemistry 48.5 (2011): 1181-1186.
 3504. Viridi, Amardeep Singh, and Narpinder Singh. "Antimicrobial Peptides and Polyphenols: Implications in Food Safety and Preservation." Microbial Control and Food Preservation. Springer, New York, NY, 2017. 117-152.
 3505. Voltolini, B.G. Obtenção de chalconas heterocíclicas via condensação de claisen-schmidt e avaliação do seu potencial como inibidores da enzima YopH. MS Thesis, Univ. Fed. de Santa Catarina, 2012.
 3506. Wang Z., Yang Q., Zhang H., He Y., Wang R., Lu X. Isolation, Identification, and Antibacterial Activities of Flavonoids from Jujube (Zizipus Jujuba Mill.) Fruit. International Journal of Fruit Science, 23 (1), pp. 51 – 61, 2023. DOI: 10.1080/15538362.2023.2186149.
 3507. Wu, Jianzhang, et al. "Synthesis and crystal structure of chalcones as well as on cytotoxicity and antibacterial properties." Medicinal Chemistry Research 21.4 (2012): 444-452.
 3508. Xia, Y. & C. Dongmei. Progress in the synthesis and biological activity of chalcones. Zhejiang Chem. Industry, 2011, 42 (9), 11-13.
 3509. Xiao, Yingcong. "The crystal structure of (3Z, 3' Z)-4, 4'-((1, 4-phenylenebis (methylene)) bis (azanediyl)) bis (pent-3-en-2-one), C₁₈H₂₄N₂O₂." Zeitschrift für Kristallographie-New Crystal Structures (2021).
 3510. Xu, Man, et al. "Chalcone derivatives and their antibacterial activities: Current development." Bioorganic chemistry 91 (2019): 103133.
 3511. Xu, Wang, et al. "Synthesis, Crystal Structures and Antioxidant Activities of 1, 5-Diketone Derivatives." CHINESE JOURNAL OF STRUCTURAL CHEMISTRY 39.9 (2020): 1655-1661.
 3512. Yadav, Khushbu, Abha Sharma, and J. N. Srivastava. "Microwave Assisted Synthesis and Antibacterial Activity of Chalcone Derivatives." Asian Journal of Chemistry 24.12 (2012): 5779.
 3513. Yosef, Hisham Abdallah A., et al. "Synthesis, stereochemistry and antitumor evaluation of some novel chalcone derivatives." Journal of Chemical and Pharmaceutical Research 8.7 (2016): 943-957.
 3514. You, T., F. Liu, L. Wen, et al. Advance in studies on antibacterial effect of flavonoids. China J. Chinese Mat. Med., 2013, 38(21), 3645. DOI : 10.4268/cjcmm20132109
 3515. Zhang, Weilian, et al. "The crystal structure of (Z)-3-((2-(2-(2-aminophenoxy) ethoxy) phenyl) amino)-1-phenylbut-2-en-1-one, C₂₄H₂₄N₂O₃." Zeitschrift für Kristallographie-New Crystal Structures 236.3 (2021): 601-603.
 - 97). Al-Saftawy, M.M., Moawad, A.A., et al. (2024). Brucellosis Seroprevalence, REP-PCR-

Based Genotyping, and Virulence-Associated Genes Distribution Among *Brucella melitensis* Strains Isolated from Ruminants in Kafr Elsheikh Governorate of Egypt. *Egyptian Journal of Veterinary Sciences*, 55(4), 979-990.

3516. Al Dahouk, S., Le Flèche, P., Nöckler, K. Et al. (2007). Evaluation of *Brucella* MLVA typing for human brucellosis. *Journal of microbiological methods*, 69(1), 137-145.
3517. Amoupour, M., et al. "Differentiation of *Brucella abortus* and *B. melitensis* biovars using PCR-RFLP and REP-PCR." *New microbes and new infections* 32 (2019): 100589.
3518. Amoupour, Moein, et al. "Differentiation of *Brucella* species by repetitive element palindromic PCR." *Reviews in Medical Microbiology* 30.3 (2019): 155-160.
3519. Baruta, D., Ardoino, S.M., Riesco, S. et al. (2000). Evaluación de un método serológico para la detección de anticuerpos contra *Brucella canis*. *Ciencia Veterinaria*, 2(1), 62-65.
3520. Behrouzikhah, A., Keyvanfar, H., Tabatabayi, A., Alamian, S., & Feyzabadi, M. (2005). Differentiation Of Iranian Strains Of *Brucella* Spp. By Random Amplification Of Polymorphic Dna.
3521. Bricker, B.J. (2002). PCR as a diagnostic tool for brucellosis. *Veterinary microbiology*, 90(1-4), 435-446.
3522. Bricker, B.J., Ewalt, D.R., Halling, S.M. (2003). *Brucella* 'HOOF-Prints': strain typing by multi-locus analysis of variable number tandem repeats (VNTRs). *BMC microbiology*, 3, 1-13.
3523. Bricker, B.J. (2005). Molecular diagnostics of animal brucellosis: a review of PCR-based assays and approaches. *Brucella: Molecular & Cell Biol*, 25-51.
3524. Bricker, B.J., & Ewalt, D.R. (2006). HOOF prints: *Brucella* strain typing by PCR amplification of multilocus tandem-repeat polymorphisms. *Diagnostic Bacteriology Protocols*, 141-173.
3525. Bricker, B.J., & Ewalt, D.R. (2008). HOOF Prints. *Diagnostic Bacteriology Protocols*, 345, 141.
3526. Cloeckaert, A., Verger, J. M., Grayon, M. Et al. (2001). Classification of *Brucella* spp. isolated from marine mammals by DNA polymorphism at the *omp2* locus. *Microbes and Infection*, 3(9), 729-738.
3527. Cloeckaert, A., & Vizcaíno, N. (2005). DNA polymorphism and taxonomy of *Brucella* species. In *Brucella: Molecular & Cell Biol* (pp. 1-24). Taylor & Francis.
3528. Cui Buyun, Yin Jiming, Li Lanyu et al. (2005). Study on Rep-PCR typing of *Brucella*. *Disease Surveillance*, 20(8), 397-400.
3529. De Santis, R., Ciammaruconi, A., Faggioni, G. et al. (2009). Lab on a chip genotyping for *Brucella* spp. based on 15-loci multi locus VNTR analysis. *BMC microbiology*, 9, 1-5.
3530. De Santis, R., Ciammaruconi, A., Faggioni, G. et al. (2011). High throughput MLVA-16 typing for *Brucella* based on the microfluidics technology. *BMC microbiology*, 11, 1-9.
3531. Del Pozo, L., Nazario, S., Valencia, A. Et al. (2006, October). Estudio de un brote intrahospitalario por *Salmonella typhimurium* productora de beta-lactamasa de espectro extendido SHV-5. In *Anales de la Facultad de Medicina* (Vol. 67, No. 4, pp. 318-326). UNMSM. Facultad de Medicina.
3532. Gallien, P., Dorn, C., Alban, G. et al. (1998). Detection of *Brucella* species in organs of naturally infected cattle by polymerase chain reaction. *Veterinary record*, 142(19), 512-514.
3533. Garin-Bastuji, B., Blasco, J. M., Marin, C., Albert, D. (2006). The diagnosis of

- brucellosis in sheep and goats, old and new tools. *Small Ruminant Research*, 62(1-2), 63-70.
3534. He, Y. (1999). Induction of protection, antibodies and cell mediated immune responses by *Brucella abortus* strain RB51, *Ochrobactrum anthropi* and recombinants thereof (Doctoral dissertation, Virginia Tech).
3535. Hollender, D. (2016). *Brucella abortus*: tipificación molecular por MLVA en aislamientos locales, análisis in silico de sus lipoproteínas e implicancia de las lipoproteínas en virulencia (Doctoral dissertation, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires).
3536. Human, T. Evaluation of a Multilocus Variable-Number.
3537. Jiang, H., Cui, B., Li, L., Zhao, H., & Piao, D. (2009). Genotyping of *Brucella canis* by MLVA technique and multiplex PCR. *Biotechnology Letters*, 20(3), 336-338.
3538. Jiang H, Cui Buyun, Zhao Hongyan, Park Dongri, & Li Lanyu. (2009). Differential diagnosis of *Brucella suis* biotype 1 field strain and *Brucella suis* vaccine type 2. *Chinese Journal of Vector Biology and Control*, 20(3), 237-239.
3539. Jiang Hai, Rong Rong, Tian Guozhong et al. (2015). Analysis of variable number tandem repeat sequences in *Brucella* multilocus using multiplex polymerase chain reaction and multicolor capillary electrophoresis. *Chinese Journal of Zoonoses*, 31(4), 303-310.
3540. Jones, S.W., Dobson, M.E., Francesconi, S.C. et al. (2005). DNA assays for detection, identification, and Individualization of select agent microorganisms. *Croatian Medical Journal*, 46(4).
3541. Kattar, M.M., Jaafar, R.F., Araj, G.F. et al. (2008). Evaluation of a multilocus variable-number tandem-repeat analysis scheme for typing human *Brucella* isolates in a region of brucellosis endemicity. *Journal of clinical microbiology*, 46(12), 3935-3940.
3542. Kim, J.W., Lee, Y.J., & Tak, R.B. (2003). Occurrence of canine brucellosis in large kennels and characterization of *Brucella canis* isolates by PCR-RFLP. *Korean Journal of Veterinary Research*, 43(1), 67-75.
3543. Kim, S.G., Kim, Y.H., Lee, H.Y. et al. (2011). Genotyping of *Brucella abortus* isolated from cattle in Gyeongbuk province by MLVA. *Korean Journal of Veterinary Service*, 34(3), 227-234.
3544. Konstantinidis, A.P. (2007). Evaluation of the fluorescence polarization assay (FPA) serological method in the diagnosis of human brucellosis.
3545. Kumari, G., Reddy, A., Gunalan, R. Et al. (2021). Molecular differentiation of *Brucella* isolates by repetitive sequence-based PCR. *Indian Journal of Comparative Microbiology, Immunology and Infectious Diseases*, 42(2spl), 99-109.
3546. Leal-Klevezas, D.S., Martínez-Vázquez, I.O., García-Cantú, J. et al. (2000). Use of polymerase chain reaction to detect *Brucella abortus* biovar 1 in infected goats. *Veterinary Microbiology*, 75(1), 91-97.
3547. Liu, H. (2003). Expression and Localization of Green Fluorescent Protein in *B. abortus* strain RB51 (Doctoral dissertation, Virginia Tech).
3548. Liu, D. (Ed.). (2011). Molecular detection of human bacterial pathogens. CRC press.
3549. López-Goñi, I., & Moriyón, I. (2004). *Brucella*: molecular and cellular biology. Horizon Bioscience.
3550. Madkour, M.M., & Young, E.J. (2001). Immunology of brucellosis. Madkour's *Brucellosis*, 39-50.
3551. Marianelli, C., Ciuchini, F., Tarantino, M., Pasquali, P., & Adone, R. (2006).

- Molecular characterization of the *rpoB* gene in *Brucella* species: new potential molecular markers for genotyping. *Microbes and infection*, 8(3), 860-865.
3552. Martinov, S.P. (2018). *Chlamydiae and chlamydial infections*. River Publishers.
 3553. Martinov, S. P. (2022). *Q fever*. River Publishers.
 3554. Moussa, I.M., Omnia, M.E., Amin, A.S. et al. (2011). Evaluation of the currently used polymerase chain reaction assays for molecular detection of *Brucella* species. *Afr J Microbiol Res*, 5(12), 1511-20.
 3555. Mustafa, A. S., Habibi, N., Osman, A., Shaheed, F., & Khan, M. W. (2017). Species identification and molecular typing of human *Brucella* isolates from Kuwait. *PLoS One*, 12(8), e0182111.
 3556. Ocampo-Sosa, A.A., Agüero-Balbín, J., García-Lobo, J.M. (2005). Development of a new PCR assay to identify *Brucella abortus* biovars 5, 6 and 9 and the new subgroup 3b of biovar 3. *Veterinary microbiology*, 110(1-2), 41-51.
 3557. Padilla, C., & Ventura, G. (2003). Genotipificación de aislamientos de *Bartonella bacilliformis* por amplificación de elementos repetitivos mediante el uso de REP-PCR y ERIC-PCR. *Revista Peruana de Medicina Experimental y Salud Pública*, 20(3), 128-131.
 3558. Rademaker, J.L.W., Louws, F.J., Versalovic, J.A.M.E.S. et al. (2004). Characterization of the diversity of ecologically important microbes by rep-PCR genomic fingerprinting. *Molecular microbial ecology manual*, 1(2), 611-643.
 3559. Saito, S., Kobayashi, M., Kimoto-Nira, H. Et al. (2011). Intraspecies discrimination of *Lactobacillus paraplantarum* by PCR. *FEMS microbiology letters*, 316(1), 70-76.
 3560. Scholz, H.C., & Vergnaud, G. (2013). Molecular characterisation of *Brucella* species. *Rev Sci Tech*, 32(1), 149-62.
 3561. Seongguk Kim, Younghwan Kim, Hongyoung Lee et al. (2011). Genotyping of *Brucella abortus* isolated from cattle in the Gyeongbuk region using MLVA. *Korean Journal of Livestock Hygiene Society (KOJVS)*, 34(3), 227-234.
 3562. Shang Deqiu. (2004). Research progress on brucellosis. *Chinese Journal of Endemic Disease Control*, 19(4), 204-212.
 3563. Singh, Maninder, et al. "Multilocus variable number tandem repeat analysis (MLVA)-typing of *Brucella abortus* isolates of India reveals limited genetic diversity." *Tropical animal health and production* (2019): 1-8.
 3564. Sriranganathan, N., Seleem, M.N., Olsen, S.C. et al. (2009). *Brucella*. Genome mapping and genomics in animal-associated microbes, 1-64.
 3565. Suh DongKyun, S.D. (2005). DNA fingerprinting of *Brucella abortus* isolated from bovine brucellosis outbreaks by repetitive element sequence (rep)-PCR.
 3566. Tantillo, G.M., Di Pinto, A., Buonavoglia, C. (2003). Detection of *Brucella* spp. in soft cheese by semi-nested polymerase chain reaction. *Journal of dairy research*, 70(2), 245-247.
 3567. van Dijk, M.A., Engelsma, M.Y., Visser, V.X., et al. (2021). Transboundary spread of *Brucella canis* through import of infected dogs, the Netherlands, November 2016–December 2018. *Emerging Infectious Diseases*, 27(7), 1783.
 3568. Vemulapalli, R., McQuiston, J.R., Schurig, G.G. et al. (1999). Identification of an IS 711 element interrupting the *wboA* gene of *Brucella abortus* vaccine strain RB51 and a PCR assay to distinguish strain RB51 from other *Brucella* species and strains. *Clinical diagnostic laboratory immunology*, 6(5), 760-764.
 3569. Vizcaíno, N., Cloeckaert, A., Verger, J.M., Grayon, M., & Fernández-Lago, L. (2000).

- DNA polymorphism in the genus *Brucella*. *Microbes and infection*, 2(9), 1089-1100.
3570. Wang, C., Yu, Q., Ma, L. et al. (2009). Optimization of *Salmonella* REP-PCR molecular typing technology and its application in drug-resistant strains. *Journal of Health Research*, (6), 736-739.
3571. Wang, Y., Wang, Z., Zhang, Y. et al. (2014). Polymerase chain reaction–based assays for the diagnosis of human brucellosis. *Annals of clinical microbiology and antimicrobials*, 13, 1-8.
3572. Weimer, P.J. (2001). Microbiology of the dairy animal. In *Applied dairy microbiology* (pp. 21-78). CRC Press.
3573. Whatmore, A.M. (2009). Current understanding of the genetic diversity of *Brucella*, an expanding genus of zoonotic pathogens. *Infection, Genetics and Evolution*, 9(6), 1168-1184.
3574. Zhang Hongli, Cui Buyun, Fan Weixing et al. (2008). Current status of canine brucellosis and related research in my country. *Chinese Journal of Endemiology*, 27(5), 584-586.
3575. Zhao Hongyan, Yang Jie, Zhang Xu et al. (2012). Application of multi-locus variable number tandem repeat sequence analysis method in brucellosis surveillance. *Chinese Journal of Endemiology*, 31(4), 441-447.
3576. Савченко, С.С. (2008). Генотипирование штаммов *Burkholderia mallei* и *Burkholderia pseudomallei* (Doctoral dissertation, Савченко Сергей Сергеевич).
- 98). Kamenarska, Z., Gasic, M. J., Zlatovic, M., Rasovic, A., Sladic, D., Kljajic, Z., Stefanova, K., Seizova, K., NAJDENSKI, H., Kujumgiev, A., Tsvetkova, I., Popov, S.. Chemical Composition of the Brown Alga *Padina pavonia* (L.) Gaill. from the Adriatic Sea. *Botanica Marina*, 45, Walter de Gruyter, 2002, ISSN:1437-4323, 339-345. ISI IF:0.976**
3577. Abdelaal, N.S.M.M., El Seedy, G.M., Elhassaneen, Y.A.E.E. (2021). Chemical composition, nutritional value, bioactive compounds content and biological activities of the brown alga (*sargassum subrepandum*) collected from the mediterranean sea, egypt. *Alexandria Science Exchange Journal*, 42(4), 893-906.
3578. Abdel-Aal, E.I., Haroon, A.M., Mofeed, J. (2015). Successive solvent extraction and GC–MS analysis for the evaluation of the phytochemical constituents of the filamentous green alga *Spirogyra longata*. *The Egyptian journal of aquatic research*, 41(3), 233-246.
3579. Abdel-Hamid, Mohammad I., Eman I. Abdel-Aal, and Mamdouh Abdel-Mogib. "Isolation and characterization of new *Botryococcus braunii* (Trebouxiophyceae) isolates." *Renewable Energy* 141 (2019): 782-790.
3580. Acharjee, M. (2022). Purification and characterization of water soluble natural compounds from the marine algae *Padina* that interact with the membrane progesterone receptor (mPR).
3581. Akbary, Paria, et al. "Sterol and fatty acid profiles of three macroalgal species collected from the Chabahar coasts, southeastern Iran." *Aquaculture International* (2020): 1-11
3582. Arafa, N.M., Aly, U.I., Elsayed, M.M. et al. (2020). New approach for production of saturated fatty acids in cassava cell cultures as antibreast cancer agent. *Egyptian Pharmaceutical Journal*, 19(4), 361-370.
3583. Arrieche, Dioni, et al. "Secondary Metabolites Isolated from Chilean Marine Algae: A

- Review." *Marine Drugs* 20.5 (2022): 337.
3584. Arrieche, Dioni, et al. "Secondary Metabolites Isolated from Chilean Marine Algae: A Review." *Marine Drugs* 20.5 (2022): 337.
3585. Awad, N.E., Selim, M.A., Metawe, H.M. et al. (2008). Cytotoxic xenicane diterpenes from the brown alga *Padina pavonia* (L.) Gaill. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 22(12), 1610-1613.
3586. Awad, N.E., Motawe, H.M., Selim, M.A. et al. (2009). Volatile constituents of the brown algae *Padina pavonia* (L.) Gaill. And *Hydroclathrus clathratus* (C. Agardh) Howe and their antimicrobial activity. *Medicinal and Aromatic Plant Science and Biotechnology*, 3(1), 12-15.
3587. Batteau, M., Bouju, E., Ramirez-Romero, A. et al. (2024). Resolving phytosterols in microalgae using offline two-dimensional reversed phase liquid chromatography-supercritical fluid chromatography coupled with quadrupole time-of-flight mass spectrometry. *Analytical Methods*, 16(15), 2278-2285.
3588. Belattmania, Z., Reani, A., Barakate, M., Zrid, R., Elatouani, S., Hassouani, M., ... & Sabour, B. ISSN 0975-413X CODEN (USA): PCHHAX.
3589. Ben Ali, A.I., Ktari, L., Boudabbous, A. et al. (2010). Seasonal variation of antibacterial activity of the brown alga *Padina pavonica* (L) thivy collected from northern coast of Tunisia.
3590. Bernardini, G., Minetti, M., Polizzotto, G. et al. (2018). Pro-apoptotic activity of French Polynesian *Padina pavonica* extract on human osteosarcoma cells. *Marine Drugs*, 16(12), 504.
3591. Bogolitsyn, K., Parshina, A., Mamatmyrodov, K. et al. (2023). Recent advances in biochemistry of marine phaeophyta: chemical analysis, structural studies and applications. *Studies in Natural Products Chemistry*, 79, 435-486.
3592. Brajica, L. (2020). Odabrane vrste smeđih alga porodice Dictyotacea: pregled istraživanja isparljivih organskih spojeva.
3593. Cobongela, S.Z.Z. (2023). Algal Essential Oils and Their Importance in the Ecosystem. *Essential Oils: Extraction Methods and Applications*, 551-564.
3594. da Silva, I.M. (2007). Desenvolvimento de uma Metodologia para a Análise da Composição Volátil do Sal Marinho Por Microextração em Fase Sólida e Cromatografia em Fase Gasosa Acoplada à Espectrometria de Massa (SPME-GC-MS) (Doctoral dissertation, Universidade de Aveiro (Portugal)).
3595. Elbanna, S.M., & Hegazi, M.M. (2011). Screening of some seaweeds species from South Sinai, Red Sea as potential bioinsecticides against mosquito larvae; *Culex pipiens*. *Egyptian Academic Journal of Biological Sciences. A, Entomology*, 4(2), 21-30.
3596. Elbanna, S.M., & Hegazi, M.M. *Egypt. Acad. J. biolog. Sci.*, 4 (2): 21-30 (2011) A. Entomology Email: egyptianacademic@yahoo. com ISSN: 1687-8809 Received: 13/12/2011 www. eajbs. eg. net Screening of some seaweeds species from South Sinai, Red Sea as potential bioinsecticides against mosquito larvae; *Culex pipiens*.
3597. El Hattab, M. (2020). Algae essential oils: Chemistry, ecology, and biological activities. *Essential Oils—Bioactive Compounds, New Perspectives and Applications*, 1-20.
3598. El Shoubaky, G.A., & Salem, E.A. (2014). Active ingredients fatty acids as antibacterial agent from the brown algae *Padina pavonica* and *Hormophysa*

