

1.3. ЦИТИРАНИЯ НА НАУЧНИ ТРУДОВЕ (БЕЗ АВТОЦИТАТИ) В НАУЧНИ ПУБЛИКАЦИИ И В ПАТЕНТИ ЗА ИЗОБРЕТЕНИЯ У НАС И В ЧУЖБИНА

на чл.-кор. д.н. Иван Александров Илиев,
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Забележка: Представен е доказателствен материал на хартиен носител и копия от цитиранията на флаш памет.

Общ брой на цитиранията от други автори: 591

- от български автори: 171
- от чуждестранни автори: 420

Разпределение на цитиранията след получаване на академична длъжност „член-кореспондент“: общо – 189.

В реферирани списания (Web of Science and SCOPUS): 102 (84, 85, 86, 87, 88, 89, 90, 97, 127, 131, 133, 135, 138, 151, 187, 190, 204, 205, 206, 222, 246, 247, 248, 250, 251, 253, 254, 256, 257, 269, 270, 286, 289, 297, 303, 315, 316, 317, 322, 324, 326, 329, 340, 341, 345, 346, 347, 348, 350, 351, 352, 354, 355, 357, 358, 363, 365, 366, 367, 368, 398, 399, 402, 403, 406, 407, 408, 410, 412, 414, 415, 417, 423, 424, 461, 470, 473, 475, 477, 478, 480, 481, 482, 484, 485, 486, 489, 490, 491, 493, 496, 497, 498, 501, 576, 577, 583, 584, 585, 586, 587, 591).

В нереферирани списания: 56 (82, 124, 125, 126, 129, 130, 132, 134, 136, 140, 188, 191, 192, 209, 221, 249, 252, 255, 287, 298, 318, 321, 323, 325, 336, 337, 339, 353, 356, 359, 360, 361, 362, 364, 400, 404, 405, 409, 413, 416, 425, 462, 466, 468, 469, 476, 483, 487, 494, 499, 500, 554, 572, 574, 575, 581).

В глави от книги и монографии: 13 (207, 271, 285, 288, 320, 327, 465, 471, 472, 474, 492, 495, 573, 590).

В сборници от научни конференции: 9 (139, 319, 401, 426, 463, 479, 582, 588, 589).

В дисертации за получаване на научни степени: 9 (83, 137, 189, 203, 284, 397, 464, 467, 488).

Националност на авторите цитирали публикации на кандидата (68 държави):

Австрия (90, 206, 346, 452, 565), **Австралия** (90, 251, 352), **Албания** (442), **Алжир** (359, 500), **Аржентина** (326, 590), **Беларус** (123, 160, 302, 422), **Белгия** (138), **Босна и Херцеговина** (207), **Бразилия** (99, 104, 117, 190, 250, 404, 455, 456, 469, 491), **България** (1, 2, 4, 5, 6, 7, 8, 10, 11, 13, 14, 19, 22, 23, 25, 27, 30, 31, 32, 34, 36, 37, 44, 45, 51, 52, 58, 64, 65, 66, 69, 71, 74, 75, 76, 77, 90, 95, 96, 110, 116, 119, 120, 141, 144, 146, 148, 149, 153, 154, 158, 162, 163, 165, 167, 173, 180, 181, 182, 193, 208, 219, 233, 236, 237, 238, 239, 258, 259, 265, 268, 272, 276, 281, 285, 290, 292, 293, 301, 304, 307, 309, 312, 313, 327, 330, 331, 334, 335, 338, 339, 340, 372, 389, 419, 421, 431, 432, 433, 435, 436, 438, 439, 440, 442, 445, 447, 448, 449, 471, 478, 504, 505, 506, 507, 508, 513, 517, 518, 523, 524, 525, 527, 528, 529, 530, 534, 535, 536, 540, 542, 543, 544, 545, 546, 547, 548, 549, 550, 551, 552, 555, 556, 557, 560, 561, 562, 566, 567, 568, 569, 570, 571), **Венецуела** (108), **Виетнам** (85, 131, 354, 357, 365, 501), **Германия** (56, 200, 206, 274, 275, 289, 416, 472), **Грузия** (161, 317), **Гърция** (53, 68, 89, 111, 112, 263, 273, 279, 283, 284, 287, 367, 451, 493), **Египет** (24, 176, 178, 186, 223, 226, 231, 232, 235, 243, 246, 379, 384, 394, 583, 584, 585, 586, 587), **Еквадор** (78), **Естония** (565), **Иемен** (246), **Израел** (59, 143, 170, 355), **Индия** (43, 60, 109, 126, 132, 145, 150, 157, 170, 183, 187, 214, 215, 240, 291, 311, 320, 324, 325, 348, 376, 377, 385, 388, 392, 396, 410, 411, 412, 413, 465, 482, 487, 492, 498, 573, 578, 581), **Индонезия** (139, 365, 366, 464, 473, 475, 479, 483, 494), **Ирак** (192, 255, 318, 425, 499), **Иран** (46, 103, 159, 211, 249, 328, 398, 450, 459, 466, 468, 481), **Ирландия** (245),

Испания (12, 100, 397, 398, 476, 478), **Италия** (205, 270, 289, 317, 355, 363, 378, 403, 416, 418, 576, 577, 591), **Камерун** (495), **Канада** (63, 156, 212, 289, 409, 485), **Кения** (138), **Китай** (26, 42, 59, 61, 92, 185, 188, 204, 209, 252, 254, 269, 300, 303, 310, 353, 356, 360, 368, 373, 400, 402, 407, 408, 415, 441), **Корея** (194, 220, 229, 344, 358, 423, 490, 581), **Латвия** (72, 83, 151), **Либия** (223, 226), **Литва** (203), **Малави** (444), **Малайзия** (73, 118, 351, 385, 406, 467, 480, 484), **Мароко** (364), **Мексико** (33, 70, 81, 101, 102, 104, 105, 106, 107, 113, 115, 134, 323, 350, 382, 453, 457, 461), **Нигерия** (187), **Нидерландия** (98, 140), **Нова Зеландия** (174, 185, 371), **Обединено кралство** (206, 242, 247, 251, 346, 393), **Полша** (248, 256, 277, 286, 314, 329, 332, 345, 347, 348, 446, 458, 497, 498, 516), **Португалия** (142, 164, 412, 470, 477), **Румъния** (29, 48, 49, 50, 115, 166, 206, 294, 295, 296, 297, 335, 341, 361, 362, 420, 427, 428, 429, 430, 496, 519, 520, 521, 522, 537, 538, 539, 541), **Русия** (3, 9, 20, 21, 39, 40, 57, 82, 86, 93, 124, 129, 136, 155, 221, 241, 280, 315, 316, 319, 333, 336, 342, 374, 395, 399, 401, 417, 509, 510, 511, 512, 526, 533, 572, 574, 575), **Саудитска арабия** (235, 246, 257, 412, 583, 584, 585, 586, 587), **САЩ** (47, 62, 91, 115, 121, 122, 133, 195, 211, 224, 225, 227, 230, 244, 260, 262, 264, 269, 271, 343, 369, 380, 383, 393, 405, 454, 583, 584, 585, 586, 587), **Словакия** (28, 169, 559, 579, 580), **Словения** (208), **Сърбия** (29, 177, 184, 261, 266, 278, 282, 349, 474, 553, 554), **Тайван** (147, 179, 234, 386), **Тайланд** (486), **Танзания** (463), **Турция** (67, 84, 87, 88, 114, 127, 135, 137, 140, 171, 175, 189, 191, 253, 266, 270, 288, 299, 337, 424, 426, 462, 488, 589), **Уганда** (490), **Украйна** (130, 202, 322, 381, 387), **Унгария** (15, 16, 17, 18, 38, 41, 54, 194, 206, 210, 298, 434, 437, 531, 532, 558, 563), **Финландия** (94), **Франция** (128, 206, 470), **Хърватия** (172, 261, 266, 267, 460, 502, 503, 514, 515, 564), **Чехия** (97, 185, 205, 213, 216, 217, 218, 222, 228, 305, 306, 308, 588), **Швейцария** (275, 285, 289), **Швеция** (152, 385, 393, 565), **Шри Ланка** (582), **Южно-Африканска Република** (108, 226, 246, 369, 391), **Япония** (26, 35, 55, 188, 196, 197, 198, 199, 201, 321, 370, 375, 390, 400, 407, 414).

Публикация: Илиев Ив. (1988). Проучване на естествените популации от обикновена бреза (*Betula pendula* Roth.) в западна България и отбор на ценни декоративни форми. Дисертационен труд за придобиване на научна степен „кссн“, ВЛТИ, София, 181 стр.

Ц и т и р а н а в:

1. Юруков Ст. (2003). Дендрология, СД “Лотус”, 212 стр.
2. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр.
3. Ветчинникова Л. В. (2005). Карельская береза и другие редкие представители рода *Betula* L. Рецензенты: Марковская Е. Ф., Новицкая Л. Л. Российская академия наук, Карельский научны центр, Института леса, Москва, Издательство Наука, 264 стр.
4. Alexandrov A., Dobrev R. (2011). Country Report for The State of World's Forest Genetic Resources – Bulgaria, 63 pp.
5. Димитров М. (2012). 25G1 Гори от бяла бреза (*Betula pendula*). В: Бисерков В. и др. (ред.) 2011. Червена книга на Република България. Том 3. Природни местообитания. ИБЕИ – БАН & МОСВ, София: 330-331
6. Zhelev P., Angelov V. (2012). Variation in a silver birch locality near Ardino (eastern rhodopes). Forestry Ideas, vol. 18, No 2 (44): 125-131 ISSN: 1314-3905 (Print), ISSN: 26-03-2996 (Online). https://forestry-ideas.info/issues/issues_Index.php?journalFilter=45
7. Alexandrov A., Dobrev R. (2012). State of forest genetic resources in Bulgaria. Forest Science, No 1/2: 2-22.
8. Янгъзов Б. (2013). Еколого-лесовъдска оценка на залесяванията в района на Кържали и насоки за стопанисване. Дисертация за присъждане на образователна и научна степен “доктор”. ИГ при БАН, София, 124 стр.

9. Ветчинникова Л. В., Титов А. Ф., Кузнецова Т. Ю. (2013). Карельская береза: биологические особенности, динамика ресурсов и воспроизводство. Петрозаводск, 310 стр. ISBN: 978-5-9274-0608-1.
10. Alexandrov A. H., Dobrev R. D. (2014). The State of Forest Genetic Resources in Bulgaria. Sofia, 84 pp. ISBN: 978-3-639-66981-7
11. Юруков Ст., Панайотов М. (2015). Дендрология, РУТА - ХБ", 212 стр. ISBN: 978-954-9557-21-3.

Публикация: Iliev I., Chavdarov Iv. (1990). *In vitro* propagation of silver birch (*Betula pendula* Roth.) through induced adventitious buds from leaf cuttings. News, IPPS, autumn.

Цитирана в:

12. Vieitez F. J. (1995). Somatic embryogenesis in chesnut. In: Jain S. M., Gupta P. K., Newton R. J. (Eds.). Somatic embryogenesis in woody plants, vol. 2 – Angiosperms, Kluwer Acad. Publ.: 375-407.

Публикация: Iliev I., Chavdarov Iv., Alexandrov P. (1993). *In vitro* propagation of *Albizzia julibrissin* Dur. through hipocotyl, cotyledon and epicotyl segments. -Annual vol. of International Plant Propagators' Society, June: 23-29.

Цитирана в:

13. Калмуков К. (2000). Проучвания върху *Albizzia julibrissin* Dur. В района на Свищов. II. Производство на посадъчен материал. В: Propagation of Ornamental Plants, Fourth International Scientific Conference: 223-228. ISBN: 954-8783-34-7.

Публикация: Iliev I., Alexandrov P., Barashka E. (1993). The possibilities for vegetative propagation of *Thujaopsis dolabrata* Sieb. et Zucc. 'Variegata'. Folia Dendrologica, vol. 20: 155-163.

Цитирана в:

14. Милев М., Илиев Н., Глушкова М. (2000). Проучване върху автовететативното размножаване на сребристия смърч (*Picea pungens* Engelm.). В: Propagation of Ornamental Plants, Fourth International Scientific Conference: 248-254. ISBN: 954-8783-34-7.

Публикация: Iliev I., Zhelev P. (1994). Morphological and biochemical studies of the *in vitro* rhizogenesis of regenerants of *Betula pendula* Roth. A Kerteszeti es Elelmiszeripari Egyetem, Kozlemenyei, Publicationes Universitatis Horticulturae Industriaeque Alimentariae, Budapest, vol. LIV: 84-90.

Цитирана в:

15. Schmidt G., Marta K., Pal J. (1995). Effect of age of micropropagated and cutting-propagated stockplants on the rooting of 3 *Sorbus* hybrids. Horticultural Science, 27 (1-2): 77-79
16. Schmidt G. (1996). Propagation of some hard-to-root cultivars of Cupressaceae family with juvenile microcuttings. В: Размножаване на декоративни растения, 5-7.10.1996: 135-142. ISBN: 954-8783-10-X (Т.2).
17. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences).

18. Ördögh M., Jámbo-Benczúr E., Tilly-Mándy A., Lelik L. (2009). Effects of different cytokinins on proliferation of *Sorbus borbasii* 'Herkulesfüdrö'. Propagation of Ornamental Plants, 9, (1): 43-46. ISSN: 1311-9109.
19. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

Публикация: Iliev I. (1996). *In vitro* propagation of *Betula pendula* Roth 'jougii'. In: "IPPS in Bulgaria - Propagation of Ornamental Plants", International Conference, 5-7 October, Sofia: 44-54. ISBN: ISBN: 954-8783-10-X (T. 2).

Цитирания:

20. Ветчинникова Л. В. (2002). Морфо-физиологические и биохимические особенности различных видов и разновидностей березы семенного и вегетативного происхождения в условиях Восточной Финляндии. Диссертация для получения ученой степени доктор биологических наук, Петрозаводск, 366 стр. (код специальности ВАК: 03.00.12 Физиология и биохимия растений).
21. Ветчинникова Л. В. (2005). Карельская береза и другие редкие представители рода *Betula* L. Рецензенты: Марковская Е. Ф., Новицкая Л. Л. Российская академия наук, Карельский научный центр, Института леса, Москва, Издательство Наука, 264 стр. ISBN: 5-02-033684-X.
22. Илиев Н., Петракиева А. (2007). Хетеровегетативно размножаване на обикновената череша (*Prunus avium* L.). Наука за гората, 4: 3-15.
23. Петракиева А. (2008). Възможности за развъждане на обикновената череша (*Prunus avium* L.) в България. Диссертация, ЛТУ, София, 161 стр.
24. Omar A. F. Dewir Y. H., El-Mahrouk M. E. (2014). Molecular identification of phytoplasmas in fasciated cacti and succulent species and associated hormonal perturbation. Journal of Plant Interactions, 9 (1): 632-639. DOI: 10.1080/17429145.2014.882421, <http://dx.doi.org/10.1080/17429145.2014.882421>
25. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Диссертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
26. Hanzhi Liang, Yuping Xiong, Beiyi Guo, Haifeng Yan, Shuguang Jian, Hai Ren, Xinhua Zhang, Yuan Li, Songjun Zeng, Kunlin Wu, Feng Zheng, Jaime A. Teixeira da Silva, Youhua Xiong, Guohua Ma (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. Environmental and Experimental Biology, 17: 169-177 DOI: 10.22364/eeb.17.17

Публикация: Iliev I., Trifonov T. (1996). Influence of donor plant's age upon the *in vitro* cloning of *Sequoia sempervirens* Endl. Nauka za gorata, No 2: 3-11.

Цитирания:

27. Илиев Н. (1996). Възможности за повишаване използването на гигантската секвоя (*Sequoiadendron giganteum* (Lindl.) Buchh.) чрез семенно и вегетативно размножаване. Диссертация, ЛТУ, София: 123 стр.
28. Kamenicka A., Lanakova M. (2000). Effect of medium composition and type of vessel closure on axillary shoot production of magnolia *in vitro*. Acta Physiologiae Plantarum, 22 (2): 129-134.
29. Halmagyi A., Kevresan S., Kovacevic B., Orlovic S., Miladinovic D., Cirin-Novto V., Kuhajda K. (2010). Effects of naphthalenic acids on rooting *in vitro* grown *Sequoia sempervirens*. Acta Horticulturae, 885: 139-143. ISBN: 90 6605 210 4, ISSN: 0567-7572.

30. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
31. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Iliev I., Iliev N. (1996). Influence of donor plant's age on the *in vitro* cloning of gaint sequoia (*Sequoiadendron giganteum* (Lindl.) Buchh. In: Schmidt G., Batiz E. (Eds). “Lippay Janos” Scientific Symposium, 17-18 October, Budapest: 22-28.

Цитирана в:

32. Илиев Н. (2000). Производство на посадъчни материали от гигантска секвоя (*Sequoiadendron giganteum* (Lindl.) Buchh.). B: Propagation of Ornamental Plants. Fourth International Scientific Conference: 203-211. ISBN: 954-8783-34-7.
33. González-Perez E., Juárez-Muñoz J., Ayala-Garay O. J., De Jesús Yáñez-Morales. (2014). *Ex vitro* acclimatization of Gladiolus plantlets. Propagation of Ornamental Plants, 14 (3): 125-132. ISSN: 1311-9109.
34. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Iliev I., Iliev N. (1997). Az anyanövény korának hatása a mammutfenyő (*Sequoiadendron giganteum* (Lindl.) Buchh) *in vitro* klonozására. Uj Kertgazdasag, 3 (3): 26-31 (на Унгарски).

Цитирана в:

35. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences)
36. Dancheva D. (2014). Rooting and acclimatization of micropropagated shoots of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, vol. 30 (1): 75-80.
37. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.

Публикация: Iliev I., Besendorfer V., Peskan T. (1998). *In vitro* propagation of *Betula pendula* ‘Dalecarlica’ In: Tsekos I., Moustakas M. (Eds). Progress in Botanical Research, Kluwer Academic Publishers: 513-516. eBook ISBN: 978-94-011-5274-7. ISBN: 978-0-7923-5305-8. doi: 10.1007/978-94-011-5274-7_117 . DOI: <https://doi.org/10.1007/978-94-011-5274-7>

Цитирана в:

38. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences)
39. Ветчинникова Л. В. (2002). Морфо-физиологические и биохимические особенности различных видов и разновидностей березы семенного и вегетативного происхождения в условиях Восточной Фенноскандии. Диссертация для получения

ученая степень доктор биологических наук, Петрозаводск, 366 стр. (код специальности ВАК: 03.00.12 Физиология и биохимия растений).

40. Ветчинникова Л. В. (2005). Карельская береза и другие редкие представители рода *Betula* L. Рецензенты: Марковская Е. Ф., Новицкая Л. Л. Российская академия наук, Карельский научный центр, Института леса, Москва, Издательство Наука, 264 стр.
41. Ördögh M., Jámbor-Benczúr E., Tilly-Mándy A., Lelik L. (2009). Effects of different cytokinins on proliferation of *Sorbus borbasii* 'Herkulesfüdrö'. Propagation of Ornamental Plants, (9), 1: 43-46. ISSN: 1311-9109.
42. Chen G. D., Yang J., Wang L. Q., Liu H., Shen S., Qin F. (2010). *In vitro* adventitious shoot formation from petiole explants of *Swainsonia salsula* Taubert. Propagation of Ornamental Plants, 10 (3): 122-128. ISSN: 1311-9109.
43. Zaki M. (2011). *In vitro* studies on regeneration of some rare trees of Kashmir Himalayas. PhD Thesis. Department of Botany, University of Kashmir, Hazratbal, Srinagar, India, 190 pp.
44. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-964-332-127-8.
45. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
46. Payamnoor Vahide, Khodadadi Negar, Jafari Hajati Razieh (2021). Bio nanoparticles as elicitors increase accumulation of betulin and betulinic acid in callus cultures. South African Journal of Botany, 141: 431-439. ISSN: 0254-6299.
https://www.sciencedirect.com/science/article/pii/S0254629921001769?casa_token=yk_3Kf0PUhwAAAAA:3HwJz4MCchGjjroWvIM8JYWEc6gESkvIsBGUtMJp10rpWbG1DNd1iJBTiCuma76SnIarXgfBq6Et

Публикация: Iliev I., Tzvetkov Iv., Denkova St., Chavdarov Iv. (1998). Micropropagation of decorative plants in Bulgaria. Proceedings of International Plant Propagators Society, vol. 48: 188-191. ISSN: 0538-9143. <https://www.pubhort.org/ipps/48/39.htm>

Цитирана в:

47. Goodhart F. W. (2000). Propagation aspects of *Sciadopitys verticillata* and *Sequoiadendron giganteum* 'Hazel Smith'. Combined proceedings of International Plant Propagators' Society, vol. 50: 222-227.

Публикация № 15: Scaltsoyiannes A., Iliev I., Tsaksira M., Pasayiannes G., Tsoulpha P., Moulalis D. (1998). Rejuvenation and micropropagation of wild cherry tree (*Prunus avium* L.). In: Plant Breeding and Genetics for Friendly Agriculture to the Environment, Proceedings of 7th PanHellenik Congress, Eraklion, Creta: 272-280 (на Гръцки).

Цитирана в:

48. Corneanu M., Mindrila G., Corneanu G. C., Cristea C., Atyim P., Minea R. (2004). The influence of humic acids and magnetic fluids on shoot elongation and *in vitro* rooting processes in *Prunus avium* L. Buletinul USAMV, Cluj-Napoca, Seria Zootehnie si Biotehnologii, vol. 60: 313-318. ISSN-1454-2382.
49. Bălan V., Tudor V., Topor E., Corneanu M. (2007). Apricot genetics and biotechnology in Romania. The European Journal of Plant Science and Biotechnology, 1 (2): 152-169.
50. Corneanu M., Bălan V. (2008). Botehnologia la cais. In: Popa C., Dumitru A. (Eds). Caisul și caisele, Editura Ceres, București: 189-226. ISBN: 978-973-40-0797-4.
51. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISSN: 978-954-332-127-8.

52. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Iliev I., Kitin P., Funada R. (2001). Morphological and anatomical study of *in vitro* root formation of Silver birch (*Betula pendula* Roth.). Propagation of Ornamental Plants, vol. 1: 10-19. ISSN: 1311-9109. https://www.journal-pop.org/2001_1_1_10-19.html

Цитирани в:

53. Hatzilazarou S., Economou A., Rifaki N. (2002). Changes in sugar content of *Gardenia jasminoides* Ellis. Microcuttings in relation to *in vitro* and *ex vitro* rooting. Propagation of Ornamental Plants, vol. 2, No 2: 42-46. ISSN: 1311-9109.
54. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences).
55. Muruyama E. (2003). *In vitro* propagation of Jacaranda (*Jacaranda mimosaeifolia* D. Don.). Propagation of Ornamental Plants, 3(1): 27-33. ISSN: 1311-9109.
56. Pinker I., Wagner T. (2004). Stem development of *Amelanhier lamarckii* (F.G. Shroeder) *in vitro* and its importance for *in vitro* rooting. Propagation of Ornamental Plants, 4 (1): 53-57. ISSN: 1311-9109.
57. Ветчинникова Л. В. (2005). Карельская береза и другие редкие представители рода *Betula* L. Рецензенты: Марковская Е. Ф., Новицкая Л. Л. Российская академия наук, Карельский научны центр, Института леса, Москва, Издательство Наука, 264 стр. ISBN: 5-02-033684-X.
58. Данчева Д. (2005). Автовегетативно размножаване на планинския ясен (*Fraxinus excelsior* L.). Наука да гората, 2: 17-30.
59. Ziv M., Chen J. (2008). The anatomy and morphology of tissue cultured plants. In: George E. F., Hall M., De Klerk G-J. (Eds.). Plant propagation by tissue culture, 3rd edition, Springer, Dordrecht, NL: 465-477.
60. Suthar R. K., Purohit S. D. (2010). Anatomy of root development in tissue culture raised *Terminalia bellerica* Roxb. and its significance in hardening. Acta Horticulturae, 865: 123-127. ISBN: 90 6605 210 4, ISSN: 0567-7572.
61. Chen G. D., Yang J., Wang L. Q., Liu H., Shen S., Qin F. (2010). *In vitro* adventitious shoot formation from petiole explants of *Swainsonia salsula* Taubert. Propagation of Ornamental Plants, 10 (3): 122-128. ISSN: 1311-9109.
62. Hartmann H. T., Kester D. E., Davies F. E., Geneve R. (2010). Propagation of Ornamental Trees, Shrubs, and Woody Vines. In: Hatrman & Kester's Plant Propagation: Principles and Practices. Prentice Hall, 928 pp.
63. Oinam G., Yeung E., Kurepin L., Haslam T., Lopez-Villalobos A. (2011). Adventitious root formation of ornamental plants: I. General overview and recent successes. Propagation of Ornamental Plants, 11 (2): 78-90. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_11_2\(78-90\).pdf](https://www.journal-pop.org/References/Vol_11_2(78-90).pdf)
64. Zhelev P., Angelov V. (2012). Variation in a silver birch locality near Ardino (eastern rhodopes). Forestry Ideas, Forestry Ideas, vol. 18, No 2 (44): 125-131. ISSN: 1314-3905 (Print), ISSN: 26-03-2996 (Online). https://forestry-ideas.info/issues/issues_Index.php?journalFilter=45

65. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.
66. Тихомирова Л. И. (2012). Анатомия границы корня и побега у ириса в культуре тканей *in vitro*. Вестник Удмуртского Университета, 4: 154-157.
67. Acemi A., Özen F., Kiran R. (2013). *In vitro* propagation of *Amsonia orientalis* Dence. from nodal segments of adult plants. Propagation of Ornamental Plants, 13 (1): 25-32. ISSN: 1311-9109.
68. Economou A. S. (2013). From microcutting rooting to microplant establishment: key points to consider for maximum success in woody plants. Acta Horticulturae, 988: 43-56. ISBN: 90 6605 210 4, ISSN: 0567-7572.
69. Тихомирова Л. И. (2013). Изучение влияния БАП в средах размножения на показатели этапа укоренения у *Iris ensata* Thunb. Биологические науки, 106-110. DOI: 10.14258/izvasu(2013)3.2-23.
70. González-Perez E., Juárez-Muñoz J., Ayala-Garay O. J., De Jesús Yáñez-Morales. (2014). *Ex vitro* acclimatization of Gladiolus plantlets. Propagation of Ornamental Plants, 14 (3): 125-132. ISSN: 1311-9109.
71. Dancheva D. (2014). Rooting and acclimatization of micropropagated shoots of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, vol. 30 (1): 75-80. ISSN: 1454-6914.
72. Dokane K., Megre D., Lazdane M., Uldis Kondratovics U. (2014). Does shoot anatomical heterogeneity influence *ex vitro* adventitious root formation in rhododendron microcuttings? Propagation of Ornamental Plants, 14 (4): 171-176. ISSN: 1311-9109.
73. Eng W. H., Aziz M. A., Sinniah U. R. (2015). *In vitro* regeneration of *Citrus hystrix* DC. Brazilian Journal of Botany, 38: 235-242. DOI 10.1007/s40415-014-0127-5
74. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
75. Parvanova P., Lyubomirova T., Tzvetkova N., Ilinkin V. (2015). Effect of auxin on protein content during *in vitro* phizogenesis of *Syringa vulgaris* L. Annuaire de l'Université de Sofia “St. Kliment Ohridski”, Faculte de Biologie, vol. 4: 208-215.
76. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.
77. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
78. Humberto Estrella-Maldonado, Gabriela Fuentes Ortíz, Arianna C. Chan León, Luis C. Rodríguez Zapata, Carlos Talavera May, Francisco Espadas y Gil, Felipe Barredo Pool, Fabio Marcelo Idrovo Espín, Jorge M. Santamaría (2016). The papaya CpAUX1/LAX and CpPIN genes: structure, phylogeny and expression analysis related to root formation on *in vitro* plantlets. Plant Cell, Tissue and Organ Culture, 126 (2): 187-204. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. DOI: 10.1007/s11240-016-0989-2
79. West T. P., Coronado R. F. (2016). Micropropagation of *Ostrya virginiana* (Mill.) K. Kock. An underutilized ornamental landscape tree. Propagation of Ornamental Plants, 16 (2): 56-61. ISSN: 1311-9109.
80. Dokane K., Megre D., Lazdane M., Kondratovics U. (2017). Influence of the *Calcarisporium arbuscula* on *in vitro* growth and *ex vitro* adventitious rooting of two elepidote rhododendron cultivars. Propagation of Ornamental Plants, 17 (2): 39-47. ISSN: 1311-9109.

81. Estrella Maldonado H. J. (2017). Caracterización molecular de genes que modulan la ytranscripción y el transporte de auxinas y su papel en la rizogénesis en vitro plants de *Carica papaya* L. cv. Maradol. Tesis que presenta. En opción al; titulu de Doctor en ciencias. Centro de Investigación Científica de Yucatán, A. C. Postgrado en Ciencias Biológicas, Merida, Yucatán, Mexico, 117 pp.
82. Машкина О. С., Табацкая Т. М., Внукова Н. И. (2019). Технология долгосрочного хранения в культуре *in vitro* ценных генотипов березы и выращивание на ее основе посадочного материала. Биотехнология, Т. 35, № 3: 57-67. ISSN 0234-2758. <http://csli.isc.irk.ru/bd/%D0%96%D1%83%D1%80%D0%BD%D0%B0%D0%BB%D1%8B/%D0%91%D0%B8%D0%BE%D1%82%D0%B5%D1%85%D0%BD%D0%BE%D0%BB%D0%BE%D0%B3%D0%B8%D1%8F%202019%20%D0%A235/%E2%84%96%203/57-67.pdf>
83. Dokane K. (2019). Rododendru un to sakņu endofītisko sēņu mijiedarbības anatomiski fizioloģiskie aspekti un izmantošana rododendru pavairošanā. PhD thesis. Latvijas Universitātes Bioloģijas fakultātē Augu fizioloģijas, 114 pp.
84. Ozdemir S., Ozdemir M., Yetilmezsoy K. (2020). Effects of acclimatization soil prepared from hazelnut husk and gyttja on *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 2: 63-71. ISSN: 1311-9109.
85. Vu Quoc Luan, Le Kim Cuong, Hoang Thanh Tung, Vu Thi Hien, Do Manh Cuong, Phan Le Ha Nguyen, Bui Van The Vinh, Duong Tan Nhut (2020). Highly effective micropropagation and induction of early flowering by gibberellic acid in *Paphiopedilum delenatii* Guillaumin. Propagation of Ornamental Plants, 20 (1): 3-11. ISSN: 1311-9109.
86. Tikhomirova L. (2021). Features of morphogenesis of *Iris ensata* Thunb. *in vitro* culture. Russian Journal of Plant Physiology, 68: 669-677. ISSN: 1608-3407 (Online). <https://doi.org/10.1134/S1021443721040191>
87. İsfendiyaroğlu M. (2021). Anatomy of adventitious root development in mastic tree (*Pistacia lentiscus* var. *chia* Duham.) cuttings. Propagation of Ornamental Plants, 21 (2): 58-64. ISSN: 1311-9109.
88. Turp G. A., Turp S. M., Ozdemir S. (2021). The role of pyroligneous acid for *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 21 (4): 116-122. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_21_4\(116-122\).pdf](https://www.journal-pop.org/References/Vol_21_4(116-122).pdf)

Публикация: Iliev I., Dakov A. (2002). Country report: Bulgaria. In: EUROFORGEN. Mediterranean Oaks Network, Gozo, Malta: 28-31. ISBN: 92-9043-592-5. https://www.euforgen.org/fileadmin/templates/euforgen.org/upload/Publications/Network_Reports/903_Mediterranean_Oaks_Network_Report_Second_Meeting.pdf

Цитирана в:

89. Siam A. M. J., Radoglou K. M., Noitsakis B., Smiris P. (2008). Physiological and growth responses of three Mediterranean oak species to different water availability regimes. Journal of Arid Environments, 72: 583-592. ISSN: 0140-1963. doi:10.1016/j.jaridenv.2007.11.001
90. Wolkerstorfer S. V., Wonisch, A. Stankova T., Tsvetkova N., Tausz M. (2011). Seasonal variations of gas exchange, photosynthetic pigments, and antioxidants in Turkey oak (*Quercus cerris* L.) and Hungarian oak (*Quercus frainetto* Ten.) of different age. Trees, 25: 1043–1052. ISSN: 1432-2285. Print ISSN: 0931-1890. DOI 10.1007/s00468-011-0579-1. https://link.springer.com/epdf/10.1007/s00468-011-0579-1?author_access_token=-IUagJGqjxpaC6oMfbli3_e4RwlQNchNByi7wbcMAY7Txx2mlTOSSQHXYgWrCiJZVKzvmvHdJqJBfYbkNHI7C1hGlrXvJ-mBb73GjFU6dbsh7kfXj8JOJH2qed7MosWoJALiqqwJ1GOYqiRBsYJmFA%3D%3D

Публикация: Iliev I., Scaltsoyiannes A., Rubos A. (2003). Shoot organogenesis and plant regeneration from leaf callus cultures of black bark silver birch (*Betula pendula* Roth. 'Melanocortea'). Acta Horticulturae, No 616: 321-326. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Ц и т и р а н а в:

91. Magnusson V. A., Castillo C. M., Dai W. (2009). Micropropagation of two elite birch species through shoot proliferation and regeneration. Acta Horticulturae, 812: 223-229. ISBN: 90 6605 210 4, ISSN: 0567-7572.
92. Zhai Qiaoli, Fan Guizhi, Zhan Yaguang (2010). Research progress of biotechnology on birch. Chinese Agricultural Science Bulletin, 26 (4): 93-98
93. Shadrina T. E., Sjhstibratov K. A. (2010). Efficient shoot regeneration from leaf explants of *Betula pendula*, *B. pubescens* and *B. nigra*. Acta Horticulturae, 885: 349-353. ISBN: 90 6605 210 4, ISSN: 0567-7572.
94. Niskanen A.-M. (2013). Clonal variation in Scots pine (*Pinus sylvestris*) and in transgenic silver birch (*Betula pendula*). Dissertation, University of Helsinki, 62 pp.
95. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 122 стр.
96. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
97. Vítámvás J., Kuneš I., Viehmannová I., Linda R., Balá M. (2020). Conservation of *Betula oycoviensis*, an endangered rare taxon, using vegetative propagation methods. iForest-Biogeosciences and Forestry, vol. 13: 107-113. ISSN: 1971-7458. doi: 10.3832/for3243-013

Публикация: Rohr R., Iliev I., Scaltsoyiannes A., Tsoulouha P. (2003). Acclimatization of micropropagated forest trees. Acta Horticulturae, No 616: 59-69. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Ц и т и р а н а в:

98. Marcelis L., Grashoff K., van der Zweerde W., Kempkes F., Stanghellini (2005). Reductie van verdamping door verhoging van bladweerstand middels anti-transpiranten. Plant Research International, Wageningen UR, 76 pp.
99. Da Silva Costa F. H. (2007). Micropropagacao da bananiera: caracteristicas fitotecnicas, fisiologicas e anatomicas, Dissertação, Brazil, 113 pp.
100. Sánchez I. H. (2007). Regeneración clonal de alcornoques adultos (*Quercus suber* L.) mediante embriogénesis somática. Tesis Doctoral. Universidad de Alcalá (Madrid), 215 pp.
101. Martínez Hernández M. de J. (2007). Cultivo *in vitro* de portainjertos tolerantes al virus de la tristeza en cítricos: utilización de sustratos inertes. Tesis presentada como requisito parcial para obtener el grado de: doctor en ciencias. Institución de enseñanza e investigación en ciencias agrícolas, Mexico, 80 pp.
102. Santacruz-Ruvalcaba F., Morán M. I. T., Portillo L. (2008). Micropropagación de *Agave tequilana* Weber cv. Azul: Problemas y perspectivas. Scientia-cucba, 10 (1-2): 7-20.
103. Sharifian S., Vahdati K., Mirmasoumi M., Ghaem Maghami S. A. (2009). Assesment of phloroglucinol effect on rooting of tissue cultured Persian walnut. Acta Horticulturae, 812: 189-195. ISBN: 90 6605 210 4, ISSN: 0567-7572.
104. Da Silva Costa F. H., De Castro E. M., Pasqual M., Everson J., Pereira S., De Oliveira C. (2009). Alterações anatômicas de bananieras micropropagades em resposta a aclimatização *ex vitro*. Ciência Rural, vol. 39, No 2: 386-392. ISSN: 0103-8478.