- triquetra. Journal of coastal life medicine, 2(7), 535-542.
3599. Ferraces-Casais, P., Lage-Yusty, M.A., de Quirós, A.R.B. et al. (2013). Rapid identification of volatile compounds in fresh seaweed. Talanta, 115, 798-800.
3600. Garzoli, L., Poli, A., Prigione, V. et al. (2018). Peacock's tail with a fungal cocktail: first assessment of the mycobiota associated with the brown alga *Padina pavonica*. Fungal Ecology, 35, 87-97.
3601. Generalić Mekinić, I., Šimat, V., Botić, V., Crnjac, A., Smoljo, M., Soldo, B., ... & Skroza, D. (2021). Bioactive phenolic metabolites from Adriatic brown algae *Dictyota dichotoma* and *Padina pavonica* (Dictyotaceae). Foods, 10(6), 1187.
3602. Germoush, Mousa O., et al. "Consumption of Terpenoids-Rich *Padina pavonia* Extract Attenuates Hyperglycemia, Insulin Resistance and Oxidative Stress, and Upregulates PPAR γ in a Rat Model of Type 2 Diabetes." Antioxidants 9.1 (2020): 22.
3603. Gharib, M.A.A., Elhassaneen, Y.A.E.E., Radwan, H. (2022). Nutrients and nutraceuticals content and in vitro biological activities of reishi mushroom (*ganoderma lucidum*) fruiting bodies. Alexandria Science Exchange Journal, 43(2), 301-316.
3604. Goecke, F., Escobar, M., Collantes, G. (2012). Chemical composition of *Padina fernandeziana* (Phaeophyceae, Dictyotales) from Juan Fernandez Archipelago, Chile. Rev Latinoam Biotechnol Amb Algal, 3(2), 95-104.
3605. Hu, J., Yang, B., Lin, X. et al. (2011). Bioactive metabolites from seaweeds. Handbook of Marine Macroalgae: biotechnology and applied phycology, 262-284.
3606. Ismail, A., Ktari, L., Ben Redjem Romdhane, Y. et al. (2018). Antimicrobial fatty acids from green alga *Ulva rigida* (Chlorophyta). BioMed Research International, 2018.
3607. Jerković, I., Marijanović, Z., Roje, M. et al. (2018). Phytochemical study of the headspace volatile organic compounds of fresh algae and seagrass from the Adriatic Sea (single point collection). PLoS One, 13(5), e0196462.
3608. Jerković, Igor, et al. "Chemical diversity of headspace and volatile oil composition of two brown algae (*Taonia atomaria* and *Padina pavonica*) from the Adriatic sea." Molecules 24.3 (2019): 495.
3609. Kerzabi-Kanoun, K., Belyagoubi-Benhammou, N., Belyagoubi, L. et al. (2021). Antioxidant Activity Of Brown Seaweed (*Padina Pavonica* (L.) Extracts From The Algerian Mediterranean Coast. Journal Of Natural Product Research And Applications, 1(02), 54-62.
3610. Klimjit, A., Praiboon, J., Tiengrim, S. et al. (2021). Phytochemical composition and antibacterial activity of brown seaweed, *Padina australis* against human pathogenic bacteria.
3611. Koçoğlu, Z.G. (2010). Çanakkale Boğazındaki (Çanakkale, Türkiye) bazı kahverengi alglerde alginat miktarının yıllık değişimi (Master's thesis, Fen Bilimleri Enstitüsü).
3612. Kokilaramani, S., Rajasekar, A., AlSalhi, M.S. et al. (2021). Characterization of methanolic extract of seaweeds as environmentally benign corrosion inhibitors for mild steel corrosion in sodium chloride environment. Journal of Molecular Liquids, 340, 117011.
3613. Koteska, Diana, et al. "Identification of Volatiles of the Dinoflagellate *Prorocentrum cordatum*." Marine Drugs 20.6 (2022): 371.
3614. Kranjac, M., Zekić, M., Radonić, A. et al. (2020). BioProspecting Jadranskog mora: Pregled dosadašnjih istraživanja isparljivih organskih spojeva. Kemija u industriji:

- Časopis kemičara i kemijskih inženjera Hrvatske, 69(9-10 (special issue)), 521-529.
3615. Lopes, G., Sousa, C., Bernardo, J. et al. (2011). Sterol profiles in 18 macroalgae of the portuguese coast 1. *Journal of phycology*, 47(5), 1210-1218.
 3616. Lopes, G.L.L. (2014). Seaweeds from the Portuguese Coast: Chemistry, antimicrobial and anti-inflammatory capacity (Doctoral dissertation, Universidade do Porto (Portugal)).
 3617. Makhlof M.E.M., El-Sheekh M.M., EL-Sayed A.I.M. In vitro antibiofilm, antibacterial, antioxidant, and antitumor activities of the brown alga *Padina pavonica* biomass extract. *International Journal of Environmental Health Research* 2023. DOI: 10.1080/09603123.2023.2165045
 3618. Malcata, F.X., Pinto, I.S., Guedes, A.C. (2018). *Marine macro-and microalgae: an overview*. CRC Press.
 3619. Mohedin, Nasreen. Phytochemical investigation of species of native marine algae in Oman and their biological effects
 3620. Milović, S., Kundaković, T., Macić, V. et al. (2017). Anti α -glucosidase, antitumour, antioxidative, antimicrobial activity, nutritive and health protective potential of some seaweeds from the Adriatic coast of Montenegro. *Farmacia*, 65(5), 731-740.
 3621. Omezzine, F., Haouala, R., El Ayeb, A. et al. (2009). Allelopathic and antifungal potentialities of *Padina pavonica* (L.) extract. *Journal of Plant Breeding and Crop Science*, 1(4), 194-203.
 3622. Patra, J.K., Lee, S.W., Kwon, Y.S. et al. (2017). Chemical characterization and antioxidant potential of volatile oil from an edible seaweed *Porphyra tenera* (Kjellman, 1897). *Chemistry Central Journal*, 11, 1-10.
 3623. Pereira, C.M., Nunes, C.F., Zambotti-Villela, L. et al. (2017). Extraction of sterols in brown macroalgae from Antarctica and their identification by liquid chromatography coupled with tandem mass spectrometry. *Journal of Applied Phycology*, 29, 751-757.
 3624. Pereira R.C., Paradas W.C., de Carvalho R.T., de Lima Moreira D., Kelecom A., Passos R.M.F., Atella G.C., Salgado L.T. Chemical Defense against Herbivory in the Brown Marine Macroalga *Padina gymnospora* Could Be Attributed to a New Hydrocarbon Compound. *Plants*, 12 (5), art. no. 1073, 2023. DOI: 10.3390/plants12051073.
 3625. Polat, S., & Ozogul, Y. (2008). Biochemical composition of some red and brown macro algae from the Northeastern Mediterranean Sea. *International journal of food sciences and nutrition*, 59(7-8), 566-572.
 3626. Polat, S., & Ozogul, Y. (2013). Seasonal proximate and fatty acid variations of some seaweeds from the northeastern Mediterranean coast. *Oceanologia*, 55(2), 375-391.
 3627. Radha, M., & Murugesan, A.G. (2017). Enhanced dark fermentative biohydrogen production from marine macroalgae *Padina tetrastomatica* by different pretreatment processes. *Biofuel Research Journal*, 4(1), 551-558.
 3628. Radman, Sanja, et al. "Seasonal Variability of Volatilome from *Dictyota dichotoma*." *Molecules* 27.9 (2022): 3012.
 3629. Rahman, A.U. (Ed.). (2023). *Studies in natural products chemistry*. Elsevier.
 3630. Ravi, P., Manivannan, M., Subramanian, G., Ashwathaman, S. (2018). Proximate Composition Of *Padina Boergesenii*-A Brown Alga From Vadakkadu Coastal Regions, Southeast Coast Of India.
 3631. Reese, K.L. (2020). Profiles of Volatile Compounds as Microbial Markers in

3632. Rodríguez, A., Clemente, S., Hernández, J.C. et al. (2017). Nutritional, structural and chemical defenses of common algae species against juvenile sea urchins. *Marine Biology*, 164, 1-14.
3633. Sangha, J.S., Ross, R.E., Subramanian, S. et. al. (2017). Potential use of extracts of seaweeds against plant pathogens. In *Marine Macro-and Microalgae: An Overview* (pp. 177-197). CRC Press.
3634. Samar, Juveria, et al. "Phytochemical and Biological Activities From Different Extracts of *Padina antillarum* (Kützinger) Piccone." *Frontiers in Plant Science* 13 (2022).
3635. Shanab, S.M.M. (2007). Bioactive allelo-chemical compounds from *Oscillatoria* species (Egyptian isolates). *Int. J. Agric. Biol*, 9(4), 617-621.
3636. Silva, I., Rocha, S.M., Coimbra, M.A. (2009). Headspace solid phase microextraction and gas chromatography–quadrupole mass spectrometry methodology for analysis of volatile compounds of marine salt as potential origin biomarkers. *Analytica Chimica Acta*, 635(2), 167-174.
3637. Silva, I., Rocha, S.M., Coimbra, M.A. (2010). Quantification and potential aroma contribution of β -ionone in marine salt. *Flavour and fragrance journal*, 25(2), 93-97.
3638. SILVA, M.V.D.D.B. (2022). Estudo preliminar de propriedades biológicas de extratos da macroalga *Padina pavonica* (linnaeus) thivy 1960 e sua incorporação na produção de revestimento comestível ativo para a conservação pós-colheita de mangas Tommy atkins.
3639. Stanojković, T.P., Šavikin, K., Zdunić, G. et al. (2013). In vitro antitumoral activities of *Padina pavonia* on human cervix and breast cancer cell lines. *J. Med. Plant Res*, 7, 419-424.
3640. Sultana F., Wahab M.A., Nahiduzzaman M., Mohiuddin M., Iqbal M.Z., Shakil A., Mamun A.-A., Khan M.S.R., Wong L., Asaduzzaman M. Seaweed farming for food and nutritional security, climate change mitigation and adaptation, and women empowerment: A review. *Aquaculture and Fisheries*, 8 (5), pp. 463 – 480, 2023. DOI: 10.1016/j.aaf.2022.09.001.
3641. Sultana, F., Wahab, M.A., Nahiduzzaman, M., et al. "Seaweed farming for food and nutritional security, climate change mitigation and adaptation, and women empowerment: A review". *Aquaculture and Fisheries*, 2022
3642. Tarakhovskaya, E., Lemesheva, V., Bilova, T. et al. (2017). Early embryogenesis of brown alga *Fucus vesiculosus* L. is characterized by significant changes in carbon and energy metabolism. *Molecules*, 22(9), 1509.
3643. Teixeira, Thaiz Rodrigues, et al. "Characterization of the lipid profile of Antarctic brown seaweeds and their endophytic fungi by gas chromatography–mass spectrometry (GC–MS)." *Polar Biology* 42.8 (2019): 1431-1444.
3644. Uslu, L., Sayin, S., Naz, M. et al. (2021). Proximate Analysis And Fatty Acid Profile Of Some Brown Macroalgae Collected From The Northeastern Mediterranean Coast. *Fresenius Environmental Bulletin*, 30(7).
3645. Valentão, P., Trindade, P., Gomes, D. et al. (2010). *Codium tomentosum* and *Plocamium cartilagineum*: Chemistry and antioxidant potential. *Food chemistry*, 119(4), 1359-1368.
3646. Vaseghi, Golnaz, et al. "Evaluation of the Anti-tuberculosis and Cytotoxic Potential of the Seaweed *Padina australis*." *Iranian Journal of Pharmaceutical Sciences* 15.1 (2020):

3647. Vimali, Elamathi, et al. "Comparative study of different catalysts mediated FAME conversion from macroalga *Padina tetrastomatica* biomass and hydrothermal liquefaction facilitated bio-oil production." *Chemosphere* 292 (2022): 133485.
3648. Wichachucherd, B., Liddle, L.B., Prathep, A. (2010). Population structure, recruitment, and succession of the brown alga, *Padina boryana* Thivy (Dictyotales, Heterokontophyta), at an exposed shore of Sirinart National Park and a sheltered area of Tang Khen Bay, Phuket Province, Thailand. *Aquatic Botany*, 92(2), 93-98.
3649. Yu, K.X., Wong, C.L., Ahmad, R. et al. (2015). Larvicidal activity, inhibition effect on development, histopathological alteration and morphological aberration induced by seaweed extracts in *Aedes aegypti* (Diptera: Culicidae). *Asian Pacific journal of tropical medicine*, 8(12), 1006-1012.
3650. Солоненко, А.М. (2015). Водорості гіпергалійних водойм північно-західного узбережжя Азовського моря та їх участь в утворенні мулових сульфідних пелоїдів: дис.... доктора біологічних наук: 03.00. 05 (Doctoral dissertation, Міністерство освіти і науки України).
- 99). Popova, M., Trusheva, B., Antonova, D., Cutajar, S., Mifsud, D., Farrugia, C., Tsvetkova, I., NAJDENSKI, H., Bankova, V. The specific chemical profile of Mediterranean propolis from Malta. *Food Chemistry*, 126, 3, Elsevier, 2011, ISSN:0308-8146, DOI:10.1016/j.foodchem.2010.11.130, 1431-1435. ISI IF:3.458**
3651. Aditya, J., Madhyastha, S., Sahu, S.S. et al. *Research journal of pharmaceutical, biological and chemical sciences*.
3652. Afata, Tariku Neme, et al. "Phytochemical investigation, physicochemical characterization, and antimicrobial activities of Ethiopian propolis." *Arabian Journal of Chemistry* 15.7 (2022): 103931.
3653. Afata, T.N., Dekebo, A. (2023). Chemical composition and antimicrobial effect of western Ethiopian propolis. *Chemistry & Biodiversity*, 20(2), e202200922.
3654. Ahangari, Z., Naseri, M., Vatandoost, F. (2018). Propolis: Chemical composition and its applications in endodontics. *Iranian endodontic journal*, 13(3), 285.
3655. AIM, T.U.N.A.N., SMAIL, T., ULAIMAN, S.I.A.M.S. et al. (2018). Chemical Constituents of Malaysian *Apis mellifera* Propolis (Juzuk Kimia Propolis *Apis mellifera* Malaysia). *Sains Malaysiana*, 47(1), 117-122.
3656. Akbulut, K. (2014). Türk propolisinin sulu ekstraktının insan laringeal epidermoid karsinoma (HEp-2) hücre serilerinde hücre içi serbest kalsiyum ve hidrojen peroksit düzeylerine etkisi (Master's thesis, Sağlık Bilimleri Enstitüsü).
3657. Al-Ghamdi, A.A., Bayaqoob, N.I., Rushdi, A.I. et al. (2017). Chemical compositions and characteristics of organic compounds in propolis from Yemen. *Saudi journal of biological sciences*, 24(5), 1094-1103.
3658. Alqarni, A.S., Rushdi, A.I., Owayss, A.A. et al. (2015). Organic tracers from asphalt in propolis produced by urban honey bees, *Apis mellifera* Linn. *PLoS One*, 10(6), e0128311.
3659. Aminimoghadamfarouj, N., Nematollahi, A. (2017). Propolis diterpenes as a remarkable bio-source for drug discovery development: A review. *International journal of molecular sciences*, 18(6), 1290.

3660. Aminimoghadamfarouj, N., Nematollahi, A. (2017). Structure elucidation and botanical characterization of diterpenes from a specific type of bee glue. *Molecules*, 22(7), 1185.
3661. An, S.H., Ban, E., Chung, I.Y. et al. (2021). Antimicrobial activities of propolis in poloxamer based topical gels. *Pharmaceutics*, 13(12), 2021.
3662. Anđelković, B.D. (2017). Примена rezultata NMR и FTIR спектроскопских техника у мултиваријантној анализи за класификацију прополиса. Универзитет у Београду.
3663. Archin, T., Onagh, A.G., Tehrani, A.A. et al. (2020). Evaluation of the antibacterial effect of Propolis Nanomaterials, Propolis Ethanolic extract, ciprofloxacin and their combination against *Pseudomonas aeruginosa*. *Armaghane Danesh*, 25(1), 23-39.
3664. Avula, Bharathi, et al. "Quantification and characterization of phenolic compounds from northern Indian propolis extracts and dietary supplements." *Journal of AOAC International* 103.5 (2020): 1378-1393.
3665. Bāk, B., Wilk, J., Artiemjew, P. et al. (2023). The Identification of Bee Comb Cell Contents Using Semiconductor Gas Sensors. *Sensors*, 23(24), 9811.
3666. Béji-Srairi, Raja, et al. "Ethanolic extract of Tunisian propolis: chemical composition, antioxidant, antimicrobial and antiproliferative properties." *Journal of Apicultural Research* (2020): 1-11.
3667. Benhanifia, M., & SOLTANI, A. (2022). Biological Activity of Propolis: An Update.
3668. Bertotto, Carlize, et al. "Development of a biodegradable plastic film extruded with the addition of a Brazilian propolis by-product." *LWT* 157 (2022): 113124.
3669. Buitrago, D.M., Perdomo, S.J., Silva, F.A. et al. (2024). Physicochemical Characterization, Antioxidant, and Proliferative Activity of Colombian Propolis Extracts: A Comparative Study. *Molecules*, 29(7), 1643.
3670. Danert, F.C., Zampini, C., Ordoñez, R. et al. (2014). Nutritional and functional properties of aqueous and hydroalcoholic extracts from Argentinean propolis. *Natural Product Communications*, 9(2), 1934578X1400900209.
3671. da Silva, Caroline Cristina Fernandes, et al. "Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from Ceará state." *Revista Brasileira de Farmacognosia* 29.3(2019) 309-318.
3672. Çakir, Oğuz. "Mustafa Abdullah Yilmaz."
3673. Çakiroğlu, T.N. (2017). Türk propolisinin sulu ve etanollü ekstraktının HPLC ve GC-MS ile karakterizasyonu.
3674. Cardinault, N., Cayeux, M.O., Percie du Sert, P. (2012). Propolis: origin, composition and properties. *Phytothérapie*, 10, 298-304.
3675. Cheng, H., Qin, Z.H., Guo, X F. et al. (2013). Geographical origin identification of propolis using GC–MS and electronic nose combined with principal component analysis. *Food research international*, 51(2), 813-822.
3676. Dantas Silva, R.P., Machado, B.A.S., Barreto, G.D.A. et al. (2017). Antioxidant, antimicrobial, antiparasitic, and cytotoxic properties of various Brazilian propolis extracts. *Plos one*, 12(3), e0172585.
3677. de Groot, A.C. (2013). Propolis: a review of properties, applications, chemical composition, contact allergy, and other adverse effects. *Dermatitis*, 24(6), 263-282.
3678. El-Guendouz, Soukaina, Badiia Lyoussi, and Maria Graça Costa Miguel. "Insight on propolis from Mediterranean countries chemical composition, biological activities and

- application fields." *Chemistry & biodiversity* 16.7(2019): e1900094.
3679. El-Khamsa, M. S. (2017). *Caractérisation et activités biologiques de substances naturelles, cas de la propolis* (Doctoral dissertation, UNIVERSITE FERHAT ABBAS–SETIF).
 3680. Erler, S., Cotter, S.C., Freitak, D. et al. (2024). Insects' essential role in understanding and broadening animal medication. *Trends in Parasitology*.
 3681. Essongue, C.F., Talla, E., Céline, H. et al. Evaluation of Radical Scavenging and Metal Chelating Potential of Cameroonian Propolis and Isolation of Some Chemical Constituents.
 3682. Falcão, S.I., Vale, N., Gomes, P. et al. (2013). Phenolic profiling of Portuguese propolis by LC–MS spectrometry: Uncommon propolis rich in flavonoid glycosides. *Phytochemical Analysis*, 24(4), 309-318.
 3683. Falcao, S.I.D.M. (2013). *Chemical composition of Portuguese propolis boactive properties* (Doctoral dissertation, Universidade do Porto (Portugal)).
 3684. Falcão, S.I., Vale, N., Cos, P. et al. (2014). In vitro evaluation of Portuguese propolis and floral sources for antiprotozoal, antibacterial and antifungal activity. *Phytotherapy research*, 28(3), 437-443.
 3685. Ferreira, A.R. (2016). *Avaliação da atividade antifúngica de ésteres benzoicos estruturalmente relacionados frente a espécies de Candida*.
 3686. Gameiro, A.F.P. (2020). *Utilização de própolis em medicamentos e produtos de saúde* (Master's thesis).
 3687. Guaraca Merchán, A.L., Palomino Calderón, D.L. (2018). *Estudio de la composición química y actividad antibacteriana de muestras de propóleos de diferente localización geográfica* (Master's thesis).
 3688. Guerreiro, P.I.M. (2020). *O poder anti-inflamatório do Própolis* (Doctoral dissertation).
 3689. Gültekin Subaşı, B., Ozdal, T., Capanoglu, E. “Propolis and gastrointestinal tract diseases” (Book Chapter), *Bee Products and Their Applications in the Food and Pharmaceutical Industries*, 2022, pp. 139-158.
 3690. Höche, J., 2023. *Infektionserreger als potenzielle Ursache morphologisch erfassbarer Gehirnveränderungen bei Wildkarnivoren in Sachsen-Anhalt*
 3691. Höche, Jennifer, et al. "Pathogen Screening for Possible Causes of Meningitis/Encephalitis in Wild Carnivores From Saxony-Anhalt." *Frontiers in veterinary science* 9 (2022)
 3692. Hochheim, Sabrina, et al. "A Bioguided Approach for the Screening of Antibacterial Compounds Isolated From the Hydroalcoholic Extract of the Native Brazilian Bee's Propolis Using Mollicutes as a Model." *Frontiers in Microbiology* 11 (2020): 558.
 3693. Hossain, Rajib, et al. "Propolis: An update on its chemistry and pharmacological applications." *Chinese medicine* 17.1 (2022): 1-60.
 3694. Hu, Hao, et al. "Two novel markers to discriminate poplar-type propolis from poplar bud extracts: 9-oxo-ODE and 9-oxo-ODA." *Journal of Food Composition and Analysis* 105 (2022): 104196.
 3695. Huang, S., Zhang, C.P., Wang, K. et al. (2014). Recent advances in the chemical composition of propolis. *Molecules*, 19(12), 19610-19632.
 3696. Isidorov, V. A., Bakier, S., Pirožnikow, E. et al. (2016). Selective behaviour of

- honeybees in acquiring European propolis plant precursors. *Journal of chemical ecology*, 42, 475-485.
3697. Ismail, T.N.N.T., Sulaiman, S.A., Ponnuraj, K.T. et al. (2018). Chemical constituents of Malaysian *Apis mellifera* propolis. *Sains Malaysiana*, 47(1), 117-122.
3698. Jug, M., Karas, O., Kosalec, I. (2017). The influence of extraction parameters on antimicrobial activity of propolis extracts. *Natural product communications*, 12(1), 1934578X1701200113.
3699. Kalia, P., Kumar, N.R., Harjai, K. (2015). The therapeutic potential of propolis against damage caused by *Salmonella typhimurium* in mice liver: A biochemical and histological study. *Archives of Biological Sciences*, 67(3), 807-816.
3700. Kasiotis, K.M. (2014). Propolis non-volatile constituents: A Review. *Hygeia. JD Med*, 6(1), 111-121.
3701. King, D.I. (2017). Kangaroo Island Propolis: Improved Characterisation and Assessment of Chemistry and Botanical Origins through Metabolomics (Doctoral dissertation).
3702. Kljenak, A. (2017). Fitokemijske karakteristike i biološka aktivnost propolisa (Doctoral dissertation, University of Split. Faculty of Chemistry and Technology. University of Split. School of Medicine).
3703. Kubalová, M. (2015). In vivo hodnocení účinnosti kosmetických formulací s obsahem včelích produktů na pokožku.
3704. Kumar, R., Srivastava, P. (2023). Comparative analysis of physicochemical and antioxidant activity of propolis. *Journal of Experimental Zoology India*, 26(2).
3705. Luis-Villaroya, A., Espina, L., García-Gonzalo, D. et al. (2015). Bioactive properties of a propolis-based dietary supplement and its use in combination with mild heat for apple juice preservation. *International journal of food microbiology*, 205, 90-97.
3706. Maragou, N.C., Strati, I.F., Gialouris, P.L. et al. (2023). Honey and bee products. In *Emerging Food Authentication Methodologies Using GC/MS* (pp. 137-213). Cham: Springer International Publishing.
3707. Maraschin, M., Somensi-Zeggio, A., Oliveira, S.K. et al. (2016). Metabolic profiling and classification of propolis samples from Southern Brazil: an NMR-based platform coupled with machine learning. *Journal of natural products*, 79(1), 13-23.
3708. Meira, H.C. (2017). Avaliação in-vitro da atividade antifúngica e da citotoxicidade da própolis de copaíba contra espécies do gênero *Candida* spp.
3709. Mendonça, R.Z., Nascimento, R.M., Fernandes, A.C O. et al. (2022). Antiviral action of aqueous extracts of propolis from *Scaptotrigona aff. postica* (Hymenoptera; Apidae) against Zica, Chikungunya, and Mayaro virus. *bioRxiv*, 2022-11.
3710. Mezaache-Aichour, Samia, et al. "Antiviral and anti-quorum sensing activities of lyophilized aqueous extract of propolis from sétif." *Journal of Microbiology, Biotechnology and Food Sciences* 11.5 (2022): e5222-e5222.
3711. Miguel, M.G., Antunes, M.D. (2011). Is propolis safe as an alternative medicine?. *Journal of pharmacy and bioallied sciences*, 3(4), 479-495.
3712. Miguel, M.G. (2013). Chemical and biological properties of propolis from the western countries of the Mediterranean basin and Portugal. *Int. J. Pharm. Pharm. Sci*, 5(3), 403-409.
3713. Mohameda, S.M., Elokely, K.M., Bachkeetb, E.Y. et al. (2015). NPC Natural Product Communications 2015. *Natural Product Communications*, 10, 11.

3714. Mohiuddin, I., Kumar, T.R., Zargar, M.I. et al. (2022). GC-MS analysis, phytochemical screening, and antibacterial activity of *Cerana indica* propolis from Kashmir region. *Separations*, 9(11), 363.
3715. Nada, A.A., Metwally, A.M., Asaad, A.M. et al. (2024). Synergistic effect of potential alpha-amylase inhibitors from Egyptian propolis with acarbose using in silico and in vitro combination analysis. *BMC Complementary Medicine and Therapies*, 24(1), 65.
3716. Najarnezhad, V., Jalilzadeh-Amin, G., Ownagh, A. et al. (2020). Treatment of experimental vaginal candidiasis with ethanolic extract of propolis in goat. *Iranian Veterinary Journal*, 16(1), 105-113.
3717. Netíková, L., Bogusch, P., Heneberg, P. (2013). Czech ethanol-free propolis extract displays inhibitory activity against a broad spectrum of bacterial and fungal pathogens. *Journal of Food Science*, 78(9), M1421-M1429.
3718. Oliveira, S.C. (2017). Relatório de Estágio realizado na Farmácia Santo Antônio.
3719. Oryan, A., Alemzadeh, E., Moshiri, A. (2018). Potential role of propolis in wound healing: Biological properties and therapeutic activities. *Biomedicine & pharmacotherapy*, 98, 469-483.
3720. Ownagh, A., Adibhesami, M. (2013). Treatment of *Candidia Albicans* cutaneous infection by ethanol extract of propolis in an experimental model. *Medical Journal of Tabriz University of Medical Sciences*, 35(2), 6-11.
3721. Pant, K., Sharma, A., Chopra, H.K. et al. (2023). Impact of biodiversification on propolis composition, functionality, and application in foods as natural preservative: A review. *Food Control*, 110097.
3722. Ong, Abdul Ghafar, Adib Hosami, Massoud. (2012). Treatment of experimental vaginal candidiasis in rabbits with propolis ethanolic extract. *Armaghane Danesh*, 17(3), 233-242
3723. Okińczyc, P. Charakterystyka fitochemiczna oraz aktywność biologiczna różnych rodzajów propolisów oraz ich źródeł roślinnych.
3724. Ouahab, A., Grara, N., Debbi, A. (2023). Antifungal property of bee propolis collected from the province of Souk Ahras (northeast Algeria). *Annals of Oradea University, Biology Fascicle/Analele Universității din Oradea, Fascicula Biologie*, 30(1).
3725. Radulović, N.S., Zlatković, D.B., Randjelović, P.J. et al. (2013). Chemistry of spices: bornyl 4-methoxybenzoate from *Ferula ovina* (Boiss.) Boiss.(Apiaceae) induces hyperalgesia in mice. *Food & Function*, 4(12), 1751-1758.
3726. Rebaza, R., Amaya, L., Gutiérrez, A. (2017). Aplicación del propóleo en envasado activo.
3727. Reis, T.C., Emiliano, S.A., de Carvalho Costa, F.E. et al. (2021). Atividade antimicrobiana de própolis de diferentes origens. *Brazilian Journal of Natural Sciences*, 4(1), 630-645.
3728. Righi, A.A., Negri, G., Salatino, A. (2013). Comparative chemistry of propolis from eight Brazilian localities. *Evidence-Based Complementary and Alternative Medicine*, 2013.
3729. Rizzolo, A., Bianchi, G., Povoio, M. et al. (2016). Volatile compound composition and antioxidant activity of cooked ham slices packed in propolis-based active packaging. *Food Packaging and Shelf Life*, 8, 41-49.
3730. Rodríguez, Y., Sánchez-Catalán, F., Rojano, B. et al. (2012). Caracterización

- fisicoquímica y evaluación de la actividad antioxidante de propóleos recolectados en el departamento del Atlántico, Colombia. *Revista UDCA Actualidad & Divulgación Científica*, 15(2), 303-311.
3731. Rushdi, A.I., Adgaba, N., Bayaqoob, N.I. et al. (2014). Characteristics and chemical compositions of propolis from Ethiopia. *SpringerPlus*, 3, 1-9.
 3732. Salah, N.M., Mettwally, W.S., Afifi, A.H. et al. (2023). Anti-candida Effect of Saudi Propolis: GC/MS Analysis, In-silico Study and Nano Encapsulation. *Egyptian Journal of Chemistry*, 66(8), 25-39.
 3733. Santos, H.C.D., Vieira, D.S., Yamamoto, S.M. et al. (2019). Antimicrobial activity of propolis extract fractions against *Staphylococcus* spp. isolated from goat mastitis. *Pesquisa Veterinária Brasileira*, 39(12), 954-960.
 3734. Sharma, S., Rana, A., Khakhary, D. (2024). GC-MS analysis of ethanolic extract of propolis from Jhajjar district in Haryana, India. *Journal of Environmental Biology*.
 3735. Silva, E.C.D. (2012). Estudo da composição química e atividade biológica da geoprópolis de *Melipona interrupta* e *Melipona seminigra*.
 3736. Silva, E.C.C.D., Muniz, M.P., Nunomura, R.D.C.S. et al. (2013). Constituintes fenólicos e atividade antioxidante da geoprópolis de duas espécies de abelhas sem ferrão amazônicas. *Química Nova*, 36, 628-633.
 3737. Silva-Carvalho, R., Baltazar, F., Almeida-Aguiar, C. (2015). Propolis: a complex natural product with a plethora of biological activities that can be explored for drug development. *Evidence-based complementary and alternative medicine*, 2015.
 3738. Silva, C.C.F.D., Salatino, A., Motta, L.B.D. et al. (2019). Chemical characterization, antioxidant and anti-HIV activities of a Brazilian propolis from Ceará state. *Revista Brasileira de Farmacognosia*, 29, 309-318.
 3739. Silva, H., Francisco, R., Saraiva, A. et al. (2021). The cardiovascular therapeutic potential of propolis—A comprehensive review. *Biology*, 10(1), 27.
 3740. Siripatrawan, U., Vitchayakitti, W., Sanguandeeikul, R. (2013). Antioxidant and antimicrobial properties of Thai propolis extracted using ethanol aqueous solution. *International Journal of Food Science & Technology*, 48(1), 22-27.
 3741. Sirma, E. Pervari (Siirt) Bölgesi (Çemikari, Kovanagzi, Sariyaprak) Propolislerinin Kimyasal İçerikleri Ve Botanik Orjininin Belirlenmesi (Master's thesis, Fen Bilimleri Enstitüsü).
 3742. Soltani, E.K. (2018). Caractérisation et activités biologiques de substances naturelles, cas de la propolis (Doctoral dissertation).
 3743. Soltani, A., Benhanifia, M. (2023). Propolis, Plant Sources and Antimicrobial Activity: An Overview. *Anti-Infective Agents*, 21(5), 20-39.
 3744. Spulber, R., COLȚA, T., BĂBEANU, N. et al. (2017). Chemical diversity of polyphenols from bee pollen and propolis. *AgroLife Scientific Journal*, 6(2).
 3745. Subaşı, B.G., Ozdal, T., Capanoglu, E. (2022). Propolis and gastrointestinal tract diseases. In *Bee Products and Their Applications in the Food and Pharmaceutical Industries* (pp. 139-158). Academic Press.
 3746. Suleman, T. (2016). The antimicrobial and chemical properties of South African propolis (Doctoral dissertation).
 3747. Sulaeman, A., Kalsum, N., Mahani, M. (2019). Trigona propolis and its potency for health and healing process. In *The role of functional food security in global health* (pp.

- 425-448). Academic Press.
3748. Tran, Trong D., et al. "Lessons from exploring chemical space and chemical diversity of propolis components." *International journal of molecular sciences* 21.14 (2020): 4988.
3749. Tumbarski, Y., Todorova, M., Topuzova, M. et al. (2023). Comparative Study on Physicochemical, Antioxidant and Antimicrobial Properties of Propolis Collected from Different Regions of Bulgaria. *Journal of Apicultural Science*, 67(1), 37-56.
3750. Tumbarski, Y., Petkova, N., Ivanov, I. et al. (2024). Design of functional yogurts with microencapsulated biologically active compounds. *Natural and Life Sciences Communications*, 23(1), e2024011.
3751. Vilorio, J.D., Gil, J.H., Durango, D.L. et al. (2012). Caracterización físicoquímica del propóleo de la región del bajo cauca antioqueño (Antioquia, Colombia). *Biotecnología en el sector agropecuario y agroindustrial*, 10(1), 77-86.
3752. Vranješ, M. (2023). *Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa* (Doctoral dissertation, University of Zagreb. Faculty of Veterinary Medicine).
3753. Wieczorek, Piotr Paweł, et al. "Chemical Variability and Pharmacological Potential of Propolis as a Source for the Development of New Pharmaceutical Products." *Molecules* 27.5 (2022): 1600.
3754. Womble, M., Cabot, M.L., Harrison, T., Watanabe, T.T.N. "Outbreak in African lions of *Yersinia pseudotuberculosis* infection, with aberrant bacterial morphology. *Journal of Veterinary Diagnostic Investigation*. 2022, 34(2), pp. 334-338.
3755. Zabaoui, N., Fouache, A., Trousson, A. et al. (2017). Biological properties of propolis extracts: Something new from an ancient product. *Chemistry and physics of lipids*, 207, 214-222.
3756. Zammit, E.J., Theuma, K.B., Darmanin, S. et al. 2013). Totarol content and cytotoxicity varies significantly in different types of propolis.
3757. Zlatković, D.B. (2019). Nova biološki aktivna sintetska azaheterociklična jedinjenja i novi sekundarni metaboliti iz odabranih biljnih vrsta: sinteza, izolovanje i spektralna karakterizacija. Универзитет у Нишу.
3758. Zulhendri, Felix, et al. "Antiviral, Antibacterial, Antifungal, and Antiparasitic Properties of Propolis: A Review." *Foods* 10.6 (2021): 1360.
- 100). NAJDENSKI, H., Speck, S.. *Yersinia Infections. Infectious Diseases of Wild Mammals and Birds in Europe, Wiley-Blackwell, 2012, DOI:https://doi.org/10.1002/9781118342442.ch21, 10, 293-302***
3759. Balestrieri, A. (2023). Pine Marten *Martes martes*.
3760. Common, S., Donald, H., Sainsbury, A. (2024). Revised Disease Risk Analysis for the Conservation Translocation of the Eurasian Beaver (*Castor fiber*) to England [NECR540]. *Natural England*, 5.
3761. Höche, J., House, R.V., Heinrich, A. et al. (2022). Pathogen Screening for Possible Causes of Meningitis/Encephalitis in Wild Carnivores From Saxony-Anhalt. *Frontiers in Veterinary Science*, 9, 826355.
3762. Höche, J. (2023). Infektionserreger als potenzielle Ursache morphologisch erfassbarer Gehirnveränderungen bei Wildkarnivoren in Sachsen-Anhalt.
3763. Krmpotić Đerek, A. (2020). Patogeneza crijevnih disbioza srne obične (*Capreolus capreolus* L.) (Doctoral dissertation, University of Zagreb. Faculty of Veterinary

- Medicine. Department of Pathophysiology).
3764. Lawson, B., Best, D. (2016). Passerines and other small birds. In BSAVA manual of wildlife casualties (pp. 421-438). BSAVA Library.
 3765. Torrontegi, Olalla, et al. "Naturally Avian Influenza Virus–Infected Wild Birds Are More Likely to Test Positive for Mycobacterium spp. and Salmonella spp." *Avian diseases* 63.1 (2018): 131-137.
 3766. Tsokana, Constantina N., et al. "European Brown hare (*Lepus europaeus*) as a source of emerging and re-emerging pathogens of Public Health importance: A review." *0 Veterinary Medicine and Science* 6(3), (2020), 550-564.
 3767. Varga, M. (2016). Deer. In BSAVA Manual of Wildlife Casualties (pp. 275-298). BSAVA Library.
 3768. Womble, M., Cabot, M.L., Harrison, T. et al. (2022). Outbreak in African lions of *Yersinia pseudotuberculosis* infection, with aberrant bacterial morphology. *Journal of Veterinary Diagnostic Investigation*, 34(2), 334-338.
 3769. Гончарук, М., Альшинецкий, М., Аржанова, Т., Короткова, И., Найдено, С., & Льюис, Д. Анализ рисков развития заболеваний для программы реинтродукции амурских (дальневосточных) леопардов (*Panthera pardus orientalis*).
- 101). Stein, E., Inic-Kanada, A., Belij, S., Monta, J., Bintner, N., Schlacher, S., Mayr, U. B., Lubitz, W., Stojanovic, M., NAJDENSKI, H., Barisani-Asenbauer, T. In vitro and in vivo uptake study of Escherichia coli Nissle 1917 bacterial ghosts: cell-based delivery system to target ocular surface diseases Escherichia coli Nissle 1917 bacterial ghosts. 54, 9, Investigative ophthalmology & visual science, 2013, 6326-6333. ISI IF:4.12**
3770. Alanazi, Fars K., et al. "Vision of bacterial ghosts as drug carriers mandates accepting the effect of cell membrane on drug loading." *Drug Development and Industrial Pharmacy* 46.10 (2020): 1716-1725.
 3771. Amara, A.A. (2016). Lysozymes, proteinase K, bacteriophage E lysis proteins, and some chemical compounds for microbial ghosts preparation: a review and food for thought. *SOJ Biochem*, 2(1), 16.
 3772. Amara, A.A.A.F. (2023). Bacterial and Microbial Ghosts Preparation. *Biomedical Journal of Scientific & Technical Research*, 48(2), 39607-39616.
 3773. Batah, Ali M., and Tarek A. Ahmad. "The Development of Ghosts Vaccine Trials." *Expert Review of Vaccines* (2020), 19, 6, 549-562.
 3774. Chen, H., Ji, H., Kong, X. et al. (2021). Bacterial ghosts-based vaccine and drug delivery systems. *Pharmaceutics*, 13(11), 1892.
 3775. Chi, Y., Zhang, S., Ji, S. (2024). Delivery of a Hepatitis C Virus Vaccine Encoding NS3 Linked to the MHC Class II Chaperone Protein Invariant Chain Using Bacterial Ghosts. *Biomedicines*, 12(3), 525.
 3776. Farjadian, F., Moghoofoei, M., Mirkiani, S. et al. (2018). Bacterial components as naturally inspired nano-carriers for drug/gene delivery and immunization: Set the bugs to work?. *Biotechnology advances*, 36(4), 968-985.
 3777. Fijan, S. (2014). Microorganisms with claimed probiotic properties: an overview of recent literature. *International journal of environmental research and public health*, 11(5), 4745-4767.
 3778. Goyal, A., Garg, T., & Rath, G. (2016). Probiotics in human health. *Onco*

Therapeutics, 7(1-2).