105. María de Jesús Martínez-Hernández, Alejandro Alonso López, Francisco Osorio-Acosta, Felipe Gallardo López, Héctor López Moctezuma, Martín Mata Rosas (2009). Aclimación del porta injerto tolerante al VTC *Citrus volkameriana* mediante la regulación de la intensidad de luz. *Advances en la Investigación Agrícola, Forestal y Acuicícola en el Trópico Mexicano* 2009: 95-101. ISBN: 978-607-425-214-9.
106. Torres-Moran M. I., Escoto-Delgadillo M., Molina-Moret S., Rivera-Rodriguez D. M., Velasco-Ramirez A. P., Infante D., Portillo L. (2010). Assessment of genetic fidelity among *Agave tequilana* plants propagated asexually via rhizomes versus *in vitro* culture. *Plant Cell, Tissue and Organ Culture*, 103 (3): 403-409. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. DOI: 10.1007/s11240-010-9777-6
107. González M. R. C. (2011). Proceso de compostaje de bagazo de agave tequilero a gran escala y uso de la composta para el cultivo del *Agave tequilana* en contenedor. Tesis que para obtener el grado de Doctor en Ciencias en Biosistemática, Ecología y Manejo de Recursos Naturales y Agrícolas, Universidad de Guadalajara, 202 pp.
108. Hajari E. (2011). Development of strategies towards the cryopreservation of germplasm of *Ekebergia capensis* Sparrm. – an indigenous species that produces recalcitrant seeds. Submitted in fulfillment of the academic requirements for the degree of Doctor of Philosophy in Science in the School of Biological and Conservation Sciences, University of KwaZulu-Natal, Durban, 273 pp.
109. Chariya L. D. (2012). Physiological, Biochemical and Molecular Marker Studies in *Musa* spp. PhD Thesis. Centre for Advanced Studies in plant Biotechnology & Genetic Engineering (CPBGE) Department of Biosciences Saurashtra University Rajkot-360 005, India, 167 pp.
110. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.
111. Martini A. N., Papafotiou M. (2013). Effects of plant growth regulators and environmental factors on *in vitro* propagation of *Malosorbus florentina*. *Propagation of Ornamental Plants*, 13 (3): 112-122. ISSN: 1311-9109.
112. Economou A. S. (2013). From microcutting rooting to microplant establishment: key points to consider for maximum success in woody plants. *Acta Horticulturae*, 988: 43-56. ISBN: 90 6605 210 4, ISSN: 0567-7572.
113. González-Perez E., Juárez-Muñoz J., Ayala-Garay O. J., De Jesús Yáñez-Morales. (2014). *Ex vitro* acclimatization of *Gladiolus* plantlets. *Propagation of Ornamental Plants*, 14 (3): 125-132. ISSN: 1311-9109.
114. Acemi A., Özen F., Kiran R. (2013). *In vitro* propagation of *Amsonia orientalis* Dence. from nodal segments of adult plants. *Propagation of Ornamental Plants*, 13 (1): 25-32. ISSN: 1311-9109.
115. Clapa D., Fira A., Joshee N (2013). An efficient *ex vitro* rooting and acclimatization method for horticultural plants using floral hydroculture. *HortScience* 48 (9): 1159-2013.
116. Dancheva D. (2014). Rooting and acclimatization of micropropagated shoots of *Fraxinus excelsior* L. *Oltenia Journal for Studies in Natural Sciences*, vol. 30 (1): 75-80. ISSN: 1454-6914.
117. Silva de Oliveira L. (2014). Propagação de *Eucalyptus cloeziana* F. Muell. Tese apresentada para obtenção do título de Doutor em Ciências, Universidade de São Paulo Escola Superior de Agricultura “Luiz de Queiroz”, Piracicaba, 144 p.
118. Eng W. H., Aziz M. A., Sinniah U. R. (2015). *In vitro* regeneration of *Citrus hystrix* DC. *Brazilian Journal of Botany*, 38: 235-242. DOI 10.1007/s40415-014-0127-5
119. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.

120. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
121. Adelberg J., Adelberg J. N., Rapaka V. (2015). A novel rooting matrix and vessel system resulted in larger plants and faster growth during greenhouse acclimatization of *Hydrangea quercifolia* ‘Sikes Dwarf. Propagation of Ornamental Plants, 15 (3): 89-94. ISSN: 1311-9109.
122. West T. P., Coronado R. F. (2016). Micropropagation of *Ostrya virginiana* (Mill.) K. Kock. An underutilized ornamental landscape tree. Propagation of Ornamental Plants, 16 (2): 56-61. ISSN: 1311-9109.
123. Кодун-Иванова М. А. (2017). Показатели водного стресса микроклонально размноженных растений осины *Populus tremula* при их выращивании в условиях *ex vitro*. Труды БГТУ, серия 1, no 1: 146-155.
124. Маляровская В. И., Самарина Л. С. (2018). Особенности адаптации *Lilium caucasicum* (Misch.) к условиям *ex vitro*. Субтропическое и декоративное садоводство, 66: 120-125. ISSN: 2225-3068, DOI: 10.31360/2225-3068-2018-66-120-125.
125. Valencia Juárez M. C., López D. E., Díaz L. F., Espino, Pérez E. G. (2019). Aclimatación *ex vitro* de plántulas de *Fragaria* × *ananassa* Duch. Revista Mexicana de Ciencias Agrícolas, 10 (1): 91-100. ISSN: 2007-0934, ISSN-e: 2007-9230 https://www.researchgate.net/publication/330943464_Aclimatacion_ex_vitro_de_plantula_s_de_Fragaria_x_ananassa_Duch
126. G. Indravathi, P. Suresh Babu (2019). Enhancing acclimatization of tissue cultured plants of *Albizia amara* by Biotization. International Journal of Scientific Research in Biological Sciences, 6 (4): 43-50, E-ISSN: 2347-7520 DOI: <https://doi.org/10.26438/ijsrbs/v6i4.4350>
127. Ozdemir S., Ozdemir M., Yetilmezsoy K. (2020). Effects of acclimatization soil prepared from hazelnut husk and gyttja on *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 2: 63-71. ISSN: 1311-9109.
128. Mortier E., Lamotte O., Martin-Laurent F., Recorbet G. (2020). Forty years of study on interactions between walnut tree and arbuscular mycorrhizal fungi. A review. Agronomy for Sustainable Development, 40: 43. Electronic ISSN: 1773-0155, Print ISSN: 1774-0746. <https://doi.org/10.1007/s13593-020-00647-y>
129. Маляровская В. И., Самарина А. С. (2020). Адаптация культивируемых *in vitro* регенерантов *Hydrangea macrophylla* к нестерильным условиям. Садоводство и виноградарство, 6: 12-17. <https://doi.org/10.31676/0235-2591-2020-6-12-17>
130. Зеленианська Н. М., Теслюк Н. І., Гоголінська О. І. Подуст Н. В. (2020). Розробка структурованого поживного середовища для адаптації вегетативної маси і кореневої системи мікроклонів винограду до умов *in vivo*. Таврійський науковий вісник, 116, Частина 1: 64-75. УДК 634.836:631.532:631.544 DOI <https://doi.org/10.32851/2226-0099.2020.116.1.8>
131. Vu Quoc Luan, Le Kim Cuong, Hoang Thanh Tung, Vu Thi Hien, Do Manh Cuong, Phan Le Ha Nguyen, Bui Van The Vinh, Duong Tan Nhut (2020). Highly effective micropropagation and induction of early flowering by gibberellic acid in *Paphiopedilum delenatii* Guillaumin. Propagation of Ornamental Plants, 20, (1): 3-11. ISSN: 1311-9109.
132. Singh P. R., Singh L. J. (2021). *In vitro* propagation for improvement of medicinal plants: a review. Journal of Pharmacognosy and Phytochemistry, 10 (1): 1484-1489 E-ISSN: 2278-4136 P-ISSN: 2349-8234
133. Murphy R., Adelberg J. (2021). Physical factors increased quantity and quality of micropropagated shoots of *Cannabis sativa* L. in a repeated harvest system with *ex vitro* rooting. In Vitro Cellular & Developmental Biology - Plant, 57: 923-931. <https://doi.org/10.1007/s11627-021-10166-4>, ISSN: 1054-5476 (print); 1475-2689 (web)

134. José Raymundo Enríquez-del Valle, Gerardo Rodríguez-Ortiz, Vicente Arturo Velasco-Velasco, Edmundo López-Hernández (2021). Fertilización durante la aclimatación en invernadero de plantas de *Agave potatorum* micropropagadas. Revista Mexicana de Agroecosistemas, Vol. 8 (1): 36-45. ISSN: 2007-9559 https://www.voaxaca.tecnm.mx/revista/docs/RMAE%20vol%208_1_2021/4-RMAE_2021-36-Fertilizacion-Resumen.pdf
135. Turp G. A., Turp S. M., Ozdemir S. (2021). The role of pyroligneous acid for *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 4: 116-122. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_21_4\(116-122\).pdf](https://www.journal-pop.org/References/Vol_21_4(116-122).pdf)
136. Шуркина Е. С., Маляровская В. И. (2021). Особенности адаптации *Galanthus woronowii* Losinsk. к условиям *ex vitro*. Субтропическое и декоративное садоводство, 78: 82-88. ISSN: 2225-3068. УДК 578:581.5(470.62). doi: 10.31360/2225-3068-2021-78-82-88
137. Rahma Bejaoui (2022). Kalanşo (*Kalanchoe blossfeldiana* Poelln.)'Nun *in vitro* koşullarda mikroçoğaltimi. Ankara üniversitesi fen bilimleri enstitüsü, Doktora tezi, 102 pp.
138. Constantin Dushimimana, Michael Ajanja Sakha, Mercy Jebiwott Korir, Joyce Mnyazi Jefwa, Jan Vandenabeele, Titus Magomere, Eunice Wanjiru Mutitu, Jackson Mulatya, Florence Olubayo, Guy Smagghe, and Stefaan P. O. Werbrouck (2022). Early growth performance of *in vitro* raised *Melia volkensii* Gürke plantlets in response to beneficial microorganisms under semi-arid conditions. Plants, 11: Article 1300. ISSN: 2223-7747. <https://doi.org/10.3390/plants11101300>, <https://www.mdpi.com/journal/plants>
139. Yohannes Wibisono, Toni Herawan, Asri Insiana Putri, Fithry Ardhany, Liliek Haryjanto (2023). Clonal propagation of Sandalwood (*Santalum album* L.) through tissue culture by using branch-soaked explants. Proceedings of the 1st Unhas International Conference on Agricultural Technology (UICAT 2021) AIP Conference Proceedings, Volume 2596 (1), 100021-1–100021-12; Published by AIP Publishing. On line ISSN: 1551-7616, Print ISSN: 0094-243X, <https://doi.org/10.1063/5.0119100>
140. Rahma Bejaoui, Gökem Eren Özdemir, Ş. Şebnem Ellialtıoğlu (2023). *In vitro* or *ex vitro* Rooting and Acclimatization of *Kalanchoe blossfeldiana* Poelln. Shoots Propagated by Tissue Culture Technique. Türk Tarım ve Doğa Bilimleri Dergisi (Turkish Journal of Agricultural and Natural Sciences), 10(4): 843–853. ISSN: 2148-3647. <https://doi.org/10.30910/turkjans.1288919>

Публикация: Iliev I., Rubos A., Scaltsoyiannes A., Nellas H., Kitin P. (2003). Anatomical study of *in vitro* obtained fasciated shoots from *Betula pendula* Roth. Acta Horticulturae, No 616: 481-484. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

141. Илиев Н., Петракиева А. (2007). Хетеровегетативно размножаване на обикновената череша (*Prunus avium* L.). Наука за гората, 4: 3-15.
142. Reboredo F., Silves C. (2007). Fasciation phenomena and mineral balance in *Spartium junceum* L. Phyton, vol. 47, No 1-2: 123-132.
143. Ziv M., Chen J. (2008). The anatomy and morphology of tissue cultured plants. In: George E. F., Hall M., De Klerk G-J. (Eds.). Plant propagation by tissue culture, 3rd edition, Springer, Dordrecht, NL: 465-477.
144. Петракиева А. (2008). Възможности за развъждане на обикновената череша (*Prunus avium* L.) в България. Дисертация, ЛТУ, София, 161 стр.
145. Zaki M. (2011). *In vitro* studies on regeneration of some rare trees of Kashmir Himalayas. PhD Thesis. Department of Botany, University of Kashmir, Hazratbal, Srinagar, India, 190 pp.
146. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.

147. Liao Y. K., Ji Y. Y. (2014). Mass propagation of *Drosera burmannii* Vahl via induction of shoot fasciation and recovery of morphologically normal plantlets. *Propagation of Ornamental Plants*, 4: 158-170. ISSN: 1311-9109.
148. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
149. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
150. Isah T. (2015). Adjustments to *in vitro* culture conditions and associated anomalies in plants. *Acta Biologica Cracoviensia, Series Botanica* 57/2, DOI: 10.1515/abcsb-2015-0026
151. Gailis A., Samsone I., Šēnhofa S., Girgžde E., Kāpostiniš R., Jansons Ā. (2021). Silver birch (*Betula pendula* Roth.) culture initiation *in vitro* and genotype determined differences in micropropagation. *New Forests*, 52 (5): 1-16. Electronic ISSN: 1573-5095, Print ISSN: 0169-4286. <https://doi.org/10.1007/s11056-020-09828-9>, <https://link.springer.com/article/10.1007/s11056-020-09828-9>

Публикация: Пиев И., Tomita M. (2003). Micropropagation of *Betula pendula* Roth. ‘Fastigiata’ by leaf callus cultures. *Propagation of Ornamental Plants*, vol. 3, No 1: 20-26. ISSN: 1311-9109. https://www.journal-pop.org/2003_3_1_20-26.html

Цитирана в:

152. Haggman H., Sutela S., Welander M. (2007). Micropropagation of *Betula pendula* Roth including genetically modified plants. In: S. M. Jain, Haggman H. (Eds.). *Protocols for micropropagation of woody trees and fruits*, Springer: 153-162.
153. Илиев Н., Петракиева А. (2007). Хетеровегетативно размножаване на обикновената череша (*Prunus avium* L.). *Наука за гората*, 4: 3-15.
154. Петракиева А. (2008). Възможности за развъждане на обикновената череша (*Prunus avium* L.) в България. Дисертация, ЛТУ, София, 161 стр.
155. Shadrina T. E., Shestibratov K. A. (2010). Efficient shoot regeneration from leaf explants of *Betula pendula*, *B. pubescens* and *B. nigra*. *Acta Horticulturae*, 885: 349-353. ISBN: 90 6605 210 4, ISSN: 0567-7572.
156. Oinam G., Yeung E., Kurepin L., Haslam T., Lopez-Villalobos A. (2011). Adventitious root formation of ornamental plants: I. General overview and recent successes. *Propagation of Ornamental Plants*, 11 (2): 78-90. ISSN: 1311-9109.
157. Zaki M., Sofi M. S., Kalo Z. A. (2011). A reproducible protocol for raising clonal plants from leaf segments excised from mature trees of *Betula utilis* a threatened tree species of Kashmir Himalayas. *International Multidisciplinary Research Journal*, 1 (5): 7-13. ISSN: 2231-6302.
158. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.
159. Nazari J., Alizadeh M., Payamnoor V., Bezdi K. G. (2012). Sterilization treatments optimizing of *Betula pendula* seeds for *in vitro* culture. *Proceedings of the 12th Iranian Congress of Genetics* (in Persian).
160. Кильчевский А. В., Хотылева Л. В. (2012). Генетические основы селекции растений. Биотехнология и селекции растений. Том 3: Клеточная инженерия. Минск, Беларусь, „Беларуская навука”, 489 стр., ISBN: 978-985-08-1392-3. <https://www.iprbookshop.ru/29441.html>
161. Gaidamashvili M., Khurtsidze E., Barblishvili T. (2015). Micropropagation of threatened *Betula* species for *in vitro* conservation. *International Conference on Plant, Marine and Environmental Sciences (PMES-2015)* Jan. 1-2, 2015 Kuala Lumpur (Malaysia): 12-16.

162. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
163. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Kitin P., Iliev I., Scaltsoyiannes A., Nellas Ch., Rubos A., Funada R. (2005). A comparative histological study between normal and fasciated shoots of *Prunus avium* L. generated *in vitro*. Plant Cell, Tissue and Organ Culture, 82: 141-150. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. <https://link.springer.com/article/10.1007/s11240-004-8153-9>

Цитирани в:

164. Reboredo F., Silveiras C. (2007). Fasciation phenomena and mineral balance in *Spartium junceum* L. Phyton, vol. 47, No1-2: 123-132.
165. Илиев Н., Петракиева А. (2007). Хетеровегетативно размножаване на обикновената череша (*Prunus avium* L.). Наука за гората, 4: 3-15.
166. Păunescu A. (2008). *In vitro* and *in vivo* variability of histological traits of *Dianthus callizonus* (Cariophyllaceae) aerial vegetative organs. Phytologia Balcanica, 14 (3): 417-423.
167. Петракиева А. (2008). Възможности за развъждане на обикновената череша (*Prunus avium* L.) в България. Дисертация, ЛТУ, София, 161 стр.
168. Tunjit P. (2008). Micropropagation of *Euphorbia muilli* Desmoul via Cyathium Explants. Dissertation, Taiwan, 97 pp.
169. Đurković J., Mišalová A. (2008). Micropropagation of temperate noble hardwoods: an overview. Functional Plant Science and Biotechnology, 2 (1): 1-19.
170. Amutha S., Kathiravan K., Singer S., Jashi L., Shomer I., Steinitz B., Gaba V. (2009). Adventitious shoot formation in decapitated dicotyledonous seedlings starts with regeneration of abnormal leaves from cells not located in a shoot apical meristem. In vitro Cellular & Developmental Biology-Plant, 45: 758-768. ISSN: 1054-5476 (print); 1475-2689 (web). DOI 10.1007/s11627-009-9232-8
171. Aktürk Z. (2009). Kirazi (*Prunus avium* L.) *in vitro* mikroçoğaltimi. T.C. Dicle Üniversitesi Fen, Bilimleri Enstitüsü, Doktora tezi biyoloji, Anabilim Dalı Diyarbakir Aralık, 174 pp.
172. Cramarić O. T., Štambuk S., Kajba D. (2011). Genotipska raznolikost divlje trešnje (*Prunus avium* L.) u dijelu prirodne rasprostranjenosti u hrvatskoj. Šumarski list, 135 (11-12): 543-555.
173. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.
174. Samarakoon U. C. (2012). The physiology and control of crown bud formation and development in gentians. PhD thesis. Massey University, Palmerston North, New Zealand, 326 pp.
175. İsfendiyaroğlu M. (2021). Anatomy of adventitious root development in mastic tree (*Pistacia lentiscus* var. *chia* Duham.) cuttings. Propagation of Ornamental Plants, 21 (2): 58-64. ISSN: 1311-9109.
176. El-Banna A. N., El-Nady M. F., Dewir Y. H., El-Mahrouk M. E. (2013). Stem fasciation in cacti and succulent species - tissue anatomy, protein pattern and RAPD polymorphisms. Acta Biologica Hungarica, 64 (3): 305-318. DOI: 10.1556/ABiol.64.2013.3.4
177. Нонић М., Поповић В., Керкез И., Шијачић-Николић М. (2013). Варијабилност морфометријских карактеристика семена различитих тест стабала дивље трешње (*Prunus avium* L.) са подручја Београда. Шумарство, 1-2: 113-123.

178. Omar A. F. Dewir Y. H., El-Mahrouk M. E. (2014). Molecular identification of phytoplasmas in fasciated cacti and succulent species and associated hormonal perturbation. *Journal of Plant Interactions*, 9 (1): 632-639. DOI: 10.1080/17429145.2014.882421, <http://dx.doi.org/10.1080/17429145.2014.882421>
179. Liao Y. K., Ji Y. Y. (2014). Mass propagation of *Drosera burmannii* Vahl via induction of shoot fasciation and recovery of morphologically normal plantlets. *Propagation of Ornamental Plants*, 4: 158-170. ISSN: 1311-9109.
180. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
181. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
182. Корнова К., Попов С. (2015). Оптимизиране процеса на вкореняване при *in vitro* размножаване на черешовата подложка Gisela 6 (*P. cerasus* × *P. canescens* sp.). *Растениевъдни науки*, LII, 2: 29-33.
183. Isah T. (2015). Adjustments to *in vitro* culture conditions and associated anomalies in plants. *Acta Biologica Cracoviensia, Series Botanica* 57/2: 9–28. DOI: 10.1515/abcsb-2015-0026
184. Popović V., Kerkez I. (2016). Varijabilnost populacija divlje trešnje (*Prunus avium* L.) u Srbiji prema morfološkim svojstvima listova. *Šumarski list*, 7-8: 347-355.
185. Dhandapani P., Song J., Novak O., Jameson P. E. (2016). Infection by *Rhodococcus fascians* maintains cotyledons as a sink tissue for the pathogen. *Annals of Botany*, 119: 841-852. DOI 10.1093/aob/mcw202
186. Dewir Y. H. (2016). Cacti and succulent plant species as phytoplasma hosts: a review. *Phytopathogenic Mollicutes*, 6 (1): 1-9.
187. Adedapo Adedayo Adeniran, Mubo Adeola Sonibare, Girish Halemirle Rajacharya, Shashi Kuma (2018). Assessment of genetic fidelity of *Dioscorea bulbifera* L. and *Dioscorea hirtiflora* Benth. and medicinal bioactivity produced from the induced tuberous roots. *Plant Cell, Tissue and Organ Culture*, 132 (2): 343-357. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. <https://doi.org/10.1007/s11240-017-1334-0>.
188. Liang H., Xiong Y., Guo B., Yan H., Jian Sh., Ren H., Zhang X., Li Y., Zeng S., Wu K., Zheng F., Teixeira da Silva J. A., Xiong Y., Ma G. (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. *Environmental and Experimental Biology*, 17: 169–177. eISSN: 2255-9582pISSN: 1691-8088. DOI: 10.22364/eeb.17.17
189. Aydin E. (2019). Anaç adayı kiraz, vişne ve mahlep genotiplerinin doku kültürü yöntemiyle çoğaltılması. T.C. Ordu Üniversitesi fen bilimleri enstitüsü, Doktora tezi, Bahçe bitkileri anabilim dalı, Ordu, 101 pp.
190. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Livia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. *Rodriguésia* 72: e00372019. 2021 <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>. ISSN: 03706583, ISSN: 21757860.
191. Aydin E., Yarılgac T. (2021). In Vitro Propagation of Some Mahaleb Genotypes as Candidate Rootstock for Sweet Cherries. *Yuzuncu Yil University Journal of Agricultural Sciences*, 31 (4): 847 – 857. ISSN: 1308-7576, EISSN: 1308-7584, DOI: 10.29133/yyutbd.892027, <https://doi.org/10.29133/yyutbd.892027>, <https://dergipark.org.tr/en/download/article-file/1621019>.
192. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). *Science Archeives*, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>

Публикация: Илиев Н., Илиев И., Парк Y.-G. (2005). Black locust (*Robinia pseudoacacia* L.) in Bulgaria. *Journal of Korean Forest Society*, 94 (5): 291-301. ISSN: 0445-4650.

https://www.researchgate.net/publication/323836661_Black_locust_Robinia_pseudoacacia_L_in_Bulgaria#fullTextFileContent https://www.researchgate.net/profile/Nasko-Iliev-3/publication/312530134_Black_Locust_Robinia_pseudoacacia_L_in_Bulgaria/links/5880ceffaca272de1a410007/Black-Locust-Robinia-pseudoacacia-L-in-Bulgaria.pdf

Цитирани в:

193. Панайотов П., Калмуков К., Панайотов М. (2006). Фактори оказващи влияние на устойчивото развитие на бялата акация. Управление и устойчиво развитие, том 14, № 1-2: 194-202.
194. Redei K., Osvath-Bujtas Z., Keseru Zs., Park Y.-G. (2007). Clonal Selection of Black Locust (*Robinia pseudoacacia* L.) in Hungary : a Review. Korean Journal of Apiculture, 22 (2): 189-193.
195. Corneanu M., Corneanu G., Hernea C., Niculescu M., Netoiu C., Posta D., Sarac I., Petcov A., Gradinaru M.-M. (2008). Studies on the flower and leaves variability in *Robinia pseudoacacia* genotypes of south Oltenia. Anales of the University of Craiova, vol. XXXVIII/B: 111-119.
196. Kurokochi H., Toyama K., Hogetsu T. (2010). Regeneration of *Robinia pseudoacacia* riparian forests after clear-cutting along the Chikumagawa river in Japan. Plant Ecology, 210 (1): 31-41. DOI 10.1007/s11258-010-9735-8
197. Kirokouci H. (2011). Dynamic state of *Robinia pseudoacacia* forests at dry riverbed of Chikuma river. PhD thesis. University of Tokyo, Graduate School of Agricultural and Life Sciences, Faculty of Agriculture, Department of Forest Science, 90 pp. (in Japanese).
198. Masaka K. (2013). Option and its scientific evidence concerning the exitic tree species *Robinia pseudoacacia*. Journal of Japanese Forest Society, 95: 332-341.
199. Kurokochi H., Toyama K. (2015). Invasive Tree Species *Robinia pseudoacacia*: A Potential Biomass Resource in Nagano Prefecture, Japan. Small-scale Forestry, 14: 205-215. DOI 10.1007/s11842-014-9282-6
200. Guse T., Schneck V., von Wühlisch G., Liesebach M. (2015). Untersuchungen der Ertragsleistung und -stabilität bei Robinien-Jungpflanzen verschiedener Herkunft auf einem Standort im Land Brandenburg. In: Liesebach M. (Ed.): FastWOOD II: Züchtung schnellwachsender Baumarten für die Produktion nachwachsender Rohstoffe im Kurzumtrieb - Erkenntnisse aus 6 Jahren FastWOOD, Thünen Report, No. 26: 85-97. ISBN: 978-3-86576-131-6, ISSN: 2196-2324, <http://hdl.handle.net/10419/108517>. DOI: 10.3220/REP_26_2015
201. Masaka K., Torita H., Kon H., Fukuchi M. (2015). Seasonality of sprouting in the exotic tree *Robinia pseudoacacia* L. in Hokkaido, northern Japan. Journal of Forest Research, 20: 386-395.
202. Зайцева І. А., Ситникова А. К. (2017). Аналіз ушкодження насіння *Robinia* L. комахами у паркових і вуличних насадженнях м. Дніпро. Питання біоіндикації та екології. Вип. 22, № 2: 111-127. ISSN: 2312-2056.
203. Černulienė S. (2018). Baltažiedės robinijos (*Robinia pseudoacacia* L.) plitimas ir ekologiniai ypatumai lietuvis sąlygomis, Daktaro disertacija Žemės ūkio mokslų sritis, Miškotyros kryptis (04A). Aleksandro Stulginskio Universitetas Lietuvos Agrarinių Ir Miškų Mokslų Centras (Aleksandras Stulginskis University Lithuanian Research Centre For Agriculture And Forestry), 136 pp. ISBN 978-609-449-145-0 https://www.lammc.lt/data/public/uploads/2018/11/s.cernuliene_disertacija.pdf
204. Zhang K., Shen Z., Yang X., Ma L., Duan J., Li Y. (2019). Spatial Patterns in Different Stages of Regeneration after Clear-Cutting of a Black Locust Forest in Central China. Forests, 10, 1066. ISSN: 1999-4907. doi: 10.3390/f10121066

205. Vítková M., Sádlo J., Roleček J., Petřík P., Sitzia T., Müllerová J., Pyšek P. (2020). *Robinia pseudoacacia*-dominated vegetation types of Southern Europe: species composition, history, distribution and management. *Science of the Total Environment*, 707, 134857. ISSN: 0048-9697. doi: <https://doi.org/10.1016/j.scitotenv.2019.134857> (IF).
206. Nicolescu V.-N., Rédei K., Mason W. L., Vor T., Pöetzelsberger E., Bastien J.-Ch., Brus R., Benčať T., Đodan M., Cvjetković B., Andrašev S., La Porta N., Lavnyy V., Mandžukovski D., Petkova K., Roženberger D., Waşık R., Mohren G. M. J., Monteverdi M. C., Musch B., Klisz M., Perić S., Keça L., Bartlett D., Hernea C., Pástor M. (2020). Ecology, growth and management of black locust (*Robinia pseudoacacia* L.), a non-native species integrated into European forests. *J. For. Res.*, 31 (4):1081–1101. Online ISSN: 1993-0607, Print ISSN: 10007-662X, <https://doi.org/10.1007/s11676-020->
207. S. Barudanović E. Zečić A. Macanović B. Duraković, E. Mašić (2021). Invasive Alien Plant Species in Global Perspectives with Special References to Bosnia and Herzegovina. In: T. Pullaiah, Michael R. Ielmini (Eds). *Invasive Alien Species: Observations and Issues from Around the World, Volume 3: Issues and Invasions in Europe*: 215-252. Print ISBN: 9781119607021 Online ISBN: 9781119607045, <https://doi.org/10.1002/9781119607045.ch31>

Публикация: Karanikas Ch., Scaltsoyiannes A., Iliev I., Tsoulpha P., Tsaktsira M. (2005). **Micropropagation of mature *Morus alba* ‘Pendula’ trees for ornamental use. Proceedings of the 12th Greek Forest Society Conference, 2-5 Oct. 2005, Drama, Greece: 87-94 (in Greek).**

Цитирана в:

208. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

Публикация: Iliev I. (2008). E. F. George, M. A. Hall, and G.-J. De Klerk (Eds). **Plant propagation by tissue culture. Volume 1. The background. 3rd edn. Book review. Plant Cell, Tissue and Organ Culture, 93: 353-355. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044.**

Цитирана в:

209. Yanping Lu, Liejian Huang, Hong Muhammad, Qasim Shahid (2021). *Acacia mangium* × *A. auriculiformis* micropropagation in a non-sterile environment. Research Square: 1-23. <https://assets.researchsquare.com/files/rs-690162/v1/a6f8d17b-2103-459c-8b4d-f3104193c53d.pdf?c=1631886833>

Публикация: Nikolaou P., Zagaz D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). **Advances in the micropropagation of service tree *Sorbus domestica*. Propagation of Ornamental Plants, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html**

Цитирана в:

210. Ördögh M., Jámor-Benczúr E., Tilly-Mándy A., Lelik L. (2009). Effects of different cytokinins on proliferation of *Sorbus borbasii* ‘Herkulesfüdrö’. *Propagation of Ornamental Plants*, 9 (1): 43-46. ISSN: 1311-9109.
211. Ali S. B. G. M., Kouros V., Hassan B. S., Siamak K., Charles L. (2010). Enhancement of maturation and germination of somatic embryos in Persian walnut (*Juglans regia* L.) using osmolites, hormones and cold treatments. *African Journal of Food Science*, 4 (12): 735-743. ISSN 1996-0794. <http://www.academicjournals.org/ajfs>

212. Oinam G., Yeung E., Kurepin L., Haslam T., Lopez-Villalobos A. (2011). Adventitious root formation of ornamental plants: I. General overview and recent successes. *Propagation of Ornamental Plants*, 11 (2): 78-90. ISSN: 1311-9109.
 213. Malá J., Cvrčková H., Machova P., Dostal J. (2011). Micropropagace jeřábu okeruše (*Sorbus domestica* L.). *Lesnický průvodce*, 4: 1-17. ISBN 978-80-7417-083-6, ISSN 0862-7657, <http://www.vulhm.cz>
 214. Ördögh M. (2011). Táptalaj kiegészítők morfológiai, anatómyai és fiziológiai hatása *Sorbus* taxonok mikroszaporítása során. Doktori (PhD) élekezés. Budapesti Corvinus Egyetem, 219 pp.
 215. Dobránszki J., Magyar-Tábori K., Jevcsák M., Ördögh M., Jámor-Benczúr E. (2012). Improving the *in vitro* rooting of micro-shoots of *Sorbus rotundifolia* 'Bükk szépe' by the sequential application of Humus® FW and Wuxal® Super organic and chemical fertilisers *Journal of Horticultural Science & Biotechnology* 87 (5): 509-513.
 216. Máchová P., Malá J., Cvrčková H., Dostál J., Buriánek V. (2013). *In vitro* reproduction of rare and endemic species of rowan tree *Journal of Forest Science*, 59 (10): 386-390
 217. Malá J., Cvrčková H., Máchová P., Buriánek V. (2014). Mikropropagace endemitních jeřábů (*Sorbus* spp.). *Lesnický průvodce*, 4: 1-20. ISBN 978-80-7417-083-6, ISSN 0862-7657. <http://www.vulhm.cz>
 218. Hrdoušek V., Špišek Z., Krška B., Šediva J., Bakay L. (2014). Oskeruše strom pro novou Evropu. In: Paganova V., Řezníček V., Vašut R. J., Benediková M. (Eds). *Petr Brázda – vydavatelství*, 240 pp. ISBN: 978-80-87387-28-3.
 219. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
 220. Jeong B. R., Sivanesan I. (2015). Micropropagation of *Sorbus commixta* Hedl. *Propagation of Ornamental Plants*, 15 (4): 142-146. ISSN: 1311-9109
 221. Máté Ördögh (2022). *In vitro* propagation results of *Sorbus aria* 'Gran Sasso'. Review on Agriculture and Rural Development, vol. 11 (1-2): 204-222. ISSN: 2677-0792. DOI: 10.14232/rard.2022.1-2.204-214.
 222. Jana Šedivá, Jiří Velebil, Daniel Zahradník (2023). Micropropagation as a tool for the conservation of autochthonous *Sorbus* species of Czechia. *Plants*, 12, 488. ISSN: 2223-7747. <https://doi.org/10.3390/plants12030488>
- Публикация: Mitras D., Kitin P., Iliev I., Dancheva D., Scaltsoyiannes A., Tsaktsira M., Nellas C., Rohr R. (2009). *In vitro* propagation of *Fraxinus excelsior* L. by epycotyls. *Journal of Biological Research*, 11: 37-48. ISSN: 2241-5793. <https://ikee.lib.auth.gr/record/233121/files/Mitras%20et%20al.pdf>**

Цитирана в:

223. El-Mahrouk M. E. S., Dewir Y. H., Omar A. M. K. (2010). *In vitro* propagation of adult strawberry tree (*Arbutus unedo* L.) through adventitious shoots and somatic embryogenesis. *Propagation of Ornamental Plants*, 10 (2): 93-98. ISSN: 1311-9109.
224. Pijut P. M., Lawson S. S., Michler C. H. (2010). Biotechnological efforts for preserving and enhancing temperate hardwood tree biodiversity, health, and productivity. *In vitro Cellular & Developmental Biology-Plant*, 47: 123-147. ISSN: 1054-5476 (print); 1475-2689 (web). DOI 10.1007/s11627-010-9332-5
225. Palla K. J., Pijut P. M. (2011). Regeneration of plants from *Fraxinus americana* hypocotyls and cotyledons. *In vitro Cellular & Developmental Biology-Plant*, 47 (2): 250-256. ISSN: 1054-5476 (print); 1475-2689 (web). DOI 10.1007/s11627-011-9360-9
226. Dewir Y. H., Singh N., Mngomezulu, Omar A. M. K. (2011). Micropropagation and detection of important triterpenes in *in vitro* and field grown plants of *Syzygium cordatum*. *Journal of Medicine Plants Research*, vol. 5(14): 3078-3083. ISSN 1996-0875

227. Stevens M. E., Pijut P. M. (2012). Hypocotyl derived *in vitro* regeneration of pumpkin ash (*Fraxinus profunda*). Plant Cell, Tissue and Organ Culture, 108: 129-135. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. DOI 10.1007/s11240-011-0021-9
228. Šedivá J. (2012). *In vitro* root formation in *Pulsatilla vernalis* (L.) Mill. Propagation of Ornamental Plants, 12: 96-101. ISSN: 1311-9109.
229. Sivanesan I., Jeong B. R. (2013). Direct adventitious shoot regeneration from cotyledonary explants of *Jeffersonia dubia* Benth et. Hook. Propagation of Ornamental Plants, 13 (1): 46-48. ISSN: 1311-9109.
230. Beasley R. R., Pijut P. M. (2013). Regeneration of plants from *Fraxinus nigra* Marsh. hypocotyls. HortScience, 48 (7): 887-890.
231. El-Banna A. N., El-Nady M. F., Dewir Y. H., El-Mahrouk M. E. (2013). Stem fasciation in cacti and succulent species - tissue anatomy, protein pattern and RAPD polymorphisms. Acta Biologica Hungarica, 64 (3): 305-318. DOI: 10.1556/ABiol.64.2013.3.4
232. Omar A. F. Dewir Y. H., El-Mahrouk M. E. (2014). Molecular identification of phytoplasmas in fasciated cacti and succulent species and associated hormonal perturbation. Journal of Plant Interactions, 9 (1): 632-639. DOI: 10.1080/17429145.2014.882421, <http://dx.doi.org/10.1080/17429145.2014.882421>
233. Dancheva D. (2014). Rooting and acclimatization of micropropagated shoots of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, vol. 30 (1): 75-80. ISSN 1454-6914
234. Liao Y. K., Ji Y. Y. (2014). Mass propagation of *Drosera burmannii* Vahl via induction of shoot fasciation and recovery of morphologically normal plantlets. Propagation of Ornamental Plants, 4: 158-170. ISSN: 1311-9109.
235. Dewir Y. H., El-Mahrouk M. E., El-Banna. (2015). *In vitro* propagation and preliminary results of *Agrobacterium*-mediated genetic transformation of *Cordyline fruticosa*. South African Journal of Botany, 98: 45-51. ISSN: 0254-6299. <http://dx.doi.org/10.1016/j.sajb.2015.01.017>
236. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
237. Parvanova P., Lyubomirova T., Tzvetkova, N., Ilinkin V. (2015). Effect of auxin on protein content during *in vitro* phizogenesis of *Syringa vulgaris* L. Annuaire de l'Université de Sofia "St. Kliment Ohridski", Faculte de Biologie, vol. 4: 208-215.
238. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 122 стр.
239. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
240. Isah T. (2015). Adjustments to *in vitro* culture conditions and associated anomalies in plants. Acta Biologica Cracoviensia, Series Botanica 57/2, DOI: 10.1515/abcsb-2015-0026
241. Lebedev V., Shestibratov K. (2016). Large-scale micropropagation of common ash Biotechnology ISSN: 1682-296X (print), ISSN: 1682-2978 (online). DOI: 10.3923/biotech.2016.1.9
242. Thomas P. A. (2016). Biological flora of the British isles: *Fraxinus excelsior*. Journal of Ecology, 281. DOI 10.1111/1365-2745.12566.
243. Dewir Y. H. (2016). Cacti and succulent plant species as phytoplasma hosts: a review. Phytopathogenic Mollicutes Vol. 6 (1): 1-9
244. West T. P., Coronado R. F. (2016). Micropropagation of *Ostrya virginiana* (Mill.) K. Kock. An underutilized ornamental landscape tree. Propagation of Ornamental Plants, 16 (2): 56-61. ISSN: 1311-9109.