3779. Groza, D., Gehrig, S., Kudela, P., Holcman, M. et al. (2018). Bacterial ghosts as adjuvant to oxaliplatin chemotherapy in colorectal carcinomatosis. *Oncoimmunology*, 7(5), e1424676.
3780. Grumezescu, A.M., Prakash, B., Cazarin, C.B.B. et al. Probiotics and Prebiotics in Foods: Challenges, Innovations, and Advances.
3781. Hajam, I.A., Dar, P.A., Won, G. et al. (2017). Bacterial ghosts as adjuvants: mechanisms and potential. *Veterinary Research*, 48, 1-13.
3782. Haq, Nazrul, et al. "Greenness estimation of chromatographic assay for the determination of anthracycline-based antitumor drug in bacterial ghost matrix of *Salmonella typhimurium*." *Sustainable Chemistry and Pharmacy* 26 (2022): 100642.
3783. Hosseini Doust, Z., Mostaghaci, B., Yasa, O. et al. (2016). Bioengineered and biohybrid bacteria-based systems for drug delivery. *Advanced drug delivery reviews*, 106, 27-44.
3784. Hou, W., Li, J., Cao, Z. et al. (2021). Decorating bacteria with a therapeutic nanocoating for synergistically enhanced biotherapy. *Small*, 17(37), 2101810.
3785. Jiang Yinbo, & Zeng Tiebing. (2016). Research progress of bacterial ghost vaccines. *Progress in Microbiology and Immunology*, 44(3), 72-75.
3786. Lei, P., Yu, H., Ma, J. et al. (2023). Cell membrane nanomaterials composed of phospholipids and glycoproteins for drug delivery in inflammatory bowel disease: A review. *International Journal of Biological Macromolecules*, 126000.
3787. Moghimipour, E., Abedishirehjin, S., Baghbadorani, M.A. et al. (2021). Bacteria and Archaea: A new era of cancer therapy. *Journal of Controlled Release*, 338, 1-7.
3788. Mohamed, Sara, et al. "Antibacterial effects of antibiotics and cell-free preparations of probiotics against *Staphylococcus aureus* and *Staphylococcus epidermidis* associated with conjunctivitis." *Saudi Pharmaceutical Journal*, 2020, 28, 12, 1558-1565.
3789. Ou, B., Yang, Y., Tham, W.L. et al. (2016). Genetic engineering of probiotic *Escherichia coli* Nissle 1917 for clinical application. *Applied microbiology and biotechnology*, 100, 8693-8699.
3790. Pengyu, L.; Haiyang, Yu; Jiahui, Ma; Jiao, Du; Yimeng F.; Qinsi, Y.; Kun, Z.; Li, L.; Libo, J.; Wei, Wu; Da, Sun. Cell membrane nanomaterials composed of phospholipids and glycoproteins for drug delivery in inflammatory bowel disease: A review. *International Journal of Biological Macromolecules*, Vol. 249. 2023, 126000. ISSN 0141-8130. <https://doi.org/10.1016/j.ijbiomac.2023.126000>.
3791. Rabea, Sameh, et al. "Salmonella-innovative targeting carrier: Loading with doxorubicin for cancer treatment." *Saudi Pharmaceutical Journal* 28.10 (2020): 1253-1262.
3792. Saleh, N., Mahmoud, H.E., Eltaher, H., et al. "Prodigiosin-Functionalized Probiotic Ghosts as a Bioinspired Combination Against Colorectal Cancer Cells". *Probiotics and Antimicrobial Proteins*, 2022.
3793. Schiemann, M., Sonnenborn, U., Schulze, J., Müller, H.E. coli.
3794. Shende, Pravin, and Vasavi Basarkar. "Recent trends and advances in microbe-based drug delivery systems." *DARU Journal of Pharmaceutical Sciences* 27.2 (2019): 799-809.
3795. Shrivastava, S., Bhatu, N. (2023). A Study on the Usage of Probiotics as a Safer Antipyretic. *Journal of Cellular Signaling*, 4(2), 73-77.
3796. Shui, Z. (2020). Novel Cell-Derived Delivery Systems for Anti-Obesity

- Treatment (Doctoral dissertation, National University of Singapore (Singapore)).
3797. Singh, A.V., & Sitti, M. (2016). Bioengineered and biohybrid bacteria-based systems for drug delivery☆.
 3798. Thanuja, M.Y., Anupama, C., Ranganath, S.H. (2018). Bioengineered cellular and cell membrane-derived vehicles for actively targeted drug delivery: so near and yet so far. *Advanced drug delivery reviews*, 132, 57-80.
 3799. Vinderola, G., & Burns, P. (2021). The biotics family. *Probiotics and Prebiotics in Foods*, 1.
 3800. Wu, Hong-Hui, et al. "Mesenchymal stem cell-based drug delivery strategy: from cells to biomimetic." *Journal of controlled release* 294 (2019): 102-113.
 3801. Wu, Jie, and Guanghui Ma. "Imitation of nature: Bionic design in the study of particle adjuvants." *Journal of Controlled Release* 303 (2019): 101-108.
 3802. Youssof, Abdullah ME, et al. "Bacterial Ghosts Carrying 5-Fluorouracil: A Novel Biological Carrier for Targeting Colorectal Cancer." *AAPS PharmSciTech* 20.2 (2019): 48.
 3803. Zhang, Chen, et al. "Bacterial ghost as delivery vehicles loaded with DNA vaccine induce significant and specific immune responses in common carp against spring viremia of carp virus." *Aquaculture* 504 (2019): 361-368.
 3804. Zhao, D., Zheng, W.-H., Jiao, H.-M., Li, G.-C., Progress in research on potential and mechanism of bacterial ghosts as adjuvant., *Chinese Journal of Biologicals*, 2020, 33, 4, 461-464.
 3805. Zheng, Y.Y., Zhang, C., Li, Y. et al. (2021). Immersion immunization of common carp with bacterial ghost-based DNA vaccine inducing prophylactic protective immunity against spring viraemia of carp virus. *Journal of Fish Diseases*, 44(12), 2021-2029.
 3806. Zhu, W., Hao, L., Liu, X. et al. (2018). Enhanced anti-proliferative efficacy of epothilone B loaded with *Escherichia coli* Nissle 1917 bacterial ghosts on the HeLa cells by mitochondrial pathway of apoptosis. *Drug Development and Industrial Pharmacy*, 44(8), 1328-1335.
- 102). Pavlova Hristova I., Danova S., NAIDENSKI H., Tropcheva R., Milanova A.. Effect of probiotics on enrofloxacin disposition in gastrointestinal tract of poultry. *Journal of Veterinary Pharmacology and Therapeutics*, 38, 6, 2015, DOI:10.1111/jvp.12232, SJR (Scopus):0.651**
3807. Atta, A.H., Darwish, A.S., Atef, M. (2021). Effect of Probiotics on the Pharmacokinetic Aspects and Tissue Residues of Difloxacin in Broiler Chickens. *Pakistan Veterinary Journal*, 41(2).
 3808. Gutiérrez, L., Monroy-Barreto, M., García-Guzmán, P. et al. (2023). Pharmaceutical Design of a Formulation of Enrofloxacin-Alginate and its Strategic Dosage to Achieve Mutant Prevention PK/PD Ratios in Broiler Chickens. *Current Pharmaceutical Design*, 29(21), 1701-1709.
 3809. Hend, M.F., Atta, A.H., Darwish, A.S. & Atef, M. 2021, "Effect of probiotics on the pharmacokinetic aspects and tissue residues of difloxacin in broiler chickens", *Pakistan Veterinary Journal*, vol. 41, no. 3, pp. 269-273.
 3810. Matuskova, Z., Anzenbacher, P., Vecera, R. et al. (2017). Effect of *Lactobacillus casei* on the pharmacokinetics of amiodarone in male Wistar rats. *European journal of*

drug metabolism and pharmacokinetics, 42, 29-36.

3811. Mohamed, A., Atta, A.H., Darwish, A., Mohamed, H.F. Pharmacokinetics of Difloxacin in Probiotics-Treated Mycoplasma Gallisepticum Infected Chickens (2022) Acta Veterinaria Eurasia, 48 (2), pp. 87-93.
3812. Polaka, S., Rajpoot, K., Tekade, M., Sharma, M.C., Tekade, R.K. Food-drug interactions and their implications on oral drug bioavailability (2022) Pharmacokinetics and Toxicokinetic Considerations - Vol II, pp. 263-289
3813. Selig, D. J., Deluca, J. P., Li, Q., Lin, H., Nguyen, K., Scott, S. M., . . . Livezey, J. R. Saccharomyces boulardii CNCM I-745 probiotic does not alter the pharmacokinetics of amoxicillin. Drug Metabolism and Personalized Therapy, 35(1) doi:10.1515/dmpt-2019-0032.
3814. Zhao, H., Lu, Z., Lu, Y. (2022). The potential of probiotics in the amelioration of hyperuricemia. Food & Function, 13(5), 2394-2414.

103). Tsvetanova Z., NAJDENSKI H. Pathogenic bacteria in water and drinking water-associated biofilms. Ecological Engineering and Environment Protection, 1, National Society of Ecological Engineering and Environment Protection, 2017, ISSN:1311-8668, 50-61

3815. Flores-Encarnación M., Acosta-Báez, L.A., Aguilar-Gutiérrez G.R., Cabrera-Maldonado C., Gallardo-Soto H.M., Sánchez-Herrera A.J. "The Effect of Essential Oil of Thymus vulgaris on the Growth of Bacterial Environmental and Clinical Isolates", International Journal of Research Studies in Biosciences (IJRSB) Vol, 7 (5), 2019, 26-33 ISSN No. (Online) 2349-0365.
3816. Uguru, H. (2024). The Impact of Soak Away Pit Leachate on the Spatial Distribution of the Groundwater Quality. NIPES-Journal of Science and Technology Research, 6(1).
3817. Waluyo, L. (2018, July). Antagonism of Microbial Consortium Decomposers in Deadly Water-borne Pathogens in Domestic Wastewater. In 2018 3rd International Conference on Education, Sports, Arts and Management Engineering (ICESAME 2018) (pp. 606-609). Atlantis Press.

104). Poehlein, Anja, NAJDENSKI, Hristo, Simeonova, Dilianna D.. Draft Genome Sequence of Flavobacterium succinicans strain DD5b. Genome Announcements, 5, 2, ASM, 2017, ISSN:2169-8287, DOI:doi:10.1128/genomeA.01492-16, SJR (Scopus):0.217

3818. Evaluation of microbial communities of bottled mineral waters and preliminary traceability analysis using ngs microbial fingerprints Carraturo, F., Carmela, C., Compagnone, M., et .al., 2021 Water (Switzerland) 13(20), 2824.
3819. Leigh, Samantha C., Caitlyn Catabay, and Donovan P. German. "Sustained changes in digestive physiology and microbiome across sequential generations of zebrafish fed different diets." Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology 273 (2022): 111285
3820. Kumru, S., Tekedar, H.C., Blom, J., Lawrence, M.L., Karsi, A., „Genomic diversity in flavobacterial pathogens of aquatic origin“ Microbial Pathogenesis 142, 104053.
3821. MITREA, L., Monica, T.R.I.F., Alexandru, R.U.S.U. et al. (2019). Klebsiella pneumoniae—a useful pathogenic strain for biotechnological purposes: 1, 3-propanediol biosynthesis.
3822. Sverdrup, Anthony Elias, Ludmila L. Frolova, and Stella Sagdeeva. "The Monitoring

- of Sredniy Kaban Lake by 16s Rrna Gene Amplicon Data Set-Based Bacterial Diversity.", *Journal of Environmental Treatment Techniques*, 7(Special Issue), 1016-1025, (2019).
3823. Swiatek, L.S., Surmann, K., Eger, E. et al. (2023). Multi-omics investigations uncover unique pathogenic markers in clinical *Klebsiella pneumoniae* that could be leveraged as novel antimicrobial targets. *bioRxiv*, 2023-12.
- 105). Zheleva-Dimitrova, D., Gevrenova, R., Zaharieva, M. M., NAJDENSKI, H., Ruseva, S., Lozanov, Valentin, Balabanova, V., Yagi, S., Momekov, G., Mitev, V.. HPLC-UV and LC-MS Analyses of Acylquinic Acids in *Geigeria alata* (DC) Oliv. & Hiern. and their Contribution to Antioxidant and Antimicrobial Capacity. Phytochemical analysis, John Wiley & Sons Ltd, 2017, ISSN:1099-1565, DOI:10.1002/pca.2658, n/a-n/a-n/a-n/a. JCR-IF (Web of Science):2.337**
3824. Alcázar Magaña, A., Kamimura, N., Soumyanath, A. et al. (2021). Caffeoylquinic acids: Chemistry, biosynthesis, occurrence, analytical challenges, and bioactivity. *The plant journal*, 107(5), 1299-1319.
3825. Atxitia, U., Chubykalo-Fesenko, O., Chantrell, R.W. et al. Ultrafast Spin Dynamics: The Effect of Colored Noise.
3826. Bernard, Guillaume, et al. "MeJA Elicitation of Chicory Hairy Roots Promotes Efficient Increase of 3, 5-diCQA Accumulation, a Potent Antioxidant and Antibacterial Molecule." *Antibiotics* 9.10 (2020): 659.
3827. Birsa, M.L., Sarbu, L.G. (2023). Health Benefits of Key Constituents in *Cichorium intybus* L. *Nutrients*, 15(6), 1322.
3828. Chew, J., Peh, S.C., Sin Yeang, T. (2019). Non-microbial natural products that inhibit drug-resistant *Staphylococcus aureus*. *Staphylococcus aureus*, 10.
3829. Clifford, M.N. (2017). Some notes on the chlorogenic acids. Part 4. Botanical distribution of the chlorogenic acids.
3830. Elbashir, S.M.I., Devkota, H.P., Wada, M. et al. (2018). Free radical scavenging, α -glucosidase inhibitory and lipase inhibitory activities of eighteen Sudanese medicinal plants. *BMC complementary and alternative medicine*, 18, 1-12.
3831. Fadul, E., Nizamani, A., Rasheed, S. et al. (2020). Anti-glycating and anti-oxidant compounds from traditionally used anti-diabetic plant *Geigeria alata* (DC) Oliv. & Hiern. *Natural product research*, 34(17), 2456-2464.
3832. Ganzera, M., & Sturm, S. (2018). Recent advances on HPLC/MS in medicinal plant analysis—An update covering 2011–2016. *Journal of Pharmaceutical and Biomedical Analysis*, 147, 211-233.
3833. Gronbach, M. (2020). Charakterisierung der Inhaltsstoffe von Pflanzenextrakten, deren Untersuchung auf ihre ER-Stress-lindernde Aktivität und Anwendung der Sublimation auf Fruchtpulver zur Identifizierung von Marker-Substanzen und Isolierung von Naturstoffen (Doctoral dissertation, Dissertation, Rostock, Universität Rostock, 2021).
3834. Heena, Kaushal, S., Kaur, V. et al. (2024). Isolation of quinic acid from dropped *Citrus reticulata* Blanco fruits: its derivatization, antibacterial potential, docking studies, and ADMET profiling. *Frontiers in Chemistry*, 12, 1372560.
3835. Ibrahim, S., Osman, W., Maaz, M.A. et al. *Geigeria alata*-a potential source for anti-Alzheimer's constituents: In vitro and computational investigations. *Indonesian Journal of*

Pharmacy.

3836. Islam, S., Adam, Z., Akanda, J.H. (2024). Quinic and caffeic acids derivatives: Affecting antioxidant capacities and phenolics contents of certain therapeutic and specialty crops employing water and ethanolic extracts. *Food Chemistry Advances*, 4, 100693.
3837. Kłeczek, N., Michalak, B., Malarz, J. et al. (2019). *Carpesium divaricatum* Sieb. & Zucc. revisited: Newly identified constituents from aerial parts of the plant and their possible contribution to the biological activity of the Plant. *Molecules*, 24(8), 1614.
3838. Lee, C.D., Cho, H., Shim, J. et al. (2023). Characteristics of Phenolic Compounds in *Peucedanum japonicum* According to Various Stem and Seed Colors. *Molecules*, 28(17), 6266.
3839. Li, K., Yao, Q., Zhang, M. et al. (2023). Exploring the effective components and potential mechanisms of Zukamu granules against acute upper respiratory tract infections by UHPLC-Q-Exactive Orbitrap-MS and network pharmacology analysis. *Arabian Journal of Chemistry*, 16(8), 104875.
3840. Liu, Y., Wang, C., Wu, J. et al. (2023). Study on the Comprehensive Phytochemicals and the Anti-Ulcerative Colitis Effect of *Saussurea pulchella*. *Molecules*, 28(4), 1526.
3841. Milutinović, V., Niketić, M., Ušjak, L. et al. (2018). Methanol extracts of 28 *Hieracium* species from the Balkan Peninsula—Comparative LC–MS analysis, chemosystematic evaluation of their flavonoid and phenolic acid profiles and antioxidant potentials. *Phytochemical analysis*, 29(1), 30-47.
3842. Mohamed, M.A.M. (2021). Phytochemical, Antioxidant and Anti-acetylcholinesterase Screening of Some Selected Plants from Sudan (Doctoral dissertation, University of Khartoum).
3843. Mohammed, S.I.H., EL-Kamali, H.H., EL-Tahir, A. S. (2022). Haematological Effects of Aqueous Extracts of *Geigeria alata* in Male Albino Rats. *Journal of The Faculty of Science and Technology*, (9 (2)), 125-133.
3844. Nowak, J., Kiss, A.K., Wambebe, C. et al. (2022). Phytochemical analysis of polyphenols in leaf extract from *Vernonia amygdalina* Delile plant growing in Uganda. *Applied Sciences*, 12(2), 912.
3845. Orlando, G., Zengin, G., Ferrante, C. et al. (2019). Comprehensive chemical profiling and multidirectional biological investigation of two wild *Anthemis* species (*Anthemis tinctoria* var. *pallida* and *A. cretica* subsp. *tenuiloba*): focus on neuroprotective effects. *Molecules*, 24(14), 2582.
3846. Osman, W., Maaz, M.A., Ali, A. et al. (2023). *Geigeria Alata*-A Potential Source for Anti-Alzheimer's Constituents: In Vitro And Computational Investigations. *Indonesian Journal of Pharmacy/Majalah Farmasi Indonesia*, 34(4).
3847. Pereira, I.D.S.P., Vega, M.R.G., da Silva Mathias, M. et al. (2019). Phytochemical and biological studies on *Piptocarpha axillaris* (Less.) Baker (Asteraceae). *Biochemical systematics and ecology*, 85, 24-30.
3848. Quinty, V., Nasreddine, R., Colas, C. et al. (2023). Antioxidant and anti-lipase capacities from the extracts obtained from two invasive plants: *Ambrosia artemisiifolia* and *Solidago canadensis*. *Food Bioscience*, 55, 103069.
3849. Singla, R., Jaitak, V. (2018). Recent Advances in Plant Metabolites Analysis, Isolation, and Characterization. *Recent Trends and Techniques in Plant Metabolic*

Engineering, 75-115.

3850. Varvouni, E.F., Graikou, K., Gortzi, O. et al. (2021). Chemical and biological evaluation of the oil and seedcake from seeds of a Greek cardoon cultivar as potential functional vegetable oil. Comparison with sesame, flaxseed and extra virgin olive oils. *Foods*, 10(11), 2665.
3851. Xu, Z., Li, K., Pan, T. et al. (2019). Lonicerin, an anti-algE flavonoid against *Pseudomonas aeruginosa* virulence screened from Shuanghuanglian formula by molecule docking based strategy. *Journal of ethnopharmacology*, 239, 111909.
3852. Zengin, G., Aktumsek, A., Ceylan, R. et al. (2017). Shedding light on the biological and chemical fingerprints of three *Achillea* species (*A. biebersteinii*, *A. millefolium* and *A. teretifolia*). *Food & function*, 8(3), 1152-1165.
- 106). Trusheva, B., Petkov, H., Popova, M., Dimitrova, L., Zaharieva, M., Tsvetkova, I., NAJDENSKI, H., Bankova, V.. "Green" approach to propolis extraction: natural deep eutectic solvents.. 72, 7, Comptes rendus de l'Académie bulgare des Sciences, 2019, DOI:10.7546/CRABS.2019.07.06, 897-905. SJR (Scopus):0.22, JCR-IF (Web of Science):0.321**
3853. Alpat, U., Nar, T., Karasu, S. and Sagdic, O., 2023. Optimization of propolis extraction with natural deep eutectic solvents using central composite design. *Phytochemistry Letters*, 58, pp.49-59.
3854. Atan, T., 2023. Propolis ile fonksiyonel maden suyu üretimi (Master's thesis, Bursa Uludağ Üniversitesi).
3855. Aydin, M., Eyiz, V., Tontul, İ. Value Addition of Fruit Processing Industrial Waste. In *Wealth out of Food Processing Waste* (pp. 84-146). CRC Press.
3856. Aydın, M., Danacıoğlu, D.A., Türker, S. Propolisin Genel Özellikleri Ve Kullanımı. *Gıda*, 46(1), 69-81.
3857. Bragagnolo, F.S., Strieder, M.M., Pizani, R.S. et al. (2024). REVISITING NATURAL DEEP EUTECTIC SOLVENTS (NADES) AS EXTRACTION MEDIA AND READY-TO-USE PURPOSES. *TrAC Trends in Analytical Chemistry*, 117726.
3858. Çakır, B., Yılmaz, Ş.M., & Güzel, N. Propolis Ekstraksiyonunda Uygulanan Yeşil Ve Yenilikçi Yöntemler. *Uludağ Arıcılık Dergisi*, 24(1), 153-166.
3859. Cea-Pavez, I., Manteca-Bautista, D., Morillo-Gomar, A. et al. (2024). Influence of the Encapsulating Agent on the Bioaccessibility of Phenolic Compounds from Microencapsulated Propolis Extract during In Vitro Gastrointestinal Digestion. *Foods*, 13(3), 425.
3860. Contieri, L.S., Ribeiro, T.B., Sosa, F.H., Vaz, B.M., Pizani, R.S., Pintado, M., Ventura, S.P., Mesquita, L.M.D.S. and Rostagno, M.A., 2023. Unlocking the Full Potential of Green Propolis: A Novel Extraction Approach Using Eutectic Solvents for Improved Phenolic Compound Recovery. *ACS Sustainable Chemistry & Engineering*, 11(36), pp.13470-13482.
3861. Contieri, Letícia S., et al. "Recent progress on the recovery of bioactive compounds obtained from propolis as a natural resource: Processes, and applications." *Separation and Purification Technology* (2022): 121640.
3862. Dos Santos, C.M., de Souza Mesquita, L.M., Braga, A.R.C. et al. (2021). Red propolis as a source of antimicrobial phytochemicals: extraction using high-performance

- alternative solvents. *Frontiers in Microbiology*, 12, 659911.
3863. Farooq, Muhammad Qamar, Nabeel Mujtaba Abbasi, and Jared L. Anderson. "Deep eutectic solvents in separations: Methods of preparation, polarity, and applications in extractions and capillary electrochromatography." *Journal of Chromatography A* 1633 (2020): 461613.
3864. Fuente-Ballesteros, A., Privolos, I., Ares, A.M., Samanidou, V. and Bernal, J., 2023. GREEN SAMPLE PREPARATION METHODS FOR THE ANALYSIS OF BIOACTIVE COMPOUNDS IN BEE PRODUCTS: A REVIEW. *Advances in Sample Preparation*, p.100060.
3865. Maria Nichitoi, Madalina, et al. "Do Ultrasonic Field Effects Upon the Polyphenolics Profile of Propolis Extracts Improve Their Antioxidant and Antimicrobial Activity?" *Ultrasonics Sonochemistry* 92 (2022): 106274.
3866. Nichitoi, M.M., Josceanu, A.M., Isopescu, R.D., Isopencu, G.O., Geana, E.I., Ciucure, C.T. and Lavric, V., 2023. Do ultrasonic field effects upon the polyphenolics profile of propolis extracts improve their antioxidant and antimicrobial activity?. *Ultrasonics Sonochemistry*, 92, p.106274.
3867. Özdal, H.R., Nakilcioğlu, E., Ötleş, S. (2023). Propolisin Biyoaktif Bileşenleri Üzerine Ekstraksiyon Yöntemlerinin Ve Ekstraksiyon Değişkenlerinin Etkileri. *Gıda*, 48(6), 1123-1131.
3868. Rendueles, E., Mauriz, E., Sanz-Gómez, J., Adanero-Jorge, F. and García-Fernandez, C., 2023. Antimicrobial Activity of Spanish Propolis against *Listeria monocytogenes* and Other *Listeria* Strains. *Microorganisms*, 11(6), p.1429.
3869. Sankaran, S., Dubey, R. and Lohidasan, S., 2023. Optimization of Extraction Conditions using Response Surface Methodology and HPTLC Fingerprinting Analysis of Indian Propolis. *Journal of Biologically Active Products from Nature*, 13(1), pp.76-93.
3870. Tzani, A., Pitterou, I., Divani, F. et al. (2022). Green extraction of Greek propolis using natural deep eutectic solvents (NADES) and incorporation of the NADES-extracts in cosmetic formulation. *Sustainable Chemistry*, 4(1), 8-25.
3871. Vranješ, M., 2023. Mehanizmi antibakterijskog djelovanja inovativnog ekstrakta propolisa (Doctoral dissertation, University of Zagreb. Faculty of Veterinary Medicine).
- 107). HRISTO NAJDENSKI, Venelin Hubenov, Ivan Simeonov, Veselin Kussovski, Lyudmila Dimitrova, Penka Petrova, Plamen Angelov, Lyudmila Kabaivanova. Microbial Biodegradation as an Option for Waste Utilization during Long Term Manned Space Missions. Bulgarian Chemical Communications, 52, 3, Bulgarian Academy of Sciences, 2020, ISSN:0324-1130, DOI:10.34049/bcc.52.3.5227, 379-386. SJR (Scopus):0.179**
3872. Alla G. Dyachenko, Olena V. Ischenko, Mykola G. Zhudenko, Snizhana V. Gaidai, Tetiana M. Zakharchova, Andrii V. Yatsymyrskiy, Vladyslav V Lisnyak CO₂ methanation over Co–Ni/Al₂O₃ and Co–Ni/SiC catalysts. *Bulgarian Chemical Communications* 52(3):342-347; doi 10.34049/bcc.52.3.5134.
- 108). Diukendjieva, A, Zaharieva, MM, Mori, M, Alov, P, Tsakovska, I, Pencheva, T, NAJDENSKI, H, Kren, V, Felici, C, Bufalieri, F, Di Marcotullio, L, Botta, B, Botta, M, Pajeva, I. Dual SMO/BRAF Inhibition by Flavonolignans from *Silybum marianum*. Antioxidants, 9, 5, MDPI, 2020, ISSN:2076-3921, DOI:10.3390/antiox9050384, 1-13. SJR**

(Scopus):1.1, JCR-IF (Web of Science):6.313

3873. Albohy, A., Said, M.A., Abdelrahman, M.A. et al. (2022). Remdesivir analog as SARS-CoV-2 polymerase inhibitor: virtual screening of a database generated by scaffold replacement.
3874. Aslam, Andleeb, et al. "Quercetin ameliorates thioacetamide-induced hepatic fibrosis and oxidative stress by antagonizing the Hedgehog signaling pathway." *Journal of Cellular Biochemistry* 123.8 (2022): 1356-1365
3875. Chettri, Sumiran, et al. "Computational Analysis of Natural Product B-Raf Inhibitors." *Journal of Molecular Graphics and Modelling* 118 (2022): 108340.
3876. Gerasimova, E., Gazizullina, E., Radosteva, E., & Ivanova, A. (2021). Antioxidant and antiradical properties of some examples of flavonoids and coumarins—Potentiometric studies. *Chemosensors*, 9(5), 112.
3877. Malik, V., Kumar, V., Kaul, S.C. et al. (2021). Potential of Withaferin-A, Withanone and Caffeic Acid Phenethyl ester as ATP-competitive inhibitors of BRAF: A bioinformatics study. *Current Research in Structural Biology*, 3, 301-311.
3878. Nawaz, Aisha, et al. "Isolation and Characterization of a Flavonoid and a Neolignan from *Silybum marianum*: In-vitro Cytotoxic Evaluation." *ChemistrySelect* 7.15 (2022): e202200502
3879. Said, M. A., Albohy, A., Abdelrahman, M. A., Ibarhim, H. S. Remdesivir analog as SARS-CoV-2 polymerase inhibitor: virtual screening of a database generated by scaffold replacement. // *RSC Advances*, 2022, Vol. 12, Issue 35, p. 22448-22457, ISSN 20462069 (ISSN). doi:10.1039/d2ra00486k
3880. Tomko, Andrea M., et al. "Anti-cancer potential of cannabinoids, terpenes, and flavonoids present in cannabis." *Cancers* 12.7 (2020): 1985. 1-81.
3881. Valentová, Kateřina. "Cytoprotective Activity of Natural and Synthetic Antioxidants." (2020): 713. 1-4.
3882. Ye, Wenjun, et al. "Design, Synthesis and Biological Evaluation of Novel Triazoloquinazolinone and Imidazoquinazolinone Derivatives as Allosteric Inhibitors of Shp2 Phosphatase." *Journal of Enzyme Inhibition and Medicinal Chemistry* 37.1 (2022): 1495-513
- 109). Ivanova, A., Kostova, I., Tsvetkova, I. and NAJDENSKI, H., 2001. GC-MS investigation of *Haplophyllum suaveolens* extracts. *Comptes Rendus de l'Academie Bulgare des Sciences*, vol. 54, p. 6: 35, 54(6), pp.6-35.**
3883. Karahan, F., 2023. Evaluation of trace element and heavy metal levels of some ethnobotanically important medicinal plants used as remedies in Southern Turkey in terms of human health risk. *Biological Trace Element Research*, 201(1), pp.493-513.
3884. Subramonian, K., Ganthi, A.S., Namasivayam, K. (2023). GC-MS analysis of *Orthosiphon comosus* Wight ex Benth. *Biochemical & Cellular Archives*, 23(2).
- 110). Amerikova, M.N., Zaharieva, M.M., Andonova, L.A., Pencheva-El Tibi, I.P., Maslarska, V. and NAJDENSKI, H.M., 2020. Analytical study and antimicrobial activity of alpha-defensin 2 dissolved in pharmacopoeia buffers with different pH. *Acta Poloniae Pharmaceutica-Drug Research*, 77(1), pp.3-10.**
3885. Habibie, A., Raharjo, T.J., Swasono, R.D. and Retnaningrum, E., 2023. Antibacterial

activity of active peptide from marine macroalgae *Chondrus crispus* protein hydrolysate against *Staphylococcus aureus*. *Pharmacia*, 70(4), pp.983-992

111). Lyudmila V. Kabaivanova, Hristo M. NAJDENSKI, Venelin N. Hubenov, Elena I. Chorukova, Ivan Simeonov, Juliana G. Ivanova. Biotechnological exploitation of lignocellulosic wastes for biomethane production and algae cultivation in the digestate. International Journal of Pharma Medicine and Biological Sciences, 9, 4, 2020, ISSN:ISSN 2278-5221, 152-157. SJR (Scopus):0.24

3886. Mittal, V., Talapatra, K.N. & Ghosh, U.K. A comprehensive review on biodiesel production from microalgae through nanocatalytic transesterification process: lifecycle assessment and methodologies. *Int Nano Lett*, 12, 351–378, (2022).

112). Dimitrova, L., Kaleva, M., Zaharieva, MM, Stoykova, C., Tsvetkova, I., Angelovska, M., Ilieva, Y., Kussovski, V., Naydenska, S., NAJDENSKI, H.. Prevalence of Antibiotic-Resistant *Escherichia coli* Isolated from Swine Faeces and Lagoons in Bulgaria. Antibiotics, 10(8), 940, PubMed PMID, 2021, ISSN:20796382, DOI:<https://doi.org/10.3390/antibiotics10080940>, 1-15. JCR-IF (Web of Science):5.222

3887. Duangurai, T., Rungruengkitkul, A., Kong-Ngoen, T., Tunyong, W., Kosoltanapiwat, N., Adisakwattana, P., Vanaporn, M., Indrawattana, N., Pumirat, P. Phylogenetic analysis and antibiotic resistance of *Escherichia coli* isolated from wild and domestic animals at an agricultural land interface area of Salaphra wildlife sanctuary, Thailand. // *Veterinary World*, 2022, Vol. 15, Issue 12, p. 2800-2809, ISSN 09728988 (ISSN). doi:10.14202/vetworld.2022

3888. Mustafa, S. S., Batool, R., Kamran, M., Javed, H., Jamil, N. Evaluating the Role of Wastewaters as Reservoirs of Antibiotic-Resistant ESKAPEE Bacteria Using Phenotypic and Molecular Methods. // *Infection and Drug Resistance*, 2022, Vol. 15, p. 5715-5728, ISSN 11786973 (ISSN). doi:10.2147/idr.s368886

3889. Paneri, M. and Sevt, P., 2023. Overview of Antimicrobial Resistance: An Emerging Silent Pandemic. *Global Journal of Medical, Pharmaceutical, and Biomedical Update*, 18

113). Ilieva, YE, Dimitrova, LL, Zaharieva, MM, Kaleva, M, Alov, P, Tsakovska, I, Pencheva, T, Pencheva-El Tibi, I, NAJDENSKI, H, Pajeva, I. Cytotoxicity and Microbicidal Activity of Commonly Used Organic Solvents A Comparative Study and Application to a Standardized Extract from *Vaccinium macrocarpon*. Toxics, 9, 5, MDPI, 2021, ISSN:2305-6304, DOI:10.3390/toxics9050092, 92-108. SJR (Scopus):0.786, JCR-IF (Web of Science):4.472

3890. Adiningrat, A., Maulana, I., Fadhlurrahman, A.G. et al. (2023). Evaluation of biocompatibility and effectiveness of propolis *Tetragonula* sp. as dental anti-microbial agent. *Journal of Stomatology*, 76(2), 94-100.

3891. Afonso, J., Mezzetta, A., Marrucho, I.M. and Guazzelli, L., 2023. History repeats itself again: Will the mistakes of the past for ILs be repeated for DESs? From being considered ionic liquids to becoming their alternative: the unbalanced turn of deep eutectic solvents. *Green Chemistry*.

3892. Alberdi, L. V., Rosso, G., Velóz, L., Romeo, C., Farias, J., Di Tomaso, M. V., Calero, M., Kun, A. Curcumin and Ethanol Effects in Trembler-J Schwann Cell Culture. // *Biomolecules*, 2022, Vol. 12, Issue 4 C7 - 515, ISSN 2218273X (ISSN).

doi:10.3390/biom12040515

3893. Alberdi, L.V., Martínez-Busi, M., Echeverry, C., Calero, M. and Kun, A., 2023. Low hormetic dose of curcumin-PDA nanoparticles improves viability and proliferation in cell culture.
3894. Banerjee, J., Hasan, S. N., Samanta, S., Giri, B., Bag, B. G., Dash, S. K. Self-Assembled Maslinic Acid Attenuates Doxorubicin Induced Cytotoxicity via Nrf2 Signaling Pathway: An In Vitro and In Silico Study in Human Healthy Cells. // Cell Biochemistry and Biophysics, 2022, Vol. 80, Issue 3, p. 563-578, ISSN 10859195 (ISSN) C2 - 35849306. doi:10.1007/s12013-022-01083-3
3895. Dusi, Renata Garcia. "Desenvolvimento de protótipos de biopesticidas a partir do alho para o controle vetorial de *Aedes aegypti*." (2022)
3896. Gavanji, S., Bakhtari, A., Famurewa, A.C. and Othman, E.M., 2023. Cytotoxic activity of herbal medicines as assessed in vitro: A review. Chemistry & Biodiversity, 20(2), p.e202201098.
3897. Ghasemzadeh, F., Darzi, G. N., Mohammadi, M. Extraction and Purification of Ursolic Acid from the Apple Peel and in vitro Assessment of the Biochemical Antibacterial, Antioxidant and Wound Healing Characteristics. // Applied Food Biotechnology, 2022, Vol. 9, Issue 1, p. 17-30, ISSN 23455357 (ISSN). doi:10.22037/afb.v9i1.34756
3898. Kamalia, A.Z., Tunjung, W. A. S. (2023). Efficacy of Different Solvents in the Extraction of Bioactive Compounds and Anti-cancer Activities of *Thymus vulgaris* Leaves and Twigs. Indonesian Journal of Pharmacy/Majalah Farmasi Indonesia, 34(3).
3899. Koc, A., Karabay, A. Z., Ozkan, T., Buyukbingol, Z., Aktan, F. Time and Concentration Dependent Effects of Different Solvents on Proliferation of K562, HL60, HCT-116 and H929 Cell Lines. // Journal of Research in Pharmacy, 2022, Vol. 26, Issue 3, p. 494-501, ISSN 26306344 (ISSN). doi:10.29228/jrp.146
3900. Korankye, M., 2023. Cytotoxicity Profiling and Apoptotic Potential of Carboxylic Acids-Based Deep Eutectic Solvents on Mammalian Cell Lines (Doctoral dissertation, South Dakota State University).
3901. Mahmoud, D.M., Ali, M.R., Aldosari, B.N. et al. (2024). Functional candesartan loaded lipid nanoparticles for the control of diabetes-associated stroke: In vitro and in vivo studies. International Journal of Pharmaceutics: X, 7, 100227.
3902. Masaryk, L., Zoufalý, P., Słoczyńska, K., Zahradníková, E., Milde, D., Koczurkiewicz-Adamczyk, P., Štarha, P. New Pt(II) diiodido complexes containing bidentate 1, 3, 4-thiadiazole-based ligands: Synthesis, characterization, cytotoxicity. // Inorganica Chimica Acta, 2022, Vol. 536 C7 - 120891, ISSN 00201693 (ISSN). doi:10.1016/j.ica.2022.120891
3903. Mathew, A.A., Antony, M., Thomas, R., Sarojini, S. and Balachandran, M., 2023. Fluorescent PVDF dots: from synthesis to biocidal activity. Polymer Bulletin, 80(1), pp.411-428.
3904. Mathew, Aleena Ann, et al. "Fluorescent PVDF dots: from synthesis to biocidal activity." Polymer Bulletin (2022): 1-18.
3905. Nedeljkovic, I., Doulabi, B. Z., Abdelaziz, M., Feilzer, A. J., Exterkate, R. A. M., Szafert, S., Gulia, N., Krejci, I., Kleverlaan, C. J. Cytotoxicity and anti-biofilm properties of novel hybrid-glass-based caries infiltrant. // Dental Materials, 2022, Vol. 38, Issue 12,