245. Douglas G. C., Namara J. M., O'connell K., Dunne L., Grant J. (2017). Vegetative propagation of dieback-tolerant *Fraxinus excelsior* on commercial scale. In: R. Vasaitis & R. Enderle (Eds). Dieback of European Ash (*Fraxinus* spp.): Consequences and Guidelines for Sustainable Management, Swedish University of Agricultural Sciences. Printed in Sweden: 288-299. ISBN: (print version) 978-91-576-8696-1, ISBN: (electronic version) 978-91-576-8697-8
246. Dewir Y. H., Aldubai A. A., El-Hendawy S., Alsadon, Kadry Seliem A. A. M., Naidoo Y. (2018). Micropropagation of Buttonwood Tree (*Conocarpus erectus*) through Axillary Shoot Proliferation. HortScience, 53 (5): 687–691. Print ISSN: 0018-5345, Online ISSN: 2327-9834. DOI: <https://doi.org/10.21273/HORTSCI12987-18>
247. Fenning T. M. (2019). The use of tissue culture and in-vitro approaches for the study of tree diseases. Plant Cell Tissue and Organ Culture, 136: 415-430. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. <https://doi.org/10.1007/s11240-018-01531-0>
248. Pacholczak A., Nowakowska K. (2019). Micropropagation of February daphne (*Daphne mezereum* L.). Propagation of Ornamental Plants, 19: 106-112. ISSN: 1311-9109.
249. H. R. Islami Rostami, S. M. Hosseini Nasr, S. K. Kazemitabar, F. Zafarian (2019). Effect of provenances and culture media on seed germination of ash (*Fraxinus excelsior* L.) in embryo *in vitro* culture. Iranian Journal of Forest and Poplar Research, 27 (2): 159-168. Printing ISSN: 1735-0883, Electronic ISSN: 2383-1146. DOI: 10.22092/ijfpr.2019.120122
250. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Livia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. Rodriguésia 72: e00372019. 2021 ISSN printed version: 0370-6583, ISSN online version: 2175-7860. <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>
251. Trevor Fenning, Margaret O'Donnell, Katharine Preedy, Aurélia Bézanger, David Kenyon, Gustavo Lopez. (2022). The rooting ability of in-vitro shoot cultures established from a UK collection of the common ash (*Fraxinus excelsior* L.) and their *ex-vitro* survival. Annals of Forest Science, 79: 30. ISSN: 1297-966X, <https://doi.org/10.1186/s13595-022-01146-8>
252. Haifeng Y., Anyua L., Jiaqia W., Guoshenga Z., Shusenb W., Xina Z. (2021). Identification of fasciation phenotype and anatomic structure analysis in *Salix psammophila*. Journal of Desert Research, vol. 41, No. 4: 45-50. ISSN: 1000-694X. DOI: 10.7522/j.issn.1000-694X.2021.00031
253. İsfendiyaroğlu M. (2021). Anatomy of adventitious root development in mastic tree (*Pistacia lentiscus* var. *chia* Duham.) cuttings. Propagation of Ornamental Plants, 21 (2): 58-64. ISSN: 1311-9109.
254. Xu L., Cheng F., Zhong Y. (2021). Micropropagation of *Paeonia ostii* 'Feng Dan' via *in vitro* mature embryo culture. Propagation of Ornamental Plants, 3: 78-87. ISSN: 1311-9109
255. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). Science Archeives, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>
256. Katarzyna Nawrot-Chorabik, Bartosz Pluciński, Dariusz Latowski (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. New Forests. ISSN: 0169-4286, eISSN: 1573-5095. <https://doi.org/10.1007/s11056-023-09981-x>
257. Ali Mohsen Al-Ali, Yaser Hassan Dewir and Rashid Sultan Al-Obeed (2023). Influence of Cytokinins, Dark Incubation and Air-Lift Bioreactor Culture on Axillary Shoot Proliferation of Al-Taif Rose (*Rosa damascena* trigintipetala (Diek) R. Keller). Horticulturae, 9, 1109. ISSN 2311-7524. <https://doi.org/10.3390/horticulturae9101109>

Публикация: Scaltsoyiannes A., Tsulpha P., Iliev I., Theriou K., Tsaktsira M., Mitras D., Karanikas C., Mahmoud S., Christopoulos V., Scaltsoyiannes V., Zaragotas D., Tzouvara A. (2009). Vegetative propagation of ornamental genotypes of *Prunus avium* L. Propagation of Ornamental Plants, 9(4): 198-206. ISSN: 1311-9109. https://www.journal-pop.org/2009_9_4_198-206.html

Цитирани в:

258. Илиев Н., Петракиева А. (2007). Хетеровегетативно размножаване на обикновената череша (*Prunus avium* L.). Наука за гората, 4: 3-15.
259. Петракиева А. (2008). Възможности за развъждане на обикновената череша (*Prunus avium* L.) в България. Дисертация, ЛТУ, София, 161 стр.
260. Cheong E. J. (2012). Biotechnological approaches for improvement and conservation of *Prunus* species. Plant Biotechnology Reports, 6 (1): 17-28. DOI 10.1007/s11816-011-0195-y
261. Bošnjaković D., Ognjanov V., Barać G., Ljubojević M., Pranjić A., Dugalić K. (2013). Mikropropagacija selekcija slabobujnih pologa za višnju i trešnju. Voćarstvo, 47 (183-184): 121-128.
262. Wang Y., Pijut P. M. (2014). Can *Prunus serotina* be genetically engineered for reproductive sterility and insect pest resistance? Springer Science Reviews, 2:75-93. DOI 10.1007/s40362-014-0024-0
263. Tchatchouai D. T., Barbas E., Filippou A. (Phil) Aravanopoulos F. A. (2014). Micropropagation of elite genotypes of *Castanea sativa* (Mill.) Journal of Advances in Biotechnology, Vol 3, No 2: 200-209. ISSN: 2348-6201.
264. Cheong E. J. (2014). Biotechnological approaches for improvement and conservation of *Prunus* species. In: Kishan Gopal Ramawat, Jean-Michel Mérillon, M. R. Ahuja (Eds). Tree Biotechnology. CRC Press: 446-468 pages. ISBN: 978-1-4665-9714-3. https://books.google.bg/books?id=spA-AwAAQBAJ&pg=PA466&lpg=PA466&dq=prunus+avium,+iliev+i.&source=bl&ots=ykQymu8vQc&sig=ACfU3U38YISBOP9aSXEWpeusSmXzZodz7Q&hl=en&sa=X&ved=2ahUKEwjEyM6x2v_0AhWOQ_EDHTxnBE04ChDoAXoECBAQAw#v=onepage&q=prunus%20avium%2C%20iliev%20i.&f=false
265. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
266. Dorić D., Ognjanov V., Barać G., Ljubojević M., Pranjić A., Dugalić K., Ercişli S. (2015). Use of *in vitro* propagation of 'Oblacinska' sour cherry in rootstock breeding. Turkish Journal of Biology, 39: 575-581. doi:10.3906/biy-1412-85
267. Crnarić O. T., Kajba D. (2016). Micropropagation of wild cherry (*Prunus avium* L.) from a clonal seed orchard. Šumarski list, 5-6: 273-282.
268. Tomov V. (2017). Rooting of norway maple (*Acer platanoides* L.) cuttings. Forestry Ideas, 23, No 1 (53): 57-64.
269. Sun Q., Sun H., Bell R. L., Guan Q. (2018). Plant regeneration from *in vitro* leaf explants of Chinese wild cherry (*Prunus pseudocerasus* Lindl). Propagation of Ornamental Plants, 18: 3-11. ISSN: 1311-9109
270. Selin Gül Ünsal, Angelo Gismondi, Gabriele Di Marco, Antonella Canini, Burcu Uzan Eken, Yelda Özden Çiftçi (2019). Intraspecific discrimination study of wild cherry populations from North-Western Turkey by DNA barcoding approach. Tree Genetics & Genomes, 15: 16. ISSN: 1614-2942 (print), ISSN: 1614-2950 (online) <https://doi.org/10.1007/s11295-019-1323-z>
271. Pijut P. (2020). *Prunus* spp. Cherry. In: Richard E. Litz, Fernando Pliego-Alfaro, Jose Ignacio Hormazap (Eds). Biotechnology of Fruit and Nut Crops, 2nd Edition: 532-560. ISBN-13:

Публикация: Scaltsoyiannes A., Tsaktsira M., Pasagiannis G., Tsoulpha P., Zhelev P., Iliev I., Rohr R. (2009). Allozyme variation of European black (*Pinus nigra* Arnold) and scots pine (*Pinus sylvestris* L.) populations and implications on their evaluation: a comparative study. *Journal of Biological Research*, 11: 95-106. ISSN: 2241-5793. https://www.researchgate.net/publication/279622910_Allozyme_variation_of_European_Black_Pinus_nigra_Arnold_and_Scots_pine_Pinus_sylvestris_L_populations_and_implications_on_their_evolution_A_comparative_study
<https://ikee.lib.auth.gr/record/233302/files/Skaltsogiannis.pdf>

Цитирани е:

272. Alexandrov A., Dobrev R. (2011). Country Report for The State of World's Forest Genetic Resources – Bulgaria, 63 pp.
273. Raptis D., Zagas D., Karamanolis D., Karnouskos P., Zagas Th. (2011). Structure analysis of Black Pine (*Pinus nigra* Arn.) stands in the Southeast Mt. Olympus ΠΡΑΚΤΙΚΑ 15ου ΠΑΝΕΛΛΗΝΙΟΥ ΔΑΣΟΛΟΓΙΚΟΥ ΣΥΝΕΔΡΙΟΥ: 567-578. (in Greek).
274. Thiel D., Nagy L., Beierkuhnlein C., Huber G., Jentsch A., Konnert M., Kreyling J. (2012). Uniform drought and warming responses in *Pinus nigra* provenances despite specific overall performances. *Forest Ecology and Management*, 270: 200-208.
275. Swzymczak S., Joachimski M. M., Bräuning A., Hetzer T., Kuhlemann J. (2012). Are pooled ring $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ series reliable climate archives? - a case study of *Pinus nigra* spp. *laricio* (Corsica/France). *Chemical Geology*, 308-309: 40-49. doi:10.1016/j.chemgeo.2012.03.013
276. Alexandrov A., Dobrev R. (2012). State of forest genetic resources in Bulgaria. *Forest Science*, No 1/2: 2-22. ISSN 0861-007X
277. Chudzińska E., Diatta J. B., Wojnicka-Półtorak A. (2013). Adaptation strategies and referencing trial of Scots and black pine populations subjected to heavy metal pollution. *Environmental Science and Pollution Research*, 21(3): 2165-2177. DOI 10.1007/s11356-013-2081-3.
278. Zorica S. Šarac (2014). Utvrđivanje varijabilnosti i diferencijacije prirodnih populacija *Pinus nigra* arnold u srbiji korišćenjem fitohemijskih i molekularnih markera. Doktorska disertacija. Univerzitet u Beogradu, Biološki Fakultet, 268 pp.
279. Christopoulou A., Fyllas N. M., Andriopoulos P., Koutsias N., Dimitrakopoulos P. G., Arianoutsou M. (2014). Post-fire regeneration patterns of *Pinus nigra* in a recently burned area in Mount Taygetos, Southern Greece: The role of unburned forest patches. *Forest Ecology and Management*, 327 (1): 148-156.
280. Лебедев А. Г. (2014). Анализ изменчивости количественных признаков хвои сосны обыкновенной (*Pinus sylvestris* L.) в связи с дифференциацией популяций. Диссертация на соискание ученой степени кандидата биологических наук, 03.02.08 – Экология, Министерство образования и науки российской федерации, Федеральное государственное бюджетное образовательное, Учреждение высшего профессионального образования, «Вятская государственная сельскохозяйственная академия», Киров, 146 стр.
281. Alexandrov A. H., Dobrev R. D. (2014). The State of Forest Genetic Resources in Bulgaria. Sofia, 84 pp. ISBN: 978-3-639-66981-7
282. Šarac Z., Aleksic J. M., Dodoš T., Rajcevic N., Bojovic S., Marin P. D. (2015). Cross-species amplification of nuclear est-microsatellites developed for other *Pinus* species in *Pinus nigra*. *Genetika*, 47 (1): 205-217. DOI: 10.2298/GENSR1501205S

283. Papaioannou A. G. (2015). Ecological and soil conditions of black pine (*Pinus nigra* Arn.) stands in the area of the Russian Monastery at Mount Athos. Russian Journal of Ecology, 46 (5): 438-443. ISSN 1067_4136.
284. Κωστας Ε. Ιωαννίδης (2018). Γενετική βελτίωση πολλαπλών γνωρισμάτων στη μαυρη πευκή (*Pinus nigra* Arn.). Διδακτορική Διατριβή. ΑΡΙΣΤΟΤΕΛΕΙΟ ΠΑΝΕΠΙΣΤΗΜΙΟ ΘΕΣΣΑΛΟΝΙΚΗΣ ΣΧΟΛΗ ΓΕΩΠΟΝΙΑΣ, ΔΑΣΟΛΟΓΙΑΣ ΚΑΙ ΦΥΣΙΚΟΥ ΠΕΡΙΒΑΛΛΟΝΤΟΣ ΤΜΗΜΑ ΔΑΣΟΛΟΓΙΑΣ ΚΑΙ ΦΥΣΙΚΟΥ ΠΕΡΙΒΑΛΛΟΝΤΟΣ, Thessaloniki, 242 pp.
285. Momchil Panayotov, Nickolay Tsvetanov, Evgeni Tsavkov, Georgi Gogushev, Peter Bebi, Petar Zhelev, and Stefan Yurukov (2019). Effect of Climate Change on the High-Mountain Tree Species and Their Genetic Resources in Bulgaria. Springer Nature Switzerland AG 2019. In: M. Šijačić-Nikolić et al. (Eds.), Forests of Southeast Europe Under a Changing Climate, Advances in Global Change Research 65, https://doi.org/10.1007/978-3-319-95267-3_35
286. Przybylski P., Masternak K., Jastrzębowski S. (2020). Isozyme polymorphism and seed and cone variability of Scots pine (*Pinus sylvestris* L.) in relation to local environments in Poland. Folia Forestalia Polonica, Series A – Forestry, Vol. 62 (2), 88–99. ISSN: 0071-6677, eISSN: 2199-5907. DOI: 10.2478/ffp-2020-0010
287. Ράπτης Δημήτριος, Καζάνα Βασιλική, Καζακλής Άγγελος, Σταματίου Χρήστος (2020). Ανάπτυξη γραμμικού μοντέλου μικτών επιδράσεων για την πρόβλεψη της διαμέτρου της κόμης ομήλικων συστάδων μαύρης Πεύκης (*Pinus nigra* Arn.) στην περιοχή του Εθνικού Δρυμού της Πίνδου. Γεωτεχνικά επιστημονικά θέματα - Σειρά VI – Τόμος 29 – Τεύχος 1: 4-13. ISSN 1105-9478 (*Raptis D., Kazana V., Kazaklis A., Stamatiou Ch. (2020). Development of a linear mixed effect crown width model for Black Pine (Pinus nigra Arn.) even aged natural forest stands in Pindos National Park Raptis. Geotechnical Scientific Issues. Geotechnical Chamber Of Greece, Vol. 29, Issue VI - No 1: 4-13. ISSN 1105-9478*)
288. Yetkin Usta, Ertuğrul Bilgili Kadir, Alperen Coşkuner (2020). Karaçam ekosistemlerinde orman yangınlarının ekolojik rolü: 252-272. https://www.researchgate.net/profile/Yetkin-Usta/publication/371911618_Karacam_Ekosistemlerinde_Orman_Yanginlarinin_Ekolojik_Rolu/links/649bdf0295bbbe0c6efc140b/Karacam-Ekosistemlerinde-Orman-Yanginlarinin-Ekolojik-Rolue.pdf
289. S. Versace, A. Bräuning, P. Cherubini M. DiFebbraro, M. Häusser, F. Lombardi, M. Marchetti, P. A. Marziliano, F. Salbitano, S. Szymczak, R. Tognetti. (2022). New evidence for population-specific responses to drought events from tree ring chronologies of *Pinus nigra* ssp. *laricio* across the entire distribution range. Agricultural and Forest Meteorology, Volume 323: 109076. ISSN: 0168-1923. <https://doi.org/10.1016/j.agrformet.2022.109076>

Публикация: Iliev I., Kitin P., Scaltsoyiannes A., Tsaktsira M., Corneanu M., Nellas C. (2009). Comparative study of the variability of *Betula pendula* Roth cultivars during the propagation *in vitro*. In: Soare M., Călina A., Panzaru Radu L., Niculescu M., Alexandru T., Stancu I., Cola M., Netiou C., Dimitru I. (Eds). Analele Universitatii din Craiova, vol. XXXIX/B : 163-171. ISSN 1841-8317. https://cis01.central.ucv.ro/analele_universitatii/agricultura/2009/vol_XXXIX_B_2009.pdf

Цитирана е:

290. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.

291. Gogoi G., Borua P. K. (2014). Standardization parameters for critical problems encountered in plant *in vitro* culture technique. International Journal of Current Research. 6, (12): 10964-10973. <http://www.journalcra.com>
292. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8
293. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Corneanu M., Corneanu G., Iliev I., Danci O., Stefanescu I., Popa M. (2010). Micropropagation of *Robinia pseudoacacia* var. *oltenica* selected stress resistant clones on media with deuterium depleted water. Journal of Horticulture, Forestry and Biotechnology, 14 (1): 141-144. ISSN: 2066-1797. https://journal-hfb.usab-tm.ro/romana/Lista%20lucrari%202010_PDF/JHFB%20-%20vol.1/29Corneanu%20Mihaela.pdf

Цитирани в:

294. Enescu C. M., Dănescu. A. (2013). Black locust (*Robinia pseudoacacia* L.) – an invasive neophyte in the conventional land reclamation flora in Romania. Bulletin of the Transilvania University of Braşov, Series II: Forestry • Wood Industry • Agricultural Food Engineering, Vol. 6 (55) No. 2: 23-30.
295. Ruben B., Adrian T. (2013). Study regarding the diameter at the base ring of some acacia varieties in the first years of vegetation. Analele Univ. din Oradea, Fascicula Protectia Mediului, vol. XXI: 329-333.
296. Petruş-Vancea A., Fordon A. (2015). Deuterium depleted water effect on *Euphorbia canariensis* L. micropropagation Studia Universitatis Babeş-Bolyai Biologia, LX, 2: 31-38.
297. Ciuvăt A. L., Abrudan I. V., Ciuvăt C. G., Marcu C., Lorent A., Dincă L., Szilard B. (2022). Black locust (*Robinia pseudoacacia* L.) in Romanian forestry. Diversity 2022, 14, 780. ISSN 1424-2818. <https://doi.org/10.3390/d14100780>
298. Ábri T., Cseke K., Keserű Z., Porcsin A., Szabó F. M., Rédei K. (2023). Breeding and improvement of black locust (*Robinia pseudoacacia* L.) with a special focus on Hungary: a review. iForest 16: 290-298. ISSN: 1971-7458. doi: 10.3832/ifor4254-016.

Публикация: Dancheva D., Iliev I., Scaltsoyiannes A., Tsaktsitra M., Tsoulpha P. (2010). Effect of medium on *in vitro* germination of embryos of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, Tom XXVI/1: 34-38. ISSN: 1454-6914. https://www.researchgate.net/publication/312937948_EFFECT_OF_MEDIUM_ON_I_N_VITRO_GERMINATION_OF_EMBRYOS_OF_FRAXINUS#fullTextFileContent https://biozoojournals.ro/oscsn/cont/26_1/B06-Dancheva.pdf

Цитирани в:

299. Yilmaz M., Tonguc F. (2013). Dormancy and dormancy-breaking pretreatments in seeds of *Fraxinus ornus* subsp. *Cilicica*. Propagation of Ornamental Plants, 13 (1): 40-45. ISSN: 1311-9109.
300. Yao D., Jin Y., Liu W., Wang X., Guo H., Xie X. (2013). Plant regeneration from mature zygotic embryo explants of *Acacia crassicaarpa* A. Cunn ex Benth. via adventitious shoots. Propagation of Ornamental Plants, 13 (2): 86-92. ISSN: 1311-9109.
301. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
302. Кудряшова О. А., Чалей А. В., Буй А. В., Луконина Ю. Д., Водчиц Н. В., Борисевич Т. А., Волотович А. А. (2017). Сравнительный анализ асептического введения и

стабілізації *in vitro* різних типів експлантів ясеня обыкновенного. Веснік Палескага дзяржаўнага універсітэта. Серыя прыродазнаўчых навук: навучна-практычны журнал. - Пінск: ПолесГУ, № 2: 14-21.
<https://rep.polessu.by/handle/123456789/13188> (BAK)

303. Xu L., Cheng F., and Zhong Y. (2021). Micropropagation of *Paeonia ostii* 'Feng Dan' via *in vitro* mature embryo culture. Propagation of Ornamental Plants, 3: 78-87. ISSN: 1311-9109.

Публикация: Iliev I., Scaltsoyiannes A., Tsaktsira M., Gajdosova A. (2010). Micropropagation of *Betula pendula* Roth cultivars by adventitious shoot induction from leaf callus. Acta Horticulturae, 885: 161-173. ISBN: 90 6605 210 4, ISSN: 0567-7572.

DOI: https://www.researchgate.net/publication/284350138_Micropropagation_of_Betula_pendula_roth_cultivars_by_adventitious_shoot_induction_from_leaf_callus

Цитирана е:

304. Alexandrov A., Dobrev R. (2011). Country Report for The State of World's Forest Genetic Resources – Bulgaria, 63 pp.
305. Šedivá J., Vejsadová H., Vlašínová H., Mertelík J., Kloudová K. (2011). Způsoby *in vitro* regenerace u *Aesculus hippocastanum*. Acta Pruhoniana 99: 127–130. ISBN 978-80-85116-89-2 (VÚKOZ, v.v.i. Průhonice), ISBN 978-80-7415-053-1 (Nová Tiskárna Pelhřimov, s.r.o. Pelhřimov), ISSN 0374-5651
306. Haviček M., Krejčíková B., Chrudina Z., Borovec R., Svoboda J. (2011). Změny ve využití krajiny na vodních tocích v povodí veličky a v horních povodích kyjovská svratky. Acta Pruhoniana, 99: 5-17.
307. Zhelev P., Angelov V. (2012). Variation in a silver birch locality near Ardino (eastern rhodopes). Forestry Ideas, 18, 2 (44): 125-131. ISSN: 1314-3905 (Print), ISSN: 26-03-2996 (Online). https://forestry-ideas.info/issues/issues_Index.php?journalFilter=45
308. Šedivá J. (2012). *In vitro* root formation in *Pulsatilla vernalis* (L.) Mill. Propagation of Ornamental Plants, 12: 96-101. ISSN: 1311-9109.
309. Alexandrov A., Dobrev R. (2012). State of forest genetic resources in Bulgaria. Forest Science, No 1/2: 2-22.
310. Li N., Shu X., Tang S., Tang G., Lu X. (2013). *In vitro* propagation of Nanjing linden (*Tilia miquellana* Maxim.). Propagation of Ornamental Plants, 13 (4): 168-173. ISSN: 1311-9109.
311. Gogoi G., P. K Borua P. K. (2014). Standardization parameters for critical problems encountered in plant *in vitro* culture technique. International Journal of Current Research, 6 (12): 10964-10973. ISSN: 0975-833X. <http://www.journalcra.com>
312. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.
313. Томов В. (2015). Анализ на семеипроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.
314. Świerczyński S., Malinowska A., Golcz-Polaszewska M. (2017). Porównanie wzrostu drzew brzozy brodawkowatej (*Betula pendula* Roth) odmiany 'Purpurea' rozmnażanych przez szczepienie oraz metodą kultur *in vitro*. Nauka Przyroda Technologie, 11 (2): 197-205. ISSN: 1897-7820. <http://dx.doi.org/10.17306/J.NPT.00183>.
315. Mashkina O. S., Tabatskaya T. M. (2020). Morphogenesis of a dissected birch leaf *in vitro* culture. Russian Journal of Developmental Biology, 51: 397-409. ISSN: 1062-3604 <https://doi.org/10.1134/S1062360420060053>

316. Машкина О., Табацкая Т. (2020). Морфогенез раззеченного листа березы в культуре *in vitro*. Онтогенез, 51 (6): 460-472. ISSN (Print): 0475-1450. УДК 575:581.143.6:582.632.1
http://ontogenez.org/archive/2020/6/%D0%9C%D0%B0%D1%88%D0%BA%D0%B8%D0%BD%D0%B0_6_2020.pdf
317. Gaidamashvili M., Benelli G. (2021). Threatened woody plants of Georgia and micropropagation as a tool for *in vitro* conservation. Agronomy 11, Article 1082. ISSN: 2073-4395. <https://doi.org/10.3390/agronomy11061082>.
<https://www.mdpi.com/journal/agronomy>
318. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). Science Archeives, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>
319. Sergeev R., Kleva O., Timakov A., Zontikov D., Khusainova A. (2022). Features of *Betula* spp. cultivation through *in vitro* culture. IOP Conf. Ser.: Earth Environment Science 949 012007, doi:10.1088/1755-1315/949/1/012007

Публикация: Gajdošová A., Libiaková G., Iliev I., Hricová A. (2013). Adventitious shoots induction of *Amaranthus cruentus* L. *in vitro*. Propagation of Ornamental Plants, 13 (1): 33-39. ISSN: 1311-9109. [HTTPS://WWW.JOURNAL-POP.ORG/2013_13_1_33-39.HTML](https://www.journal-pop.org/2013_13_1_33-39.html)

Цитирана в:

320. Kumar S. S., Aruna M. C., Parvatam Giridhar P. (2016). Improvement of Green Leafy Vegetables: The Role of Plant Tissue Culture and Biotechnology. In: Anis M., Ahmad N. (Eds). Plant Tissue Culture: Propagation, Conservation and Crop Improvement, Springer: 547-582. Print ISBN978-981-10-1916-6, Online ISBN978-981-10-1917-3, https://doi.org/10.1007/978-981-10-1917-3_24
321. Kiryu K. (2019). Evaluating the physiological potential of amaranth seeds. Caribbean Journal of Science, Vol. 52, No. 1: 491-494. Print ISSN: 0008-6452, Online ISSN: 0008-6452
322. Yaroshko O. (2021). Achievements in genetic engineering of *Amaranthus* L. representatives. International Journal of Secondary Metabolite, 8 (2): 172–185 e-ISSN: 2148-6905, <https://dx.doi.org/10.21448/ijsm.925737>, <https://dergipark.org.tr/en/pub/ijsm>
323. Magali Ruíz-Rivas, Catarino Eduardo Téllez-Valerio, Marcelino Martínez-Núñez, Pedro Fernando Vera-Hernández, Esperanza Martínez-Romero, Flor de Fátima Rosas-Cárdenas (2022). Influencia de la luz en la generación de callos y el cultivo *in vitro* de plantas. Revista Mexicana de Ciencias Agrícolas. Special publication number 27 August 15 - September 30: 11-21. ISSN en línea 2007-9230, ISSN impreso 2007-2934. <https://cienciasagricolas.inifap.gob.mx/index.php/agricolas/article/view/3156/5090>
324. Anuradha, Manisha Kumari, Gaurav Zinta, Ramesh Chauhan, Ashok Kumar, Sanatsujat Singh and Satbeer Singh (2023). Genetic resources and breeding approaches for improvement of amaranth (*Amaranthus* spp.) and quinoa (*Chenopodium quinoa*). Frontiers in Nutrition, 10: 1129723. ISSN: 2296-861X (Online). doi: 10.3389/fnut.2023.1129723
325. Magali Ruíz-Rivas, Catarino Eduardo Téllez-Valerio, Marcelino Martínez-Núñez, Pedro Fernando Vera-Hernández, Esperanza Martínez-Romero, Flor de Fátima Rosas-Cárdenas (2022). Influence of light on callus generation and *in vitro* plant culture. Revista Mexicana de Ciencias Agrícolas. Special publication number 27: 11-21. ISSN: 2007-0934, ISSN-e: 2007-9230. <https://cienciasagricolas.inifap.gob.mx/index.php/agricolas/article/view/3156/5091>

Публикация: Iliev N., Dancheva D., Iliev I. (2013). Effect of selective herbicides in the production of turkey oak seedlings (*Quercus cerris* L.). Oltenia Journal for Studies in

Цитирана е:

326. Laclau P., Murilo N. L., Bertoli B., Osvaldo V. (2020). Tolerance of pedunculate oak (*Quercus robur*) saplings to herbicides tolerancia de plantines de roble pedunculado (*Quercus robur*) a herbicidas. Revista de Investigaciones Agropecuarias, 46 (3): 387-396. ISSN en linea 1669-2314, ISSN impreso 0325-8718, <http://ppct.caicyt.gov.ar/index.php/ria/article/view/15358/45454575771041>
327. Савев С. (2023). Култивиране на висока американска боровинка в горските територии. Рецензенти: Петкова К., Стоянова М. Издателство Интел Ентранс, 174 стр. ISBN: 978-619-7703-16-0.