- p. 2052-2061, ISSN 01095641 (ISSN) C2 - 36437129. doi:10.1016/j.dental.2022.11.018
3906. Qiming, Z., Inagaki, N.F., Hirabayashi, Y., Kamihira, M., & Ito, T. (2024). Development of rapid hypoxia-detectable artificial oxygen carriers with a core-shell structure and erythrocyte mimetic shape. *Materials Advances*.
3907. Shafie, A. S., Rashid, A. H. A., Masilamani, T., Zin, N. S. N. M., Azmi, N. A. S., Goh, Y. M., Samsulrizal, N. IN-VITRO MELANOGENESIS, CYTOTOXICITY, AND ANTIOXIDANT ACTIVITIES OF *Peltophorum pterocarpum* LEAF EXTRACTS. // *Malaysian Applied Biology*, 2022, Vol. 51, Issue 4, p. 201-211, ISSN 01268643 (ISSN). doi:10.55230/mabjournal.v51i4.29
3908. Shirazi-Fard, S., Zolghadr, A.R., Klein, A. (2023). How does aggregation of doxorubicin molecules affect its solvation and membrane penetration?. *New Journal of Chemistry*, 47(48), 22063-22077.
3909. Vázquez Alberdi, L., Rosso, G., Velóz, L. et al. (2022). Curcumin and Ethanol Effects in Trembler-J Schwann Cell Culture. *Biomolecules*, 12(4), 515.
3910. Vázquez Alberdi, L., Martínez-Busi, M., Arrarte, E. et al. (2024). A low dose of curcumin-PDA nanoparticles improves viability and proliferation in endoneurial fibroblasts and Schwann cell cultures. *Discover Nano*, 19(1), 81.
3911. Wen, L., Fan, C., Cao, X. (2024). One-step extraction and hydrolysis of crocin into crocetin by recyclable deep eutectic solvents. *Industrial Crops and Products*, 209, 117969.
3912. Wutke, N., 2023. Amphiphilic block copolymers in aqueous and nonaqueous emulsion systems for biomedical applications (Doctoral dissertation, Johannes Gutenberg-Universität Mainz).
- 114). Georgieva, A, Ilieva, Y, Kokanova-Nedialkova, Z, Zaharieva Margaritova, M, Nedialkov, P, Dobрева, A, Kroumov, A, NAJDENSKI, H, Mileva, M. Redox-Modulating Capacity and Antineoplastic Activity of Wastewater Obtained from the Distillation of the Essential Oils of Four Bulgarian Oil-Bearing Roses. *Antioxidants*, 10, 10, MDPI, 2021, ISSN:20763921, DOI:<https://doi.org/10.3390/antiox10101615>, 1615. JCR-IF (Web of Science):7.675**
3913. Butnariu, M., Fernández Ochoa, Á., Segura Carretero, A. and Cádiz Gurrea, M.D.L.L., 2022. A Review on Tradescantia: Phytochemical Constituents, Biological Activities and Health-Promoting Effects. *Front. Biosci. (Landmark Ed)* 2022, 27(6), 197; <https://doi.org/10.31083/j.fbl2706197>
3914. Felicia, W.X.L., Rovina, K., Aqilah, N.M.N., Jaziri, A.A. (2024). Optimisation of supercritical fluid extraction of orange (*Citrus sinensis* L.) peel essential oil and its physicochemical properties. *Current Research in Green and Sustainable Chemistry*, 8, 100410.
3915. Mskhiladze, L., Kakhetlidze, M. and Yavich, P., 2022. Development of a formula for a cosmetic day cream using by-products of oil production from the petals of *Rosa x damascena*, which grows in Georgia. *GEORGIAN SCIENTISTS*, 4(4), pp.21-28.
3916. Niu, Y., Liao, J., Zhou, H., Wang, C. C., Wang, L., Fan, Y. Flavonoids from *Lycium barbarum* Leaves Exhibit Anti-Aging Effects through the Redox-Modulation. // *Molecules*, 2022, Vol. 27, Issue 15 C7 - 4952, ISSN 14203049 (ISSN). doi:10.3390/molecules27154952
3917. Oargă, D.P., Cornea-Cipcigan, M., Cordea, M. I. (2024). Unveiling the mechanisms

- for the development of rosehip-based dermatological products: an updated review. *Frontiers in Pharmacology*, 15, 1390419.
3918. Osman, E.E., Bazaid, S.A. and Abdel-Hameed, E.S.S., 2023. Chemo-profiling and bioactivities of Taif rose (*Rosa damascena* Mill.) industrial by-products after hydrodistillation. *Journal of Applied Pharmaceutical Science*, 13(10), pp.119-131.
3919. Selamovska, A., Elizabeta Miskoska-Milevska, K.N. (2022). Genotype Expression Of Traditional Pear Variety Sinec Depending On Ecological Factors. *Eco-Conference, Ecological Movement Of Novi Sad*.
3920. Trendafilova, A., Staleva, P., Petkova, Z. et al. (2023). Phytochemical Profile, Antioxidant Potential, Antimicrobial Activity, and Cytotoxicity of Dry Extract from *Rosa damascena* Mill. *Molecules*, 28(22), 7666.
3921. Zapata-Zapata, C., Loaiza-Oliva, M., Martínez-Pabón, M.C., Stashenko, E.E. and Mesa-Arango, A.C., 2022. In Vitro Activity of Essential Oils Distilled from Colombian Plants against *Candida auris* and Other *Candida* Species with Different Antifungal Susceptibility Profiles. *Molecules*, 27(20), p.6837.
- 115). Yoncheva K, Benbassat N, Zaharieva MM, Dimitrova LL, Kroumov A, Spassova I, Kovacheva D, NAJDENSKI H. Improvement of the Antimicrobial Activity of Oregano Oil by Encapsulation in Chitosan-Alginate Nanoparticles. *Molecules*, 26, 22, MDPI, 2021, ISSN:1420-3049, DOI:10.3390/molecules26227017, 7017. JCR-IF (Web of Science):4.412**
3922. Aleksandra Such, Anna Wisła-Świder, Ewelina Węsierska, Ewelina Nowak, Piotr Szatkowski, Joanna Kopcińska, Aneta Koronowicz, Edible chitosan-alginate based coatings enriched with turmeric and oregano additives: Formulation, antimicrobial and non-cytotoxic properties, *Food Chemistry*, 426, 2023, 136662.
3923. Baysal, Gülay, et al. "The antioxidant and antibacterial properties of chitosan encapsulated with the bee pollen and the apple cider vinegar." *Journal of Biomaterials Science, Polymer Edition* 33.8 (2022): 995-1011.
3924. Bora, Larisa, et al. "Phytochemical Characterization and Biological Evaluation of *Origanum Vulgare* L. Essential Oil Formulated as Polymeric Micelles Drug Delivery Systems." *Pharmaceutics* 14.11 (2022): 2413.
3925. Caciandone-Cringureanu, M., Anghel, A.G., Anghel, I. (2023). Biofilms on Voice Prosthesis—Challenges and Therapeutic Insights. *Maedica*, 18(3), 498.
3926. de Souza, R.L., Dantas, A.G.B., de Oliveira Melo, C. et al.. (2022). Nanotechnology as a tool to improve the biological activity of carvacrol: a review. *Journal of Drug Delivery Science and Technology*, 76, 103834.
3927. El Fawal, Gomaa, and Marwa M. Abu-Serie. "Bioactive properties of nanofibers based on poly (vinylidene fluoride) loaded with oregano essential oil: Fabrication, characterization and biological evaluation." *Journal of Drug Delivery Science and Technology* 69 (2022): 103133.
3928. Fabrikov, D., Varga, Á.T., García, M.C V. et al. (2024). Antimicrobial and antioxidant activity of encapsulated tea polyphenols in chitosan/alginate-coated zein nanoparticles: a possible supplement against fish pathogens in aquaculture. *Environmental Science and Pollution Research*, 31(9), 13673-13687.
3929. Ghosh, S., Sett, U., Pal, A. et al. (2024). Antibiofilm potential of nanonized eugenol

- against *Pseudomonas aeruginosa*. *Journal of Applied Microbiology*, 135(1), 1xad305.
3930. KABANOVA, S., SHAKHMATOV, P., BORTSOV, V., DANCHENKO, M., SCOTT, S., KABANOV, A., KREKOVA, Y. and KOCHEGAROV, I., 2023. Cultivation experiment of *Origanum vulgare* L. in Northern Kazakhstan using nitrogen fertilizer. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 51(1), pp.13077-13077.
 3931. Khan, Z., Maqsood, Q., Baradoke, A., Ferreira, L.F.R., Franco, M., Schmidt, J.E. and Hussain, N., 2023. Environmental and toxicological implications of chitosan nanostructures. In *Advances in Chemical Pollution, Environmental Management and Protection*. Elsevier
 3932. Kowalczyk, T., Merecz-Sadowska, A., Ghorbanpour, M., Szemraj, J., Piekarski, J., Bijak, M., Śliwiński, T., Zajdel, R. and Sitarek, P., 2023. Enhanced Natural Strength: Lamiaceae Essential Oils and Nanotechnology in In Vitro and In Vivo Medical Research. *International Journal of Molecular Sciences*, 24(20), p.15279.
 3933. Kučuk, N., Primožič, M., Knez, Ž. and Leitgeb, M., 2023. Sustainable Biodegradable Biopolymer-Based Nanoparticles for Healthcare Applications. *International Journal of Molecular Sciences*, 24(4), p.3188.
 3934. Li, Shangyong, et al. "Application of chitosan/alginate nanoparticle in oral drug delivery systems: prospects and challenges." *Drug Delivery* 29.1 (2022): 1142-1149.
 3935. Mercan, D. A., Niculescu, A. G., Grumezescu, A. M. Nanoparticles for Antimicrobial Agents Delivery—An Up-to-Date Review. // *International Journal of Molecular Sciences*, 2022, Vol. 23, Issue 22 C7 - 13862, ISSN 16616596 (ISSN) C2 - 36430343. doi:10.3390/ijms232213862
 3936. Niculescu, A. G., Grumezescu, A. M. Applications of Chitosan-Alginate-Based Nanoparticles— An Up-to-Date Review. // *Nanomaterials*, 2022, Vol. 12, Issue 2 C7 - 186, ISSN 20794991 (ISSN). doi:10.3390/nano12020186
 3937. Osorio-Alvarado, Carlos Enrique, et al. "Immobilization Systems of Antimicrobial Peptide Ib– M1 in Polymeric Nanoparticles Based on Alginate and Chitosan." *Polymers* 14.15 (2022): 3149.
 3938. Rahaiee, S., Shojaosadati, S.A. and Hashemi, M., 2023. An efficient ionic gelation based nano-delivery system to improve the stability and controlled release of saffron extracts. *Biocatalysis and Agricultural Biotechnology*, 52, p.102831.
 3939. Rajasekaran, J., Viswanathan, P. (2023). Anti-bacterial and antibiofilm properties of seaweed polysaccharide-based nanoparticles. *Aquaculture International*, 31(5), 2799-2823.
 3940. Rigopoulos, N., Gkaliouri, C.M., Sakavitsi, V. and Gournis, D., 2023. Full Factorial Design Synthesis of Silver Nanoparticles Using *Origanum vulgare*. *Reactions*, 4(3), pp.505-517.
 3941. Romero-Montero, A., Melgoza-Ramírez, L.J., Ruíz-Aguirre, J.A. et al. (2023). Essential-Oils-Loaded Biopolymeric Nanoparticles as Strategies for Microbial and Biofilm Control: A Current Status. *International Journal of Molecular Sciences*, 25(1), 82.
 3942. Roshandel-Hesari, Narjes, et al. "Investigation of physicochemical properties, antimicrobial and antioxidant activity of edible films based on chitosan/casein containing *Origanum vulgare* L. essential oil and its effect on quality maintenance of cherry tomato." *Food Chemistry* 396 (2022): 133650.
 3943. San, Htet Htet Moe, et al. "Development of Turmeric Oil—Loaded Chitosan/Alginate Nanocapsules for Cytotoxicity Enhancement against Breast Cancer." *Polymers* 14.9

(2022): 1835.

3944. Shinde, M.M., Malik, M., Kaur, K. et al. (2024). Formularization and characterization of guar gum and almond gum based composite coating and their application for shelf-life extension of okra (*Hibiscus esculentus*). *International Journal of Biological Macromolecules*, 129630.
3945. Souza, R. L. D., Dantas, A. G. B., Melo, C. D. O., Felício, I. M., Oliveira, E. E. Nanotechnology as a tool to improve the biological activity of carvacrol: A review. // *Journal of Drug Delivery Science and Technology*, 2022, Vol. 76 C7 - 103834, ISSN 17732247 (ISSN). doi:10.1016/j.jddst.2022.103834
3946. Such, A., Wisła-Świder, A., Węsierska, E. et al. (2023). Edible chitosan-alginate based coatings enriched with turmeric and oregano additives: Formulation, antimicrobial and non-cytotoxic properties. *Food Chemistry*, 426, 136662.
3947. Yang, X., Zhao, D., Ge, S., Bian, P., Xue, H. and Lang, Y., 2023. Alginate-based edible coating with oregano essential oil/ β -cyclodextrin inclusion complex for chicken breast preservation. *International Journal of Biological Macromolecules*, 251, p.126126.
3948. Yuan, Yanghua, et al. "Sodium alginate/gum arabic/glycerol multicomponent edible films loaded with natamycin: Study on physicochemical, antibacterial, and sweet potatoes preservation properties." *International Journal of Biological Macromolecules* 213 (2022): 1068-1077.
3949. Wathoni, N., Herdiana, Y., Suhandi, C. et al. (2024). Chitosan/Alginate-Based Nanoparticles for Antibacterial Agents Delivery. *International Journal of Nanomedicine*, 5021-5044.
- 116). Nikolov, A.S., N.E. Stankova, D.B. Karashanova, N.N. Nedyalk, E.L. Pavlov, K.Tz. Koev, Hr. NAJDENSKI, V. Kussovski, L.A. Avramov, C. Ristoscu, M. Badiceanu, I.N. Mihailescu. Synergistic effect in a two-phase laser procedure for production of silver nanoparticles colloids applicable in ophthalmology. *Optics and Laser Technology*, 138, Elsevier, 2021, ISSN:0030-3992, DOI:https://doi.org/10.1016/j.optlastec.2020.106850, 1-8. SJR (Scopus):3.867**
3950. Alhajj, M., Abd Aziz, M.S., Salim, A.A. et al. (2023). Customization of structure, morphology and optical characteristics of silver and copper nanoparticles: Role of laser fluence tuning. *Applied Surface Science*, 614, 156176.
3951. Gruszka, J. (2023). Badania nanocząstek srebra i tlenku tytanu (IV) w próbkach środowiskowych i biologicznych z zastosowaniem techniki single particle ICP MS.
3952. Jianing, Liao, Zhang Dongshi, and Li Zhuguo. "Advance in femtosecond laser fabrication of flexible electronics." *Opto-Electronic Engineering* 49.2 (2022): 210388-1.
3953. Kasáľková, N.S., Juřicová, V., Rimpelová, S. et al. (2024). LIPSS pattern induced by polymer surface instability for myoblast cell guidance. *Polymer Degradation and Stability*, 221, 110667.
3954. Liao Jianing, Zhang Dongshi, Li Zhuguo. (2022). Progress in femtosecond laser fabrication of flexible electronic devices. *Optoelectronic Engineering*, 49(2), 210388-1.
3955. López-Álvarez, M., González-Rodríguez, L., Gontad, F., et al. "Dual pulsed laser deposition of Ag nanoparticles on calcium phosphate coatings for biomedical applications" *Biomedical Physics and Engineering Express*, 2022, 8(6), 065019
3956. Mahmood Alhajj, Md. Safwan Abd Aziz, A.A. Salim, Sunita Sharma, W.H.A.

- Kamaruddin, S.K. Ghoshal. Customization of structure, morphology and optical characteristics of silver and copper nanoparticles: Role of laser fluence tuning. *Applied Surface Science*, Volume 614, 2023, 156176, ISSN 0169-4332, <https://doi.org/10.1016/j.apsusc.2022.156176>
3957. Marquis, M., Musino, D., Gemin, V. et al. (2023). Alginate microgels encapsulation strategy of silver nanoparticles active against *Candida albicans*. *Carbohydrate Polymer Technologies and Applications*, 6, 100405.
3958. Nazha, H.M., Darwich, M.A., Ammar, B. et al. (2024). Determination of Laser Parameters in Thermomechanical Treatment of Skin Based on Response Surface Methodology. *Applied Sciences*, 14(6), 2619.
3959. Nene, A., Galluzzi, M., Hongrong, L. et al. (2021). Synthetic preparations and atomic scale engineering of silver nanoparticles for biomedical applications. *Nanoscale*, 13(33), 13923-13942.
3960. Pérez-Tanoira, R., Fernández-Arias, M., Potel, C., et al. "Silver Nanoparticles Produced by Laser Ablation and Re-Irradiation Are Effective Preventing Peri-Implantitis Multispecies Biofilm Formation". *International Journal of Molecular Sciences*, 2022, 23(19), 12027.
3961. Suliman, E.M., Nayef, U.M., Mutlak, F.A. (2022). Synthesis of Au: TiO₂ nanoparticles via laser ablation in liquid deposited on porous-Si for improved spectral responsivity. *Journal of Applied Sciences and Nanotechnology*, 2(3), 147-156.
- 117). Ivanova, DI, Nedialkov, PT, Tashev, AN, Olech, M, Nowak, R, Ilieva, YE, Nedialkova-Kokanova, ZK, Atanasova, TN, Angelov, G, NAJDENSKI, HM. Junipers of Various Origins as Potential Sources of the Anticancer Drug Precursor Podophyllotoxin. *Molecules*, 26 (17), 5179, MDPI Multidisciplinary Digital Publishing Institute, 2021, ISSN:1420-3049, DOI:10.3390/ molecules26175179, SJR (Scopus):0.782, JCR-IF (Web of Science):3.267**
3962. Bohórquez-Moreno C.D., Öksüz K.E., Dinger E., Hepokur C., Şen İ. Plant-inspired adhesive and injectable natural hydrogels: in vitro and in vivo studies. (2023) *Biotechnology Letters*, 45 (9), pp. 1209 – 1222. DOI: 10.1007/s10529-023-03400-z.
3963. El-Banna, A.A., Shawky, E., Celik, I. et al. (2024). Deciphering the putative bioactive metabolites and the underlying mechanism of *Juniperus horizontalis* Moench (Creeping juniper) in the treatment of inflammation using network pharmacology and molecular docking. *Journal of Pharmacy and Pharmacology*, 76(5), 514-533.
3964. Hao, M. and Xu, H., 2023. Chemistry and Biology of Podophyllotoxins– An Update. *Chemistry–A European Journal*, p.e202302595.
3965. Kaswan P. Natural resources as cancer-treating material. (2023) *South African Journal of Botany*, 158, pp. 369 – 392. DOI: 10.1016/j.sajb.2023.05.028.
3966. Motyka S., Jaferník K., Ekiert H., Sharifi-Rad J., Calina D., Al-Omari B., Szopa A., Cho W.C. Podophyllotoxin and its derivatives: Potential anticancer agents of natural origin in cancer chemotherapy. (2023) *Biomedicine and Pharmacotherapy*, 158, art. no. 114145. DOI: 10.1016/j.biopha.2022.114145.
3967. Rana, Anita, et al. "Functionalized graphene oxide based nanocarrier for enhanced cytotoxicity of *Juniperus squamata* root essential oil against breast cancer cells." *Journal of Drug Delivery Science and Technology* 72 (2022): 103370.