Публикация: Dancheva D., Iliev N., Iliev I. (2013). *In vitro* propagation of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, 29 (1): 78-84. P-ISSN: 1454-6914. https://www.researchgate.net/publication/312937765_IN_VITRO_PROPAGATION_OF_Fraxinus_excelsior_L_DANCHEVA_Desislava_ILIEV_Nasko_ILIEV_Ivan

Цитирана е:

328. Islami Rostami H. R., Hosseini Nasr S. M., Kazemitabar S. K., F. Zafarian (2019). Effect of provenances and culture media on seed germination of ash (*Fraxinus excelsior* L.) in embryo *in vitro* culture. Iranian Journal of Forest and Poplar Research, 27 (2): 159-168. Printing ISSN: 1735-0883, Electronic ISSN: 2383-1146. DOI: 10.22092/IJFPR.2019.120122
329. Nawrot-Chorabik K., Pluciński B., Latowski D. (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. New Forests. Electronic ISSN: 1573-5095, Print ISSN: 0169-4286. <https://doi.org/10.1007/s11056-023-09981-x>

Публикация: Lyubomirova T., Iliev I. (2013). *In vitro* propagation of *Syringa vulgaris* L. Forestry Ideas. 19: 173-185. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online). https://forestry-ideas.info/files/issue/Forestry_Ideas_BG_2013_19_2_7.pdf

Цитирана е:

330. Dancheva D. (2014). Rooting and acclimatization of micropropagated shoots of *Fraxinus excelsior* L. Oltenia Journal for Studies in Natural Sciences, vol. 30 (1): 75-80. ISSN 1454-6914
331. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
332. Pczuk A., Jagiełło-Kubiec K. (2015). The effect of plant growth regulators and sucrose on the micropropagation of common lilac (*Syringa vulgaris* L.). Annals of Warsaw University of Life Sciences – SGGW, Horticulture and Landscape Architecture No 36, 2015: 3-12.
333. Жакова С. Н. (2015). Репродуктивная биология некоторых видов и культиваров рода сирень (*Syringa* L.). Диссертации на соискание ученой степени кандидата биологических наук, Пермский государственный национальный исследовательский университет, Пермь, 03.02.01 – ботаника, Биологические науки, Д 212.189.02, 196 стр. <https://www.dissercat.com/content/reproduktivnaya-biologiya-nekotorykh-vidov-i-kultivarov-roda-siren-syringa>
334. Tomov V. (2017). Rooting of Norway maple (*Acer platanoides* L.) cuttings. Forestry Ideas, 23, No 1 (53): 57-64.
335. Borsai Orsolya, Doina Clapa, Alexandru Fira, Monica Hârta, Adelina Dumitraș, Rodica Pop, Doru Pamfil (2017). The effect of gelling agent on the micropropagation of common lilac (*Syringa vulgaris* L.). Agriculture - Science and Practice, no. 3 – 4 (103-104): 63-71.

336. Лободина Е. В., Супрун И. И., Леонидовна Е. Т., Анатольевна Е. Б. (2020). Влияние сроков отбора эксплантов сирени (*Syringa vulgaris* L.) на жизнеспособность и контаминацию при введении в культуру *in vitro*. Плодоводство и виноградарство Юга России, № 61(1): 98-107. DOI 10.30679/2219-5335-2020-1-61-98-107. <http://journalkubansad.ru/pdf/20/01/08.pdf>

Публикация: Lyubomirova T., Iliev I. (2014). Factors affecting the rooting of cuttings of *Syringa vulgaris* L. cultivars. Oltenia Journal for Studies in Natural Sciences, 30 (1): 67-74. P-ISSN: 1454-6914. https://biozoojournals.ro/oscsn/cont/30_1/11_Lyubomirova_pp67-74.pdf

Ц и т и р а н а в:

337. Erken K., Özzambak M. E. (2020). Effects of cutting times and doses of some auxins on rooting of the hairy broom cuttings (*Chamaecytisus hirsutus* (L.) LINK). International Journal of Agricultural and Natural Sciences 13(2): 53-61. E-ISSN: 2651-3617. <https://www.ijans.org/index.php/ijans/article/view/495/476>

Публикация: Tomov V., Iliev N., Iliev I. (2014). Analyzis of the forestry seed production base of *Acer platanoides* L. in Bulgaria. Forestry Ideas, 20, 1 (47): 67-76. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online). file:///C:/Users/ivan/Downloads/Forestry_Ideas_BG_2014_20_1_5.pdf

Ц и т и р а н а в:

338. Качова В., Донкова Р., Попов Е., Стефанова П. (2017). Изменения в микробиологичните съобщества на почвата след прилагане на органични почвени подобрители при отглеждане на фиданки от шестил. Наука за Гората, 2: 59-68.

339. Kachova V. (2018). Growth increment of juvenile *Acer platanoides* L. and soil organic matter after application of organic fertilizers. Natural Resources and Sustainable Development, 8 (2): 166-174. eISSN: 2601-5676, pISSN: 2066-6276. DOI: 10.31924/nrsd.v8i2.017

340. Kachova V. (2020). Effect of organic and mineral fertilizers on the growth of *Acer platanoides* L. Bulgarian Journal of Soil Science, Volume 5, Issue 1: 50-63. (ISSN-online 2367-9212, ISSN-print 2534-8787). https://www.bsss.bg/issues/Issue1_2020/BJSS_2020_1_5.pdf. www.bsss.bg

341. Budău R., Apăfăian A., Caradaică M., Bratu I. A., Timofte C. S. C., Enescu C. M. (2023). Expert-Based Assessment of the Potential of Agroforestry Systems in Plain Regions across Bihor County, Western Romania. Sustainability, 15, EISSN 2071-1050. 15724. <https://doi.org/10.3390/su152215724>

Публикация: Dancheva D., Iliev I. (2015). Factors affecting adventitious shoot formation in *Fraxinus excelsior* L. Propagation of Ornamental Plants, 15 (1): 10-20. ISSN: 1311-9109. https://www.journal-pop.org/2015_15_1_10-20.html

Ц и т и р а н а в:

342. Lebedev V., Shestibratov K. (2016). Large-scale micropropagation of common ash. Biotechnology. ISSN 1682-296X. DOI: 10.3923/biotech.: 1-9

343. West T. P., Coronado R. F. (2016). Micropropagation of *Ostrya virginiana* (Mill.) K. Kock. An underutilized ornamental landscape tree. Propagation of Ornamental Plants, 16 (2): 56-61. ISSN: 1311-9109.

344. Kim D. H., Kang K. W., Sivanesan I. (2017). Micropropagation of *Haworthia retusa* Duval. Propagation of Ornamental Plants, 17 (3): 77-82. ISSN: 1311-9109.

345. Pacholczak A., Nowakowska K. (2019). Micropropagation of February daphne (*Daphne mezereum* L.). Propagation of Ornamental Plants, 19: 106-112. ISSN: 1311-9109.

346. Fenning T., O'Donnell M., Preedy K., Bézanger A., Kenyon D., Lopez G. (2022). The rooting ability of *in vitro* shoot cultures established from a UK collection of the common ash (*Fraxinus excelsior* L.) and their *ex vitro* survival. *Annals of Forest Science*, 79: 30. ISSN: 1286-4560. <https://doi.org/10.1186/s13595-022-01146-8>
347. Nawrot-Chorabik K., Pluciński B., Latowski D. (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. *New Forests*. Electronic ISSN: 1573-5095. Print ISSN: 0169-4286. Electronic ISSN: 1573-5095. <https://doi.org/10.1007/s11056-023-09981-x>.
348. Nowakowska K., Nongdam P., Samsurizal N. A., Pacholczak A. (2023). An efficient micropropagation protocol for the endangered European shrub February Daphne (*Daphne mezereum* L.) and identification of bacteria in culture. *Agriculture*, 13, 1692. ISSN 2077-0472. <https://doi.org/10.3390/agriculture13091692>

Публикация: Iliev I. (2017). Factors affecting the axillary and adventitious shoots formation in woody plants *in vitro*. Acta Horticulturae, 1155: 15-27. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана е:

349. Antonić D., Trajković M., Cingel A., Subotić A., Jevremović S. (2017). Plant regeneration from *in vitro*-derived leaf and petiole explants of *Viola cornuta* L. 'Lutea Splendens'. *Propagation of Ornamental Plants*, 17 (3): 95-102. ISSN: 1311-9109.
350. Zarate-Díaz Y., Barba-Gonzalez R., Aguilera-Cauich E. A., Tapia-Campos E., Herrera-Cool G. J., Uc-Varguez A., Lopez-Puc G. (2019). *In vitro* organogenic behavior and molecular characterization of *Eustoma exaltatum* × *Eustoma grandiflorum* hybrids. *Propagation of Ornamental Plants*, 19 (4): 118-124. ISSN: 1311-9109.
351. Anis Adilah Mustafa, Mohammad Rahmat Derise, Wilson Thau Lym Yong, Kenneth Francis Rodrigues (2021). A Concise Review of *Dendrocalamus asper* and Related Bamboos: Germplasm Conservation, Propagation and Molecular Biology. *Plants*, 10, 1897. ISSN: 2223-7747. <https://doi.org/10.3390/plants10091897>
352. Burrows G. E. (2021). Gymnosperm resprouting - a review. *Plants*, 10, Article 2551. ISSN: 2223-7747. <https://doi.org/10.3390/plants10122551>
353. Xingmei Ai, Yonghui Wen, Chao Wang. (2021). *In Vitro* propagation and rejuvenation of senescent maternal plant of *Ardisia crenata* var. Bicolor (Primulaceae). *Research Square*. DOI:10.21203/rs.3.rs-992973/v1
354. Duc Tran Minh, Quynh Thi Nguyen, Thanh Pham (2021). Regeneration of plants via callus-mediated organogenesis from leaf, petiole and stem explants of *Ardisia silvestris* Pitard. *Propagation of Ornamental Plants*, 21 (3): 96-103. ISSN: 1311-9109.
355. Maurizio Capuana, Werther Guidi Nissim and Joshua D. Klein. (2022). Protocol for In Vitro Propagation of *Salix acmophylla* (Boiss.). *Studies on Three Ecotypes*. *Forests*, 13, 1124. ISSN: 1999-4907. <https://doi.org/10.3390/f13071124>, <https://www.mdpi.com/journal/forests>
356. Miaomiao Guo, Qiuying Yu, Daijun Li, Kexin Xu, Zexin Di, Yong Zhang, Yang Yu, Jian Zheng and Yan Zhang (2023). *In vitro* propagation, shoot regeneration, callus induction, and suspension from lamina explants of *Sorbus caloneura*. *Forestry Research*, 3: 7. <https://doi.org/10.48130/FR-2023-0007>. ISSN 2767-3812.
357. Pham Thi Diem Thi, Nguyen Thi Nguyen Man, Nguyen Thi Khanh Quynh, Trieu Thy Hoa, Pham Mai Thu Thuy, Hoang Tan Quang (2023). Micropropagation of long-leaved paperbark (*Melaleuca leucadendron* (L.) L.). *Propagation of Ornamental Plants*, 23: 91-98. ISSN: 1311-9109.

Публикация: Tsaktsira M., Alevropoulos A., Tsoulpha P., Scaltsoyiannes V., Scaltsoyiannes A., Iliev I. (2018). Inter- intra- genetic variation on rooting ability of *Ilex aquifolium* L. varieties and cultivars. *Propagation of Ornamental Plants*, 18 (4): 131-138. ISSN: 1311-9109. ISSN: 1311-9109. https://www.journal-pop.org/2018_18_4_131-138.html

Цитирани в:

358. Wang M., Hu J., Guo G., Park Y. G., Jeong B. R. (2021). Effect of auxins and their concentrations, immersion time, and rooting medium on rooting of cutting-propagated *Pyracantha angustifolia* C. K. Schneid. *Propagation of Ornamental Plants*, 21 (1): 3-10. ISSN: 1311-9109.
359. Khater N., Benahmed A., Zereg N., Cherouana K. (2021). Callogenesis induction of *Ilex aquifolium* L. (Aquifoliales Aquifoliaceae). *Biodiversity Journal*, 12 (1): 265-272. ISSN: 2039-0408 (Online). <https://doi.org/10.31396/Biodiv.Jour.2021.12.1.265.272>
360. Wang X., Lv Y., Xing S., Liu G., Sun J., Wang Y., Wang C., Yu X. (2021). Study on the efficient cutting propagation technology for *Ilex* “China Girl”. *American Journal of Plant Sciences*, 12: 1459-1467. ISSN Online: 2158-2750 ISSN Print: 2158-2742. <https://www.scirp.org/journal/ajps>
361. Kentelky E., Jucan D., Cantor M., Szekely-Varga Z. (2021). Efficacy of different concentrations of NAA on selected ornamental woody shrubs cuttings. *Horticulturae*, 7, 464. ISSN: 2311-7524. <https://doi.org/10.3390/horticulturae7110464>, file:///C:/Users/ivan/Downloads/Efficacy_of_Different_Concentrations_of_NAA_on_Sel.pdf

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Nianiou-Obeidat, Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* ‘Agrifoglio Commune’. *Sustainability*, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирани в:

362. Kentelky E., Jucan D., Cantor M., Szekely-Varga Z. (2021). Efficacy of Different Concentrations of NAA on Selected Ornamental Woody Shrubs Cuttings. *Horticulturae*, 7, 464. ISSN: 2311-7524. <https://doi.org/10.3390/horticulturae7110464>, file:///C:/Users/ivan/Downloads/Efficacy_of_Different_Concentrations_of_NAA_on_Sel.pdf
363. Loconsole D., Cristiano G., De Lucia B. (2022). Image analysis of adventitious root quality in wild sage and glossy abelia cuttings after application of different indole-3-butyric acid concentrations. *Plants* 2022, 11, Article 290. <https://doi.org/10.3390/plants11030290> <https://www.mdpi.com/journal/plants>
364. Aithida A., Tahiri A., Azlay L., L. Amina Idrissi Hassani, Mokhtari M. (2022). Effects of cutting origin and exogenous auxin treatment on the rooting of *Rosa damascena* (Mill) cuttings from the M’goun-Dades valleys in Morocco. *Arabian Journal of Medicinal & Aromatic Plants*, V8N1: 134-155. ISSN 2458-5920, file:///C:/Users/ivan/Downloads/30674-81635-1-PB.pdf, DOI: 10.48347/IMIST.PRSM/ajmap-v8i1.30674
365. Thi Kim Phuong Tran, Minh Hac Pham, Thi Huong Trinh, Sasanti Widiarsih, Viet The Ho (2022). Investigation of the genetic diversity of jewel orchid in Vietnam using RAPD and ISSR markers (2022). *Biodiversitas*, Volume 23, Number 9: 4816-4825. ISSN: 1412-033X, E-ISSN: 2085-4722, DOI: 10.13057/biodiv/d230950
366. Siti Sofiah, Luchman Hakim, Serafinah Indriyani, Iyan Robiansyah (2022). Acclimation study of *Smilax nageliana* A.DC., a climber species endemic to East Java, Indonesia.

Biodiversitas, 23 (8): 4082-4089. E-ISSN: 2085-4722 ISSN: 1412-033X, DOI: 10.13057/biodiv/d230828

367. Pipinis E., Kostas S., Hatzilazarou S., Petropoulos S., Mitsi D., Stampoulidis A., Milios E., Smiris P. (2023). Effects of donor tree age, cutting collection time and K-IBA application on rooting ability of *Taxus baccata* L. stem cuttings: preliminary results. *Folia Oecologica*, 50 (1): 97-103, ISSN: 1338-7014. doi: 10.2478/foecol-2023-0009 (SCOPUS)
368. Huanyu Chen, Lizhou Hong, Angyan Ren, Kai Yu, Kai Wang, Sunan He, Chong Liu, Jincheng Xing (2023). Growth regulators on the shooting and rooting of *Tamarix chinensis* stem cuttings. *Rhizosphere*, 100679, ISSN: 2452-2198, <https://doi.org/10.1016/j.rhisph.2023.100679>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. *Plant Growth Regulation*, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Цитирани в:

369. Bairu M. W., Kane M. E. (2011). Physiological and developmental problems by *in vitro* cultured plants. *Plant Growth Regulation*, 62 (2): 101-103. DOI 10.1007/s10725-011-9565-2
370. Fujita H., Kawaguchi M. (2011). Strategy for shoot meristem proliferation in plants. *Plant Signaling & Behavior*, 6 (11): 1851-1854. ISSN: (Print) 1559-2324 (Online), DOI: 10.4161/psb.6.11.17656, <https://doi.org/10.4161/psb.6.11.17656>
371. Jin-Hu Wu, A. Ross Ferguson, Brian G. Murray, Yilin Jia, Paul M. Datson, Jingli Zhang (2012). Induced polyploidy dramatically increases the size and alters the shape of fruit in *Actinidia chinensis*. *Annals of Botany*, 109 (1): 169-179. doi:10.1093/aob/mcr256
372. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.
373. Fu C., Li L., Wu W., Li M., Yu X., Yu L. (2012). Assessment of genetic and epigenetic variation during long-term *Taxus* cell culture. *Plant Cell Reports*, 31 (7): 1321-1331. DOI 10.1007/s00299-012-1251-y
374. Майоров С. Н., Маолчанова А. В., Бондарева Л. Л., Старцев В. И. (2012). Типы фасциаций и факторы, влияющие на её проявление. *Овощи России*, 2 (15): 54-59. УДК 631.524.8, ISSN: 2072-9146, <https://readera.ru/14024947>
375. Fujita H., Kawaguchi M. (2013). Pattern formation by two-layer Turing system with complementary synthesis. *Journal of Theoretical Biology*, 322: 33-45. <http://dx.doi.org/10.1016/j.jtbi.2013.01.008>
376. Bhojwani S. S., Dantu P. K. (2013). Micropropagation. *In: Plant Tissue Culture: an introductory text*. Springer: 245-274. ISBN 978-81-322-1025-2, ISBN 978-81-322-1026-9 (eBook), DOI 10.1007/978-81-322-1026-9
377. Gupta S., Kaliamoorthy S., Mao A. A., Sarma S. (2013). *In vitro* regeneration of *Gymnocladus assamicus* Kanjilal ex P.C. Kanjilal, an ethnomedical critically endangered and endemic tree species from Northeast India. *Int. J. Med. Arom. Plants*, 3 (2): 204-213. ISSN: 2249-4340. <http://creativecommons.org/licenses/by-nc-nd/3.0>, <http://www.accessscience.com>
378. Ruffoni B., Savona M. (2013). Physiological and biochemical analysis of growth abnormalities associated with plant tissue culture. *Horticulture, Environment, and Biotechnology*, 54 (3): 191-205. ISSN: 22113460, 22113452. DOI 10.1007/s13580-013-0009-y
379. El-Banna A. N., El-Nady M. F., Dewir Y. H., El-Mahrouk M. E. (2013). Stem fasciation in cacti and succulent species — tissue anatomy, protein pattern and RAPD polymorphisms. *Acta Biologica Hungarica*, 64 (3): 305-318. DOI: 10.1556/ABiol.64.2013.3.4

380. Reed B. M., Wada S., DeNoma J., Niedz R. P. (2013). Mineral nutrition influences physiological responses of pear *in vitro*. In *Vitro Cellular & Developmental Biology - Plant*, 49 (6): 699-709. ISSN: 1054-5476 (print); 1475-2689 (web). DOI 10.1007/s11627-013-9556-2
381. Maliarenko V. M., Mudrak T. P. (2013). Cactus viruses in fasciated plants. *Biologija*, 59 (2): 213-218.
382. Arellano-Perusquia A., Lopez-Peralta M. C. G., Chable-Moreno F., Estrada-Luna A. A. (2013). Effect of growth regulators on the organogenesis and multiplication of *Ortegocactus macdougallii* Alexander. *Propagation of Ornamental Plants*, 13 (4): 160-167. ISSN: 1311-9109.
383. Pavlovic N. B., Korte M. K., Kathryn McEachern A. K., Grunde R. (2013). First report of fasciation in pitcher's thistle, *Cirsium pitcheri* (Asteraceae). *The Michigan Botanist*, 52: 52-66.
384. Omar A. F., Dewir Y. H., El-Mahrouk M. E. (2014). Molecular identification of phytoplasmas in fasciated cacti and succulent species and associated hormonal perturbation. *Journal of Plant Interactions*, 9 (1): 632-639. DOI: 10.1080/17429145.2014.882421, <http://dx.doi.org/10.1080/17429145.2014.882421>
385. Chiruvella K. K., Mohammed A., Ghanta R. G. (2014). Phenotypic aberrations during micropropagation of *Soymida febrifuga* (Roxb.) Adr. Juss. *Not Sci Biol*, 6(1): 99-104. www.notulaebiologicae.ro Print ISSN: 2067-3205; Electronic ISBN: 2067-3264
386. Liao Y. K., Ji Y. Y. (2014). Mass propagation of *Drosera burmannii* Vahl via induction of shoot fasciation and recovery of morphologically normal plantlets. *Propagation of Ornamental Plants*, 14 (4): 158-170. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_14_4\(158-170\).pdf](https://www.journal-pop.org/References/Vol_14_4(158-170).pdf)
387. Маляренко В. А., Гайдаржи М. М. (2014). Аномальні форми росту сукулентних рослин та перевірка гіпотез їх походження. *Інтродукція рослин*, № 3: 46-52.
388. Gogoi G., Borua P. K. (2014). Standardization parameters for critical problems encountered in plant *in vitro* culture technique. *International Journal of Current Research*, 6 (12): 10964-10973.
389. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
390. Fujinami R., Imaichi R. (2015). Developmental morphology of flattened shoots in *Dolzellia ubonensis* and *Indodolzellia gracilis* with implications for the evaluation of diversified shoot morphologies in subfamily Tristichoidae (Todostemaceae). *American Journal of Botany*, 102 (6): 848-859. <http://www.amjbot.org/cgi/doi/10.3732/ajb.1500206>, doi:10.3732/ajb.1500206
391. Moyo M., Aremu A. O., Van Staden J. (2015). Insights into the multifaceted application of microscopic techniques in plant tissue culture systems. *Planta*, 242: 773-790. DOI 10.1007/s00425-015-2359-4
392. Isah T. (2015). Adjustments to *in vitro* culture conditions and associated anomalies in plants. *Acta Biologica Cracoviensia, Series Botanica* 57/2, DOI: 10.1515/abcsb-2015-0026
393. Byoung Il Je, Jeremy Gruel, Young Koun Lee, Peter Bommert, Edgar Demesa Arevalo, Andrea L Eveland, Qingyu Wu, Alexander Goldshmidt, Robert Meeley, Madelaine Bartlett, Mai Komatsu, Hajime Sakai, Henrik Jönsson, David Jackson (2016). Signaling from maize organ primordia via fasciated EAR3 regulates stem cell proliferation and yield traits. *Nature of Genetics*, 48: 785-791. doi:10.1038/ng.3567
394. Dewir Y. H. (2016). Cacti and succulent plant species as phytoplasma hosts: a review. *Phytopathogenic Mollicutes Vol. 6* (1): 1-9

395. Цаценко Л. В., Савиченко Д. Л. (2016). Фасциация в природе и эксперименте. Научный журнал КубГАУ, № 123 (09): 1-15. УДК 635.63:631.527 UDC 635.63:631.527 03.00.00, Doi: 10.21515/1990-4665-123-120.
396. Mehta A. (2017). Growth and development of rooted plantlets of *Bacopa monnieri* L. in auxin-free tissue culture medium. International Journal of Botany and Research, 7 (3): 11-16. ISSN(P): 2277-4815; ISSN(E): 2319-4456
397. Nezami-Alanagh E. (2018). Design of suitable culture media for pistachio rootstocks using computer-based tools. Doctoral Dissertation, University de Vigo, EIDO International Doctoral School, 126 pp. + XVI appendixes.
398. Nezami-Alanagh E., Garoosi G.-A., Landín M., Gallego P. P. (2019). Computer-based tools provide new insight into the key factors that cause physiological disorders of pistachio rootstocks cultured *in vitro*. Scientific Reports, 9, Article number: 9740, ISSN: 2045-2322. <https://doi.org/10.1038/s41598-019-46155-2>.
399. Nabieva Alexandra Yurievna, Gerasimovich Lyudmila Vladimirovna (2019). The application of different reproduction techniques for rare species waterlily tulip (*Tulipa kaufmanniana* Regel.) propagation under *ex situ* conditions. Ornamental Horticulture, 25 (4): 450-460. ISSN: 2447536X, <https://ornamentalthorticulture.emnuvens.com.br/rbho/article/viewFile/2035/1620>
400. Liang H., Xiong Y., Guo B., Yan H., Jian Sh., Ren H., Zhang X., Li Y., Zeng S., Wu K., Zheng F., Teixeira da Silva J. A., Xiong Y., Guohua Ma G. (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. Environmental and Experimental Biology, 17: 169–177. ISSN : 2255-9582, DOI: 10.22364/eeb.17.17
401. Brailko V., Ivanova N., Zhdanova I., Mitrofanova O. (2020). Morphological and anatomical features of narrow-leaved lavender plants with prolonged conservation under *in vitro* genebank. BIO Web of Conferences 24, 00015 (2020) International Conferences “Plant Diversity: Status, Trends, Conservation Concept” 2020: 1-5. <https://doi.org/10.1051/bioconf/20202400015>
402. Zheng-Wei Zhang, Xiang-Hua Li, Xu-Dong Wang, Ke-Jing Wang (2020). Discovering adventitious roots in *Glycine tomentella* Hayata: lack of adventitious roots as a morphological and taxonomic species indicator is worth considering. Genetic Resources and Crop Evolution, 67 (6): 1345-1350. ISSN: 1573-5109 (Online). DOI: <https://doi.org/10.1007/s10722-020-00929-0>
403. Pace L., Pellegrini M., Pannunzio G., Gianfranco Pirone G. (2020). First report of fasciation symptom in *Artemisia eriantha* (Asteraceae), a typical orophyte of high-altitude cliffs, in Central Apennines (Italy). Plant Sociology, 57(1): 23-28. ISSN 1120-4605. DOI 10.3897/pls2020571/03
404. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Livia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. Rodriguésia 72: e00372019. 2021 ISSN printed version: 0370-6583, ISSN online version: 2175-7860. <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>
405. Egger J. M. (2021). Stem fasciation in the genus *Castilleja* (Orobanchaceae). Phytoneuron 2021-16: 1-29. ISSN: 2153-733X. <https://www.phytoneuron.net/wp-content/uploads/2021/08/16PhytoN-CastillejaFasciation.pdf>
406. Sriskanda D., Liew Y. X., Khor S. P., Merican F. M. M. S., Sreeramanan Subramaniam, Chew B. L. (2021). An efficient micropropagation protocol for *Ficus carica* cv. Golden Orphan suitable for mass propagation. Biocatalysis and Agricultural Biotechnology, 38 (2021) 102225 ISSN: 1878-8181 (print), <https://doi.org/10.1016/j.bcab.2021.102225>
407. Xincheng Yu, Xiaohong Chen, Yuping Xiong, Yujie Zeng, Zhenpeng Wei, Jinhui Pang, Xinhua Zhang, Yuan Li, Kunlin Wu, Songjun Zeng, Jaime A. Teixeira da

- Silva & Guohua Ma (2022). Shoot organogenesis from leaf and stem explants of *Heliotropium foertherianum* Diane and Hilger. In *Vitro Cellular & Developmental Biology - Plant*. 58(4):559-566 ISSN: 1054-5476 (print); 1475-2689 (web), DOI: <https://doi.org/10.1007/s11627-022-10257-w>
408. Feng Cheng, Mengfei Song, Mengru Zhang, Chunyan Cheng, Jinfeng Chen, Qunfeng Lou (2022). A SNP mutation in the *CsCLAVATA1* leads to pleiotropic variation in plant architecture and fruit morphogenesis in cucumber (*Cucumis sativus* L.). *Plant Science*, 323, 111397. ISSN 0168-9452. <https://doi.org/10.1016/j.plantsci.2022.111397>
409. Aurora M. Nedelcu (2022). Evo-devo perspectives on cancer. *Essays in Biochemistry*, EBC20220041. Online ISSN: 1744-1358. Print ISSN: 0071-1365. <https://doi.org/10.1042/EBC20220041>
410. Manokari M., Priyadharshini S., Shekhawat M. S. (2022). Repairing mechanism of foliar micro-morphological anomalies during acclimatization and field transfer of *in vitro* raised plantlets of *Aerva lanata* (L.) Juss. ex Schult.: a medicinally important plant. *Vegetos*, 35: 520–526. ISSN: 2229-4473, Electronic ISSN: 2229-4473, <https://doi.org/10.1007/s42535-021-00317-8>
411. Sant Saran Bhojwani, Prem Kumar Dantu. (2013). *Plant Tissue Culture: An Introductory Text*, 390 pp. ISBN-13: 978-8132210252, ISBN: 978-81-322-1026-9
412. Manokari M., Cokulraj M., Badhepuri M. K., Abhijit Dey, Mohammad Faisal, Abdulrahman A. Alatar, Rupesh Kumar Singh, Mahipal S. Shekhawat (2023). Microstructural and histochemical modifications in leaves at successive stages of *in vitro* development of the terrestrial orchid *Spathoglottis plicata* Blume. *Horticulture Environment, and Biotechnology*. Vol. 64(3): 497-510. ISSN: 22113460, 22113452, <https://doi.org/10.1007/s13580-022-00485-9>
413. Yadav R. S., Mewada Ketan, R. K Sugoor (2023). A record of stem fasciation phenomenon in *Cocculus hirsutus* (L.). *Zoos' Print Journal*, 38(3): 8-10. ISSN: 0971-6378. <https://zoosprint.zooreach.org/index.php/zp/article/view/7581/6837>
414. Tetsuya K. Matsumoto, Ryohei Fujisato, Munetaka Sugiyama, Yuko Miyazaki, Jin Murata (2023). A malformation of sex-changing plant *Arisaema serratum* (Araceae) produces both male and female inflorescences. *Botany Letters*. ISSN: 2381-8107, Journal homepage: <https://www.tandfonline.com/loi/tabg21>, <https://doi.org/10.1080/23818107.2023.2234973>, DOI: 10.1080/23818107.2023.2234973
415. Yimei Zang, Lei Xie, Jiaxian Su, Zuliang Luo, Xunli Jia, Xiaojun Ma (2023). Advances in DNA methylation and demethylation in medicinal plants: a review. *Molecular Biology Reports*, 50: 7783-7796, ISSN: 0301-4851 (print); 1573-4978 (web). <https://doi.org/10.1007/s11033-023-08618-8>
416. Li K., Tassinari A., Giuliani S., Rosignoli S., Urbany C., Tuberosa R. and Salvi S. (2023). QTL mapping identifies novel major loci for kernel row number-associated ear fasciation, ear prolificacy and tillering in maize (*Zea mays* L.). *Frontiers in Plant Science*, 13: 1017983. ISSN: 1664-462X (Online). doi: 10.3389/fpls.2022.1017983. <https://europepmc.org/backend/ptpmcrender.fcgi?accid=PMC9871824&blobtype=pdf>
417. Iliya Bulavin, A. I. Sidiyakin (2023). Structural and functional characterization of the root of *Arabidopsis thaliana in vitro*. *Biol Bull Russ Acad Sci*, 50(6): 1241-1249. ISSN 1062-3590 https://www.researchgate.net/publication/374847861_Structural_and_Functional_Characterization_of_the_Root_of_Arabidopsis_thaliana_In_Vitro/references

Публикация: Iliev I., Iliev N., Dancheva D., Corneanu M., Tsaktsira M., Gajdošova A., Mladenova S. (2010). Factors affecting the rooting of cuttings from cultivars of *Chamaecyparis lawsoniana* Parl. In: Soare M., Călina A., Panzaru Radu L., Niculescu

M., Alexandru T., Stancu I., Cola M., Netiou C., Dimitru I. (Eds). **Durable Agriculture – Agriculture of the Future. Analele Universitatii din Craiova**, vol. XL, No 2: 174-181. ISSN 1841-8317.
https://www.researchgate.net/publication/340265023_Factors_affecting_the_rooting_of_cuttings_from_cultivars_of_Chamaecyparis_lawsoniana_Parl_In_Soare_M_Calina_A_Panzaru

Цитирани е:

418. Fascella G., Militello M., Carrubba A. (2012). Propagation of *Artemisia arborescens* L. by stem-cutting : adventitious root formation under different conditions. *Propagation of Ornamental Plants*, 12: 171-177. ISSN: 1311-9109.
419. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
420. Szász-Len A-M., Holonec L., Truța A., Reborean F. A. (2015). Effect of cutting size on the rooting of *Thuja occidentalis* ‘Columna’. *Bulletin UASVM Horticulture* 72 (1): 233-234. Print ISSN 1843-5254, Electronic ISSN 1843-5394, DOI: 10.15835/buasvmcn-hort: 10999.
421. Tomov V. (2017). Rooting of norway maple (*Acer platanoides* L.) cuttings. *Forestry Ideas*, 23, No 1 (53): 57-64.
422. Торчик В. И., Келько А. Ф., Холопук Г. А. (2017). Ризогенез у декоративных садовых форм хвойных растений и способы его интенсификации. Рецензенты: Гетко Н. В., Сарнацкий В. В. Минск, Издательский дом „Беларуская навука“, 218 с. УДК [635.92:582.47]:631.535. ISBN: 978-985-08-2103-4.
https://books.google.bg/books?id=GwNTDwAAQBAJ&pg=PA200&lpg=PA200&dq=Chamaecyparis,Iliev+I.&source=bl&ots=B8anI7pz-P&sig=ACfU3U1iYlQ0cjaEMcL_Dw0ssGENzvZj6g&hl=en&sa=X&ved=2ahUKEwjDzbuloJntAhUNlxQKHeESCqM4ChDoATAHegQIBRAC#v=onepage&q&f=true
423. Wang M., Hu J., Guo G., Park Y. G., Byoung Ryong Jeong. (2021). Effect of auxins and their concentrations, immersion time, and rooting medium on rooting of cutting-propagated *Pyracantha angustifolia* C. K. Schneid. *Propagation of Ornamental Plants*, 21 (1): 3-10. ISSN: 1311-9109.
424. Güney D., Bayraktar A., Atar F., Turna I. (2021). The effects of different factors on propagation by hardwood cuttings of some coniferous ornamental plants. *Şumarski list, CXLV*, 9–10: 467–477. ISSN: 0373-1332 (Print). UDK: 630* 232 (001). <https://doi-org/10.31298/sl.145.9-10.5>
425. Mashkhal Mohammed Amin Qadir, Othman Kamil Aref Hawramee. (2022). The rooting capacity of red tip photinia; *Photinia* × *fraseri* hardwood cuttings under the effects of cutting time and IBA concentration. *Journal of Kerbala for Agricultural Sciences Issue* (4), Volume (9): 118 – 133. ISSN: 2309-5199 (Print), ISSN: 2616-6933 (Electronic). <https://journals.uokerbala.edu.iq/index.php/Agriculture/article/view/1068/478>
426. Güney D., İ. Turna. (2022). *Chamaecyparis lawsoniana* kültivarlarının çelikle üretilmesi üzerine köklendirme ortamı sıcaklığı ile fitohormonların etkileri. In: Dr. Öğr. Üyesi Mehmet Çolak (Ed.). *Ziraat, Orman VE Su Ürünleri. Alanında Uluslararası Araştırmalar IV*. Yayınevi Türkiye Ofis: İstanbul: Eğitim Yayınevi Tic. Ltd. Şti., Atakent mah. Yasemen sok. №: 4/B, Ümraniye, İstanbul, Türkiye, 87-104. (In Turkish). ISBN: 978-625-8223-21-7

Публикация: Iliev I. (1992). *In vitro* clonning of *Robinia pseudoacacia* L. 'Rectissima' Raber. In: International Symposium at the occasion of 100-th Anniversary of the Arboretum Mlynany: "Woody plants biology commersial and landscape forming importance", Nitra: 463-471. ISBN: 80-224-0398-9.

Цитирани в:

427. Corneanu M., Corneanu G. C., Blejoi I.-S., Atyi, P., Netoiu C., Bica D. (2001). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica* on new culture media. Acta Horti Bot. Bucurest, 29: 307-315.
428. Corneanu M., Marinescu G., Corneanu G. C., Badea E., Babeanu C., Atyim P., Bica D., Cristea C. (2003). *In vitro* development of the *Robinia pseudoacacia* var. *Oltetica* subculture, on different media. Annales of the University of Craiova, vol. VIII (XLIV): 220-226.
429. Corneanu M., Mandrila G., Corneanu G. C., Cristea C., Atyim P., Netoiu C., Bica D. (2004). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica*. In: Maksimovic I., Cuvardic M., Duric S. (Eds). Proceedings of the XXXIV Annual ESNA Meeting, 29.08-2.09, Novi Sad, Serbia and Montenegro: 231-234. ISBN: 86-7520-034-X.
430. Corneanu M., Corneanu G. C., Cristea C., Bercu R., Roşca A., Ştefănescu I. (2005). The effects of the deuterium-depleted water on *in vitro* organogenesis process in *Robinia pseudoacacia* var. *oltetica*. Revue de Cytologie et de Biologie Végétales le Botaniste, 28: 236-245. ISBN: 0181-7582.
431. Димитрова П. (2012). Оценка на интродукцията на *Robinia pseudoacacia* L. в България. Наука за гората, No 1/2: 71-82.
432. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

Публикация: Tzvetkova N., Iliev I., Zhelev P. (1995). Morphological and biochemical investigations of the rhizogenesis *in vitro* of *Sequoia sempervirens* Endl. In: Recent Advances in plant biotechnology, Nitra: 40-46.

Цитирани в:

433. Илиев Н. (1996). Възможности за повишаване използването на гигантската секвоя (*Sequoiadendron giganteum* (Lindl.) Buchh.) чрез семенно и вегетативно размножаване. Дисертация, ЛТУ, София: 123 стр.
434. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences)
435. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
436. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.

Публикация: Iliev I., Tzvetkova N. (1995). Peroxidase activity during the *in vitro* rhizogenesis of *Betula pendula* Roth. 'jougii'. In: Recent Advances in plant biotechnology, Nitra: 52-57.

Цитирани в:

437. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and

introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences)

438. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
439. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Iliev I., Tzvetkov Iv. (1995). Possibilities for *in vivo* and *in vitro* propagation of *Metasequoia glyptostroboides* Hu. et Cheng. In: Recent Advances in plant biotechnology, Nitra: 64-72.

Цитирани в:

440. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирани в:

441. Chen G. D., Yang J., Wang L. Q., Liu H., Shen S., Qin F. (2010). *In vitro* adventitious shoot formation from petiole explants of *Swainsonia salsula* Taubert. Propagation of Ornamental Plants, 10 (3): 122-128. ISSN: 1311-9109.
442. Ginova A., Tsvetkov I., Kondakova V. (2012). *Rosa damascena* Mill. – an overview for evaluation of propagation methods. Bulgarian Journal of Agricultural Science, 18 (4): 545-556.
443. Gashi B (2013). Karakterizimi biomorfologjik, fiziologjik dhe ruajtja ex situ e gjermoplazmës së specieve të “ringjalljes” sëgjinisë ramonda në kosivë dhe në maqedoni. Disertacioni për marrjen e gradës shkoncore “Doctor”, Universiteti i Tiranës, Fakulteti i shkencave natyrore, Departamenti i bioteknologjisë, Tiranë, 199 pp.
444. Nyaika J. A., Njoloma J. P., Zimba S., Mwase W. F., Maliro M. F., Bokosi J. M., Kwapata M. B. (2014). The effectiveness of repeated shoot tip culture on pathogens load reduction in different local potato genotypes in Mmalawi. The International Journal of Biotechnology, 3 (7): 91-103.
445. Alexandrov A. H., Dobrev R. D. (2014). The State of Forest Genetic Resources in Bulgaria. Sofia, 84 pp. ISBN: 978-3-639-66981-7.
446. Andrys D. (2014). Kultury kalusa i zawiesinowe oraz możliwości ich wykorzystania. Dokonania Młodych Naukowców, 2: 18-20. ISSN 2300-4436.
447. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
448. Томов В. (2015). Анализ на семеипроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.
449. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.
450. Kaviani B. (2015). Some useful information about micropropagation. Journal of Ornamental Plants, 5 (1): 29-40. ISSN (Print): 2251-6433, ISSN (Online): 2252-6441.
451. Kintzios S. (2015). Introduction to plant tissue culture. Association of Greek Academic Libraries, 111 pp, ISBN: 978-960-603-033-8 (in Greek).

452. Bado S. (2015). Advances in plant mutation breeding. Dissertation for obtaining a doctorate degree at the University of Natural Resources and Life Sciences Vienna, 254 pp.
453. Pequeño-Granado I. L., Vázquez-Alvarado R. E., Santos-Haliscak J. A., Luna-Maldonado A. I., Moreno-Degollado G., Iracheta-Donjuan L., López-Gómez P., Castellanos-Juárez M., y Ojeda-Zacarías M. C. (2015). Inducción organogénica de *Jatropha curcas* L. a partir de hojas jóvenes. *Polibotánica*, 39: 79-89. ISSN 1405-2768.
454. Hart D. S., Keightley A. M., Sappington D., Nguyen P. T. M., Chritton C., Seckinger G. R., Torres K. C. (2016). Stability of adenine-based cytokinins in aqueous solution. *In vitro Cellular and Developmental Biology-Plant*, 52 (1): 1-9. ISSN: 1054-5476 (print); 1475-2689 (web). DOI 10.1007/s11627-015-9734-5
455. Silveira S. S., Cordeiro-Silva R., Degenhardt-Goldbach J., Quoirin M. (2016). Micropropagation of *Calophyllum brasiliense* (Cambess.) from nodal segments. *Brazilian Journal of Biology*, 76 (3): 656-663. <http://dx.doi.org/10.1590/1519-6984.23714>
456. Prado J. P. C. (2016). Análise bioquímica e proteômica durante a indução e regeneração de culturas nodulares em *Vriesea reitzii* Leme & A. Cost. Tese (doutorado). Universidade Federal de Santa Catarina, Centro de Ciencias Agrarias, Florianópolis, 104 pp.
457. Estrada-Zúñiga M. E., Aarland R. C., Rivera-Cabrera F., Bernabé-Antonio A., Buengía-González L., Cruz-Sosa A. (2016). Micropropagation of *Buddleja cordata* and the content of verbascoside and total phenols with antioxidant activity of the regenerated plantlets. *Revista Mexicana de Ingeniería Química*, 15(2): 333-346.
458. Kwaśniewska E., Pawłowska B. (2016). Efficient *in vitro* propagation of historical roses for biodiversity conservation. *Propagation of Ornamental Plants*, 17 (1): 3-11. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_17_1\(3-11\).pdf](https://www.journal-pop.org/References/Vol_17_1(3-11).pdf)
459. Davoudi Pahnekolayi A., Tehranifar A., Samiei L., Shoor M. (2017). Optimization of the micro-propagation protocol of two native rose species of Iran (*Rosa canina* and *Rosa beggeriana*). *Acta Horticulturae*, 1131: 87-95. ISBN: 90 6605 210 4, ISSN: 0567-7572. DOI 10.17660/ActaHortic.2016.1131.12
460. Tkalec M. (2017). Tehnologija uzgoja presadnica *Rosa canina* L. i *Pelargonium zonale* L. u kulturi tkiva i njihova adaptacija u različitim supstratima. Doktorska disertacija. Republika Hrvatska, Sveučilište Josipa Jurja Strossmayera u Osijek, u Poljoprivredni fakultet u Osijeku, 178 pp.
461. Aguirre-Alberto L., de Lourdes Martinez-Cardenas M. (2018). Production of callus and roots from lateral meristems of *Loeselia mexicana*. *Botanical Sciences*, 96 (3): 405-414. On-line ISSN: 2007-4476, Print: ISSN 2007-4298 DOI: <https://doi.org/10.17129/botsci.1910> https://www.botanicalsciences.com.mx/index.php/botanicalSciences/article/view/1910/pdf_4
462. Özzambak M. E., Zeybekoğlu E., Gün İ, Kiliç T. (2018). *Spathiphyllum*'un *in vitro* mikro çoğaltımı üzerine şeker konsantrasyonlarının etkileri. *Sakarya Üniversitesi Fen Bilimleri Enstitüsü Dergisi*, 22 (3): 1015-1023. DOI: 10.16984/saufenbilder.348480, e-ISSN: 2147-835X.
463. Maduka M. S., Laswai F. F., Njongomi E. E. (2018). Tafari experience in solving forest deforestation and degradation challenges in Tanzania. *In: J. F. Kessy, C. P. Balama, P. J. Kagosi, S. Bakengesa, N. E. Pima, M. A. Mndolwa and S.E. Nkya (Eds). Proceedings of the 1st TAFORI Scientific Conference on Forestry Research for Sustainable Industrial Economy in Tanzania: 44-53.*
464. Izmi Fahani Ilham (2018). Efektivitas pemberian kompos daun sirih hutan (*Piper aduncum* L.) dan pupuk kandang sapi terhadap pertumbuhan bibit pisang fhia-17. *Scripsi, Universitas Medan Area*, 116 pp.

465. Priyadarshan P. M. (2019). Tissue Culture. *In*: Plant Breeding: Classical to Modern. Springer, Singapore, Print ISBN: 978-981-13-7094-6, Online ISBN: 978-981-13-7095-3, DOI https://doi.org/10.1007/978-981-13-7095-3_21
466. M. Noruzpour N. Zare, P. Sheikhzadeh-Mosadegh, R. Asghari-Zakaria (2019). The effect of auxin and signaling compounds on growth and production of secondary metabolites in *in vitro* cultures of Whortleberry (*Vaccinium arctostaphylos* L.). Iranian Journal of Rangelands and Forests Plant Breeding and Genetic Research, Vol. 27, No.1, 58 (in Persian). Print ISSN: 1735-0891, Online ISSN: 2383-1448. DOI: 10.22092/ijrfpbgr.2019.121178.1296
467. Zakaria S. (2019). Cryopreservation of *Oncidium* Golden Anniversary's protocorm-like bodies using encapsulation-dehydration method. Thesis submitted in fulfillment of the requirements for the degree of Doctor of Philosophy, Universiti Sains Malaysia, 246 pp.
468. Miri S. M. (2020). Micropropagation, callus induction and regeneration of Ginger (*Zingiber officinale* Rosc.). Open Agriculture, 5: 75-84. Online ISSN: 2391-9531, <https://doi.org/10.1515/opag-2020-0008>
469. Henrique Gomes B., de Matos Oliveira F., Oliveira Nogueira A. P., de Oliveira Júnior R. J. (2020). *In vitro* callogenesis and organogenesis from *Carolina reaper* (Syn. *Capsicum chinense* Jacq.) and chromosomal analysis. Biotechnology Journal International, 24 (5): 1-11. ISSN: 2456-7051. DOI: 10.9734/BJI/2020/v24i530113
470. Cordeiro D., Rito M., Borges F., Canhoto J., Correia S. (2020). Selection and validation of reference genes for qPCR analysis of miRNAs and their targets during somatic embryogenesis in tamarillo (*Solanum betaceum* Cav.). Plant Cell, Tissue and Organ Culture, 143: 109-120. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044.
471. Rusanov K., Kovacheva N., Dobрева A., Atanassov I. (2020). *Rosa x damascena* Mill. (Rose). *In*: Novak J., Blüthner W. D. (Eds). Medicinal, Aromatic and Stimulant Plants. Handbook of Plant Breeding, vol. 12. Springer, Cham: 467-500. https://doi.org/10.1007/978-3-030-38792-1_14
472. Rodziewicz P., Kayser O. (2020). *Cannabis sativa* L. – Cannabis. *In*: Novak J., Blüthner W. D. (Eds). Medicinal, Aromatic and Stimulant Plants. Handbook of Plant Breeding, vol 12. Springer, Cham: 233-264. Online ISBN: 978-3-030-38792-1 https://doi.org/10.1007/978-3-030-38792-1_3
473. Rossa Yunita, Media Fitri Isma Nugraha, Endang Gati Lestari (2020). Micro propagation of *Bucephalandra* sp. Bioscience Research, 17 (2): 1339-1343. Print ISSN: 1811-9506 Online ISSN: 2218-3973. [https://www.isisn.org/BR17\(2\)2020/1339-1343-17\(2\)2020BR20-119.pdf](https://www.isisn.org/BR17(2)2020/1339-1343-17(2)2020BR20-119.pdf)
474. Jakovljević D., Stanković M. (2020). Application of *Teucrium* species: current challenges and further perspectives. *In*: Stanković M. (Eds). *Teucrium* Species: Biology and Applications. Springer: 413-432. Print ISBN: 978-3-030-52158-5, Online ISBN: 978-3-030-52159-2. https://doi.org/10.1007/978-3-030-52159-2_15
475. Murgayanti Murgayanti, Fatilla Nur Ramadhanti, Sumadi Sumadi (2020). Peningkatan pertumbuhan tunas kunyit putih pada perbanyakan *in vitro* melalui aplikasi berbagai jenis dan konsentrasi sitokinin. Jurnal Kultivasi, Vol. 19 (3) December: 1230-1236. ISSN: 1412-4718, eISSN: 2581-138x. DOI: <https://doi.org/10.24198/kultivasi.v19i3.29469>
476. Lozano-Milo E., García-Pérez P., Gallego P. P. (2020). Narrative review of production of antioxidants and anticancer compounds from Bryophyllum spp. (Kalanchoe) using plant cell tissue culture. Longhua Chinese Medicine, 3: 18. ISSN 2616-2806. <http://dx.doi.org/10.21037/lcm-20-46>.
477. Micaela Martins, Madalena Costa, Marta Gonçalves, Sandra Duarte, Manuel AuYong-Oliveira (2021). Knowledge Creation on Edible Vaccines. The Electronic Journal of Knowledge Management, 18(3): 285-301. ISSN 1479-4411, <https://doi.org/10.34190/ejkm.18.3.2020>

<https://academic-publishing.org/index.php/ejkm/article/view/2020/1931>. available online at www.ejkm.com

478. Jorge Juan-Vicedo, Atanas Pavlov, Segundo Ríos, Jose Luis Casas (2021). Micropropagation of five endemic, rare and/or endangered narcissus species from the Iberian peninsula (Spain and Portugal). *Acta Biologica Cracoviensia, Series Botanica*, 63/1: 55-61. PL-ISSN: 0001-5296, E-ISSN: 1898-0295. DOI: 10.24425/abcsb.2020.131674.
479. Dwie Retna Suryaningsih, Sri Arijanti, Arief Eryanto. (2020). Analysis of growth and enzyme contents of papain callus papaya (*Carica papaya* L.) through tissue culture engineering with *Saccharomyces cerevisiae* elicitors on MS and VW media (2021). International Conference on Science, Technology, and Environment 2020 Surabaya, December 2-3rd, 2020: 350-360. <https://ssrn.com/abstract=3799777>
480. Mariyam Mala, J. S. Norrizah, S. Azani (2021). *In vitro* seed germination and elicitation of phenolics and flavonoids in *in vitro* germinated *Trigonella foenum graecum* plantlets. *Biocatalysis and Agricultural Biotechnology*, Volume 32, March, Article 101907. Online ISSN: 1878-8181. <https://doi.org/10.1016/j.bcab.2021.101907>
481. Asieh Zare Khafri, Mahmood Solouki, Reza Zarghami, Baratali Fakheri, Nafiseh Mahdinezhad, Masoud Naderpour (2021). *In vitro* propagation of three Iranian apricot cultivars. *In Vitro Cellular & Developmental Biology - Plant*, 57: 102–117. ISSN: 1054-5476 (print); 1475-2689 (web), <https://link.springer.com/article/10.1007/s11627-020-10112-w>
482. Venkatachalam Balamurugan, Palanisamy Karthika, Gurusamy Dhandapani, Kumarasamy Pradeepa Veerakumari, Krishnan Vasanth (2021). Plant regeneration from direct organogenesis of *Pandanus canaranus* Warb, an endemic medicinal plant. *Vegetos*, 34: 404-412. ISSN: 2229-4473, Electronic ISSN: 2229-4473, <https://doi.org/10.1007/s42535-021-00218-w>
483. Samanhudi, Muji Rahayu, Amalia Tetrani Sakya, Edi Purwanto (2021). Pemanfaatan Pekarangan dengan Pisang Hasil Kultur Jaringan pada Gapoktan Sari Tani di Desa Gentan, Bendosari, Sukoharjo. *PRIMA: Journal of Community Empowering and Services*, Vol. 5(1): 63-68, e-ISSN 2579-5074. DOI: <https://doi.org/10.20961/prima.v5i1.44631>
484. Ankita Rajendra Parab, Bee Lynn, Sreeramanan Subramaniam (2021). Assessment of genetic stability on *in vitro* and *ex vitro* plants of *Ficus carica* var. Black Jack using SSR and DAMD markers. *Molecular Biology Reports*, 48: 7223-7231. Electronic ISSN: 1573-4978, Print ISSN: 0301-4851, <https://doi.org/10.1007/s11033-021-06714-1>, DOI: <https://doi.org/10.1007/s11033-021-06714-1>
485. Akansha Saxena, Wen-Lu Bi, Mukund R. Shukla, Syd Cannings, Bruce Bennett, Praveen K. Saxena (2021). Micropropagation and cryopreservation of Yukon Draba (*Draba yukonensis*), a special concern plant species endemic to Yukon Territory, Canada. *Plants* 2021, 10, 2093. ISSN (electronic): 2223-7747. <https://doi.org/10.3390/plants10102093>
486. Saowaros Phanomchai, Sompoch Noichinda, Yongsak Kachonpadungkitti, Kitti Bodhipadma (2021). Differing *in vitro* rooting and flowering responses of the persian violet to low and high UV-C irradiation. *Plants*, 10, Article 2671. ISSN (electronic): 2223-7747. <https://doi.org/10.3390/plants10122671>
487. Kuntze Ruchika Dani, Mahavir Gosavi (2022). *In-Vitro* Studies and Multiple Shootlet Induction in *Cyathocline purpurea* (Buch.- Ham. ex D.Don). *The Journal of Plant Science Research*, 38 (1): 111-116. ISSN: 0970-2539. DOI: <https://doi.org/10.32381/JPSR.2022.38.01.10>
488. Rahma Bejaoui (2022). Kalanço (*Kalanchoe blossfeldiana* Poelln.)’Nun *in vitro* koşullarda mikroçoğaltımı. *Ankara üniversitesi fen bilimleri enstitüsü, Doktora tezi*, 102 pp.
489. Ali Mehrban Jafarlou, Pirivatlo S. P., Salehi B., Amir Hoshang, Hoseynzadeh Mogbli (2023). The Proliferation of Cherry Dwarf Rootstocks: The Effects of Nutrient

- Media, Carbon Sources, and Genetic Fidelity Evaluation Using Simple Sequence Repeat Markers. *Biology Bulletin*, 49 (Suppl 2): 102-112. Electronic ISSN: 1608-3059, Print ISSN: 1062-3590. <https://link.springer.com/article/10.1134/S1062359022140084>
490. Gang R., Komakech R., Chung Y., Denis Okello, Wook Jin Kim, Byeong Cheol Moon, Nam-Hui Yim, Youngmin Kang (2023). *In vitro* propagation of *Codonopsis pilosula* (Franch.) Nannf. using apical shoot segments and phytochemical assessments of the maternal and regenerated plants. *BMC Plant Biol* 23, 33. <https://doi.org/10.1186/s12870-022-03950-w>. ISSN: 14712229.
 491. Dinah I. J. G. C. Pinto, Alone Lima-Brito. (2023). Micropropagation of *Vellozia seubertiana* (Velloziaceae). *Rev. Caatinga*, Mossoró, 36, (2): 271-279. ISSN: 0100-316X (impresso) ISSN: 1983-2125 (online). file:///C:/Users/ivan/Downloads/Micropropagation_of_Vellozia_seubertiana_Velloziac.pdf. <http://dx.doi.org/10.1590/1983-21252023v36n204rc>
 492. Ravi Kumar, Alamgir, Vishwajeet Yadav, Devendra Pal, Rishabh Shukla (2023). Micropropagation in Horticultural Crops. *In*: Dr. Manoj Kumar Singh, Mr. Khursheed Alam, Mr. Vibhu Pandey, Ms. Shalini Singh, Dr. Mukesh Kumar (Eds). *Advanced Technology in Horticulture*, Daya Publishing House®, A Division of Astral International Pvt. Ltd. New Delhi – 110 002: 221-229. ISBN: 978-93-5461-726-3. https://www.researchgate.net/publication/371475482_Micropropagation_in_Horticultural_Crops
 493. Virginia Sarropoulou, Eleni Maloupa, Katerina Grigoriadou (2023). *In vitro* direct organogenesis of the medicinal single-mountain local prioritized vulnerable Greek endemic *Achillea occulta* under different medium variants. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 51 (2): Article number 13124. Print ISSN: 0255-965X, Electronic ISSN: 1842-4309. DOI:10.15835/nbha51213124
 494. Camelia Andriani, Erni Suminar, Muhamad Kadapi, Anne Nuraini (2023). Perbandingan efek bap dan kinetin terhadap laju multiplikasi stroberi kultivar sweet charlie. *Jurnal Agroteknologi*, Vol. 14, No. 1: 13-18. p-ISSN: 1978-1555, e-ISSN: 2502-4906. DOI: 10.24014/ja.v14i1.18824 <https://www.semanticscholar.org/reader/d9760dd7829241b5964e5b8cb9c9c3c378503f99>
 495. Zeuko'o Menkem Elisabeth, RRufin Marie Kouipou Toghueo (2023). Plant Cell Culture for Biopharmaceuticals. *In*: Santosh Kumar Upadhyay, Sudhir P. Singh (Eds). *Plants as Bioreactors for Industrial Molecules*: 89-125. Print ISBN: 9781119875086, Online ISBN: 9781119875116 DOI:10.1002/9781119875116, <https://doi.org/10.1002/9781119875116.ch4>
 496. Ana-Maria Stoenescu, Sina Niculina Cosmulescu, Nicolae Gheorghiu. (2023). Preliminary research on *in vitro* propagation of *Ziziphus jujuba* Mill. *Horticulture*, Series B, Vol. LXVII, No. 1: 176-181. Print ISSN 2285-5653, Online ISSN: 2286-1580, ISSN-L: 2285-5653. https://horticulturejournal.usamv.ro/pdf/2023/issue_1/Art25.pdf
 497. Karolina Nowakowska, Katarzyna Kowalczyk, Anna Pawełczak and Janina Gajc-Wolska (2023). Energy Efficiency of LEDs during Micropropagation of *Helleborus* 'Molly's White'. *Agriculture*, 13, 1265. ISSN (electronic): 2077-0472. <https://doi.org/10.3390/agriculture13061265>
 498. Karolina Nowakowska, Potshangbam Nongdam, Nabilah Amany Samsurizal and Andrzej Pacholczak (2023). An efficient micropropagation protocol for the endangered European shrub February Daphne (*Daphne mezereum* L.) and identification of bacteria in culture. *Agriculture*, 13, 1692. ISSN (electronic): 2077-0472. <https://doi.org/10.3390/agriculture13091692>
 499. L. K. J. Al-Amery, M. A. Al- Shamari, I. H. Mohamme, H. S. A. Al-Jubori (2023) Developing a protocols to micropropagate Gac fruit (*Momordica cochinchinensis* Spreng.) *in vitro*.

Euphrates Journal of Agricultural Science, 15 (2): 517-526. ISSN 2072-3857.
https://www.researchgate.net/profile/Lamiaa-Al-Amery/publication/374510196_Developing_a_protocols_to_micropropagate_Gac_fruit_Momordica_cochinchinensis_Spreng_in_vitro/links/652144a4d717ef1293d31bdd/Developing-a-protocols-to-micropropagate-Gac-fruit-Momordica-cochinchinensis-Spreng-in-vitro.pdf

500. Roukia Benyammi, Soumeiya Krimat, Malika Alili, Mohamed Bekhouche, Oumaima Touari, Nawel Belalia, Amina Missoum, Lakhdar Khelifi and Abdelkader Morsli (2023). *In vitro* propagation of Algerian *Lavandula stoechas* and assessment of biochemical composition and their Antioxidant activity. Bull. Pharm. Sci., Assiut University, Vol. 46, Issue 2: 899-911. Print ISSN: 1110-0052, Online ISSN: 3009-7703.
https://journals.ekb.eg/article_327719_ac3763b1a285861d99b91dbb17ada1f9.pdf
501. Pham Thi Diem Thi, Nguyen Thi Nguyen Man, Nguyen Thi Khanh Quynh, Trieu Thy Hoa, Pham Mai Thu Thuy, Hoang Tan Quang. (2023). Micropropagation of long-leaved paperbark (*Melaleuca leucadendron* (L.) L.). Propagation of Ornamental Plants, 23: 91-98. ISSN: 1311-9109.

Публикация: Илиев Ив. (1987). Физични и физиологични качества на семената от брадавическа бреза (*Betula verrucosa* Ehrh.). Горско стопанство, горска промишленост, No 1: 17-19. ISSN: 0205-1753.

Цитирана в:

502. Kajba D. (1996). Međupopulacijska i unutarpopulacijska varijabilnost obične breze (*Betula pendula* Roth.) u dijelu prirodne rasprostranjenosti u republici Hrvatskoj. Glasnik za Šumske Pokuse, vol. 33: 53-108.
503. Kajba D. (1997). Variability of morphological characteristics of silver birch (*Betula pendula* Roth.) leaf and seed in the region of Croatia. Annales Forestales, 22/1: 1-28.
504. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр. ISBN: 954-8319-39-X.

Публикация: Илиев Ив., Чавдаров Ив. (1988). Проучване върху възможностите за размножаване на обикновената бреза (*Betula pendula* Roth.) по метода на тъканните култури. Наука за гората, No 1: 15-25. ISSN: 0681-0088.

Цитирана в:

505. Гюлева В., Гарелкова З. (1993). Микроразмножаване *in vitro* на *Paulownia tomentosa* (Stend.). Биотехнология, биотехника, 1993, № 1: 33-37. ISSN: 1310-2818 (Print) 1314-3530 (Online). <https://doi.org/10.1080/13102818.1993.10819410>
506. Томов В. (2015). Анализ на семепроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.

Публикация: Добринов Ив., Илиев Ив. (1989). Жизнеспособност на полена от обикновената бреза (*Betula pendula* Roth.). Горско стопанство, No 5: 10-11. ISSN: 0205-1753.

Цитирана в:

507. Желев П. (1992). Еколого-биологични и селекционно-генетични проучвания в бялборовите популации от родопите. Дисертация, ВЛТИ, София.

Публикация: Илиев Ив. (1992). Проучване върху кариотипа на обикновената бреза (*Betula pendula* Roth.). Научни трудове на ВЛТИ, серия “Горско стопанство”, том XXXIV: 89-94.

Цитирана в:

508. Petrova A., Zielinski J., Natcheva R. (2007). Chromosome numbers of woody plants in Bulgaria. *Phytologia Balcanica*, 13 (3): 371-378.

Публикация: Илиев Ив. (1989). Полиморфизъм по кората на обикновената бреза (*Betula pendula* Roth.) в България. Горско стопанство, No 8: 17-20. ISSN: 0205-1753.

Цитирана в:

509. Галеев Э. И. (2000). Березники южного Урала: на примере березы повислой. Диссертация на соискание ученой степени кандидата сельскохозяйственных наук. Башкирский государственных университет, 128 стр. (код специальности ВАК: 06.03.03 Лесоведение и лесоводство; лесные пожары и борьба с ними). <http://www.dissercat.com/content/bereznyaki-yuzhnogo-urala-na-primere-berezy-povisloi>
510. Коновалов В. Ф. (2003). Береза повислая на Южном Урале: структура популяций, селекция и воспроизводство. Диссертация для получения ученой степени доктор сельскохозяйственных наук, Йошкар-Ола, 503 стр. (код специальности ВАК: 06.03.01 Лесные культуры, селекция, семеноводство).

Публикация: Илиев Ив. (1990). Полиморфизъм по кората и короната на обикновената бреза (*Betula pendula* Roth.). Наука за гората, vol. XXVII, No 2: 108-116. ISSN: 0861-007X.

Цитирана в:

511. Ветчинникова Л. В. (2004). Береза: вопросы изменчивости. Отв. Редактор А. Ф. Титов, Москва, Наука, 183 стр. ISBN: 5-02-032621-6
512. Шемякина А. В. (2014). Биологически активные вещества дальневосточных представителей рода *Betula* L. Диссертация на соискание ученой степени кандидата биологических наук, «Дальневосточный научно-исследовательский институт лесного хозяйства», 03.02.14 – Биологические ресурсы, Хабаровск, 151 стр.
513. Alexandrov A. H., Dobrev R. D. (2014). The State of Forest Genetic Resources in Bulgaria. Sofia, 84 pp. ISBN: 978-3-639-66981-7

Публикация: Илиев Ив. (1990). Полиморфизъм по листната петура на обикновената бреза (*Betula pendula* Roth.). Природа, No 4: 52-54. ISSN: 0032-8731.

Цитирана в:

514. Kovačević S., Šimić D. (2001). Intrapopulational and interpopulational relations of *Betula pendula* Roth (Betulaceae) in Croatia, based on leaf morphometry. *Acta Biologica Cracoviensia, Series Botanica*, 43: 87-96. PL ISSN: 0001-5296.
515. Trinajstić I., Kovačić S., Šimić D. (2001). Are there any morphometrical differences in the leaves of two shoot types of silver birch (*Betula pendula* Roth, Betulaceae)? *Glasnik za šumske pokuse*, 38: 77 - 87. UDK: 630*164. <https://urn.nsk.hr/urn:nbn:hr:108:408731>
516. Franiel I., Wieski K. (2005). Leaf features of silver birch (*Betula pendula* Roth.) variability within and between two populations (uncontaminated vs Pb-contaminated and ZN-contaminated site). *Trees*, 19: 81-88. DOI: 10.1007/s00468-004-0366-3

Публикация: Илиев И. (1990). Възможности и предимства при размножаването на декоративните и горскостопански видове *in vitro*. Горско стопанство, No 2: 6-7. ISSN: 0205-1753.

Цитирана в:

517. Илиев Н. (1994). Физиологични качества на семената от *Sequoiadendron giganteum* (Lindl.) Buchh. В условия *in vivo* и *in vitro*. В: Международна конференция "IPPS in

Bulgaria – Размножаване на декоративни растения”, 30.10 – 1.11.1994, София: 11-18. ISBN: 954-8783-01-0.

518. Илиев Н. (1996). Възможности за повишаване използването на гигантската секвоя (*Sequoiadendron giganteum* (Lindl.) Buchh.) чрез семенно и вегетативно размножаване. Дисертация, ЛТУ, София, 123.

Публикация: Илиев Ив., Ганчев П. (1991). Микроклонално размножаване на бялата акация (*Robinia pseudoacacia* L.) чрез органични култури. Наука за гората, No 2: 38-45. ISSN: 0861-007X.

Цитирана в:

519. Corneanu M., Corneanu G. C., Blejoi I.-S., Atyi, P., Netoiu C., Bica D. (2001). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica* on new culture media. Acta Horti Bot. Bucurest, 29: 307-315.
520. Corneanu M., Marinescu G., Corneanu G. C., Badea E., Babeanu C., Atyim P., Bica D., Cristea C. (2003). *In vitro* development of the *Robinia pseudoacacia* var. *Oltetica* subculture, on different media. Annales of the University of Craiova, vol. VIII (XLIV): 220-226.
521. Corneanu M., Mandrila G., Corneanu G. C., Cristea C., Atyim P., Netoiu C., Bica D. (2004). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica*. In: Maksimovic I., Cuvardic M., Duric S. (Eds.) Proceedings of the XXXIV Annual ESNA Meeting, 29.08-2.09, Novi Sad, Serbia and Montenegro: 231-234. ISBN: 86-7520-034-X.
522. Corneanu M., Corneanu G. C., Cristea C., Bercu R., Roşca A., Ştefănescu U. (2005). The effect of the deuterium-depleted water on *in vitro* organogenesis processes in *Robinia pseudoacacia* var. *oltetica*. Revue de Cytologie et Biologie végétales-Le Botaniste, 28: 236-245. ISSN: 0181-7582.
523. Димитрова П. (2012). Оценка на интродукцията на *Robinia pseudoacacia* L. в България. Наука за гората, No 1/2 71-82. ISSN: 0861-007X.
524. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

Публикация: Илиев Ив., Ганчев П., Танева А. (1991). Научно-приложни аспекти при размножаването на обикновената бреза (*Betula pendula* Roth.) по метода *in vitro*. Биотехнология и биотехника, No 4-5: 3-9. ISSN: 0205-2067.

Цитирана в:

525. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

Публикация: Илиев Ив. (1993). Влияние на различните фактори при култивиране на обикновената бреза (*Betula pendula* Roth.) *in vitro*. Наука за гората, No 4: 73-84. ISSN: 0016-6766.

Цитирана в:

526. Ахметова А. А. (2009). Интродукция и размножение тюльпанов *in vivo* и *in vitro* в лесостепной зоне Башкирского Предуралья. Оренбург, Дисертация, 191 стр.
527. Zhelev P., Angelov V. (2012). Variation in a silver birch locality near Ardino (eastern rhodopes). Forestry Ideas, vol. 18, No 2 (44): 125-131. ISSN: 1314-3905 (Print), ISSN: 26-03-2996 (Online). https://forestry-ideas.info/issues/issues_Index.php?journalFilter=45

Публикация: Илиев Ив. (1994). Техники за аклиматизация на размножените *in vitro* растения. В: Богданов Б., Денкова С., Александров П., Желев П. (Ред.). Международна конференция "IPPS in Bulgaria – Размножаване на декоративни растения", София 30.10.-1.11: 1-6. ISBN: 954-8783-01-0.

Цитирана в:

528. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 132 стр.
529. Томов В. (2015). Анализ на семеипроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 122 стр.
530. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.

Публикация: Илиев Ив. (1995). Таксономия и синонимика на обикновената бреза (*Betula pendula* Roth.). Лесовъдска мисъл, No 1: 77-83. ISSN: 1310-5639.

Цитирана в:

531. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences).
532. Schmidt G., Sütörine Dioszegi M., Kohut I. (2006). Preformált gyökerek kialakulása lombos diszcszerje taxonokon. Kertgazdaság, 38 (4): 75-78.

Публикация: Илиев Ив., Цветкова Н. (1995). Натрупване на общ белтък при *Betula pendula* Roth. по време на ризогенезиса *in vitro*. В: Брезин В., Йовков И., Динков Б., Павлова Е., Василев В., Драганава И. (Ред.). 70 години лесотехническо образование в България, Юбилейна научна сесия, 7-9 VI, том I, Горско стопанство: 12-18.

Цитирана в:

533. Ветчинникова Л. В. (2005). Карельская береза и другие редкие представители рода *Betula* L. Рецензенты: Марковская Е. Ф., Новицкая Л. Л. Российская академия наук, Карельский научны центр, Института леса, Москва, Издательство Наука, 264 стр. ISBN: 5-02-033684-X.
534. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен "доктор". ЛТУ, София, 196 стр.
535. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.