3968. Sakamoto K., Fujimoto R., Nakagawa S., Kamiyama E., Kanai K., Kawai Y., Kojima H., Hirasawa A., Wakamatsu K., Masutani T. Juniper berry extract containing Anthricin and Yatein suppresses lipofuscin accumulation in human epidermal keratinocytes through proteasome activation, increases brightness and decreases spots in human skin. (2023) International Journal of Cosmetic Science, 45 (5), pp. 655 – 671. DOI: 10.1111/ics.12876.
3969. Schnelle, M.A. (2023). Eastern Redcedar: A United States Native Tree That Ranges from Useful, to a Nuisance, and Even Invasive in Certain Environments. HortTechnology, 33(6), 570-577.
3970. Stenmark, K.R. (2021). Extraction and Purification of Podophyllotoxin from Eastern Red Cedar (*Juniperus virginiana* L.) in Oklahoma (Master's thesis, Oklahoma State University).
3971. Turah, B.T., Acar, D.İ., Gultehin, S.K. et al. (2022). Phytochemical constituents and antileukemic effects of *Juniperus oxycedrus* extract. Biotechnologia Acta, 15(5), 64-70.
3972. Verret, C., Rakotondramavo, A., Renouard, S. (2023). Green Ultrasound-Assisted Extraction of Podophyllotoxin from *Juniperus scopulorum* Needles. Applied Sciences, 13(22), 12194.
3973. Xu, S., Li, X., Liu, S., Tian, P., Li, D. "Juniperus sabina L. as a Source of Podophyllotoxins: Extraction Optimization and Anticholinesterase Activities". International Journal of Molecular Sciences, 23(18), 2022, 10205
3974. Савельева, О.В., 2023. Определение качественного состава экстрактов можжевельника ложноказацкого методом газовой хромато-масс-спектрометрии: выпускная бакалаврская работа по направлению подготовки: 04.03. 01-Химия.
- 118). Mileva, M, Dimitrova, L, Popova, M, Bankova, V, Krastev, D, NAJDENSKI, H, Zhelev, Z, Aoki, I, Bakalova, R. Redox-modulation, suppression of “oncogenic” superoxide and induction of apoptosis in Burkitt`s lymphoma cells using *Geum urbanum* L. extracts. International Journal Bioautomation, 25, 4, 2021, ISSN:13142321, 13141902, DOI:DOI: 10.7546/ijba.2021.25.4.000795, 315-330. SJR (Scopus):0.24**
3975. Mostafa EM, Mohammed HA, Musa A, Abdelgawad MA, Al-Sanea MM, Almahmoud SA, Ghoneim MM, Gomaa HA, Rahman FE, Shalaby K, Selim S. In Vitro Anti-Proliferative, and Kinase Inhibitory Activity of Phenanthroindolizidine Alkaloids Isolated from *Tylophora indica*. Plants. 2022 May 12;11(10):1295.
3976. Wang, Fei. "A Gene-disease Association Prediction Algorithm Based on Multi-source Data Fusion." International Journal Bioautomation 26, no. 1 (2022): 19
- 119). Zaharieva, M.M., Dimitrova, L.L., Philipov, S., Nikolova, I., Vilhelmova, N., Grozdanov, P., Nikolova, N., Popova, M., Bankova, V., Konstantinov, S.M. and Zheleva-Dimitrova, D., H. NAJDENSKI 2021. In vitro antineoplastic and antiviral activity and in vivo toxicity of *Geum urbanum* L. extracts. Molecules, 27(1), p.245**
3977. El-Tanani, M., Nsairat, H., Matalka, I.I. et al. (2024). The impact of the BCR-ABL oncogene in the pathology and treatment of chronic myeloid leukemia. Pathology-Research and Practice, 155161.
3978. Kashchenko, N.I., Olennikov, D.N. and Chirikova, N.K., 2023. Metabolites of *Geum aleppicum* and *Sibbaldianthe bifurca*: Diversity and α -Glucosidase Inhibitory Potential. Metabolites, 13(6), p.689.
3979. Kozłowska, M., Ścibisz, I., Przybył, J.L. et al. (2022). Antioxidant and antibacterial

- activity of extracts from selected plant material. *Applied Sciences*, 12(19), 9871.
3980. Li, J., Liao, R., Zhang, S., Weng, H., Liu, Y., Tao, T., Yu, F., Li, G. and Wu, J., 2023. Promising Remedies for Cardiovascular Disease: Natural Polyphenol Ellagic Acid and Its Metabolite Urolithins. *Phytomedicine*, p.154867
- 120). Guimarães, T, Moniz, T, Nunes, C, Zaharieva, MM, Kaleva, M, Yoncheva, K, NAJDENSKI, H, Lima, S. A. C., Reis, S. Polymeric microneedles for transdermal delivery of rivastigmine: Design and application in skin mimetic model. *Pharmaceutics*, 14, 752, MDPI, 2022, ISSN:1999-4923, DOI:<https://doi.org/10.3390/pharmaceutics14040752>, JCR-IF (Web of Science):6.525**
3981. Aulia, N.R., Putri, A.P.D., Pratama, F.A. et al. (2023). Implantable Trilayer Microneedle Transdermal Delivery System to Enhance Bioavailability and Brain Delivery of Rivastigmine for Alzheimer Treatment: a Proof-of-concept Study.
3982. McNamee M, Wong S, Guy O, Sharma S. Microneedle technology for potential SARS-CoV-2 vaccine delivery. *Expert Opin Drug Deliv*. 2023 Jun;20(6):799-814. doi: 10.1080/17425247.2023.2209718. Epub 2023 May 8. PMID: 37128730
3983. Monou, P.K., Andriotis, E., Tzetzis, D. et al. (2024). Evaluation of 3D-printed solid microneedles coated with electrosprayed polymeric nanoparticles for simultaneous delivery of rivastigmine and N-acetyl cysteine. *ACS Applied Bio Materials*, 7(5), 2710-2724.
3984. Nunes, D., Loureiro, J. A., Pereira, M. C. Drug Delivery Systems as a Strategy to Improve the Efficacy of FDA-Approved Alzheimer's Drugs. // *Pharmaceutics*, 2022, Vol. 14, Issue 11 C7 - 2296, ISSN 19994923 (ISSN). doi:10.3390/pharmaceutics14112296.
3985. Putri, A.P.D., Ilyas, N.R.A., Abdullah, D.A.P. et al. (2024). Development and validation of UV–Vis spectrophotometric method for determination of rivastigmine in PBS and biological matrices: Application to ex vivo permeation profiles and in vivo studies from trilayer dissolving microneedle. *Chemical Data Collections*, 49, 101106.
3986. Singh, B., Day, C.M., Abdella, S. et al. (2024). Alzheimer's disease current therapies, novel drug delivery systems and future directions for better disease management. *Journal of Controlled Release*, 367, 402-424.
- 121). Zaharieva MM, Dimitrova LL, Philipov S, Nikolova I, Vilhelmova N, Grozdanov P, Nikolova N, Popova M, Bankova V, Konstantinov SM, Zheleva-Dimitrova D, NAJDENSKI H. In Vitro Antineoplastic and Antiviral Activity and In Vivo Toxicity of *Geum urbanum* L. Extracts. *Molecules*, 27, 1, MDPI, 2022, ISSN:1420-3049, DOI:10.3390/molecules27010245, 245. SJR (Scopus):0.705, JCR-IF (Web of Science):4.927**
3987. Danova, S., Yankov, D., Dobрева, L., Dobрева, A., Armenova, N., Apostolov, A. and Mileva, M., 2023. Postbiotics Production of Candidate-Probiotic *Lactiplantibacillus plantarum* AC131 with Renewable Bio Resources. *Life*, 13(10), p.2006.
3988. El-Tanani, M., Nsairat, H., Matalka, I.I. et al. (2024). The impact of the BCR-ABL oncogene in the pathology and treatment of chronic myeloid leukemia. *Pathology-Research and Practice*, 155161.
3989. Kashchenko N.I., Olennikov D.N., Chirikova N.K. Metabolites of *Geum aleppicum* and *Sibbaldianthe bifurca*: Diversity and α -Glucosidase Inhibitory Potential. *Metabolites*, 13 (6), art. no. 689, 2023. DOI: 10.3390/metabo13060689.

3990. Kozłowska, M.; Ścibisz, I.; Przybył, J.L.; Laudy, A.E.; Majewska, E.; Tarnowska, K.; Małajowicz, J.; Ziarno, M. Antioxidant and Antibacterial Activity of Extracts from Selected Plant Material. *Appl. Sci.* 2022, 12, 9871. <https://doi.org/10.3390/app12199871>
3991. Li J., Liao R., Zhang S., Weng H., Liu Y., Tao T., Yu F., Li G., Wu J. Promising remedies for cardiovascular disease: Natural polyphenol ellagic acid and its metabolite urolithins. *Phytomedicine*, 116, art. no. 154867, 2023. DOI: 10.1016/j.phymed.2023.154867.
3992. Nazlić, M., Dunkić, V., Dželalija, M., Maravić, A., Mandić, M., Srećec, S., Vrca, I., Vuko, E. and Kremer, D., 2023. Evaluation of Antiphytoviral and Antibacterial Activity of Essential Oil and Hydrosol Extracts from Five Veronica Species. *Agriculture*, 13(8), p.1517.
3993. Osman, E.E., Bazaid, S.A. and Abdel-Hameed, E.S.S., 2023. Chemo-profiling and bioactivities of Taif rose (*Rosa damascena* Mill.) industrial by-products after hydrodistillation. *Journal of Applied Pharmaceutical Science*, 13(10), pp.119-131.
3994. Rusanova, M., Rusanov, K., Butterweck, V. and Atanasov, I., 2023. Indigenous Yeasts from Rose Oil Distillation Wastewater and Their Capacity for Biotransformation of Phenolics. *Microorganisms*, 11(1), p.201.
- 122). Mantareva, V., Kussovski, V., Orozova, P., Dimitrova, L., Kulu, I., Angelov, I., Durmus, M., NAJDENSKI, H.. Photodynamic Inactivation of Antibiotic-Resistant and Sensitive *Aeromonas hydrophila* with Peripheral Pd(II)- vs. Zn(II)-Phthalocyanines. *Biomedicines*, 10, 2, 2022, DOI:<https://doi.org/10.3390/biomedicines10020384>, JCR-IF (Web of Science):6.081**
3995. Dube, E., Okuthe, G.E. (2024). Applications of Antimicrobial Photodynamic Therapy in Aquaculture: Effect on Fish Pathogenic Bacteria. *Fishes*, 9(3), 99.
3996. Mušković, M., Gobin, I., Malatesti, N. (2023). Photodynamic Inactivation of Opportunistic Premise Plumbing Pathogens and Their Biofilms. *Processes*, 11(11), 3074.
3997. Kang, K., & Bacci, S. (2022). Photodynamic Therapy. *Biomedicines*, 10(11), 2701.
3998. Liu, Xiang, et al. "The identification of polyvalent protective immunogens and immune abilities from the outer membrane proteins of *Aeromonas hydrophila* in fish." *Fish & Shellfish Immunology* 128 (2022): 101-112.
3999. Tarpaga, Lassané, et al. "Synthesis and Study of the Physicochemical Properties of a Hybrid Species: Iron Phthalocyanine–Silver Nanoparticles." *Chemistry Africa* 5.4 (2022): 811-820.
4000. Urgesa, G., Lu, L., Gao, J. et al. (2024). Natural Sunlight-Mediated Emodin Photoinactivation of *Aeromonas hydrophila*. *International Journal of Molecular Sciences*, 25(10), 5444.
4001. Xu, P., Liu, J., Yi, Y. et al. (2024). A dew-responsive pectin-based herbicide for enhanced photodynamic inactivation. *Carbohydrate Polymers*, 336, 122114.
4002. Zalevskaya O.A., Gur Y.A., Kutchin A.V. Palladium complexes as promising antimicrobial agents. (2023) *Russian Chemical Reviews*, 92 (9), art. no. RCR5093. DOI: 10.59761/RCR5093
- 123). Valcheva, V., Perea, C., Savova-Lalkovska, T., Dimitrova, A., Radulski, L., Mokrousov, I., Marinov, K., NAJDENSKI H., Bonovska M.. *Mycobacterium bovis* and *M. caprae* in Bulgaria: insight into transmission and phylogeography gained through**

whole-genome sequencing.. BMC Veterinary Research, 18, 1, BMC, 2022, ISSN:17466148, DOI:<https://doi.org/10.1186/s12917-022-03249-w>, 148-159. SJR (Scopus):0.65, JCR-IF (Web of Science):2.792

4003. Gao, G., Cui, Y. and Cheng, H., 2023. Association between retinol binding protein-4 and psoriasis vulgaris: a systematic review and meta-analysis. *Frontiers in Medicine*, 10
4004. Lorente Leal, V. (2023). Nuevas metodologías para el diagnóstico de la tuberculosis bovina.
4005. Martínez, L. (2012). Influencia de la bioestimulación en la actividad cíclica y la tasa de preñez en vaquillonas de carne sometidas a un protocolo de sincronización en base a análogos de PGF2?.
4006. Ncube, Pamela, et al. "Evidence, Challenges, and Knowledge Gaps Regarding Latent Tuberculosis in Animals." *Microorganisms* 10.9 (2022): 1845.
4007. Pereira, A.C., Lourenço, J., Themudo, G. et al. (2024). Population structure and history of *Mycobacterium bovis* European 3 clonal complex reveal transmission across ecological corridors of unrecognized importance in Portugal. *Microbiology Spectrum*, e03829-23.
4008. Zahran, M., El-Shabasy, R.M., Elrashedy, A. et al. (2023). Recent progress in the genotyping of bovine tuberculosis and its rapid diagnosis via nanoparticle-based electrochemical biosensors. *RSC advances*, 13(45), 31795-31810.

124). Gugleva, V, Michailova, V, Mihaylova, R, Momekov, G, Zaharieva, MM, NAJDENSKI, H, Petrov, P, Rangelov, S, Forys, A, Trzebicka, B, Momekova, D. Formulation and Evaluation of Hybrid Niosomal In Situ Gel for Intravesical Co-Delivery of Curcumin and Gentamicin Sulfate. *Pharmaceutics*, 14, 4, MDPI, 2022, DOI:<https://doi.org/10.3390/pharmaceutics14040747>, 747. JCR-IF (Web of Science):6.525

4009. Al-Zuhairy S.A.S., El-Sawy H.S., El-Nabarawi M.A., Teaima M.H. FOCUS ON NIOSOMAL-BASED DRUG DELIVERY SYSTEMS FOR NASAL ROUTE: APPLICATIONS AND CHALLENGES. *International Journal of Applied Pharmaceutics*, 15 (1), pp. 36 – 43, 2023. DOI: 10.22159/ijap.2023v15i1.46280
4010. Baylan, B., Erdal, B. (2024). Investigation of Antibacterial Activity of Curcumin and Synergistic Effect with Gentamicin Sulfate.
4011. Eldehna W.M., El Hassab M.A., Abdelshafi N.A., Eissa R.A., Diab N.H., Mohamed E.H., Oraby M.A., Al-Rashood S.T., Eissa R.G., Elsayed Z.M., Nocentini A., Supuran C.T., Elsabahy M., Eissa N.G. Development of potent nanosized carbonic anhydrase inhibitor for targeted therapy of hypoxic solid tumors. *Int J Pharm.* 2023 Jan 25;631:122537. doi: 10.1016/j.ijpharm.2022.122537. Epub 2022 Dec 23. PMID: 36572260.
4012. Faheem, S., Hameed, H., Paiva-Santos, A.C. et al. (2024). Niosome-based gels: a smart nano-carrier for effective and advanced transdermal drug delivery. *International Journal of Polymeric Materials and Polymeric Biomaterials*, 1-19.
4013. Fahmy S.A., Nasr S., Ramzy A., Dawood A.S., Abdelnaser A., Azzazy H.M.E. Cytotoxic and Antioxidative Effects of Geranium Oil and Ascorbic Acid Coloaded in Niosomes against MCF-7 Breast Cancer Cells. *ACS Omega*. 2023 Jun 13;8(25):22774-

22782. doi: 10.1021/acsomega.3c01681. PMID: 37396262; PMCID: PMC10308595.
4014. Fahmy, Sherif Ashraf, et al. "Ozonated Olive Oil: Enhanced Cutaneous Delivery via Niosomal Nanovesicles for Melanoma Treatment." *Antioxidants* 11.7 (2022): 1318.
4015. Fernandes S.C.M., Aguirre G. Biopolymer Micro/Nanogel Particles as Smart Drug Delivery and Theranostic Systems. *Pharmaceutics*, 15 (8), art. no. 2060, 2023. DOI: 10.3390/pharmaceutics15082060.
4016. Gopalakrishna P.K., Jayaramu R.A., Boregowda S.S., Eshwar S., Suresh N.V., Abu Lila A.S., Moin A., Alotaibi H.F., Obaidullah A.J., Khafagy E-S. Piperine-Loaded In Situ Gel: Formulation, In Vitro Characterization, and Clinical Evaluation against Periodontitis. *Gels*. 2023; 9(7):577. <https://doi.org/10.3390/gels9070577>.
4017. Hemmati, J., Chegini, Z., Arabestani, M.R. (2023). Niosomal-based drug delivery platforms: a promising therapeutic approach to fight *Staphylococcus aureus* drug resistance. *Journal of Nanomaterials*, 2023.
4018. Ibrahim, B., Shamma, R., Salama, A. et al. (2023). Magnetic targeting of lornoxicam/SPION bilosomes loaded in a thermosensitive in situ hydrogel system for the management of osteoarthritis: Optimization, in vitro, ex vivo, and in vivo studies in rat model via modulation of RANKL/OPG. *Drug Delivery and Translational Research*, 1-21.
4019. Omidian H., Wilson R.L., Chowdhury S.D. Enhancing Therapeutic Efficacy of Curcumin: Advances in Delivery Systems and Clinical Applications. *Gels*, 9 (8), art. no. 596, 2023. DOI: 10.3390/gels9080596.
4020. Pourmadadi, M., Abbasi, P., Eshaghi, M. M., Bakhshi, A., Ezra Manicum, A. L., Rahdar, A., Pandey, S., Jadoun, S., Díez-Pascual, A. M. Curcumin delivery and co-delivery based on nanomaterials as an effective approach for cancer therapy. // *Journal of Drug Delivery Science and Technology*, 2022, Vol. 78 C7 - 103982, ISSN 17732247 (ISSN). doi:10.1016/j.jddst.2022.103982
4021. Rajak P., Patra E., Karmakar A., Bhuyan B. Xanthium strumarium L. Extract Loaded Phyto-Niosome Gel: Development and In Vitro Assessment for the Treatment of Tinea corporis. *Biointerface Research in Applied Chemistry*, 13 (3), art. no. 273, 2023. DOI: 10.33263/BRIAC133.273.
4022. Roostae, M., Derakhshani, A., Mirhosseini, H. et al. (2024). Composition, preparation methods, and applications of nanoniosomes as codelivery systems: a review of emerging therapies with emphasis on cancer. *Nanoscale*.
4023. Salem, H.F., Nafady, M.M., Eissa, E.M. et al. (2024). Assembly of In-Situ Gel Containing Nano-Spanlastics of an Angiotensin II Inhibitor as a Novel Epitome for Hypertension Management: Factorial Design Optimization, In-vitro Gauging, Pharmacokinetics, and Pharmacodynamics Appraisal. *AAPS PharmSciTech*, 25(5), 115.
4024. Uboldi, M.; Perrotta, C.; Moscheni, C.; Zecchini, S.; Napoli, A.; Castiglioni, C.; Gazzaniga, A.; Melocchi, A.; Zema, L. Insights into the Safety and Versatility of 4D Printed Intravesical Drug Delivery Systems. *Pharmaceutics* 2023, 15, 757. <https://doi.org/10.3390/pharmaceutics15030757>.
4025. Vyas, D., Mukhopadhyay, S., Tamta, B. (2022). Divalproex Sodium Niosomes: Formulation and Evaluation for the Treatment of Epilepsy. *NeuroQuantology*, 20(10), 3514.
4026. Xu, Z., Gao, J., Zhang, H. et al. (2024). A thermosensitive hydrogel based arginine grafted chitosan and poloxamer 407 for wound healing. *European Polymer Journal*, 113129.

4027. Yasamineh, Saman, et al. "A state-of-the-art review on the recent advances of niosomes as a targeted drug delivery system." *International Journal of Pharmaceutics* (2022): 121878.

125). Ilieva, Y, Dimitrova, L, Georgieva, A, Vilhelmova-Ilieva, N, Zaharieva, MM, Kokanova-Nedialkova, Z, Dobрева, A, Paraskev, N, Kussovski, V, NAJDENSKI, H, Mileva, M. In Vitro Study of the Biological Potential of Wastewater Obtained after the Distillation of Four Bulgarian Oil-Bearing Roses. *Plants*, 11, 8, MDPI, 2022, ISSN:2223-7747, DOI:10.3390/plants11081073, 1073. SJR (Scopus):0.765, JCR-IF (Web of Science):4.658

4028. Antoniadou, M., Rozos, G., Vaiou, N. et al. (2023). The In Vitro Assessment of Antibacterial and Antioxidant Efficacy in *Rosa damascena* and *Hypericum perforatum* Extracts against Pathogenic Strains in the Interplay of Dental Caries, Oral Health, and Food Microbiota. *Microorganisms*, 12(1), 60.

4029. Nazlić M., Dunkić V., Dželalija M., Maravić A., Mandić M., Srećec S., Vrca I., Vuko E., Kremer D. Evaluation of Antiphytoviral and Antibacterial Activity of Essential Oil and Hydrosol Extracts from Five *Veronica* Species. (2023) *Agriculture (Switzerland)*, 13 (8), art. no. 1517. DOI: 10.3390/agriculture13081517

4030. Nazlić, M., Akrap, K., Kremer, D. and Dunkić, V., 2022. Hydrosols of *Veronica* Species—Natural Source of Free Volatile Compounds with Potential Pharmacological Interest. *Pharmaceutics*, 15(11), p.1378.

4031. Nazlić, M.; Kremer, D.; Akrap, K.; Topić, S.; Vuletić, N.; Dunkić, V. Extraction, Composition and Comparisons—Free Volatile Compounds from Hydrosols of Nine *Veronica* Taxa. *Horticulturae* 2023, 9, 16. <https://doi.org/10.3390/horticulturae9010016>.

4032. Nazlić, M., Dunkić, V., Dželalija, M. et al. (2023). Evaluation of Antiphytoviral and Antibacterial Activity of Essential Oil and Hydrosol Extracts from Five *Veronica* Species. *Agriculture*, 13(8), 1517.

4033. Osman, E.E., Bazaid, S.A. and Abdel-Hameed, E.S.S., 2023. Chemo-profiling and bioactivities of Taif rose (*Rosa damascena* Mill.) industrial by-products after hydrodistillation. *Journal of Applied Pharmaceutical Science*, 13(10), pp.119-131.

4034. Rusanova, M.; Rusanov, K.; Butterweck, V.; Atanassov, I. Indigenous Yeasts from Rose Oil Distillation Wastewater and Their Capacity for Biotransformation of Phenolics. *Microorganisms* 2023, 11, 201. <https://doi.org/10.3390/microorganisms11010201>.

4035. Trendafilova, A., Staleva, P., Petkova, Z. et al. (2023). Phytochemical Profile, Antioxidant Potential, Antimicrobial Activity, and Cytotoxicity of Dry Extract from *Rosa damascena* Mill. *Molecules*, 28(22), 7666.

126). Tsvetanova Z., Tsvetkova I., NAJDENSKI H.. Antimicrobial Resistance of Heterotrophic Bacteria in Drinking Water-Associated Biofilms. *Water*, 14, 944, MDPI, 2022, SJR (Scopus):0.716, JCR-IF (Web of Science):3.53

4036. Bosch, J., Bezuidenhout, C., Coertze, R. et al. Metal- and antibiotic-resistant heterotrophic plate count bacteria from a gold mine impacted river: the Mooi River system, South Africa. *Environ Sci Pollut Res* 30, 31605–31619 (2023). <https://doi.org/10.1007/s11356-022-24015-3>

4037. Chen, Y., Li Y., Yang S., Chiang T.Y., Zhu X., Hu J. Controlling Biofilm Growth and Its Antibiotic Resistance in Drinking Water by Combined UV and Chlorination Processes.

- Water, 2022, 14 (22), 3643.
4038. Gnimadi, C.J.I., Gawou, K., Aboah, M. et al. (2024). Assessing the Influence of Hand-Dug Well Features and Management on Water Quality. *Environmental Health Insights*, 18, 11786302241249844.
4039. Mohamad, Z. A., Bakon, S. K., Jamilan, M. A. J., Daud, N., Ciric, L., Ahmad, N., Muhamad, N. A. Prevalence of Antibiotic-Resistant Bacteria and Antibiotic-Resistant Genes and the Quantification of Antibiotics in Drinking Water Treatment Plants of Malaysia: Protocol for a Cross-sectional Study. *JMIR Research Protocols*, 2022, 11(11), e37663.
4040. Vassallo, A., Kett, S., Purchase, D., & Marvasi, M. (2022). The bacterial urban resistome: recent advances, *Antibiotics*, 11(4), 512.
4041. Wolf-Baca M, Siedlecka A. Seasonal and spatial variations of antibiotic resistance genes and bacterial biodiversity in biofilms covering the equipment at successive stages of drinking water purification. *J Hazard Mater.* 2023 Aug 15;456:131660. doi: 10.1016/j.jhazmat.2023.131660. Epub 2023 May 18. PMID: 37210784.
- 127). Zaharieva, MM, Zheleva-Dimitrova, D, Rusinova-Videva, S, Ilieva, Y, Brachkova, A, Balabanova, V, Gevrenova, R, Kim, TC, Kaleva, M, Georgieva, M, Mileva, M, Yoncheva, K, Benbassat, N, NAJDENSKI, H, Kroumov, AD. Antimicrobial and Antioxidant Potential of *Scenedesmus obliquus* Microalgae in the Context of Integral Biorefinery Concept. *Molecules*, 27, 2, MDPI, 2022, ISSN:14203049, DOI:<https://doi.org/10.3390/molecules27020519>, 519. JCR-IF (Web of Science):4.927**
4042. Akermi, Sarra, Slim Smaoui, Khaoula Elhadeif, Mariam Fourati, Nacim Louhichi, Moufida Chaari, Ahlem Chakchouk Mtibaa, Aissette Baanannou, Saber Masmoudi, and Lotfi Mellouli. "Cupressus sempervirens Essential Oil: Exploring the Antibacterial Multitarget Mechanisms, Chemcomputational Toxicity Prediction, and Safety Assessment in Zebrafish Embryos." *Molecules* 27, no. 9 (2022): 2630.
4043. Alves, G., Franceschet, E., Severgnini, D. et al. (2023). Euglena sp. CULTIVADA COM SUPLEMENTAÇÃO DE RESÍDUOS SEPARADOS DE CERVEJARIA ARTESANAL E SEU POTENCIAL ANTIOXIDANTE. *REVISTA FOCO*, 16(12), e3811-e3811.
4044. Balouch, H., Demirbag, Z., Durani, M. et al. (2024). Antibacterial activity of freshwater green microalgae from Almaty region. In *BIO Web of Conferences* (Vol. 100, p. 02014). EDP Sciences.
4045. Bolaños-Martínez, O.C., Mahendran, G., Rosales-Mendoza, S. and Vimolmangkang, S., 2022. Current Status and Perspective on the Use of Viral-Based Vectors in Eukaryotic Microalgae. *Marine Drugs*, 20(7), p.434.
4046. Dejtsakdi, W., Maneeruttanarungroj, C. INVESTIGATION OF ANTIFUNGAL ACTIVITY FROM SCENEDESMUS SP. AND CHLAMYDOMONAS SP. CRUDE EXTRACTS. //, Chevet, P. F., Scarlat, N., Grassi, A. (eds), *ETA-Florence Renewable Energies*, 2022, p. 1088-1091. ISSN:22825819 (ISSN),
4047. Fayzullayevich, Ismailov Zafar. "BIR HUYAYRALI MIKROSUVO 'TLARINING ANTIBAKTERIAL VA ANTIFUNGAL XUSUSIYATLARI." *INTERNATIONAL JOURNAL OF SCIENCE AND EDUCATION* 1.1 (2022): 22-26.
4048. Fernandes, A.S., Caetano, P.A., Jacob-Lopes, E. et al. (2024). Alternative green

- solvents associated with ultrasound-assisted extraction: A green chemistry approach for the extraction of carotenoids and chlorophylls from microalgae. *Food Chemistry*, 139939.
4049. Flores, M.L., Jiménez-Veuthey, M., Córdoba, O.L. Metabolites from Microalgal Cultures as Potential Sources for the Pharmaceutical Industry. *Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures*, 139.
4050. Georgiopoulou I, Louli V, Magoulas K. Comparative Study of Conventional, Microwave-Assisted and Supercritical Fluid Extraction of Bioactive Compounds from Microalgae: The Case of *Scenedesmus obliquus*. *Separations*. 2023; 10(5):290. <https://doi.org/10.3390/separations10050290>
4051. He Z, Nam S, Liu S, Zhao Q. Characterization of the Nonpolar and Polar Extractable Components of Glanded Cottonseed for Its Valorization. *Molecules*. 2023; 28(10):4181. <https://doi.org/10.3390/molecules28104181>
4052. He Zhongqi, Shasha Liu, Sunghyun Nam, K. Thomas Klasson, Huai N. Cheng. Molecular level characterization of the effect of roasting on the extractable components of glandless cottonseed by Fourier transform ion cyclotron resonance mass spectrometry. *Food Chemistry*, Volume 403, 2023, 134404, ISSN 0308-8146, <https://doi.org/10.1016/j.foodchem.2022.134404>.
4053. Johannesen, B.A., 2022. Optimization of green extraction conditions and determination of Fucoxanthin from *Laminaria hyperborea* (Bachelor's thesis, Høgskulen på Vestlandet).
4054. Luján Flores, M., Jiménez-Veuthey, M., León Córdoba, O. (2024). Metabolites from Microalgal Cultures as Potential Sources for the Pharmaceutical Industry. *Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures*, 139-168.
4055. Lykov, A., Salmin, A., Gevorgiz, R., Zheleznova, S., Rachkovskaya, L., Surovtseva, M. and Poveshchenko, O., 2023. Study of the Antimicrobial Potential of the *Arthrospira platensis*, *Planktothrix agardhii*, *Leptolyngbya cf. ectocarpi*, *Roholtiella mixta nov.*, *Tetraselmis viridis*, and *Nanofrustulum shiloi* against Gram-Positive, Gram-Negative Bacteria, and Mycobacteria. *Marine Drugs*, 21(9), p.492.
4056. Lykov, A., Salmin, A., Gevorgiz, R., Zheleznova, S., Rachkovskaya, L., Surovtseva, M. and Poveshchenko, O., 2023. Antimicrobial Potential of the Microalgae Extracts. *Ae*
4057. Naeem M, Imran M, Latif S, Ashraf A, Hussain N, Boczkaj G, Smulek W, Jesionowski T, Bilal M. Multifunctional catalyst-assisted sustainable reformation of lignocellulosic biomass into environmentally friendly biofuel and value-added chemicals. *Chemosphere*. 2023 Jul;330:138633. doi: 10.1016/j.chemosphere.2023.138633. Epub 2023 Apr 6. PMID: 37030343.
4058. Niu, Yinhong, et al. "Flavonoids from *Lycium barbarum* Leaves Exhibit Anti-Aging Effects through the Redox-Modulation." *Molecules* 27.15 (2022): 4952.
4059. Pezzolesi L., Samorì C., Zoffoli G., Xamin G., Simonazzi M., Pistocchi R. Semi-continuous production of polyhydroxybutyrate (PHB) in the Chlorophyta *Desmodesmus communis*. *Algal Research*, 74, art. no. 103196, 2023. DOI: 10.1016/j.algal.2023.103196
4060. Shevel'yuhina, A., Babich, O., Sukhikh, S., Ivanova, S., Kashirskih, E., Smirnov, V., Michaud, P. and Chupakhin, E., 2022. Antioxidant and Antimicrobial Activity of Microalgae of the Filinskaya Bay (Baltic Sea). *Plants*, 11(17), p.2264

4061. Songserm, R., Kaeboon, S., Suksungworn, R., Duangrisai, S., Sanevas, N. GC-MS profiling, anti-oxidant and anti-diabetic assessments of extracts from microalgae *Scenedesmus falcatus* (KU.B1) and *Chlorella sorokiniana* (KU.B2). // *Plant Science Today*, 2022, Vol. 9, Issue 3, p. 632-641, ISSN 23481900 (ISSN). doi:10.14719/pst.1560
4062. Yeshi, K., Ruscher, R., Miles, K., Crayn, D., Liddell, M., Wangchuk, P. Antioxidant and Anti-Inflammatory Activities of Endemic Plants of the Australian Wet Tropics. // *Plants*, 2022, Vol. 11, Issue 19 C7 - 2519, ISSN 22237747 (ISSN). doi:10.3390/plants11192519.
- 128). Kroumov, AD, Scheufele, FB, Zaharieva, MM, Zheleva-Dimitrova, D, NAJDENSKI, H. Optimal biomass production by cyanobacteria, mathematical evaluation and improvements in the light of bio-refinery concept. *Ecophysiology and Biochemistry of Cyanobacteria*, R. P. Rastogi (Ed.), Springer Nature, Singapore, 2022, ISBN:978-981-16-4873-1, DOI:10.1007/978-981-16-4873-1_18, 401-429**
4063. Karageorgou, Dimitra, et al. "Biomass and B-Glucosidase Production by the Cyanobacterium *Pseudanabaena* Sp. Under Heterotrophic Conditions." *Biomass* 2.4 (2022): 299-315.
- 129). Alov, P., Al Sharif, M., NAJDENSKI, H., Pencheva, T., Tsakovska, I., Zaharieva, M.M. and Pajeva, I., 2022. New Potential Pharmacological Targets of Plant-Derived Hydroxyanthraquinones from *Rubia* spp. *Molecules*, 27(10), p.3274**
4064. Cabrera Bermeo, A.E. and Vásquez Urgiles, V.E., 2023. Evolución del potencial antibacteriano y sinérgico de antraquinonas sintéticas sobre bacterias Gram-negativas resistentes a ciprofloxacina.
4065. Chen, X., Lan, W., Xie, J. (2023). Natural phenolic compounds: Antimicrobial properties, antimicrobial mechanisms, and potential utilization in the preservation of aquatic products. *Food Chemistry*, 138198.
- 130). Mantareva, V.; Kussovski, V.; Orozova, P.; Angelov, I.; Durmuş, M.; NAJDENSKI, H. Palladium Phthalocyanines Varying in Substituents Position for Photodynamic Inactivation of *Flavobacterium hydatis* as Sensitive and Resistant Species. *Curr. Issues Mol. Biol.* 2022, 44, 1950-1959. <https://doi.org/10.3390/cimb44050133>**
4066. Dube, E., Okuthe, G.E. (2024). Applications of Antimicrobial Photodynamic Therapy in Aquaculture: Effect on Fish Pathogenic Bacteria. *Fishes*, 9(3), 99.
4067. Heckman TI, Yazdi Z, Pomaranski EK, Sebastião FA, Mukkatira K, Vuglar BM, Cain KD, Loch TP, Soto E. Atypical flavobacteria recovered from diseased fish in the Western United States. *Front Cell Infect Microbiol.* 2023 Apr 19;13:1149032. doi: 10.3389/fcimb.2023.1149032. PMID: 37153143; PMCID: PMC10161732.
4068. Lee, J., Cha, I.T., Lee, K.E. et al. (2024). Complete genome sequence and potential pathogenic assessment of *Flavobacterium plurextorum* RSG-18 isolated from the gut of Schlegel's black rockfish, *Sebastes schlegelii*. *Environmental Microbiology Reports*.
4069. Zalevskaya O.A., Gur Y.A., Kutchin A.V. Palladium complexes as promising antimicrobial agents. (2023) *Russian Chemical Reviews*, 92 (9), art. no. RCR5093. DOI: 10.59761/RCR5093
- 131). Trochopoulos, A.G.X.; Ilieva, Y.; Kroumov, A.D.; Dimitrova, L.L.; Pencheva-El**

- Tibi, I.; Philipov, S.; Berger, M.R.; NAJDENSKI, H.M.; Yoncheva, K.; Konstantinov, S.M.; et al. Micellar Curcumin Substantially Increases the Antineoplastic Activity of the Alkylphosphocholine Erufosine against TWIST1 Positive Cutaneous T Cell Lymphoma Cell Lines. *Pharmaceutics* 2022, 14, 2688. <https://doi.org/10.3390/pharmaceutics14122688>**
4070. Tsai, H.-H.; Yu, J.-C.; Hsu, H.-M.; Chu, C.-H.; Chang, T.-M.; Hong, Z.-J.; Feng, A.-C.; Fu, C.-Y.; Hsu, K.-F.; Dai, M.-S.; et al. The Risk of Breast Cancer between Western and Mediterranean Dietary Patterns. *Nutrients* 2023, 15, 2057.
- 132). Kroumov, A.D., Scheufele, F.B., Zaharieva, M.M., Gevrenova, R. and NAJDENSKI, H., 2022. Cyanobacteria as a competing source of bioenergy: metabolic engineering and modeling approach for medium optimization. In *Ecophysiology and Biochemistry of Cyanobacteria* (pp. 455-478). Singapore: Springer Nature Singapore**
4071. Kamshybayeva, G.K., Kossalbayev, B.D., Sadvakasova, A.K., Kakimova, A.B., Bauenova, M.O., Zayadan, B.K., Lan, C.W., Alwasel, S., Tomo, T., Chang, J.S. and Allakhverdiev, S.I., 2023. Genetic engineering contribution to developing cyanobacteria-based hydrogen energy to reduce carbon emissions and establish a hydrogen economy. *International Journal of Hydrogen Energy*.
- 133). Angelovska, M., Zaharieva, M.M., Dimitrova, L.L., Dimova, T., Gotova, I., Urshev, Z., Ilieva, Y., Kaleva, M.D., Kim, T.C., Naydenska, S., Dimitrov, Z., and H. NAJDENSKI. 2023, Prevalence, Genetic Homogeneity, and Antibiotic Resistance of Pathogenic *Yersinia enterocolitica* Strains Isolated from Slaughtered Pigs in Bulgaria. *Antibiotics*, 12(4), p.716**
4072. Al-Rawe, A.M., Al-Jomaily, O K.G., Yousif, Y.I. et al. (2023). Comparative Genomics, Phylogenetic and Functional Analysis of *Yersinia enterocolitica*, a Gastrointestinal Pathogen, with Other Soil-Borne Bacteria Causing Diseases. *Mikrobiologichnyi Zhurnal*, 85(5), 31-41.
4073. Mabekoje, O.O., Jibril, F.L., Baba, J. and Isah, R.M., 2023. Biotyping and Serological Characterization of *Yersinia enterocolitica* Isolates In Human and Pigs in Selected Farms and Hospital in Shango Community, Minna, Niger State, Nigeria. *Journal of Applied Sciences and Environmental Management*, 27(6), pp.1319-1330.
4074. Vidaković Knežević, S., Knežević, S., Vranešević, J. et al. (2024). Using essential oils to reduce *Yersinia enterocolitica* in minced meat and in biofilms. *Foods*, 13(5), 806.
4075. Труба, О.О., 2023. Йерсиніозна інфекція котятих (епізоотологія, клініка, діагностика, лікування та профілактика).
- 134). Ilieva Y, Marinov T, Trayanov I, Kaleva M, Zaharieva MM, Yocheva L, Kokanova-Nedialkova Z, NAJDENSKI H, Nedialkov P. Outstanding Antibacterial Activity of *Hypericum rochelii* - Comparison of the Antimicrobial Effects of Extracts and Fractions from Four *Hypericum* Species Growing in Bulgaria with a Focus on Prenylated Phloroglucinols. *Life* (Basel). 2023 Jan 18;13(2):274. doi: 10.3390/life13020274. PMID: 36836632; PMCID: PMC9959064**
4076. Bejenaru, C., Radu, A., Mogoşanu, G.D. et al. Hypericaceae Juss. Family. In *Natural Products and Medicinal Properties of Carpathian (Romanian) Plants* (pp. 240-248). CRC Press.
4077. Doğan, H., Fidan, H., Baş, H. et al. (2024). Determination of essential oil and chemical composition of St. John's Wort. *Open Chemistry*, 22(1), 20240001.

4078. Gradinaru, L.M.; Barbalata-Mandru, M.; Enache, A.A.; Rimbu, C.M.; Badea, G.I.; Aflori, M. "Chitosan Membranes Containing Plant Extracts: Preparation, Characterization and Antimicrobial Properties." *International Journal of Molecular Sciences*, vol. 24, no. 10, May 2023, p. 8673.
4079. Kakouri E., Daferera D., Trigas P., Charalambous D., Pantelidou M., Tarantilis P.A., Kanakis C.D. Comparative Study of the Antibacterial Activity, Total Phenolic and Total Flavonoid Content of Nine *Hypericum* Species Grown in Greece. *Applied Sciences (Switzerland)*, 13 (5), art. no. 3305. 2023. DOI: 10.3390/app13053305
4080. Machado, A., Zamora-Mendoza, L., Alexis, F. et al. (2023). Use of plant extracts, Bee-Derived products, and probiotic-related applications to fight Multidrug-resistant pathogens in the post-antibiotic era. *Future Pharmacology*, 3(3), 535-567.
4081. Manoharan M., Ragothaman P., Selvin J., Balasubramanian T.S. Initiation of Apoptotic Pathway by the Cell-Free Supernatant Synthesized from *Weissella cibaria* Through In-Silico and In-Vitro Methods. *Applied Biochemistry and Biotechnology*. 2023. DOI: 10.1007/s12010-023-04688-3
4082. Pinho, J.O., Ferreira, M., Coelho, M. et al. (2024). Liposomal Rifabutin—A Promising Antibiotic Repurposing Strategy against Methicillin-Resistant *Staphylococcus aureus* Infections. *Pharmaceuticals*, 17(4), 470.
4083. Saleh, B., Khawajkiah, M., Almariri, A. (2023). Antibacterial activity of *Hypericum* (Hypericaceae) species against Gram-positive bacterial isolates. *Journal of Agroalimentary Processes & Technologies*, 29(4).
4084. Soyuçok, A., Kart, A., Yalçın, H. et al. (2024). Determination of in vitro antimicrobial and antibiofilm activity of *Hypericum crenulatum* against some food pathogens and its phenolic content. *Acta Alimentaria*, 53(1), 12-22.

Общ брой цитати – 4084

От тях:

- 296 дисертации
- 33 дипломни работи

За конкурса прилагам 3775 цитата (без автоцитати), от които 1976 са от последните 5 години.