Публикация: Илиев Ив., Чавдаров Ив. (1996). Физиологични качества и аномалии в кълняемостта на семената от бяла акация (*Robinia pseudoacacia* L.) при условия *in vivo* и *in vitro*. Лесотехнически университет, Научни трудове, том XXXVII: 33-40.

Цитирана в:

536. Илиев Н. (1994). Физиологични качества на семената от *Sequoiadendron giganteum* (Lindl.) Buchh. В условия *in vivo* и *in vitro*. В: Международна конференция "IPPS in

Bulgaria – Размножаване на декоративни растения”, 30.10 – 1.11.1994, София: 11-18. ISBN: 954-8783-01-0.

537. Corneanu M., Corneanu G. C., Blejoi I.-S., Atyi, P., Netoiu C., Bica D. (2001). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica* on new culture media. Acta Horti Bot. Bucurest, 29: 307-315.
538. Corneanu M., Marinescu G., Corneanu G. C., Badea E., Babeanu C., Atyim P., Bica D., Cristea C. (2003). *In vitro* development of the *Robinia pseudoacacia* var. *Oltetica* subculture, on different media. Annales of the University of Craiova, vol. VIII (XLIV): 220-226.
539. Corneanu M., Mandrila G., Corneanu G. C., Cristea C., Atyim P., Netoiu C., Bica D. (2004). *In vitro* micropropagation in *Robinia pseudoacacia* var. *Oltetica*. In: Maksimovic I., Cuvardic M., Duric S. (Eds.) Proceedings of the XXXIV Annual ESNA Meeting, 29.08-2.09, Novi Sad, Serbia and Montenegro: 231-234. ISBN: 86-7520-034-X.
540. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр. ISBN: 954-8319-39-X.
541. Cristina B., Ciobanu G., Corneanu M., Marinescu G., Corneanu C. G. (2006). Isoperoxidases pattern of *Robinia pseudoacacia* var. *oltetica* *in vitro* culture of different media. Buletin USAMV-CN, 62: 25-28. ISSN: 1454-2382.
542. Димитрова П. (2012). Оценка на интродукцията на *Robinia pseudoacacia* L. в България. Наука за гората, No 1/2: 71-82.
543. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.
544. Varbeva L., Iliev N. (2015). Seed qualities and seed dormancy breaking of sycamore maple (*Acer pseudoplatanus* L.) seeds. Forestry Ideas, 2015, vol. 21, No 1 (49): 97-105.

Публикация: Илиев Н., Китин П., Илиев Ив. (1996). Анатомично проучване върху витрификацията на клонирани *in vitro* микрорезници от *Sequoiadendron giganteum* (Lindl.) Buchh. В: IPPS in Bulgaria – Second Scientific Conference “Размножаване на декоративни растения”, 5-7. X, София: 213-218. ISBN: 954-8783-10-X (Т. 2).

Цитирана в:

545. Данчева Д. И. (2012). Възможности за клониране на планинския ясен (*Fraxinus excelsior* L.). Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 132 стр.

Публикация: Илиев Ив., Александров П. (1996). Проучване на качеството на семената от *Albizzia julibrissin* Durr. В условия *in vivo* и *in vitro*. В: “IPPS in Bulgaria - Second Scientific Conference “Propagation of Ornamental Plants”, 5-7.X, София: 219-222. ISBN: 954-8783-10-X (Т. 2).

Цитирана в:

546. Калмуков К. (2000). Проучване върху *Albizzia julibrissin* Dur. в района на Свищов. II. Производство на посадъчен материал. Propagation of Ornamental Plants, Fourth International Scientific Conference: 223-228. ISBN: 954-8783-34-7
547. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр. ISBN: 954-8319-39-X.
548. Милев П., Петкова К., Илиев Н. (2015). Горски култури – Горско семепроизводство. Издателска къща при ЛТУ, 288 стр. ISBN: 978-954-332-124-7.

549. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр.

Публикация: Илиев Ив., Борисова Д. (2002). Биотехнологични методи за подобряване и клониране на декоративните видове *in vitro*. Лесовъдска мисъл, № 3-4: 104-118. ISSN: 1310-5639.

Цитирана в:

550. Илиев Н., Милев М., Александров П., Петкова К. (2015). Производство на фиданки от дървесни и храстови видове. Издателска къща при Лесотехнически университет, София, 333 стр. ISBN: 978-954-332-127-8.

551. Томов В. (2015). Анализ на семеипроизводствените източници и размножаване на *Acer platanoides* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 122 стр.

552. Тодорова Т. Л. (2015). Размножаване на култивари от *Syringa vulgaris* L. Дисертация за получаване на научна и образователно степен “доктор”. ЛТУ, София, 196 стр.

Публикация: Milev M., Kitin P., Takata K., Iliev Iv., Nakada R. (2004). The introduction of *Larix* in Bulgaria-adaptation, growth and utilization. In: Larch Breeding and Genetic Resources, IUFRO International Symposium “Larix 2004”, 26 September-1 October, 2004, Kyoto & Nagano, Japan: 50.

Цитирана в:

553. Dražić D., Veselinović M., Nikolić B., Batos B., Čule N., Golubović-Ćurguz V., Mitrović S. (2010). Initial results of plantations of *Larix europaea* L. Established for recultivation. Forestry Ideas, vol. 16, No 2 (40): 266-271.

Публикация: Alexandrov A. H., Iliev I. (2019). Forest in South-Eastern Europe. Poplar, 203: 79-85. ISSN (print): 0563-9034, ISSN (online): 2620-1593. https://ilfe.org/sites/default/files/12Alexandrov_and_Iliev.pdf

Цитирана в:

554. Karaklić V., Cvjetičanin R., Košanin O. (2019). Florističke, sinekološke i edafske karakteristike šume bukve sa borovnicom (*Vaccinio-Fagetum moesiaceae* Fukarek 1969) na području Jelove gore u zapadnoj Srbiji, Poplar, 204: 37-49. ISSN (Print): 0563-9034, e-ISSN (Online): 2620-1593. https://journalpoplar.ilfe.org/sites/default/files/05Karaklic_et_al.pdf

Публикация: Лесотехнически университет 1925-2000. Съставители: Пухалев Г., Илиев Ив. София 2000, Издава „Сежани“, Печат „Балкан прес“ АД, 176 стр.

Цитирана в:

555. Коцева В. (2001). Минало, настояще и бъдеще на обучението по “Геодезия с вертикално планиране” на студентите от специалност “Ландшафтна архитектура” в Лесотехническият университет. В: Кулелиев Й., Щилянова Е., Илиев Ив., Василева И., Вакарелов Ил. (Ред.). 50 години специалност Ландшафтна архитектура”, юбилеен сборник, Издател „Сежани“: 92-100.

556. Колев Н. (2007). Университетската гора - Учебно-опитно горско стопанство Юндола. Михов Ив., Аспарухов К., Желев П. (Ред.), Темпора ЕООД, 318 стр. ISBN: 978-954-91851-2-6.

557. Milev M., Yurukov S., Lyubenov K., Zhelev P. (2010). Development of higher forestry education in Bulgaria. Forestry Ideas, 16 (2): 141-154. ISSN: 1314-3905 (Print), ISSN:

Публикация: Илиев Ив. (2001). Генетика и селекция на декоративните растения. Добринов Ив., Желев П. (Рецензенти), Sejani, 336 стр. + 16 стр. цветни приложения. ISBN: 954-90425-3-7.

Цитирана в:

558. Schmidt G. (2002). Juvenilitás-prekondicionálás-regenerációs képesség honosításra és nemesítésre épülő vizsgálatai fásszárú díznövényeknél. Értekezés a Magyar Tudományos Akadémia Doktora tudományos cím elnyeréséhez, Budapest, Május 2002. (Gábor Schmidt. 2002. Juvenilitas-precondicionalas-regeneration interaction studies at the breeding and introduction of woody ornamentals. Dissertation for obtaining the scientific degree Doctor of Hungarian Academy of Sciences)
559. Kormut'ak A. (2002). Genetics and breeding of ornamental plants. *In*: Forest Genetics, vol. 9, No. 1: 18. ISSN: 1335-048X.
560. Юруков Ст. (2003). Дендрология, СД “Лотус”, 212 стр. ISBN: 954-9783-70-3.
561. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр. ISBN: 954-8319-39-X.
562. Коларов Д., Цветкова Н. (2006). Физиология на дървесните растения. Изд-во Ракурс, 318 стр.
563. Schmidt G., Tóth I. (2006). Kertészeti dendrológia. Mezo Gazda, ISBN 963 286 318 6, 404 pp. ISBN: 963-286-318-6.
564. Kajba D., Ballian D. (2007). Šumarska genetika. Zagreb-Saraevo, ISBN 978-953-6307-93-7, 283 pp.
565. Simon Bell, Ingrid Sarlov Herlin, Richard Stiles (Eds.) (2011). Exploring the Boundaries of Landscape Architecture. Routlage, Taylor and Francis Group, Londaon and New York, 344 pp. ISBN: 978-0-415-67984-8 (hbk), ISBN: 978-0-415-67985-5 (pbk), ISBN: 978-0-203-80514 (ebk).
566. Милев П., Петкова К., Илиев Н. (2015). Горски култури – Горско семепроизводство. Издателска къща при ЛТУ, 288 стр. ISBN: 978-954-332-124-7
567. Юруков Ст., Панайотов М. (2015). Дендрология, РУТА - ХБ”, 212 стр. ISBN: 978-954-9557-21-3

Публикация: Батов Ив., Илиев Ив. Метод за автовегетативно размножаване на дървесни видове. Изобретение No 93 053/ 22.10.1990.

Цитирана в:

568. Batov I., Kitin P. (1994). Bioenergetics of LASER stimulation of the rooting process of Vine (*Vitis vinifera* L.) cuttings. В: Международна конференция “IPPS in Bulgaria – Размножаване на декоративни растения”, 30.10 – 1.11.1994, София: 114-120. ISBN: 954-8783-01-0.
569. Batov I., Kitin P. (1995). LASER biostimulation of mulberry (*Morus alba* L.) mature cuttings. Наука за гората, № 1: 3-11.

Публикация: Илиев Ив., Чавдаров Ив. (1996). Физиологични качества и аномалии в кълняемостта на семената от бяла акация (*Robinia pseudoacacia* L.) при условия *in vivo* и *in vitro*. Лесотехнически университет, Научни трудове, том XXXVII: 33-40.

Цитирана в:

570. Милев М., Александров П., Петкова К., Илиев Н. (2004). Посевни материали от широколистни видове. Изд-во Видолов & син ООД, 437 стр. ISBN: 954-8319-39-X.

Публикация: Илиев Ив. (1988). Проучване на естествените популации от обикновена бреза (*Betula pendula* Roth.) в западна България и отбор на ценни декоративни форми. Дисертационен труд за придобиване на научна степен „кссн“, ВЛТИ, София, 181 стр.

Ц и т и р а н а в:

571. Zhelev P., Angelov V. (2012). Variation in a silver birch locality near Ardino (Eastern Rhodopes). *Forestry Ideas*, 2(44): 125-131. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online). https://forestry-ideas.info/issues/issues_Index.php?journalFilter=45

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. *Plant Growth Regulation*, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Ц и т и р а н а в:

572. Коротцева И. Б. (2023). Фасциация у тыквенных культур. Овощи России, 6: 17-21. ISSN: 2072-9146 (print), ISSN: 2618-7132 (online). <https://doi.org/10.18619/2072-9146-2023-6-17-21>

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. *In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522, DOI: 10.1002/9780470686522.ch1*

Ц и т и р а н а в:

573. Akshay Mehta, Alkesh Yadav and Aman Kumar (2023). Micropropagation in Horticultural Crops *In: Revolutionizing Horticulture. The Green Path. Elite Publishing House: 65-75. ISBN: 978-93-58995-85-5*

Публикация: Lyubomirova T., Iliev I. (2013). *In vitro* propagation of *Syringa vulgaris* L. *Forestry Ideas*, 19: 173-185. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online). https://forestry-ideas.info/files/issue/Forestry_Ideas_BG_2013_19_2_7.pdf

Ц и т и р а н а в:

574. Тишкина Е. А., Семкина Л. А., Орехова О. Н., Григорьев А. А., Суслов А. В. (2022). Оценка роста и развития некоторых сортов *Syringa vulgaris* L. в условиях среднего Урала. Успехи Современного Естествознания, № 2: 28-33. ISSN: 1681-7494. DOI: 10.17513/use.37774 <https://natural-sciences.ru/ru/article/view?id=37774>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. *Plant Growth Regulation*, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Ц и т и р а н а в:

575. Bulavin I. V., Sidyakin A. I. (2023). An *in vitro* *Arabidopsis thaliana* Root Structural and Functional Characterization. *Izvestiya Akademii Nauk. Rossijskaya Akademiya Nauk. Seriya biologicheskaya*, 6: 616-625. ISSN: 1062-3590 (Print). doi: 10.31857/S1026347022600741. DOI: 10.1134/S106235902360174X, <https://journals.rcsi.science/1026-3470/article/view/231692>

Публикация: Nikolaou P., Zagaz D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). Advances in the micropropagation of service tree *Sorbus domestica*. *Propagation of Ornamental Plans*, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html

Цитирана в:

576. Valeria Gianguzzi and Francesco Sottile. (2024). Temporary Immersion System as an Innovative Approach for In Vitro Propagation of *Sorbus domestica* L. Horticulturae, 10, 164. EISSN 2311-7524. <https://doi.org/10.3390/horticulturae10020164>

Публикация: Iliev I. (2017). Factors affecting the axillary and adventitious shoots formation in woody plants *in vitro*. Acta Horticulturae, 1155: 15-27. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

577. Valeria Gianguzzi and Francesco Sottile. (2024). Temporary Immersion System as an Innovative Approach for In Vitro Propagation of *Sorbus domestica* L. Horticulturae, 10, 164. EISSN 2311-7524. <https://doi.org/10.3390/horticulturae10020164>

Публикация: Iliev I., Rubos A., Scaltsoyiannes A., Nellas H., Kitin P. (2003). Anatomical study of *in vitro* obtained fasciated shoots from *Betula pendula* Roth. Acta Horticulturae, No 616: 481-484. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

578. Chowdhury S., Datta A. K., Maity S. (2009). Induced stem fasciation in sesame (*Sesamum indicum* L.). Plant Archeives, vol. 9, № 1: 165-170. ISSN: 0972-5210. <https://www.cabidigitallibrary.org/doi/pdf/10.5555/20123401658>

Публикация: Dancheva D., Iliev I. (2015). Factors affecting adventitious shoot formation in *Fraxinus excelsior* L. Propagation of Ornamental Plants, 15 (1): 10-20. ISSN: 1311-9109. https://www.journal-pop.org/2015_15_1_10-20.html

Цитирана в:

579. Šedivá J., Havrdová L., Maršik P. (2017). Mikropropagace jasanu ztepilého (*Fraxinus excelsior* L.). Výzkumný ústav Silva Taroucy pro krajinu a okrasné zahradnictví, v. v. i., Průhonice, 20 pp. ISBN: 978-80-87674-24-6. <http://www.nusl.cz/ntk/nusl-451234> https://invenio.nusl.cz/record/451234/files/metodika_VUKOZ_Sediva_et al.pdf

Публикация: Mitras D., Kitin P., Iliev I., Dancheva D., Scaltsoyiannes A., Tsaktsira M., Nellas C., Rohr R. (2009). *In vitro* propagation of *Fraxinus excelsior* L. by epycotyls. Journal of Biological Research, 11: 37-48. ISSN: 2241-5793. <https://ikee.lib.auth.gr/record/233121/files/Mitras%20et%20al.pdf>

Цитирана в:

580. Šedivá J., Havrdová L., Maršik P. (2017). Mikropropagace jasanu ztepilého (*Fraxinus excelsior* L.). Výzkumný ústav Silva Taroucy pro krajinu a okrasné zahradnictví, v. v. i., Průhonice, 20 pp. ISBN: 978-80-87674-24-6. <http://www.nusl.cz/ntk/nusl-451234> https://invenio.nusl.cz/record/451234/files/metodika_VUKOZ_Sediva_et al.pdf

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирана в:

581. Santhanalakshmi Balasubramaniam, Sivanandhan Ganeshan, Selvaraj Natesan and Kapildev Gnanajothi (2024). A review on genetic diversity, micropropagation and transformations in the high-value medicinal plant of Himalayas-Seabuckthorn (*Hippophae* sp.). Journal of

Pharmacognosy and Phytochemistry, 13(1): 360-366. E-ISSN: 2278-4136 P-ISSN: 2349-8234. DOI: <https://doi.org/10.22271/phyto.2024.v13.i1e.14849>

582. R. M. P. I. Rajapaksha, R. G. J. Perera, D. D. N. Vibodhani, H. D. T. Abeyrathna, N. D. G. K. N. Jayawardhane, R. M. S. M. B. Rathnayaka (2023). A review in micropropagation of ornamental aquatic plants. International Research Conference of Sri Lanka Technology Campus Colombo, Sri Lanka. 14th -15th December, 2023: 261-265.

Публикация: Iliev I., Iliev N., Dancheva D., Corneanu M., Tsaktsira M., Gajdošova A., Mladenova S. (2010). Factors affecting the rooting of cuttings from cultivars of *Chamaecyparis lawsoniana* Parl. In: Soare M., Călina A., Panzaru Radu L., Niculescu M., Alexandru T., Stancu I., Cola M., Netiou C., Dimitru I. (Eds). Durable Agriculture – Agriculture of the Future. Analele Universitatii din Craiova, vol. XL, No 2: 174-181. ISSN 1841-8317. https://www.researchgate.net/publication/340265023_Factors_affecting_the_rooting_of_cuttings_from_cultivars_of_Chamaecyparis_lawsoniana_Parl_In_Soare_M_Calina_A_Panzaru

Цитирана в:

583. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Lyubomirova T., Iliev I. (2014). Factors affecting the rooting of cuttings of *Syringa vulgaris* L. cultivars. Oltenia Journal for Studies in Natural Sciences, 30 (1): 67-74. P-ISSN: 1454-6914. https://biozoojournals.ro/oscsn/cont/30_1/11_Lyubomirova_pp67-74.pdf

Цитирана в:

584. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Scaltsoyiannes A., Tsulpha P., Iliev I., Theriou K., Tsaktsira M., Mitras D., Karanikas C., Mahmoud S., Christopoulos V., Scaltsoyiannes V., Zaragotas D., Tzouvara A. (2009). Vegetative propagation of ornamental genotypes of *Prunus avium* L. Propagation of Ornamental Plants, 9(4): 198-206. ISSN: 1311-9109. https://www.journal-pop.org/2009_9_4_198-206.html

Цитирана в:

585. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Tsaktsira M., Alevropoulos A., Tsoulpha P., Scaltsoyiannes V., Scaltsoyiannes A., Iliev I. (2018). Inter- intra- genetic variation on rooting ability of *Ilex aquifolium* L. varieties and cultivars. Propagation of Ornamental Plants, 18 (4): 131-138. ISSN: 1311-9109. ISSN: 1311-9109. https://www.journal-pop.org/2018_18_4_131-138.html

Цитирана в:

586. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Nianiou-Obeidat, Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* 'Agrifoglio Commune'. Sustainability, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирана в:

587. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Nikolaou P., Zagaz D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). Advances in the micropropagation of service tree *Sorbus domestica*. Propagation of Ornamental Plants, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html

Цитирана в:

588. Sediva J., Businsky R., Pospiskova M., Velebil J., Drahozova H., Zyka V. (2021). Conservation methods of Czech gene pool of whitebeams. Acta Horticulturae, 1324: 41-46.

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирана в:

589. Günbeği Z., Ekinçi H., Ak B. E., Şaşkın N. (2024). Determination of proliferation performance of UCB-1 (*P. atlantica* × *P. integerrima*) pistachio rootstock in *in vitro* conditions. In: H. Turan Akkoyun Seyithan Seydoşoğlu (Eds). 7th International Cukurova Agriculture and Veterinary Congress, March 9-10, Adana, Türkiye: 463-473. ISBN: 978-625-367-680-3 DOI: <https://doi.org/10.5281/zenodo.10870563>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. Plant Growth Regulation, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Цитирана в:

590. Peretti A. V., Calbacho-Rosa L. S., Olivero P. A., Oviedo-Diego M. A., Vrech D. E. (2024). When THAT Exception Persists Almost as THAT Exception. In: Rules and Exceptions in Biology: from Fundamental Concepts to Applications. Springer, Cham.: 83-222. ISBN: 978-3-031-55380-6, ISBN: 978-3-031-55382-6, <https://doi.org/10.1007/978-3-031-55382-0>

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Nianiou-Obeidat, Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* 'Agrifoglio Commune'. Sustainability, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирана в:

591. Laconsole D., Scaltrito E., Sdao A. E., Cristiano G., De Lucua B. (2024). Application of commercial seaweed extract-based biostimulants to enhance adventitious root formation in

ornamental cutting propagation protocols: a review. *Frontiers in Horticulture*, 3: 1371090.
doi: 10.3389/fhort.2024.1371090

Подпис:

1.3. ЦИТИРАНИЯ НА НАУЧНИ ТРУДОВЕ (БЕЗ АВТОЦИТАТИ) В НАУЧНИ ПУБЛИКАЦИИ И В ПАТЕНТИ ЗА ИЗОБРЕТЕНИЯ У НАС И В ЧУЖБИНА ПРЕЗ ПОСЛЕДНИТЕ ПЕТ ГОДИНИ

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Лесотехнически университет, Факултет по Екология и ландшафтна архитектура.

Общ брой на цитиранията: 179

Забележка: Представен е доказателствен материал на хартиен носител и копия от цитиранията на флаш памет. Номерирането на цитиранията е в съответствие с номерирането на общия брой на цитиранията и номерирането на представения доказателствен материал.

Публикация: Iliev I. (1996). *In vitro* propagation of *Betula pendula* Roth 'jounzii'. In: "IPPS in Bulgaria - Propagation of Ornamental Plants", International Conference, 5-7 October, Sofia: 44-54. ISBN: ISBN: 954-8783-10-X (T. 2).

Ц и т и р а н а в:

26. Hanzhi Liang, Yuping Xiong, Beiyi Guo, Haifeng Yan, Shuguang Jian, Hai Ren, Xinhua Zhang, Yuan Li, Songjun Zeng, Kunlin Wu, Feng Zheng, Jaime A. Teixeira da Silva, Youhua Xiong, Guohua Ma (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. Environmental and Experimental Biology, 17: 169-177 DOI: 10.22364/eeb.17.17

Публикация: Iliev I., Besendorfer V., Peskan T. (1998). *In vitro* propagation of *Betula pendula* 'Dalecarlica' In: Tsekos I., Moustakas M. (Eds). Progress in Botanical Research, Kluwer Academic Publishers: 513-516. eBook ISBN: 978-94-011-5274-7. ISBN: 978-0-7923-5305-8. doi: 10.1007/978-94-011-5274-7_117 . DOI: <https://doi.org/10.1007/978-94-011-5274-7>

Ц и т и р а н а в:

46. Payamnoor Vahide, Khodadadi Negar, Jafari Hajati Razieh (2021). Bio nanoparticles as elicitors increase accumulation of betulin and betulinic acid in callus cultures. South African Journal of Botany, 141: 431-439. ISSN: 0254-6299. https://www.sciencedirect.com/science/article/pii/S0254629921001769?casa_token=yk_3Kf0PUhwAAAAA:3HwJz4MCchGjjroWvIM8JYWEc6gESkvIsBGUtMJp10rpWbG1DNd1iJBTiCuma76SnIarXgfBq6Et

Публикация: Iliev I., Kitin P., Funada R. (2001). Morphological and anatomical study of *in vitro* root formation of Silver birch (*Betula pendula* Roth.). Propagation of Ornamental Plants, vol. 1: 10-19. ISSN: 1311-9109. https://www.journal-pop.org/2001_1_1_10-19.html

Ц и т и р а н а в:

82. Машкина О. С., Табацкая Т. М., Внукова Н. И. (2019). Технология долгосрочного хранения в культуре *in vitro* ценных генотипов березы и выращивание на ее основе посадочного материала. Биотехнология, Т. 35, № 3: 57-67. ISSN 0234-2758. <http://cs.lisc.irk.ru/bd/%D0%96%D1%83%D1%80%D0%BD%D0%B0%D0%BB%D1%8B/%D0%91%D0%B8%D0%BE%D1%82%D0%B5%D1%85%D0%BD%D0%BE%D0%BB%D0%BE%D0%B3%D0%B8%D1%8F%202019%20%D0%A235/%E2%84%96%203/57-67.pdf>

83. Dokane K. (2019). Rododendru un to sakņu endofītisko sēņu mijiedarbības anatomiski fizioloģiskie aspekti un izmantošana rododendru pavairošanā. PhD thesis. Latvijas Universitātes Bioloģijas fakultātē Augu fizioloģijas, 114 pp.

84. Ozdemir S., Ozdemir M., Yetilmezsoy K. (2020). Effects of acclimatization soil prepared from hazelnut husk and gyttja on *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 2: 63-71. ISSN: 1311-9109.
85. Vu Quoc Luan, Le Kim Cuong, Hoang Thanh Tung, Vu Thi Hien, Do Manh Cuong, Phan Le Ha Nguyen, Bui Van The Vinh, Duong Tan Nhut (2020). Highly effective micropropagation and induction of early flowering by gibberellic acid in *Paphiopedilum delenatii* Guillaumin. Propagation of Ornamental Plants, 20 (1): 3-11. ISSN: 1311-9109.
86. Tikhomirova L. (2021). Features of morphogenesis of *Iris ensata* Thunb. *in vitro* culture. Russian Journal of Plant Physiology, 68: 669-677. ISSN: 1608-3407 (Online). <https://doi.org/10.1134/S1021443721040191>
87. İsfendiyaroğlu M. (2021). Anatomy of adventitious root development in mastic tree (*Pistacia lentiscus* var. *chia* Duham.) cuttings. Propagation of Ornamental Plants, 21 (2): 58-64. ISSN: 1311-9109.
88. Turp G. A., Turp S. M., Ozdemir S. (2021). The role of pyroligneous acid for *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 21 (4): 116-122. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_21_4\(116-122\).pdf](https://www.journal-pop.org/References/Vol_21_4(116-122).pdf)

Публикация: Iliev I., Scaltsoyiannes A., Rubos A. (2003). Shoot organogenesis and plant regeneration from leaf callus cultures of black bark silver birch (*Betula pendula* Roth. 'Melanocortea'). Acta Horticulturae, No 616: 321-326. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

97. Vítámvás J., Kuneš I., Viehmannová I., Linda R., Balá M. (2020). Conservation of *Betula oycoviensis*, an endangered rare taxon, using vegetative propagation methods. iForest-Biogeosciences and Forestry, vol. 13: 107-113. ISSN: 1971-7458. doi: 10.3832/ifor3243-013

Публикация: Rohr R., Iliev I., Scaltsoyiannes A., Tsoulouha P. (2003). Acclimatization of micropropagated forest trees. Acta Horticulturae, No 616: 59-69. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

125. Valencia Juárez M. C., López D. E., Díaz L. F., Espino, Pérez E. G. (2019). Aclimatación *ex vitro* de plántulas de *Fragaria* × *ananassa* Duch. Revista Mexicana de Ciencias Agrícolas, 10 (1): 91-100. ISSN: 2007-0934, ISSN-e: 2007-9230 https://www.researchgate.net/publication/330943464_Aclimatacion_ex_vitro_de_plantulas_de_Fragaria_x_ananassa_Duch
126. G. Indravathi, P. Suresh Babu (2019). Enhancing acclimatization of tissue cultured plants of *Albizia amara* by Biotization. International Journal of Scientific Research in Biological Sciences, 6 (4): 43-50, E-ISSN: 2347-7520 DOI: <https://doi.org/10.26438/ijrbs/v6i4.4350>
127. Ozdemir S., Ozdemir M., Yetilmezsoy K. (2020). Effects of acclimatization soil prepared from hazelnut husk and gyttja on *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 2: 63-71. ISSN: 1311-9109.
128. Mortier E., Lamotte O., Martin-Laurent F., Recorbet G. (2020). Forty years of study on interactions between walnut tree and arbuscular mycorrhizal fungi. A review. Agronomy for Sustainable Development, 40: 43. Electronic ISSN: 1773-0155, Print ISSN: 1774-0746. <https://doi.org/10.1007/s13593-020-00647-y>
129. Маляровская В. И., Самарина А. С. (2020). Адаптация культивируемых *in vitro* регенерантов *Hydrangea macrophylla* к нестерильным условиям. Садоводство и виноградарство, 6: 12-17. <https://doi.org/10.31676/0235-2591-2020-6-12-17>

130. Зеленьянська Н. М., Теслюк Н. І., Гоголінська О. І. Подуст Н. В. (2020). Розробка структурованого поживного середовища для адаптації вегетативної маси і кореневої системи мікроклонів винограду до умов *in vivo*. Таврійський науковий вісник, 116, Частина 1: 64-75. УДК 634.836:631.532:631.544 DOI <https://doi.org/10.32851/2226-0099.2020.116.1.8>
131. Vu Quoc Luan, Le Kim Cuong, Hoang Thanh Tung, Vu Thi Hien, Do Manh Cuong, Phan Le Ha Nguyen, Bui Van The Vinh, Duong Tan Nhut (2020). Highly effective micropropagation and induction of early flowering by gibberellic acid in *Paphiopedilum delenatii* Guillaumin. Propagation of Ornamental Plants, 20, (1): 3-11. ISSN: 1311-9109.
132. Singh P. R., Singh L. J. (2021). *In vitro* propagation for improvement of medicinal plants: a review. Journal of Pharmacognosy and Phytochemistry, 10 (1): 1484-1489 E-ISSN: 2278-4136 P-ISSN: 2349-8234
133. Murphy R., Adelberg J. (2021). Physical factors increased quantity and quality of micropropagated shoots of *Cannabis sativa* L. in a repeated harvest system with *ex vitro* rooting. In Vitro Cellular & Developmental Biology - Plant, 57: 923-931. <https://doi.org/10.1007/s11627-021-10166-4>, ISSN: 1054-5476 (print); 1475-2689 (web)
134. José Raymundo Enríquez-del Valle, Gerardo Rodríguez-Ortiz, Vicente Arturo Velasco-Velasco, Edmundo López-Hernández (2021). Fertilización durante la aclimatación en invernadero de plantas de *Agave potatorum* micropropagadas. Revista Mexicana de Agroecosistemas, Vol. 8 (1): 36-45. ISSN: 2007-9559 https://www.voaxaca.tecnm.mx/revista/docs/RMAE%20vol%208_1_2021/4-RMAE_2021-36-Fertilizacion-Resumen.pdf
135. Turp G. A., Turp S. M., Ozdemir S. (2021). The role of pyroligneous acid for *in vitro* propagated ornamental plants. Propagation of Ornamental Plants, 4: 116-122. ISSN: 1311-9109. [https://www.journal-pop.org/References/Vol_21_4\(116-122\).pdf](https://www.journal-pop.org/References/Vol_21_4(116-122).pdf)
136. Шуркина Е. С., Маляровская В. И. (2021). Особенности адаптации *Galanthus woronowii* Losinsk. к условиям *ex vitro*. Субтропическое и декоративное садоводство, 78: 82-88. ISSN: 2225-3068. УДК 578:581.5(470.62). doi: 10.31360/2225-3068-2021-78-82-88
137. Rahma Bejaoui (2022). Kalanşo (*Kalanchoe blossfeldiana* Poelln.)'Nun *in vitro* koşullarda mikroçoğaltımı. Ankara üniversitesi fen bilimleri enstitüsü, Doktora tezi, 102 pp.
138. Constantin Dushimimana, Michael Ajanja Sakha, Mercy Jebiwott Korir, Joyce Mnyazi Jefwa, Jan Vandenabeele, Titus Magomere, Eunice Wanjiru Mutitu, Jackson Mulatya, Florence Olubayo, Guy Smagghe, and Stefaan P. O. Werbrouck (2022). Early growth performance of *in vitro* raised *Melia volkensii* Gürke plantlets in response to beneficial microorganisms under semi-arid conditions. Plants, 11: Article 1300. ISSN: 2223-7747. <https://doi.org/10.3390/plants11101300>, <https://www.mdpi.com/journal/plants>
139. Yohannes Wibisono, Toni Herawan, Asri Insiana Putri, Fithry Ardhany, Liliek Haryjanto (2023). Clonal propagation of Sandalwood (*Santalum album* L.) through tissue culture by using branch-soaked explants. Proceedings of the 1st Unhas International Conference on Agricultural Technology (UICAT 2021) AIP Conference Proceedongs, Volume 2596 (1), 100021-1–100021-12; Published by AIP Publishing. On line ISSN: 1551-7616, Print ISSN: 0094-243X, <https://doi.org/10.1063/5.0119100>
140. Rahma Bejaoui, Görkem Eren Özdemir, Ş. Şebnem Ellialtıoğlu (2023). *In vitro* or *ex vitro* Rooting and Acclimatization of *Kalanchoe blossfeldiana* Poelln. Shoots Propagated by Tissue Culture Technique. Türk Tarım ve Doğa Bilimleri Dergisi (Turkish Journal of Agricultural and Natural Sciences), 10(4): 843–853. ISSN: 2148-3647. <https://doi.org/10.30910/turkjans.1288919>

Публикация: Iliev I., Rubos A., Scaltsoyiannes A., Nellas H., Kitin P. (2003). Anatomical study of *in vitro* obtained fasciated shoots from *Betula pendula* Roth. Acta Horticulturae, No 616: 481-484. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

151. Gailis A., Samsone I., Šēnhofa S., Girgžde E., Kāpostiniš R., Jansons Ā. (2021). Silver birch (*Betula pendula* Roth.) culture initiation *in vitro* and genotype determined differences in micropropagation. New Forests, 52 (5): 1-16. Electronic ISSN: 1573-5095, Print ISSN: 0169-4286. <https://doi.org/10.1007/s11056-020-09828-9>, <https://link.springer.com/article/10.1007/s11056-020-09828-9>

Публикация: Kitin P., Iliev I., Scaltsoyiannes A., Nellas Ch., Rubos A., Funada R. (2005). A comparative histological study between normal and fasciated shoots of *Prunus avium* L. generated *in vitro*. Plant Cell, Tissue and Organ Culture, 82: 141-150. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. <https://link.springer.com/article/10.1007/s11240-004-8153-9>

Цитирана в:

188. Liang H., Xiong Y., Guo B., Yan H., Jian Sh., Ren H., Zhang X., Li Y., Zeng S., Wu K., Zheng F., Teixeira da Silva J. A., Xiong Y., Ma G. (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. Environmental and Experimental Biology, 17: 169–177. eISSN: 2255-9582pISSN: 1691-8088. DOI: 10.22364/eeb.17.17
189. Aydin E. (2019). Anaç adayı kiraz, vişne ve mahlep genotiplerinin doku kültürü yöntemiyle çoğaltılması. T.C. Ordu Üniversitesi fen bilimleri enstitüsü, Doktora tezi, Bahçe bitkileri anabilim dalı, Ordu, 101 pp.
190. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Livia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. Rodriguésia 72: e00372019. 2021 <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>. ISSN: 03706583, ISSN: 21757860.
191. Aydin E., Yarılgac T. (2021). In Vitro Propagation of Some Mahaleb Genotypes as Candidate Rootstock for Sweet Cherries. Yuzuncu Yil University Journal of Agricultural Sciences, 31 (4): 847 – 857. ISSN: 1308-7576, EISSN: 1308-7584, DOI: 10.29133/yyutbd.892027, <https://doi.org/10.29133/yyutbd.892027>, <https://dergipark.org.tr/en/download/article-file/1621019>.
192. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). Science Archeives, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>

Публикация: Iliev N., Iliev I., Park Y.-G. (2005). Black locust (*Robinia pseudoacacia* L.) in Bulgaria. Journal of Korean Forest Society, 94 (5): 291-301. ISSN: 0445-4650. https://www.researchgate.net/publication/323836661_Black_locust_Robinia_pseudoacacia_L_in_Bulgaria#fullTextFileContent https://www.researchgate.net/profile/Nasko-Iliev-3/publication/312530134_Black_Locust_Robinia_pseudoacacia_L_in_Bulgaria/links/5880ceffaca272de1a410007/Black-Locust-Robinia-pseudoacacia-L-in-Bulgaria.pdf

Цитирана в:

204. Zhang K., Shen Z., Yang X., Ma L., Duan J., Li Y. (2019). Spatial Patterns in Different Stages of Regeneration after Clear-Cutting of a Black Locust Forest in Central China. Forests, 10, 1066. ISSN: 1999-4907. doi: 10.3390/f10121066
205. Vítková M., Sádlo J., Roleček J., Petřík P., Sitzia T., Müllerová J., Pyšek P. (2020). *Robinia pseudoacacia*-dominated vegetation types of Southern Europe: species composition, history, distribution and management. Science of the Total Environment, 707, 134857. ISSN: 0048-9697. doi: <https://doi.org/10.1016/j.scitotenv.2019.134857> (IF).

206. Nicolescu V-N., Rédei K., Mason W. L., Vor T., Pöetzelsberger E., Bastien J.-Ch., Brus R., Benčať T., Đodan M., Cvjetkovic B., Andrašev S., La Porta N., Lavnyy V., Mandžukovski D., Petkova K., Roženberger D., Waśik R., Mohren G. M. J., Monteverdi M. C., Musch B., Klisz M., Perić S., Keça L., Bartlett D., Hernea C., Pástor M. (2020). Ecology, growth and management of black locust (*Robinia pseudoacacia* L.), a non-native species integrated into European forests. J. For. Res., 31 (4):1081–1101. Online ISSN: 1993-0607, Print ISSN: 10007-662X, <https://doi.org/10.1007/s11676-020->
207. S. Barudanović E. Zečić A. Macanović B. Duraković, E. Mašić (2021). Invasive Alien Plant Species in Global Perspectives with Special References to Bosnia and Herzegovina. In: T. Pullaiah, Michael R. Ielmini (Eds). Invasive Alien Species: Observations and Issues from Around the World, Volume 3: Issues and Invasions in Europe: 215-252. Print ISBN: 9781119607021 Online ISBN: 9781119607045, <https://doi.org/10.1002/9781119607045.ch31>

Публикация: Iliev I. (2008). E. F. George, M. A. Hall, and G.-J. De Klerk (Eds). Plant propagation by tissue culture. Volume 1. The background. 3rd edn. Book review. Plant Cell, Tissue and Organ Culture, 93: 353-355. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044.

Цитирана в:

209. Yanping Lu, Liejian Huang, Hong Muhammad, Qasim Shahid (2021). *Acacia mangium* × *A. auriculiformis* micropropagation in a non-sterile environment. Reasearch Square: 1-23. <https://assets.researchsquare.com/files/rs-690162/v1/a6f8d17b-2103-459c-8b4d-f3104193c53d.pdf?c=1631886833>

Публикация: Nikolaou P., Zagas D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). Advances in the micropropagation of service tree *Sorbus domestica*. Propagation of Ornamental Plans, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html

Цитирана в:

221. Máté Ördögh (2022). In vitro propagation results of *Sorbus aria* 'Gran Sasso'. Review on Agriculture and Rural Development, vol. 11 (1-2): 204-222. ISSN: 2677-0792. DOI: 10.14232/rard.2022.1-2.204-214.
222. Jana Šedivá, Jiří Velebil, Daniel Zahradník (2023). Micropropagation as a tool for the conservation of autochthonous *Sorbus* species of Czechia. Plants, 12, 488. ISSN: 2223-7747. <https://doi.org/10.3390/plants12030488>

Публикация: Mitras D., Kitin P., Iliev I., Dancheva D., Scaltsoyiannes A., Tsaktsira M., Nellas C., Rohr R. (2009). In vitro propagation of *Fraxinus excelsior* L. by epycotyls. Journal of Biological Research, 11: 37-48. ISSN: 2241-5793. <https://ikee.lib.auth.gr/record/233121/files/Mitras%20et%20al.pdf>

Цитирана в:

247. Fenning T. M. (2019). The use of tissue culture and in-vitro approaches for the study of tree diseases. Plant Cell Tissue and Organ Culture, 136: 415-430. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044. <https://doi.org/10.1007/s11240-018-01531-0>
248. Pacholczak A., Nowakowska K. (2019). Micropropagation of February daphne (*Daphne mezereum* L.). Propagation of Ornamental Plants, 19: 106-112. ISSN: 1311-9109.
249. H. R. Islami Rostami, S. M. Hosseini Nasr, S. K. Kazemitabar, F. Zafarian (2019). Effect of provenances and culture media on seed germination of ash (*Fraxinus excelsior* L.) in embryo in vitro culture. Iranian Journal of Forest and Poplar Research, 27 (2): 159-168.

250. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Lívia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. *Rodriguésia* 72: e00372019. 2021 ISSN printed version: 0370-6583, ISSN online version: 2175-7860. <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>
251. Trevor Fenning, Margaret O'Donnell, Katharine Preedy, Aurélia Bézanger, David Kenyon, Gustavo Lopez. (2022). The rooting ability of in-vitro shoot cultures established from a UK collection of the common ash (*Fraxinus excelsior* L.) and their *ex-vitro* survival. *Annals of Forest Science*, 79: 30. ISSN: 1297-966X, <https://doi.org/10.1186/s13595-022-01146-8>
252. Haifeng Y., Anyua L., Jiaqia W., Guoshenga Z., Shusenb W., Xina Z. (2021). Identification of fasciation phenotype and anatomic structure analysis in *Salix psammophila*. *Journal of Desert Research*, vol. 41, No. 4: 45-50. ISSN: 1000-694X. DOI: 10.7522/j.issn.1000-694X.2021.00031
253. İsfendiyaroğlu M. (2021). Anatomy of adventitious root development in mastic tree (*Pistacia lentiscus* var. *chia* Duham.) cuttings. *Propagation of Ornamental Plants*, 21 (2): 58-64. ISSN: 1311-9109.
254. Xu L., Cheng F., Zhong Y. (2021). Micropropagation of *Paeonia ostii* 'Feng Dan' via *in vitro* mature embryo culture. *Propagation of Ornamental Plants*, 3: 78-87. ISSN: 1311-9109
255. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). *Science Archeives*, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>
256. Katarzyna Nawrot-Chorabik, Bartosz Pluciński, Dariusz Latowski (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. *New Forests*. ISSN: 0169-4286, eISSN: 1573-5095. <https://doi.org/10.1007/s11056-023-09981-x>
257. Ali Mohsen Al-Ali, Yaser Hassan Dewir and Rashid Sultan Al-Obeed (2023). Influence of Cytokinins, Dark Incubation and Air-Lift Bioreactor Culture on Axillary Shoot Proliferation of Al-Taif Rose (*Rosa damascena* trigintipetala (Diek) R. Keller). *Horticulturae*, 9, 1109. ISSN 2311-7524. <https://doi.org/10.3390/horticulturae9101109>

Публикация: Scaltsoyiannes A., Tsulpha P., Iliev I., Theriou K., Tsaktsira M., Mitras D., Karanikas C., Mahmoud S., Christopoulos V., Scaltsoyiannes V., Zaragotas D., Tzouvara A. (2009). Vegetative propagation of ornamental genotypes of *Prunus avium* L. *Propagation of Ornamental Plants*, 9(4): 198-206. ISSN: 1311-9109. https://www.journal-pop.org/2009_9_4_198-206.html

Цитирана в:

270. Selin Gül Ünsal, Angelo Gismondi, Gabriele Di Marco, Antonella Canini, Burcu Uzan Eken, Yelda Özden Çiftçi (2019). Intraspecific discrimination study of wild cherry populations from North-Western Turkey by DNA barcoding approach. *Tree Genetics & Genomes*, 15: 16. ISSN: 1614-2942 (print), ISSN: 1614-2950 (online) <https://doi.org/10.1007/s11295-019-1323-z>
271. Pijut P. (2020). *Prunus* spp. Cherry. In: Richard E. Litz, Fernando Pliego-Alfaro, Jose Ignacio Hormazap (Eds). *Biotechnology of Fruit and Nut Crops*, 2nd Edition: 532-560. ISBN-13: 178064827 9, <https://www.amazon.com/Biotechnology-Fruit-Nut-Crops-Agriculture/dp/1780648278>

Публикация: Scaltsoyiannes A., Tsaktsira M., Pasagiannis G., Tsoulpha P., Zhelev P., Iliev I., Rohr R. (2009). Allozyme variation of European black (*Pinus nigra* Arnold) and scots

pine (*Pinus sylvestris* L.) populations and implications on their evaluation: a comparative study. *Journal of Biological Research*, 11: 95-106. ISSN: 2241-5793. https://www.researchgate.net/publication/279622910_Allozyme_variation_of_Europe_an_Black_Pinus_nigra_Arnold_and_Scots_pine_Pinus_sylvestris_L_populations_and_implications_on_their_evolution_A_comparative_study
<https://ikee.lib.auth.gr/record/233302/files/Skaltsogiannis.pdf>

Цитирани:

285. Momchil Panayotov, Nickolay Tsvetanov, Evgeni Tsavkov, Georgi Gogushev, Peter Bebi, Petar Zhelev, and Stefan Yurukov (2019). Effect of Climate Change on the High-Mountain Tree Species and Their Genetic Resources in Bulgaria. Springer Nature Switzerland AG 2019. In: M. Šijačić-Nikolić et al. (Eds.), *Forests of Southeast Europe Under a Changing Climate*, *Advances in Global Change Research* 65, https://doi.org/10.1007/978-3-319-95267-3_35
286. Przybylski P., Masternak K., Jastrzębowski S. (2020). Isozyme polymorphism and seed and cone variability of Scots pine (*Pinus sylvestris* L.) in relation to local environments in Poland. *Folia Forestalia Polonica, Series A – Forestry*, Vol. 62 (2), 88–99. ISSN: 0071-6677, eISSN: 2199-5907. DOI: 10.2478/ffp-2020-0010
287. Ράπτης Δημήτριος, Καζάνα Βασιλική, Καζακλής Άγγελος, Σταματίου Χρήστος (2020). Ανάπτυξη γραμμικού μοντέλου μικτών επιδράσεων για την πρόβλεψη της διαμέτρου της κόμης ομήλικων συστάδων μαύρης Πεύκης (*Pinus nigra* Arn.) στην περιοχή του Εθνικού Δρυμού της Πίνδου. Γεωτεχνικά επιστημονικά θέματα - Σειρά VI – Τόμος 29 – Τεύχος 1: 4-13. ISSN 1105-9478 (*Raptis D., Kazana V., Kazaklis A., Stamatiou Ch. (2020). Development of a linear mixed effect crown width model for Black Pine (Pinus nigra Arn.) even aged natural forest stands in Pindos National Park Raptis. Geotechnical Scientific Issues. Geotechnical Chamber Of Greece, Vol. 29, Issue VI - No 1: 4-13. ISSN 1105-9478*)
288. Yetkin Usta, Ertuğrul Bilgili Kadir, Alperen Coşkuner (2020). Karaçam ekosistemlerinde orman yangınlarının ekolojik rolü: 252-272. https://www.researchgate.net/profile/Yetkin-Usta/publication/371911618_Karacam_Ekosistemlerinde_Orman_Yanginlarinin_Ekolojik_Rolu/links/649bdf0295bbbe0c6efc140b/Karacam-Ekosistemlerinde-Orman-Yanginlarinin-Ekolojik-Rolue.pdf
289. S. Versace, A. Bräuning, P. Cherubini M. DiFebbraro, M. Häusser, F. Lombardi, M. Marchetti, P. A. Marziliano, F. Salbitano, S. Szymczak, R. Tognetti. (2022). New evidence for population-specific responses to drought events from tree ring chronologies of *Pinus nigra* ssp. *laricio* across the entire distribution range. *Agricultural and Forest Meteorology*, Volume 323: 109076. ISSN: 0168-1923. <https://doi.org/10.1016/j.agrformet.2022.109076>

Публикация: Corneanu M., Corneanu G., Iliev I., Danci O., Stefanescu I., Popa M. (2010). Micropropagation of *Robinia pseudoacacia* var. *oltenica* selected stress resistant clones on media with deuterium depleted water. *Journal of Horticulture, Forestry and Biotechnology*, 14 (1): 141-144. ISSN: 2066-1797. https://journal-hfb.usab-tm.ro/romana/Lista%20lucrari%202010_PDF/JHFB%20-%20vol.1/29Corneanu%20Mihaela.pdf

Цитирани:

297. Ciuvăt A. L., Abrudan I. V., Ciuvăt C. G., Marcu C., Lorent A., Dincă L., Szilard B. (2022). Black locust (*Robinia pseudoacacia* L.) in Romanian forestry. *Diversity* 2022, 14, 780. ISSN 1424-2818. <https://doi.org/10.3390/d14100780>
298. Ábri T., Cseke K., Keserű Z., Porcsin A., Szabó F. M., Rédei K. (2023). Breeding and improvement of black locust (*Robinia pseudoacacia* L.) with a special focus on Hungary: a review. *iForest* 16: 290-298. ISSN: 1971-7458. doi: 10.3832/ifor4254-016.

Публикация: Dancheva D., Iliev I., Scaltsoyiannes A., Tsaktsitra M., Tsoulpha P. (2010). Effect of medium on *in vitro* germination of embryos of *Fraxinus excelsior* L. *Oltenia Journal for Studies in Natural Sciences*, Tom XXVI/1: 34-38. ISSN: 1454-6914. https://www.researchgate.net/publication/312937948_EFFECT_OF_MEDIUM_ON_I_N_VITRO_GERMINATION_OF_EMBRYOS_OF_FRAXINUS#fullTextFileContent https://biozoojournals.ro/oscsn/cont/26_1/B06-Dankeva.pdf

Цитирана е:

303. Xu L., Cheng F., and Zhong Y. (2021). Micropropagation of *Paeonia ostii* 'Feng Dan' via *in vitro* mature embryo culture. *Propagation of Ornamental Plants*, 3: 78-87. ISSN: 1311-9109.

Публикация: Iliev I., Scaltsoyiannes A., Tsaktsira M., Gajdosova A. (2010). Micropropagation of *Betula pendula* Roth cultivars by adventitious shoot induction from leaf callus. *Acta Horticulturae*, 885: 161-173. ISBN: 90 6605 210 4, ISSN: 0567-7572.

DOI: https://www.researchgate.net/publication/284350138_Micropropagation_of_Betula_pendula_roth_cultivars_by_adventitious_shoot_induction_from_leaf_callus

Цитирана е:

315. Mashkina O. S., Tabatskaya T. M. (2020). Morphogenesis of a dissected birch leaf *in vitro* culture. *Russian Journal of Developmental Biology*, 51: 397-409. ISSN: 1062-3604 <https://doi.org/10.1134/S1062360420060053>

316. Машкина О., Табацкая Т. (2020). Морфогенез раззеченного листа березы в культуре *in vitro*. *Онтогенез*, 51 (6): 460-472. ISSN (Print): 0475-1450. УДК 575:581.143.6:582.632.1

http://ontogenez.org/archive/2020/6/%D0%9C%D0%B0%D1%88%D0%BA%D0%B8%D0%BD%D0%B0_6_2020.pdf

317. Gaidamashvili M., Benelli G. (2021). Threatened woody plants of Georgia and micropropagation as a tool for *in vitro* conservation. *Agronomy* 11, Article 1082. ISSN: 2073-4395. <https://doi.org/10.3390/agronomy11061082>.

<https://www.mdpi.com/journal/agronomy>

318. Altae M., Dalas I. S. (2021). Abnormal growth in the plant (fasciation). *Science Archeives*, 2 (4): 339-342. ISSN: 2582-6697, <http://dx.doi.org/10.47587/SA.2021.2412>

319. Sergeev R., Kleva O., Timakov A., Zontikov D., Khusainova A. (2022). Features of *Betula* spp. cultivation through *in vitro* culture. *IOP Conf. Ser.: Earth Environment Science* 949 012007, doi:10.1088/1755-1315/949/1/012007

Публикация: Gajdošová A., Libiaková G., Iliev I., Hricová A. (2013). Adventitious shoots induction of *Amaranthus cruentus* L. *in vitro*. *Propagation of Ornamental Plants*, 13 (1): 33-39. ISSN: 1311-9109. [HTTPS://WWW.JOURNAL-POP.ORG/2013_13_1_33-39.HTML](https://WWW.JOURNAL-POP.ORG/2013_13_1_33-39.HTML)

Цитирана е:

321. Kiryu K. (2019). Evaluating the physiological potential of amaranth seeds. *Caribbean Journal of Science*, Vol. 52, No. 1: 491-494. Print ISSN: 0008-6452, Online ISSN: 0008-6452

322. Yaroshko O. (2021). Achievements in genetic engineering of *Amaranthus* L. representatives. *International Journal of Secondary Metabolite*, 8 (2): 172–185 e-ISSN: 2148-6905, <https://dx.doi.org/10.21448/ijsm.925737>, <https://dergipark.org.tr/en/pub/ijsm>

323. Magali Ruíz-Rivas, Catarino Eduardo Téllez-Valerio, Marcelino Martínez-Núñez, Pedro Fernando Vera-Hernández, Esperanza Martínez-Romero, Flor de Fátima Rosas-Cárdenas (2022). Influencia de la luz en la generación de callos y el cultivo *in vitro* de plantas. *Revista Mexicana de Ciencias Agrícolas*. Special publication number 27 August 15 -

- September 30: 11-21. ISSN en línea 2007-9230, ISSN impreso 2007-2934.
<https://cienciasagricolas.inifap.gob.mx/index.php/agricolas/article/view/3156/5090>
324. Anuradha, Manisha Kumari, Gaurav Zinta, Ramesh Chauhan, Ashok Kumar, Sanatsujat Singh and Satbeer Singh (2023). Genetic resources and breeding approaches for improvement of amaranth (*Amaranthus* spp.) and quinoa (*Chenopodium quinoa*). *Frontiers in Nutrition*, 10: 1129723. ISSN: 2296-861X (Online). doi: 10.3389/fnut.2023.1129723
325. Magali Ruíz-Rivas, Catarino Eduardo Téllez-Valerio, Marcelino Martínez-Núñez, Pedro Fernando Vera-Hernández, Esperanza Martínez-Romero, Flor de Fátima Rosas-Cárdenas (2022). Influence of light on callus generation and *in vitro* plant culture. *Revista Mexicana de Ciencias Agrícolas*. Special publication number 27: 11-21. ISSN: 2007-0934, ISSN-e: 2007-9230.
<https://cienciasagricolas.inifap.gob.mx/index.php/agricolas/article/view/3156/5091>

**Публикация: Iliev N., Dancheva D., Iliev I. (2013). Effect of selective herbicides in the production of turkey oak seedlings (*Quercus cerris* L.). *Oltenia Journal for Studies in Natural Sciences*, 29 (2): 97-100. P-ISSN: 1454-6914.
http://olteniastudiisicomunicaristiintelenaturii.ro/cont/29_2/13_Iliev.pdf**

Цитирана е:

326. Laclau P., Murilo N. L., Bertoli B., Osvaldo V. (2020). Tolerance of pedunculate oak (*Quercus robur*) saplings to herbicides tolerancia de plantines de roble pedunculado (*Quercus robur*) a herbicidas. *Revista de Investigaciones Agropecuarias*, 46 (3): 387-396. ISSN en linea 1669-2314, ISSN impreso 0325-8718,
<http://ppct.caicyt.gov.ar/index.php/ria/article/view/15358/45454575771041>
327. Савев С. (2023). Култивиране на висока американска боровинка в горските територии. Рецензенти: Петкова К., Стоянова М. Издателство Интел Ентранс, 174 стр. ISBN: 978-619-7703-16-0.

**Публикация: Dancheva D., Iliev N., Iliev I. (2013). *In vitro* propagation of *Fraxinus excelsior* L. *Oltenia Journal for Studies in Natural Sciences*, 29 (1): 78-84. P-ISSN: 1454-6914.
https://www.researchgate.net/publication/312937765_IN_VITRO_PROPAGATION_OF_Fraxinus_excelsior_L_DANCHEVA_Desislava_ILIEV_Nasko_ILIEV_Ivan**

Цитирана е:

328. Islami Rostami H. R., Hosseini Nasr S. M., Kazemitabar S. K., F. Zafarian (2019). Effect of provenances and culture media on seed germination of ash (*Fraxinus excelsior* L.) in embryo *in vitro* culture. *Iranian Journal of Forest and Poplar Research*, 27 (2): 159-168. Printing ISSN: 1735-0883, Electronic ISSN: 2383-1146. DOI: 10.22092/IJFPR.2019.120122
329. Nawrot-Chorabik K., Pluciński B., Latowski D. (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. *New Forests*. Electronic ISSN: 1573-5095, Print ISSN: 0169-4286. <https://doi.org/10.1007/s11056-023-09981-x>

**Публикация: Lyubomirova T., Iliev I. (2013). *In vitro* propagation of *Syringa vulgaris* L. *Forestry Ideas*. 19: 173-185. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online).
https://forestry-ideas.info/files/issue/Forestry_Ideas_BG_2013_19_2_7.pdf**

Цитирана е:

336. Лободина Е. В., Супрун И. И., Леонидовна Е. Т., Анатольевна Е. Б. (2020). Влияние сроков отбора експлантов сирени (*Syringa vulgaris* L.) на жизнеспособность и контаминацию при введении в культуру *in vitro*. *Плодоводство и виноградарство Юга России*, № 61(1): 98-107. DOI 10.30679/2219-5335-2020-1-61-98-107.
<http://journalkubansad.ru/pdf/20/01/08.pdf>

Публикация: Lyubomirova T., Iliev I. (2014). Factors affecting the rooting of cuttings of *Syringa vulgaris* L. cultivars. *Oltenia Journal for Studies in Natural Sciences*, 30 (1): 67-74. P-ISSN: 1454-6914.
https://biozoojournals.ro/oscsn/cont/30_1/11_Lyubomirova_pp67-74.pdf

Цитирана в:

337. Erken K., Özzambak M. E. (2020). Effects of cutting times and doses of some auxins on rooting of the hairy broom cuttings (*Chamaecytisus hirsutus* (L.) LINK). *International Journal of Agricultural and Natural Sciences* 13(2): 53-61. E-ISSN: 2651-3617.
<https://www.ijans.org/index.php/ijans/article/view/495/476>

Публикация: Tomov V., Iliev N., Iliev I. (2014). Analyzis of the forestry seed production base of *Acer platanoides* L. in Bulgaria. *Forestry Ideas*, 20, 1 (47): 67-76. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online).
file:///C:/Users/ivan/Downloads/Forestry_Ideas_BG_2014_20_1_5.pdf

Цитирана в:

340. Kachova V. (2020). Effect of organic and mineral fertilizers on the growth of *Acer platanoides* L. *Bulgarian Journal of Soil Science*, Volume 5, Issue 1: 50-63. (ISSN-online 2367-9212, ISSN-print 2534-8787).
https://www.bsss.bg/issues/Issue1_2020/BJSS_2020_1_5.pdf
www.bsss.bg

341. Budău R., Apăfăian A., Caradaică M., Bratu I. A., Timofte C. S. C., Enescu C. M. (2023). Expert-Based Assessment of the Potential of Agroforestry Systems in Plain Regions across Bihor County, Western Romania. *Sustainability*, 15, EISSN 2071-1050. 15724.
<https://doi.org/10.3390/su152215724>

Публикация: Dancheva D., Iliev I. (2015). Factors affecting adventitious shoot formation in *Fraxinus excelsior* L. *Propagation of Ornamental Plants*, 15 (1): 10-20. ISSN: 1311-9109. https://www.journal-pop.org/2015_15_1_10-20.html

Цитирана в:

345. Pacholczak A., Nowakowska K. (2019). Micropropagation of February daphne (*Daphne mezereum* L.). *Propagation of Ornamental Plants*, 19: 106-112. ISSN: 1311-9109.

346. Fenning T., O'Donnell M., Preedy K., Bézanger A., Kenyon D., Lopez G. (2022). The rooting ability of *in vitro* shoot cultures established from a UK collection of the common ash (*Fraxinus excelsior* L.) and their *ex vitro* survival. *Annals of Forest Science*, 79: 30. ISSN: 1286-4560. <https://doi.org/10.1186/s13595-022-01146-8>

347. Nawrot-Chorabik K., Pluciński B., Latowski D. (2023). Indirect auxiliary organogenesis of *Fraxinus excelsior* L. as a tool for ash dieback control. *New Forests*. Electronic ISSN: 1573-5095. Print ISSN: 0169-4286. Electronic ISSN: 1573-5095.
<https://doi.org/10.1007/s11056-023-09981-x>

348. Nowakowska K., Nongdam P., Samsurizal N. A., Pacholczak A. (2023). An efficient micropropagation protocol for the endangered European shrub February Daphne (*Daphne mezereum* L.) and identification of bacteria in culture. *Agriculture*, 13, 1692. ISSN 2077-0472. <https://doi.org/10.3390/agriculture13091692>

Публикация: Iliev I. (2017). Factors affecting the axillary and adventitious shoots formation in woody plants *in vitro*. *Acta Horticulturae*, 1155: 15-27. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Цитирана в:

350. Zarate-Díaz Y., Barba-Gonzalez R., Aguilera-Cauich E. A., Tapia-Campos E., Herrera-Cool G. J., Uc-Varguez A., Lopez-Puc G. (2019). *In vitro* organogenic behavior and molecular

- characterization of *Eustoma exaltatum* × *Eustoma grandiflorum* hybrids. Propagation of Ornamental Plants, 19 (4): 118-124. ISSN: 1311-9109.
351. Anis Adilah Mustafa, Mohammad Rahmat Derise, Wilson Thau Lym Yong, Kenneth Francis Rodrigues (2021). A Concise Review of *Dendrocalamus asper* and Related Bamboos: Germplasm Conservation, Propagation and Molecular Biology. Plants, 10, 1897. ISSN: 2223-7747. <https://doi.org/10.3390/plants10091897>
352. Burrows G. E. (2021). Gymnosperm resprouting - a review. Plants, 10, Article 2551. ISSN: 2223-7747. <https://doi.org/10.3390/plants10122551>
353. Xingmei Ai, Yonghui Wen, Chao Wang. (2021). *In Vitro* propagation and rejuvenation of senescent maternal plant of *Ardisia crenata* var. Bicolor (Primulaceae). Research Square. DOI:10.21203/rs.3.rs-992973/v1
354. Duc Tran Minh, Quynh Thi Nguyen, Thanh Pham (2021). Regeneration of plants via callus-mediated organogenesis from leaf, petiole and stem explants of *Ardisia silvestris* Pitard. Propagation of Ornamental Plants, 21 (3): 96-103. ISSN: 1311-9109.
355. Maurizio Capuana, Werther Guidi Nissim and Joshua D. Klein. (2022). Protocol for In Vitro Propagation of *Salix acmophylla* (Boiss.). Studies on Three Ecotypes. Forests, 13, 1124. ISSN: 1999-4907. <https://doi.org/10.3390/f13071124>, <https://www.mdpi.com/journal/forests>
356. Miaomiao Guo, Qiuying Yu, Daijun Li, Kexin Xu, Zexin Di, Yong Zhang, Yang Yu, Jian Zheng and Yan Zhang (2023). *In vitro* propagation, shoot regeneration, callus induction, and suspension from lamina explants of *Sorbus caloneura*. Forestry Research, 3: 7. <https://doi.org/10.48130/FR-2023-0007>. ISSN 2767-3812.
357. Pham Thi Diem Thi, Nguyen Thi Nguyen Man, Nguyen Thi Khanh Quynh, Trieu Thy Hoa, Pham Mai Thu Thuy, Hoang Tan Quang (2023). Micropropagation of long-leaved paperbark (*Melaleuca leucadendron* (L.) L.). Propagation of Ornamental Plants, 23: 91-98. ISSN: 1311-9109.

Публикация: Tsaktsira M., Alevropoulos A., Tsoulpha P., Scaltsoyiannes V., Scaltsoyiannes A., Iliev I. (2018). Inter- intra- genetic variation on rooting ability of *Ilex aquifolium* L. varieties and cultivars. Propagation of Ornamental Plants, 18 (4): 131-138. ISSN: 1311-9109. ISSN: 1311-9109. https://www.journal-pop.org/2018_18_4_131-138.html

Цитирана в:

358. Wang M., Hu J., Guo G., Park Y. G., Jeong B. R. (2021). Effect of auxins and their concentrations, immersion time, and rooting medium on rooting of cutting-propagated *Pyracantha angustifolia* C. K. Schneid. Propagation of Ornamental Plants, 21 (1): 3-10. ISSN: 1311-9109.
359. Khater N., Benahmed A., Zereg N., Cherouana K. (2021). Callogenesis induction of *Ilex aquifolium* L. (Aquifoliales Aquifoliaceae). Biodiversity Journal, 12 (1): 265-272. ISSN: 2039-0408 (Online). <https://doi.org/10.31396/Biodiv.Jour.2021.12.1.265.272>
360. Wang X., Lv Y., Xing S., Liu G., Sun J., Wang Y., Wang C., Yu X. (2021). Study on the efficient cutting propagation technology for *Ilex* “China Girl”. American Journal of Plant Sciences, 12: 1459-1467. ISSN Online: 2158-2750 ISSN Print: 2158-2742. <https://www.scirp.org/journal/ajps>
361. Kentelky E., Jucan D., Cantor M., Szekely-Varga Z. (2021). Efficacy of different concentrations of NAA on selected ornamental woody shrubs cuttings. Horticulturae, 7, 464. ISSN: 2311-7524. <https://doi.org/10.3390/horticulturae7110464>, file:///C:/Users/ivan/Downloads/Efficacy_of_Different_Concentrations_of_NAA_on_Sel.pdf

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Naniou-Obeidat, Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* 'Agrifoglio Commune'. Sustainability, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирана в:

362. Kentelky E., Jucan D., Cantor M., Szekely-Varga Z. (2021). Efficacy of Different Concentrations of NAA on Selected Ornamental Woody Shrubs Cuttings. Horticulturae, 7, 464. ISSN: 2311-7524. <https://doi.org/10.3390/horticulturae7110464>, file:///C:/Users/ivan/Downloads/Efficacy_of_Different_Concentrations_of_NAA_on_Sel.pdf
363. Loconsole D., Cristiano G., De Lucia B. (2022). Image analysis of adventitious root quality in wild sage and glossy abelia cuttings after application of different indole-3-butyric acid concentrations. Plants 2022, 11, Article 290. <https://doi.org/10.3390/plants11030290> <https://www.mdpi.com/journal/plants>
364. Aithida A., Tahiri A., Azlay L., L. Amina Idrissi Hassani, Mokhtari M. (2022). Effects of cutting origin and exogenous auxin treatment on the rooting of *Rosa damascena* (Mill) cuttings from the M'goun-Dades valleys in Morocco. Arabian Journal of Medicinal & Aromatic Plants, V8N1: 134-155. ISSN 2458-5920, file:///C:/Users/ivan/Downloads/30674-81635-1-PB.pdf, DOI: 10.48347/IMIST.PRSM/ajmap-v8i1.30674
365. Thi Kim Phuong Tran, Minh Hac Pham, Thi Huong Trinh, Sasanti Widiarsih, Viet The Ho (2022). Investigation of the genetic diversity of jewel orchid in Vietnam using RAPD and ISSR markers (2022). Biodiversitas, Volume 23, Number 9: 4816-4825. ISSN: 1412-033X, E-ISSN: 2085-4722, DOI: 10.13057/biodiv/d230950
366. Siti Sofiah, Luchman Hakim, Serafinah Indriyani, Iyan Robiansyah (2022). Acclimation study of *Smilax nageliana* A.DC., a climber species endemic to East Java, Indonesia. Biodiversitas, 23 (8): 4082-4089. E-ISSN: 2085-4722 ISSN: 1412-033X, DOI: 10.13057/biodiv/d230828
367. Pipinis E., Kostas S., Hatzilazarou S., Petropoulos S., Mitsi D., Stampoulidis A., Milios E., Smiris P. (2023). Effects of donor tree age, cutting collection time and K-IBA application on rooting ability of *Taxus baccata* L. stem cuttings: preliminary results. Folia Oecologica, 50 (1): 97-103, ISSN: 1338-7014. doi: 10.2478/foecol-2023-0009 (SCOPUS)
368. Huanyu Chen, Lizhou Hong, Angyan Ren, Kai Yu, Kai Wang, Sunan He, Chong Liu, Jincheng Xing (2023). Growth regulators on the shooting and rooting of *Tamarix chinensis* stem cuttings. Rhizosphere, 100679, ISSN: 2452-2198, <https://doi.org/10.1016/j.rhisph.2023.100679>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. Plant Growth Regulation, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Цитирана в:

398. Nezami-Alanagh E., Garoosi G.-A., Landín M., Gallego P. P. (2019). Computer-based tools provide new insight into the key factors that cause physiological disorders of pistachio rootstocks cultured *in vitro*. Scientific Reports, 9, Article number: 9740, ISSN: 2045-2322. <https://doi.org/10.1038/s41598-019-46155-2>.
399. Nabieva Alexandra Yurievna, Gerasimovich Lyudmila Vladimirovna (2019). The application of different reproduction techniques for rare species waterlily tulip (*Tulipa kaufmanniana* Regel.) propagation under *ex situ* conditions. Ornamental Horticulture, 25 (4): 450-460.

400. Liang H., Xiong Y., Guo B., Yan H., Jian Sh., Ren H., Zhang X., Li Y., Zeng S., Wu K., Zheng F., Teixeira da Silva J. A., Xiong Y., Guohua Ma G. (2019). *In vitro* regeneration and propagation from fasciated stems of *Vitex rotundifolia*. Environmental and Experimental Biology, 17: 169–177. ISSN : 2255-9582, DOI: 10.22364/eeb.17.17
401. Brailko V., Ivanova N., Zhdanova I., Mitrofanova O. (2020). Morphological and anatomical features of narrow-leaved lavender plants with prolonged conservation under *in vitro* genebank. BIO Web of Conferences 24, 00015 (2020) International Conferences “Plant Diversity: Status, Trends, Conservation Concept” 2020: 1-5. <https://doi.org/10.1051/bioconf/20202400015>
402. Zheng-Wei Zhang, Xiang-Hua Li, Xu-Dong Wang, Ke-Jing Wang (2020). Discovering adventitious roots in *Glycine tomentella* Hayata: lack of adventitious roots as a morphological and taxonomic species indicator is worth considering. Genetic Resources and Crop Evolution, 67 (6): 1345-1350. ISSN: 1573-5109 (Online). DOI: <https://doi.org/10.1007/s10722-020-00929-0>
403. Pace L., Pellegrini M., Pannunzio G., Gianfranco Pirone G. (2020). First report of fasciation symptom in *Artemisia eriantha* (Asteraceae), a typical orophyte of high-altitude cliffs, in Central Apennines (Italy). Plant Sociology, 57(1): 23-28. ISSN 1120-4605. DOI 10.3897/pls2020571/03
404. Claudia Simões-Gurgel, Tatiana Carvalho de Castro, Cátia Henriques Callado, Livia da Silva Cordeiro, Norma Albarello (2021). Micropropagation of *Tarenaya rosea* (Cleomaceae) from leaf explants. Rodriguésia 72: e00372019. 2021 ISSN printed version: 0370-6583, ISSN online version: 2175-7860. <http://rodriguesia.jbrj.gov.br> DOI: <http://dx.doi.org/10.1590/2175-7860202172020>
405. Egger J. M. (2021). Stem fasciation in the genus *Castilleja* (Orobanchaceae). Phytoneuron 2021-16: 1-29. ISSN: 2153-733X. <https://www.phytoneuron.net/wp-content/uploads/2021/08/16PhytoN-CastillejaFasciation.pdf>
406. Sriskanda D., Liew Y. X., Khor S. P., Merican F. M. M. S., Sreeramanan Subramaniam, Chew B. L. (2021). An efficient micropropagation protocol for *Ficus carica* cv. Golden Orphan suitable for mass propagation. Biocatalysis and Agricultural Biotechnology, 38 (2021) 102225 ISSN: 1878-8181 (print), <https://doi.org/10.1016/j.bcab.2021.102225>
407. Xincheng Yu, Xiaohong Chen, Yuping Xiong, Yujie Zeng, Zhenpeng Wei, Jinhui Pang, Xinhua Zhang, Yuan Li, Kunlin Wu, Songjun Zeng, Jaime A. Teixeira da Silva & Guohua Ma (2022). Shoot organogenesis from leaf and stem explants of *Heliotropium foertherianum* Diane and Hilger. In Vitro Cellular & Developmental Biology - Plant. 58(4):559-566 ISSN: 1054-5476 (print); 1475-2689 (web), DOI: <https://doi.org/10.1007/s11627-022-10257-w>
408. Feng Cheng, Mengfei Song, Mengru Zhang, Chunyan Cheng, Jinfeng Chen, Qunfeng Lou (2022). A SNP mutation in the *CsCLAVATA1* leads to pleiotropic variation in plant architecture and fruit morphogenesis in cucumber (*Cucumis sativus* L.). Plant Science, 323, 111397. ISSN 0168-9452. <https://doi.org/10.1016/j.plantsci.2022.111397>
409. Aurora M. Nedelcu (2022). Evo-devo perspectives on cancer. Essays in Biochemistry, EBC20220041. Online ISSN: 1744-1358. Print ISSN: 0071-1365. <https://doi.org/10.1042/EBC20220041>
410. Manokari M., Priyadharshini S., Shekhawat M. S. (2022). Repairing mechanism of foliar micro-morphological anomalies during acclimatization and field transfer of *in vitro* raised plantlets of *Aerva lanata* (L.) Juss. ex Schult.: a medicinally important plant. Vegetos, 35: 520–526. ISSN: 2229-4473, Electronic ISSN: 2229-4473, <https://doi.org/10.1007/s42535-021-00317-8>

412. Manokari M., Cokulraj M., Badhepuri M. K., Abhijit Dey, Mohammad Faisal, Abdulrahman A. Alatar, Rupesh Kumar Singh, Mahipal S. Shekhawat (2023). Microstructural and histochemical modifications in leaves at successive stages of *in vitro* development of the terrestrial orchid *Spathoglottis plicata* Blume. Horticulture Environment, and Biotechnology. Vol. 64(3): 497-510. ISSN: 22113460, 22113452, <https://doi.org/10.1007/s13580-022-00485-9>
413. Yadav R. S., Mewada Ketan, R. K Sugoor (2023). A record of stem fasciation phenomenon in *Cocculus hirsutus* (L.). Zoos' Print Journal, 38(3): 8-10. ISSN: 0971-6378. <https://zoosprint.zooreach.org/index.php/zp/article/view/7581/6837>
414. Tetsuya K. Matsumoto, Ryohei Fujisato, Munetaka Sugiyama, Yuko Miyazaki, Jin Murata (2023). A malformation of sex-changing plant *Arisaema serratum* (Araceae) produces both male and female inflorescences. Botany Letters. ISSN: 2381-8107, Journal homepage: <https://www.tandfonline.com/loi/tabg21>, <https://doi.org/10.1080/23818107.2023.2234973>, DOI: 10.1080/23818107.2023.2234973
415. Yimei Zang, Lei Xie, Jiaxian Su, Zuliang Luo, Xunli Jia, Xiaojun Ma (2023). Advances in DNA methylation and demethylation in medicinal plants: a review. Molecular Biology Reports, 50: 7783-7796, ISSN: 0301-4851 (print); 1573-4978 (web). <https://doi.org/10.1007/s11033-023-08618-8>
416. Li K., Tassinari A., Giuliani S., Rosignoli S., Urbany C., Tuberosa R. and Salvi S. (2023). QTL mapping identifies novel major loci for kernel row number-associated ear fasciation, ear prolificacy and tillering in maize (*Zea mays* L.). Frontiers in Plant Science, 13: 1017983. ISSN: 1664-462X (Online). doi: 10.3389/fpls.2022.1017983. <https://europepmc.org/backend/ptpmcrender.fcgi?accid=PMC9871824&blobtype=pdf>
417. Iliya Bulavin, A. I. Sidiyakin (2023). Structural and functional characterization of the root of *Arabidopsis thaliana* *in vitro*. Biol Bull Russ Acad Sci, 50(6): 1241-1249. ISSN 1062-3590 https://www.researchgate.net/publication/374847861_Structural_and_Functional_Characterization_of_the_Root_of_Arabidopsis_thaliana_In_Vitro/references

Публикация: Iliev I., Iliev N., Dancheva D., Corneanu M., Tsaktsira M., Gajdošova A., Mladenova S. (2010). Factors affecting the rooting of cuttings from cultivars of *Chamaecyparis lawsoniana* Parl. In: Soare M., Călina A., Panzaru Radu L., Niculescu M., Alexandru T., Stancu I., Cola M., Netiou C., Dimitru I. (Eds). Durable Agriculture – Agriculture of the Future. Analele Universitatii din Craiova, vol. XL, No 2: 174-181. ISSN 1841-8317. https://www.researchgate.net/publication/340265023_Factors_affecting_the_rooting_of_cuttings_from_cultivars_of_Chamaecyparis_lawsoniana_Parl_In_Soare_M_Calina_A_Panzaru

Цитирани в:

423. Wang M., Hu J., Guo G., Park Y. G., Byoung Ryong Jeong. (2021). Effect of auxins and their concentrations, immersion time, and rooting medium on rooting of cutting-propagated *Pyracantha angustifolia* C. K. Schneid. Propagation of Ornamental Plants, 21 (1): 3-10. ISSN: 1311-9109.
424. Güney D., Bayraktar A., Atar F., Turna I. (2021). The effects of different factors on propagation by hardwood cuttings of some coniferous ornamental plants. Şumarski list, CXLV, 9–10: 467–477. ISSN: 0373-1332 (Print). UDK: 630* 232 (001). <https://doi.org/10.31298/sl.145.9-10.5>
425. Mashkhal Mohammed Amin Qadir, Othman Kamil Aref Hawramee. (2022). The rooting capacity of red tip photinia; *Photinia* × *fraseri* hardwood cuttings under the effects of cutting time and IBA concentration. Journal of Kerbala for Agricultural Sciences Issue (4),

Volume (9): 118 – 133. ISSN: 2309-5199 (Print), ISSN: 2616-6933 (Electronic).
<https://journals.uokerbala.edu.iq/index.php/Agriculture/article/view/1068/478>

426. Güney D., İ. Turna. (2022). *Chamaecyparis lawsoniana* kültürlerinin çelikle üretilmesi üzerine köklendirme ortamı sıcaklığı ile fitohormonların etkileri. In: Dr. Öğr. Üyesi Mehmet Çolak (Ed.). Ziraat, Orman VE Su Ürünleri. Alanında Uluslararası Araştırmalar IV. Yayınevi Türkiye Ofis: İstanbul: Eğitim Yayınevi Tic. Ltd. Şti., Atakent mah. Yasemen sok. №: 4/B, Ümraniye, İstanbul, Türkiye, 87-104. (In Turkish). ISBN: 978-625-8223-21-7

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирана в:

465. Priyadarshan P. M. (2019). Tissue Culture. In: Plant Breeding: Classical to Modern. Springer, Singapore, Print ISBN: 978-981-13-7094-6, Online ISBN: 978-981-13-7095-3, DOI https://doi.org/10.1007/978-981-13-7095-3_21
466. M. Noruzpour N. Zare, P. Sheikhzadeh-Mosadegh, R. Asghari-Zakaria (2019). The effect of auxin and signaling compounds on growth and production of secondary metabolites in *in vitro* cultures of Whortleberry (*Vaccinium arctostaphylos* L.). Iranian Journal of Rangelands and Forests Plant Breeding and Genetic Research, Vol. 27, No.1, 58 (in Persian). Print ISSN: 1735-0891, Online ISSN: 2383-1448. DOI: 10.22092/ijrfpbgr.2019.121178.1296
467. Zakaria S. (2019). Cryopreservation of *Oncidium* Golden Anniversary's protocorm-like bodies using encapsulation-dehydration method. Thesis submitted in fulfillment of the requirements for the degree of Doctor of Philosophy, Universiti Sains Malaysia, 246 pp.
468. Miri S. M. (2020). Micropropagation, callus induction and regeneration of Ginger (*Zingiber officinale* Rosc.). Open Agriculture, 5: 75-84. Online ISSN: 2391-9531, <https://doi.org/10.1515/opag-2020-0008>
469. Henrique Gomes B., de Matos Oliveira F., Oliveira Nogueira A. P., de Oliveira Júnior R. J. (2020). *In vitro* callogenesis and organogenesis from *Carolina reaper* (Syn. *Capsicum chinense* Jacq.) and chromosomal analysis. Biotechnology Journal International, 24 (5): 1-11. ISSN: 2456-7051. DOI: 10.9734/BJI/2020/v24i530113
470. Cordeiro D., Rito M., Borges F., Canhoto J., Correia S. (2020). Selection and validation of reference genes for qPCR analysis of miRNAs and their targets during somatic embryogenesis in tamarillo (*Solanum betaceum* Cav.). Plant Cell, Tissue and Organ Culture, 143: 109-120. ISSN (print): 0167-6857, ISSN (electronic): 1573-5044.
471. Rusanov K., Kovacheva N., Dobрева A., Atanasov I. (2020). *Rosa x damascena* Mill. (Rose). In: Novak J., Blüthner W. D. (Eds). Medicinal, Aromatic and Stimulant Plants. Handbook of Plant Breeding, vol. 12. Springer, Cham: 467-500. https://doi.org/10.1007/978-3-030-38792-1_14
472. Rodziewicz P., Kayser O. (2020). *Cannabis sativa* L. – Cannabis. In: Novak J., Blüthner W. D. (Eds). Medicinal, Aromatic and Stimulant Plants. Handbook of Plant Breeding, vol 12. Springer, Cham: 233-264. Online ISBN: 978-3-030-38792-1 https://doi.org/10.1007/978-3-030-38792-1_3
473. Rossa Yunita, Media Fitri Isma Nugraha, Endang Gati Lestari (2020). Micro propagation of *Bucephalandra* sp. Bioscience Research, 17 (2): 1339-1343. Print ISSN: 1811-9506 Online ISSN: 2218-3973. [https://www.isisn.org/BR17\(2\)2020/1339-1343-17\(2\)2020BR20-119.pdf](https://www.isisn.org/BR17(2)2020/1339-1343-17(2)2020BR20-119.pdf)

474. Jakovljević D., Stanković M. (2020). Application of *Teucrium* species: current challenges and further perspectives. In: Stanković M. (Eds). *Teucrium Species: Biology and Applications*. Springer: 413-432. Print ISBN: 978-3-030-52158-5, Online ISBN: 978-3-030-52159-2. https://doi.org/10.1007/978-3-030-52159-2_15
475. Murgayanti Murgayanti, Fatilla Nur Ramadhanti, Sumadi Sumadi (2020). Peningkatan pertumbuhan tunas kunyit putih pada perbanyakan *in vitro* melalui aplikasi berbagai jenis dan konsentrasi sitokinin. *Jurnal Kultivasi*, Vol. 19 (3) December: 1230-1236. ISSN: 1412-4718, eISSN: 2581-138x. DOI: <https://doi.org/10.24198/kultivasi.v19i3.29469>
476. Lozano-Milo E., García-Pérez P., Gallego P. P. (2020). Narrative review of production of antioxidants and anticancer compounds from *Bryophyllum* spp. (*Kalanchoe*) using plant cell tissue culture. *Longhua Chinese Medicine*, 3: 18. ISSN 2616-2806. <http://dx.doi.org/10.21037/lcm-20-46>.
477. Micaela Martins, Madalena Costa, Marta Gonçalves, Sandra Duarte, Manuel AuYong-Oliveira (2021). Knowledge Creation on Edible Vaccines. *The Electronic Journal of Knowledge Management*, 18(3): 285-301. ISSN 1479-4411, <https://doi.org/10.34190/ejkm.18.3.2020> <https://academic-publishing.org/index.php/ejkm/article/view/2020/1931>. available online at www.ejkm.com
478. Jorge Juan-Vicedo, Atanas Pavlov, Segundo Ríos, Jose Luis Casas (2021). Micropropagation of five endemic, rare and/or endangered narcissus species from the Iberian peninsula (Spain and Portugal). *Acta Biologica Cracoviensia, Series Botanica*, 63/1: 55-61. PL-ISSN: 0001-5296, E-ISSN: 1898-0295. DOI: 10.24425/abcsb.2020.131674.
479. Dwie Retna Suryaningsih, Sri Arijanti, Arief Eryanto. (2020). Analysis of growth and enzyme contents of papain callus papaya (*Carica papaya* L.) through tissue culture engineering with *Saccharomyces cerevisiae* elicitors on MS and VW media (2021). *International Conference on Science, Technology, and Environment 2020 Surabaya*, December 2-3rd, 2020: 350-360. <https://ssrn.com/abstract=3799777>
480. Mariyam Mala, J. S. Norrizah, S. Azani (2021). *In vitro* seed germination and elicitation of phenolics and flavonoids in *in vitro* germinated *Trigonella foenum graecum* plantlets. *Biocatalysis and Agricultural Biotechnology*, Volume 32, March, Article 101907. Online ISSN: 1878-8181. <https://doi.org/10.1016/j.bcab.2021.101907>
481. Asieh Zare Khafri, Mahmood Solouki, Reza Zarghami, Baratali Fakheri, Nafiseh Mahdinezhad, Masoud Naderpour (2021). *In vitro* propagation of three Iranian apricot cultivars. *In Vitro Cellular & Developmental Biology - Plant*, 57: 102–117. ISSN: 1054-5476 (print); 1475-2689 (web), <https://link.springer.com/article/10.1007/s11627-020-10112-w>
482. Venkatachalam Balamurugan, Palanisamy Karthika, Gurusamy Dhandapani, Kumarasamy Pradeepa Veerakumari, Krishnan Vasanth (2021). Plant regeneration from direct organogenesis of *Pandanus canaranus* Warb, an endemic medicinal plant. *Vegetos*, 34: 404-412. ISSN: 2229-4473, Electronic ISSN: 2229-4473, <https://doi.org/10.1007/s42535-021-00218-w>
483. Samanhudi, Muji Rahayu, Amalia Tetrani Sakya, Edi Purwanto (2021). Pemanfaatan Pekarangan dengan Pisang Hasil Kultur Jaringan pada Gapoktan Sari Tani di Desa Gentan, Bendosari, Sukoharjo. *PRIMA: Journal of Community Empowering and Services*, Vol. 5(1): 63-68, e-ISSN 2579-5074. DOI: <https://doi.org/10.20961/prima.v5i1.44631>
484. Ankita Rajendra Parab, Bee Lynn, Sreeramanan Subramaniam (2021). Assessment of genetic stability on *in vitro* and *ex vitro* plants of *Ficus carica* var. Black Jack using SSR and DAMD markers. *Molecular Biology Reports*, 48: 7223-7231. Electronic ISSN: 1573-4978, Print ISSN: 0301-4851, <https://doi.org/10.1007/s11033-021-06714-1>, DOI: <https://doi.org/10.1007/s11033-021-06714-1>

485. Akansha Saxena, Wen-Lu Bi, Mukund R. Shukla, Syd Cannings, Bruce Bennett, Praveen K. Saxena (2021). Micropropagation and cryopreservation of Yukon Draba (*Draba yukonensis*), a special concern plant species endemic to Yukon Territory, Canada. *Plants* 2021, 10, 2093. ISSN (electronic): 2223-7747. <https://doi.org/10.3390/plants10102093>
486. Saowaros Phanomchai, Sompoch Noichinda, Yongsak Kachonpadungkitti, Kitti Bodhipadma (2021). Differing *in vitro* rooting and flowering responses of the persian violet to low and high UV-C irradiation. *Plants*, 10, Article 2671. ISSN (electronic): 2223-7747. <https://doi.org/10.3390/plants10122671>
487. Kuntze Ruchika Dani, Mahavir Gosavi (2022). *In-Vitro* Studies and Multiple Shootlet Induction in *Cyathocline purpurea* (Buch.- Ham. ex D.Don). *The Journal of Plant Science Research*, 38 (1): 111-116. ISSN: 0970-2539. DOI: <https://doi.org/10.32381/JPSR.2022.38.01.10>
488. Rahma Bejaoui (2022). Kalanço (*Kalanchoe blossfeldiana* Poelln.)'Nun *in vitro* koşullarda mikroçoğaltimi. Ankara üniversitesi fen bilimleri enstitüsü, Doktora tezi, 102 pp.
489. Ali Mehrban Jafarlou, Pirivatlo S. P., Salehi B., Amir Hoshang, Hoseynzadeh Mogbli (2023). The Proliferation of Cherry Dwarf Rootstocks: The Effects of Nutrient Media, Carbon Sources, and Genetic Fidelity Evaluation Using Simple Sequence Repeat Markers. *Biology Bulletin*, 49 (Suppl 2): 102-112. Electronic ISSN: 1608-3059, Print ISSN: 1062-3590. <https://link.springer.com/article/10.1134/S1062359022140084>
490. Gang R., Komakech R., Chung Y., Denis Okello, Wook Jin Kim, Byeong Cheol Moon, Nam-Hui Yim, Youngmin Kang (2023). *In vitro* propagation of *Codonopsis pilosula* (Franch.) Nannf. using apical shoot segments and phytochemical assessments of the maternal and regenerated plants. *BMC Plant Biol* 23, 33. <https://doi.org/10.1186/s12870-022-03950-w>. ISSN: 14712229.
491. Dinah I. J. G. C. Pinto, Alone Lima-Brito. (2023). Micropropagation of *Vellozia seubertiana* (Velloziaceae). *Rev. Caatinga, Mossoró*, 36, (2): 271-279. ISSN: 0100-316X (impresso) ISSN: 1983-2125 (online). file:///C:/Users/ivan/Downloads/Micropropagation_of_Vellozia_seubertiana_Velloziac.pdf . <http://dx.doi.org/10.1590/1983-21252023v36n204rc>
492. Ravi Kumar, Alamgir, Vishwajeet Yadav, Devendra Pal, Rishabh Shukla (2023). Micropropagation in Horticultural Crops. *In: Dr. Manoj Kumar Singh, Mr. Khursheed Alam, Mr. Vibhu Pandey, Ms. Shalini Singh, Dr. Mukesh Kumar (Eds). Advanced Technology in Horticulture*, Daya Publishing House®, A Division of Astral International Pvt. Ltd. New Delhi – 110 002: 221-229. ISBN: 978-93-5461-726-3. https://www.researchgate.net/publication/371475482_Micropropagation_in_Horticultural_Crops
493. Virginia Sarropoulou, Eleni Maloupa, Katerina Grigoriadou (2023). *In vitro* direct organogenesis of the medicinal single-mountain local prioritized vulnerable Greek endemic *Achillea occulta* under different medium variants. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 51 (2): Article number 13124. Print ISSN: 0255-965X, Electronic ISSN: 1842-4309. DOI:10.15835/nbha51213124
494. Camelia Andriani, Erni Suminar, Muhamad Kadapi, Anne Nuraini (2023). Perbandingan efek bap dan kinetin terhadap laju multiplikasi stroberi kultivar sweet charlie. *Jurnal Agroteknologi*, Vol. 14, No. 1: 13-18. p-ISSN: 1978-1555, e-ISSN: 2502-4906. DOI: 10.24014/ja.v14i1.18824 <https://www.semanticscholar.org/reader/d9760dd7829241b5964e5b8cb9c9c3c378503f99>
495. Zeuko'o Menkem Elisabeth, RRufin Marie Kouipou Toghueo (2023). Plant Cell Culture for Biopharmaceuticals. *In: Santosh Kumar Upadhyay, Sudhir P. Singh (Eds). Plants as Bioreactors for Industrial Molecules*: 89-125. Print ISBN: 9781119875086, Online ISBN:

496. Ana-Maria Stoenescu, Sina Niculina Cosmulescu, Nicolae Gheorghiu. (2023). Preliminary research on *in vitro* propagation of *Ziziphus jujuba* Mill. Horticulture, Series B, Vol. LXVII, No. 1: 176-181. Print ISSN 2285-5653, Online ISSN: 2286-1580, ISSN-L: 2285-5653. https://horticulturejournal.usamv.ro/pdf/2023/issue_1/Art25.pdf
497. Karolina Nowakowska, Katarzyna Kowalczyk, Anna Pawełczak and Janina Gajc-Wolska (2023). Energy Efficiency of LEDs during Micropropagation of Helleborus 'Molly's White'. Agriculture, 13, 1265. ISSN (electronic): 2077-0472. <https://doi.org/10.3390/agriculture13061265>
498. Karolina Nowakowska, Potshangbam Nongdam, Nabilah Amany Samsurizal and Andrzej Pacholczak (2023). An efficient micropropagation protocol for the endangered European shrub February Daphne (*Daphne mezereum* L.) and identification of bacteria in culture. Agriculture, 13, 1692. ISSN (electronic): 2077-0472. <https://doi.org/10.3390/agriculture13091692>
499. L. K. J. Al-Amery, M. A. Al-Shamari, I. H. Mohammed, H. S. A. Al-Jubori (2023) Developing a protocols to micropropagate Gac fruit (*Momordica cochinchinensis* Spreng.) *in vitro*. Euphrates Journal of Agricultural Science, 15 (2): 517-526. ISSN 2072-3857. https://www.researchgate.net/profile/Lamiaa-Al-Amery/publication/374510196_Developing_a_protocols_to_micropropagate_Gac_fruit_Momordica_cochinchinensis_Spreng_in_vitro/links/652144a4d717ef1293d31bdd/Developing-a-protocols-to-micropropagate-Gac-fruit-Momordica-cochinchinensis-Spreng-in-vitro.pdf
500. Roukia Benyammi, Soumeia Krimat, Malika Alili, Mohamed Bekhouche, Oumaima Touari, Nawel Belalia, Amina Missoum, Lakhdar Khelifi and Abdelkader Morsli (2023). *In vitro* propagation of Algerian *Lavandula stoechas* and assessment of biochemical composition and their Antioxidant activity. Bull. Pharm. Sci., Assiut University, Vol. 46, Issue 2: 899-911. Print ISSN: 1110-0052, Online ISSN: 3009-7703. https://journals.ekb.eg/article_327719_ac3763b1a285861d99b91dbb17ada1f9.pdf
501. Pham Thi Diem Thi, Nguyen Thi Nguyen Man, Nguyen Thi Khanh Quynh, Trieu Thy Hoa, Pham Mai Thu Thuy, Hoang Tan Quang. (2023). Micropropagation of long-leaved paperbark (*Melaleuca leucadendron* (L.) L.). Propagation of Ornamental Plants, 23: 91-98. ISSN: 1311-9109.

Публикация: Alexandrov A. H., Iliev I. (2019). Forest in South-Eastern Europe. Poplar, 203: 79-85. ISSN (print): 0563-9034, ISSN (online): 2620-1593. https://ilfe.org/sites/default/files/12Alexandrov_and_Iliev.pdf

Цитирани:

554. Karaklić V., Cvjetičanin R., Košanin O. (2019). Florističke, sinekološke i edafske karakteristike šume bukve sa borovnicom (Vaccinio–Fagetum moesiaceae Fukarek 1969) na području Jelove gore u zapadnoj Srbiji, Poplar, 204: 37-49. ISSN (Print): 0563-9034, e-ISSN (Online): 2620-1593. https://journalpoplar.ilfe.org/sites/default/files/05Karaklic_et_al.pdf

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. Plant Growth Regulation, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Цитирани:

572. Коротцева И. Б. (2023). Фасциация у тыквенных культур. Овощи России, 6: 17-21. ISSN: 2072-9146 (print), ISSN: 2618-7132 (online). <https://doi.org/10.18619/2072-9146-2023-6-17-21>

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. *In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522, DOI: 10.1002/9780470686522.ch1*

Ц и т и р а н а в:

573. Akshay Mehta, Alkesh Yadav and Aman Kumar (2023). Micropropagation in Horticultural Crops *In: Revolutionizing Horticulture. The Green Path. Elite Publishing House: 65-75. ISBN: 978-93-58995-85-5*

Публикация: Lyubomirova T., Iliev I. (2013). *In vitro* propagation of *Syringa vulgaris* L. Forestry Ideas, 19: 173-185. ISSN: 1314-3905 (Print), ISSN: 2603-2996 (Online). https://forestry-ideas.info/files/issue/Forestry_Ideas_BG_2013_19_2_7.pdf

Ц и т и р а н а в:

574. Тишкина Е. А., Семкина Л. А., Орехова О. Н., Григорьев А. А., Суслов А. В. (2022). Оценка роста и развития некоторых сортов *Syringa vulgaris* L. в условиях среднего Урала. Успехи Современного Естествознания, № 2: 28-33. ISSN: 1681-7494. DOI: 10.17513/use.37774 <https://natural-sciences.ru/ru/article/view?id=37774>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. Plant Growth Regulation, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Ц и т и р а н а в:

575. Bulavin I. V., Sidyakina A. I. (2023). An *in vitro* *Arabidopsis thaliana* Root Structural and Functional Characterization. Izvestiya Akademii Nauk. Rossijskaya Akademiya Nauk. Seriya biologicheskaya, 6: 616-625. ISSN: 1062-3590 (Print). doi: 10.31857/S1026347022600741. DOI: 10.1134/S106235902360174X, <https://journals.rcsi.science/1026-3470/article/view/231692>

Публикация: Nikolaou P., Zagas D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). Advances in the micropropagation of service tree *Sorbus domestica*. Propagation of Ornamental Plants, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html

Ц и т и р а н а в:

576. Valeria Gianguzzi and Francesco Sottile. (2024). Temporary Immersion System as an Innovative Approach for In Vitro Propagation of *Sorbus domestica* L. Horticulturae, 10, 164. EISSN 2311-7524. <https://doi.org/10.3390/horticulturae10020164>

Публикация: Iliev I. (2017). Factors affecting the axillary and adventitious shoots formation in woody plants *in vitro*. Acta Horticulturae, 1155: 15-27. ISBN: 90 6605 210 4, ISSN: 0567-7572.

Ц и т и р а н а в:

577. Valeria Gianguzzi and Francesco Sottile. (2024). Temporary Immersion System as an Innovative Approach for In Vitro Propagation of *Sorbus domestica* L. Horticulturae, 10, 164. EISSN 2311-7524. <https://doi.org/10.3390/horticulturae10020164>

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирана в:

581. Santhanalakshmi Balasubramaniam, Sivanandhan Ganeshan, Selvaraj Natesan and Kapildev Gnanajothi (2024). A review on genetic diversity, micropropagation and transformations in the high-value medicinal plant of Himalayas-Seabuckthorn (*Hippophae* sp.). Journal of Pharmacognosy and Phytochemistry, 13(1): 360-366. E-ISSN: 2278-4136 P-ISSN: 2349-8234. DOI: <https://doi.org/10.22271/phyto.2024.v13.i1e.14849>
582. R. M. P. I. Rajapaksha, R. G. J. Perera, D. D. N. Vibodhani, H. D. T. Abeyrathna, N. D. G. K. N. Jayawardhane, R. M. S. M. B. Rathnayaka (2023). A review in micropropagation of ornamental aquatic plants. International Research Conference of Sri Lanka Technology Campus Colombo, Sri Lanka. 14th -15th December, 2023: 261-265.

Публикация: Iliev I., Iliev N., Dancheva D., Corneanu M., Tsaktsira M., Gajdošova A., Mladenova S. (2010). Factors affecting the rooting of cuttings from cultivars of *Chamaecyparis lawsoniana* Parl. In: Soare M., Călina A., Panzaru Radu L., Niculescu M., Alexandru T., Stancu I., Cola M., Netiou C., Dimitru I. (Eds). Durable Agriculture – Agriculture of the Future. Analele Universitatii din Craiova, vol. XL, No 2: 174-181. ISSN 1841-8317. https://www.researchgate.net/publication/340265023_Factors_affecting_the_rooting_of_cuttings_from_cultivars_of_Chamaecyparis_lawsoniana_Parl_In_Soare_M_Calina_A_Panzaru

Цитирана в:

583. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Lyubomirova T., Iliev I. (2014). Factors affecting the rooting of cuttings of *Syringa vulgaris* L. cultivars. Oltenia Journal for Studies in Natural Sciences, 30 (1): 67-74. P-ISSN: 1454-6914. https://biozoojournals.ro/oscsn/cont/30_1/11_Lyubomirova_pp67-74.pdf

Цитирана в:

584. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Scaltsoyiannes A., Tsulpha P., Iliev I., Theriou K., Tsaktsira M., Mitras D., Karanikas C., Mahmout S., Christopoulos V., Scaltsoyiannes V., Zaragotas D., Tzouvara A. (2009). Vegetative propagation of ornamental genotypes of *Prunus avium* L. Propagation of Ornamental Plants, 9(4): 198-206. ISSN: 1311-9109. https://www.journal-pop.org/2009_9_4_198-206.html

Цитирана в:

585. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. Propagation of Ornamental Plants, 24: 1-10.

Публикация: Tsaktsira M., Alevropoulos A., Tsoulpha P., Scaltsoyiannes V., Scaltsoyiannes A., Iliev I. (2018). Inter- intra- genetic variation on rooting ability of *Ilex aquifolium* L.

varieties and cultivars. **Propagation of Ornamental Plants**, 18 (4): 131-138. ISSN: 1311-9109. ISSN: 1311-9109. https://www.journal-pop.org/2018_18_4_131-138.html

Цитирана в:

586. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. *Propagation of Ornamental Plants*, 24: 1-10.

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Naniou-Obeidat, Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* 'Agrifoglio Commune'. *Sustainability*, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирана в:

587. Soliman W. S., Saad-Eldeen K., Abas A. M. Gahory A.-A. (2024). Effect of IBA and NAA on improving roots formation and growth of *Sanchezia speciosa* Leonard stem cuttings. *Propagation of Ornamental Plants*, 24: 1-10.

Публикация: Nikolaou P., Zagaz D., Scaltsoyiannes V., Balas E., Xilogianni V., Tsoulpha P., Tsaktsira M., Voulgaridou E., Iliev I., Triandafilou K., Scaltsoyiannes A. (2008). Advances in the micropropagation of service tree *Sorbus domestica*. *Propagation of Ornamental Plans*, 8 (3): 154-157. ISSN: 1311-9109. https://www.journal-pop.org/2008_8_3_154-157.html

Цитирана в:

588. Sediva J., Businsky R., Pospiskova M., Velebil J., Drahozova H., Zyka V. (2021). Conservation methods of Czech gene pool of whitebeams. *Acta Horticulturae*, 1324: 41-46.

Публикация: Iliev I., Gajdosova A., Libiakova G., Jain S. M. (2010). Plant micropropagation. *In: Michael Davey and Paul Anthony (Eds). Plant Cell Culture: Essential Methods*, John Wiley & Sons Ltd.: 1-23. ISBN: 978-0470686485, DOI: 10.1002/9780470686522. DOI: 10.1002/9780470686522.ch1

Цитирана в:

589. Günbeği Z., Ekinçi H., Ak B. E., Şaşkın N. (2024). Determination of proliferation performance of UCB-1 (*P. atlantica* × *P. integerrima*) pistachio rootstock in *in vitro* conditions. *In: H. Turan Akkoyun Seyithan Seydoşoğlu (Eds). 7th International Cukurova Agriculture and Veterinary Congress*, March 9-10, Adana, Türkiye: 463-473. ISBN: 978-625-367-680-3. DOI: <https://doi.org/10.5281/zenodo.10870563>

Публикация: Iliev I., Kitin P. (2011). Origin, morphology, and anatomy of fasciation in plants cultured *in vivo* and *in vitro*. *Plant Growth Regulation*, 63: 115-129. ISSN: 0167-6903. <https://doi.org/10.1007/s10725-010-9540-3>.

Цитирана в:

590. Peretti A. V., Calbacho-Rosa L. S., Olivero P. A., Oviedo-Diego M. A., Vrech D. E. (2024). When THAT Exception Persists Almost as THAT Exception. *In: Rules and Exceptions in Biology: from Fundamental Concepts to Applications*. Springer, Cham.: 83-222. ISBN: 978-3-031-55380-6, ISBN: 978-3-031-55382-6, <https://doi.org/10.1007/978-3-031-55382-0>

Публикация: Maria Tsaktsira, Eleana Chavale, Stefanos Kostas, Elias Pipinis, Parthena Tsoulpha, Stefanos Hatzilazarou, Fotios-Theocharis Ziogou, Irini Naniou-Obeidat,

Ivan Iliev, Athanasios Economou, Apostolos Scaltsoyiannes (2021). Vegetative Propagation and ISSR-Based Genetic Identification of Genotypes of *Ilex aquifolium* ‘Agrifoglio Comune’. Sustainability, 13, 10345, ISSN: 2071-1050. <https://doi.org/10.3390/su13181034>

Цитирани:

591. Laconsole D., Scaltrito E., Sdao A. E., Cristiano G., De Lucua B. (2024). Application of commercial seaweed extract-based biostimulants to enhance adventitious root formation in ornamental cutting propagation protocols: a review. *Frontiers in Horticulture*, 3: 1371090. doi: 10.3389/fhort.2024.1371090

Подпис: